Synthesis, characterization of diorganotin(IV) complexes of *N*-(2-hydroxyarylidene)amino-acetic acid and antitumour screening *in vivo* in Ehrlich ascites carcinoma cells

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Some new diorganotin(IV) complexes have been prepared by reacting potassium N-(2-hydroxyarylidene)aminoacetate with R₂SnCl₂(R = Me, Bu,Ph). The complexes have been characterized by H, 13C, 119Sn NMR, IR and 119mSn Mössbauer spectroscopic techniques in combination with elemental analysis. In the solid state, the complexes possess penta- and hexa-coordinated tin centres. The hexa-coordinated tin complexes were found to dissociate in solution, giving rise to penta-coordinated species as revealed by 119 Sn NMR spectroscopy. Antitumour screening in vivo of the complexes L⁴SnPh₂,L⁴SnPh₂·Ph₃SnCl and L⁴Sn^tBu₂·t Bu_2SnCl_2 (L⁴ = N-(2-hydroxyacetophenone)aminoacetate) is also reported. Copyright © 2001 John Wiley & Sons, Ltd.

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INTRODUCTION

Interest in organotin(IV) complexes of the N-(2hydroxyarylidene)aminoacetic acid framework is from two perspectives, viz. putative antitumour activity and structural reasons. In this context, we have recently reported the coordination behaviour of N-(2-hydroxyacetophenone)glycine towards the R₂Sn(IV) moiety;^{3,4} the isolated monomeric complexes contain an additional potential oxygencontaining site. These complexes, on further treatment with R₂SnCl₂ or R₃SnCl, yielded mixed organotin dendritic complexes in which the two tin atoms are connected via the carbonyl atoms of the ligand molecule. Such organotin oxo clusters have potential as homogeneous catalysts⁵ and in material science.^{6,7} In addition, we have also reported a series of triorganotin(IV) derivatives of *N*-(2-hydroxyarylidene)aminoacetic acid.⁸ X-ray structural studies of two complexes, viz. Ph₃SnL¹H and Me₃SnL⁴H (ligand skeleton as described in Fig. 1), reveal a polymeric trans-O₂SnC₃ trigonalbipyramidal configuration with the R groups in the equatorial positions and the axial locations occupied by a carboxylate oxygen from the ligand and the phenolic oxygen of the ligand on an adjacent complex. In the case of Me₃SnL⁴H, the ligands coordinate in the zwitterionic form with the phenolic proton moved to the nearby nitrogen atom.

One of the goals of the present work was to explore the feasibility of using various R_2Sn groups in the synthesis of organotin complexes of potassium N-(2-hydroxyarylidene)aminoacetate (LHK, Fig. 1). A second objective was to investigate the

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Abbreviation ^a	R ¹	R ²	R ³
L ¹ HK	Н	Н	Н
L ² HK	Н	CI	Н
L ³ HK	Н	NO ₂	Н
L⁴HK	CH₃	Н	Н
L⁵HK	CH₃	Н	CH₃
L⁵HK	CH₃	CH₃	Н

^a H represents hydroxyl proton.

Figure 1 The ligands used in present work.

structural consequences of such complexes in relation to their antitumour properties. For later purposes, we selected three different types of complex (see Table 1), *viz*. L⁴SnPh₂ (14), L⁴SnPh₂·Ph₃SnCl (15) and L⁴Sn⁴Bu₂·Bu₂SnCl₂ (16), which are structurally characterized both in solution and in the solid state and subjected to antitumour activity *in vivo* towards Ehrlich ascites carcinoma (EAC) cells. The complexes 15 and 16 are expected to exhibit enhanced antitumour activity compared with complex 14, since they contain two organotin moieties.

EXPERIMENTAL

Materials

Dimethyltin dichloride (Fluka), diphenyltin dichloride (Aldrich), and di-*n*-butyltin dichloride (Merck) were used as received. The ligands (L¹HK–L⁶HK) and L⁴HNa were prepared as described in earlier reports.^{3,8} The solvents used in the reactions were dried using standard procedures.

Measurements

Carbon, hydrogen and nitrogen analyses were performed with a Perkin Elmer 2400 series II instrument. IR spectra in the range 4000–400 cm⁻¹

were obtained on a Perkin Elmer 983 spectrophotometer with samples investigated as KBr discs. The ¹H, ¹³C and ¹¹⁹Sn NMR spectra were acquired on a Bruker ACF 300 spectrometer and measured at 300.13 MHz, 75.47 MHz and 111.92 MHz respectively. The ¹H, ¹³C spectra were referenced to Me₄Si and ¹¹⁹Sn chemical shifts were referenced to Me₄Sn, all set at 0.00 ppm. ¹¹⁹Sn Mössbauer spectra of the complexes in the solid state were obtained at liquid-nitrogen temperature on a conventional constant acceleration spectrometer equipped with a CRYO cryostat and a ¹¹⁹Sn Mössbauer source. The velocity calibration was made using a ⁵⁷Co Mössbauer source, and an iron foil enriched to 95% in ⁵⁷Fe (DuPont Pharma Italia, Firenze, Italy) was used as the absorber. The Ca^{119m}SnO₃ and ⁵⁷Co sources, 10 mCi, were procured from Ritverc, St Petersburg, Russia. The NORMOS program was used to solve the Mössbauer spectrum of complex $L^4SnPh_2 \cdot Ph_3SnCl$ (15).

Syntheses

Preparation of diorganotin(IV) complexes

A typical procedure is described below, using $L^4Sn^nBu_2$ (10) as an example.

"Bu₂SnCl₂ (0.50 g, 1.64 mmol) in 50 ml of methanol was added dropwise with continuous stirring to a hot methanol solution (50 ml) containing L⁴HK (0.378 g, 1.64 mmol). The reaction mixture was stirred at room temperature for 3 h, and the solvent was removed using a rotary evaporator. The dry mass was washed thoroughly with hot hexane, extracted into chloroform and filtered. A pale yellow product was obtained upon concentration of the chloroform. This was then recrystallized from the same solvent to yield pale yellow crystals of the desired product and dried *in*

The other diorganotin complexes of *N*-(2-hydroxyarylidene)aminoacetic acids were prepared analogously using appropriate R₂SnCl₂ and LHK.

Biological tests

The organotin(IV) complexes, viz. L⁴SnPh₂ (14), L⁴SnPh₂·Ph₃SnCl (15), L⁴Sn ^tBu₂·^tBu₂SnCl₂ (16) and L⁴HNa, were dissolved in dimethyl sulfoxide (DMSO) and further diluted with DMSO in order to achieve the required working solution concentration. The animals of the control group were treated with only DMSO solutions (0.1 ml).

Male Swiss-albino mice, aged 7 weeks and weighing about 21–23 g [maintained in the labora-

Table 1 The physical and analytical data for the diorganotin(IV) complexes

	Reaction	Crystallization		Yield		Elemental ana	alysis: Found	d (calc.) (%)
Complex ^a	time (h)	solvent	Colour	(%)	M.p. (°C)	С	Н	N
1 L ¹ SnMe ₂ ·OH ₂	2 ^b	Chloroform + petroleum ether	Yellow	59	133–135	38.30 (38.41)	4.18 (4.40)	3.96 (4.07)
$2 L^1 Sn^n Bu_2 \cdot OH_2$	3 ^c	Chloroform	Pale yellow	72	$124-125$ $(123.5)^{d}$	47.72 (47.70)	6.25 (6.36)	3.29 (3.23)
3 L ¹ SnPh ₂	3 ^b	Chloroform	Yellow	70	$190-191$ $(183.5)^{d}$	56.00 (56.04)	3.78 (3.81)	3.29 (3.11)
4 L2SnMe2·OH2	2 ^b	Benzene + chloroform	Yellow	30	139–140	34.90 (34.92)	3.70 (3.73)	3.70 (3.70)
$5 L^2 Sn^n Bu_2 \cdot OH_2$	3^{b}	Chloroform	Yellow	40	120-122	44.20 (44.14)	5.52 (5.67)	3.20 (3.30)
$6 L^2 SnPh_2$	3 ^b	Chloroform	Yellow	45	130-132	52.15 (52.06)	\ /	,
$7 L^3 SnMe_2 \cdot OH_2$	2 ^b	Chloroform (large volume)	Pale yellow	47	>250	33.90 (34.15)		
8 L3SnPh2·OH2	3 ^b	Chloroform (large volume)	Pale yellow	26	226–228 (dec.)	49.22 (49.16)	3.44 (3.54)	5.50 (5.46)
$9 L^4SnMe_2 \cdot OH_2$	2^{b}	Chloroform	Pale yellow	53	210-211	40.30 (40.26)	4.80 (4.79)	3.90 (3.91)
10 $L^4Sn^nBu_2 \cdot OH_2$	3^{c}	Chloroform	Pale yellow		77–78	49.02 (48.90)	6.53 (6.61)	3.33 (3.17)
$11 L^5 SnPh_2$	3 ^b	Chloroform	Yellow	79	170-172	57.65 (57.78)	4.45 (4.43)	3.00 (2.93)
12 $L^6SnMe_2 \cdot OH_2$	2^{b}	Benzene	Pale yellow	54	138-140	42.05 (41.98)	5.15 (5.15)	3.80 (3.77)
13 L ⁶ SnPh ₂	3 ^b	Chloroform	Yellow	79	236–237 ^e	57.70 (57.78)	4.40 (4.43)	2.90 (2.93)

^a The complexes **14** L⁴SnPh₂, **15** L⁴SnPh₂·Ph₃SnCl and **16** L⁴Sn^tBu₂·tBu₂SnCl₂ are included elsewhere for convenience of discussion; for details see Ref. 3.

tory in communal cages in room under controlled temperature $(27 \pm 3 \, ^{\circ}\text{C})$ and lighting $(12 \, \text{h light/}\ 12 \, \text{h dark})$ conditions on standard mouse diet (NMC Oil Mills Ltd, Pune, India) and water *ad libitum*] were used in all experiments. One hundred mice were divided randomly into five groups of 20 each.

EAC cells were maintained in 7-week-old male Swiss mice throughout intraperitoneal (i.p.) transplantation of 1×10^6 viable tumour cells to each mouse. On day 0, five groups of mice received tumour cells (1×10^6 per mouse i.p.). On day 2, the complexes to be screened, **14**, **15**, **16** and L⁴HNa, were injected (i.p.) into four different groups of mice. One untreated group, which received only DMSO, was kept as a control. Three animals from each group were sacrificed by cervical dislocation on the 12th or 13th day of cell implantation. Ascites fluid volume, packed cell volume and number of cells were then measured. The survival of drugtreated mice over controls was studied in the remainder of the mice.

RESULTS AND DISCUSSION

Treatment of LHK with R_2SnCl_2 in equimolar ratio in methanol or methanol-benzene mixture yielded the diorganotin derivatives of composition $LSnR_2 \cdot OH_2$ similar to that of the vinyl analogue³ in most cases, except for complexes $\bf 3, 6, 11$ and $\bf 13$, which were isolated as $LSnR_2$. A typical reaction is described in the Experimental section, whereas the synthetic details and characterization data are summarized in Table 1. The complexes are soluble in chloroform, methanol, ethanol and DMSO. In our earlier communications 3,4,7 we have

In our earlier communications^{3,4,7} we have reported that the ligands are tridentate, and can coordinate to tin atom *via* an oxygen atom of a carboxylate group, a phenoxide oxygen atom and the imino nitrogen atom, as revealed by X-ray crystallography. On this basis, we discuss the spectroscopic results in relation to the structure of the complexes.

^b Method: stirring in methanol.

^c Method: reflux in methanol–benzene mixture (1:1).

^d Data taken from Ref. 10.

e Data taken from Ref. 4.

Table 2	Characteristic	IR bands	(cm^{-1})	for the diorganotin(IV) complexes
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Complex	$v(OCO)_{asym}$	$v(OCO)_{sym}$	$\Delta v(OCO)$	v(C=N)	ν(Ph—(CO))
1	1617 ^a	1389	228	1617 ^a	1286
2	1622	1394	228	1585	1249
3	1684	1433	251	1615	1214
4	1604 ^a	1382	222	1604 ^a	1241
5	1612 ^a	1372	240	1612 ^a	1239
6	1682	1441	241	1602	1238
7	1627	1398	229	1587	1245
8	1679	1431	248	1616	1226
9	1595 ^a	1375	220	1595 ^a	1235
10	1612 ^a	1372	240	1612 ^a	1239
11	1686	1451	235	1600	1233
12	1593 ^a	1369	224	1593 ^a	1226
13	1679	1450	229	1616	1226

^a Vibrations due to both $\nu(OCO)_{asym}$ and $\nu(C=N)$ appeared as an unresolved broad peak.

IR spectra

Diagnostically important IR bands and their assignments are presented in Table 2. The potassium salts of the ligands display two bands at around $1630~{\rm cm}^{-1}$ and $1605~{\rm cm}^{-1}$ that are assigned to the $\nu({\rm OCO})_{\rm asym}$ and $\nu({\rm C=\!N})$ stretching vibrations respectively, in accord with earlier reports. In organotin(IV) complexes, these vibrations are shifted considerably owing to coordination through

the carboxyl oxygen and the imino nitrogen atoms.³ The $\nu(\text{OCO})_{\text{sym}}$ stretches have been detected and the magnitude of $\Delta\nu(\text{OCO})$ is in the range 220–251 cm⁻¹, indicating unidentate bonding through the carboxylate moiety.^{8,12} Coordination through the phenolic oxygen atom is reflected in the lower wavelength shifts of the $\nu(\text{Ph}\text{---}(\text{CO}))$ vibration when compared with the sodium or potassium salts.³ A medium intensity band due to Sn--O has been detected at around 540 cm⁻¹ in the com-

Table 3 ¹H chemical shifts (ppm)^a for the diorganotin(IV) complexes

			Ligand	skeleton ^b			Si	n—R skele	eton ^c	
Complex	H-2	H-6/6′	H-7	H-8/8'	H-9	H-3'	1*	2*	3*	4*
1	4.39	6.85	7.17	6.74	7.41	8.33	0.83 (89) ^d	_	_	_
2	4.34	6.79	7.17	6.74	7.43	8.40	1.64	1.58	1.38	0.87
3	4.35	7.15	7.15	6.77	7.53	8.34	_	7.89	7.45	7.45
4	4.36	6.67	7.65	_	7.25	8.38	$0.70 (93)^{d}$	_	_	_
5	4.38	6.65	7.30	_	7.14	8.32	1.64	1.52	1.29	0.84
6	4.37	7.07	7.47	_	7.16	8.34	_	7.85	7.47	7.47
7	4.00	6.32	7.75	_	8.32	8.40	$0.40 (93)^{d}$	_	_	_
8	4.36	6.96	8.23	_	8.43	8.76		7.62	7.34	7.34
9	4.29	6.84	7.36	6.79	7.51	2.67	$0.73 (78)^{d}$	_	_	_
10	4.26	6.85	7.35	6.76	7.47	2.67	1.60	1.43	1.31	0.86
11	4.25	2.44	7.27	6.69	7.38	2.57	_	7.82	7.38	7.38
12	4.27	6.75	7.18	2.27	7.25	2.65	$0.71 (75)^{d}$	_	_	_
13	4.28	7.08	7.25	2.25	7.27	2.61		7.87	7.39	7.39

^a In CDCl₃ except for complexes **4**, **7** and **8**, which are in DMSO-d₆·

^b Refer to Fig. 1 for numbering scheme.

^c Numbering scheme for Sn—R skeleton as shown below:

 $^{^{}d} J(^{1}H_{-}^{119}Sn)$ (Hz).

 $^{13}\mathrm{C}$ and $^{119}\mathrm{Sn}$ chemical shifts (ppm) a for the diorganotin(IV) complexes Table 4

					Ligan	Ligand skeleton ^b	oue				Sn—R sk	(nJ(1)	Sn—R skeleton ^b (" $J(^{13}C_{-}^{119}Sn)$ (Hz))	(z))	
Complex	C-1	C-2	C-3	C-4	C-5	9-2	C-7	C-8	6-D	C-3//6//8/	C-1*	C-2*	C-3*	C-4*	119Sn
1	172.5	57.9	172.3	117.7		122.9	137.2	117.5	135.3	1	(-) 60.0	I	ı	ı	-203.1
7	172.9	57.7	171.4	117.3		122.7	138.0	117.2	135.6	ı	$26.7 (-^{\circ})$	$26.8 (-^{\circ})$	$22.6 (-^{\circ})$	$13.7 (-^{\circ})$	-208.0
ĸ	173.0	57.0	170.4	117.1		122.8	138.3	117.8	135.6	ı	$137.6 (-^{\circ})$	136.4 (57)	128.9 (91)	130.8 (23)	-335.4
4	171.7	9.99	170.1	119.3		122.9	135.0	119.2	132.4	I	0.70 (804)	ı	ı	ı	ام
w	171.2	58.1	171.2	117.6	167.7	124.5	137.2	121.0	133.4	ı	$26.1 (-^{\circ})$	27.0 (33)	21.1 (85)	$13.5 (-^{\circ})$	-200.0
9	172.1	57.3	170.2	117.6		124.6	138.2	122.1	133.7	I	$137.3 (-^{\circ})$	136.4 (57)	129.7 (91)	131.0 (19)	-335.8
7	171.5	56.5	170.4	135.5		121.6	131.6	115.3	129.1	ı	0.20(800)	ı	ı	ı	٦٦
œ	173.0	56.8	170.4	146.0		124.8	132.8	117.5	130.4	ı	$137.3 (-^{\circ})$	$134.9 (-^{\circ})$	$129.3 (-^{\circ})$	$130.4 (-^{\circ})$	۱۳
6	181.7	53.3	170.4	121.1		123.5	135.9	117.9	130.5	22.9	0.08 (670)	I	ı	ı	-172.6
10	181.4	53.5	170.7	121.1		123.5	135.8	117.6	130.3	22.9	$26.7 (-^{\circ})$	26.9 (35)	21.0 (85)	$13.5 (-^{\circ})$	-210.8
11	182.5	53.6	170.7	128.7		119.5	136.7	117.5	131.6	23.1/16.9	$137.9 (-^{\circ})$	136.5 (54)	128.9 (90)	130.7 (20)	-349.4
12	181.5	53.2	170.3	126.9		123.2	137.2	120.6	129.8	22.8/20.5	0.30(677)	ı	I	I	-170.1
13	181.9	53.5	170.3	128.2		123.6	136.7	119.9	130.2	22.7/20.5	$137.7 (-^{\circ})$	136.7 (56)	128.8 (93)	130.6 (20)	-349.8

^a Refer to Table 3 for the solvents.

^b Refer to Table 3 for numbering schemes.

^c Could not be detected.

^d Not determined.

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plexes. ¹³ Thus, IR data provide reasonable evidence for complexation of the potentially multidentate ligand.

NMR spectra

The ¹H and ¹³C NMR data of the diorganotin complexes are shown in Tables 3 and 4 respectively. The ¹H and ¹³C chemical shift assignments of the diorganotin moiety are straightforward from the multiplicity patterns and/or resonance intensities, whereas the ligand skeletons were assigned by multiplicity patterns and/or resonance intensities of the signals, ^{3,8} and also by standard distortionless enhancement by polarization transfer (DEPT) experiments. The ¹H and ¹³C NMR spectra of the complexes show the expected resonances and integration.

In recording ¹¹⁹Sn NMR spectra of organotin(IV) complexes in solution, non-coordinating solvents are preferrable to coordinating ones to preclude possible changes in the coordination number of the tin atom. In the present investigation, a few complexes were found to be suitable for recording ¹¹⁹Sn NMR spectra in CDCl₃ solution. These are listed in Table 4. The ¹¹⁹Sn chemical shifts of dialkyltin(IV) complexes occur between -170 and -211 ppm, which fall well within the range proposed for penta-coordinate tin centres. ¹⁴ Thus, ¹¹⁹Sn NMR results indicate that the hexa-coordinated structure of diorganotin complexes (as revealed by Mössbauer spectroscopy, vide infra) is lost upon dissolution, indicating that a water molecule is not coordinated in solution, similar to that of its divinyltin analogue.³ Diphenyltin complexes show a sharp resonance between -335 and -350 ppm indicating a penta-coordinated tin structure in solution, which is consistent with our earlier report.3

Mössbauer data

In order to obtain further structural evidence, the Mössbauer spectra of the representative complexes were recorded in the solid state (Table 5). The spectra of the complexes (except 15) all display a characteristic doublet absorption, indicating a single tin site. The R₂Sn derivatives show isomer shift δ values typical of quadrivalent organotin derivatives, similar in alkyl₂Sn moieties and higher with respect to Ph₂Sn. The quadrupole splitting Δ values of complexes 2,7,8,9 and 10 are in the range 3.74–3.99 mm s⁻¹ in alkyl₂Sn and 3.61 mm s⁻¹ in Ph₂Sn derivatives, suggesting a distorted *trans*-R₂Sn octahedral

 $\begin{array}{ll} \textbf{Table 5} & ^{119}\text{Sn M\"{o}ssbauer parameters (mm s}^{-1}) \text{ for some representative diorganotin}(IV) \text{ complexes}^a \end{array}$

Complex	δ	Δ	Γ_1	Γ_2	C—Sn—C (°)
2 7 8 9 10 14	1.33 1.29 1.16 1.21 1.40 1.15 0.75	3.79 3.99 3.61 3.74 3.87 2.88 2.20	0.92 0.95 1.00 0.90 0.95 0.89 1.00	0.92 0.92 1.00 0.92 0.98 0.98 1.00	141 170 159 154 143 128 107
13	1.14	2.50	0.86	0.86	107

^a Γ_1 and Γ_2 : line widths.

structure. 15 The C—Sn—C angles, calculated by the Parish relation, ¹⁶ vary from 141 to 170°. In contrast, complex 14 shows a lower Δ value, which agrees with a trigonal bipyramidal structure with a calculated C—Sn—C angle of 128°. The spectrum of 15 (see Fig. 2 of Ref. 3) requires special comment. The spectrum shows asymmetric absorption with large values of full-width at half height, and was fitted as two doublets with different parameter values; hence the occurrence of two different tin sites can be inferred. A doublet shows a Δ value (2.20 mm s⁻¹) that can be set as the lowest in *cis*-R₂Sn trigonal bipyramidal derivatives. ¹⁵ This Ph₂Sn—O₂N trigonal bipyramidal geometry matches with the X-ray structure³ except for the C—Sn—C calculated angle; the corresponding δ value is also low, and this can arise from the electron withdrawl of the ligands to the tin. The second doublet can be attributed to a Ph₃Sn moiety in a trigonal bipyramidal geometry, with the oxygen and chlorine atoms as axial ligands. The percentage areas of the two doublets are similar, indicating equal amounts of di- and tri-organotin moieties.

Thus, the spectroscopic data suggest a six-coordinate structure for the LSnR₂·OH₂ type complexes in the solid state, owing to coordination of a water molecule. The geometry is distorted octahedral with two R groups occupying the *trans* positions and the equatorial plane by on NO₃ donor set similar to that of the vinyl analogue.³ On the other hand, a distorted trigonal bipyramidal geometry has been suggested for LSnR₂-type complexes, with two oxygen atoms defining the axial positions and two phenyl groups and nitrogen in the trigonal plane, as in the case of complex 14.^{3,4}

Evaluation of antitumour properties

The effect of the organotin complexes 14, 15, 16

Total volume (ml)

volume (ml) Mean survival time

(MST) (days) T/C^a (%)

tumour cells + ascites fluid Total packed cell

					Organ	otin co	mpounds	S	
Parameters	Control	L^4H	INa	14		1	15	1	6
Dose (mg kg ⁻¹) Cells ($\times 10^6$) per ml	No drug	6	30	6	30	6	30	6	30
Cells $(\times 10^6)$ per ml ascites fluid	168 ± 10.20	180 ± 16.32	178.3 ± 16.5	76 ± 20.5	-	-	_	-	-

 6 ± 0.6

 3.5 ± 0.8

 29 ± 3.5

90

Table 6 Effect of organotin compounds **14, 15** and **16** and L⁴HNa on the survivability of the mouse *in vivo*

 5 ± 0.6

 3.1 ± 0.3

 30.2 ± 3.16

93

 7.3 ± 1.45

 5.2 ± 0.40

 32.5 ± 2.69

100

and L⁴HNa on the survivability of mice is shown in Table 6. L⁴HNa is found to be non-toxic to the mice at 30 mg kg⁻¹. It has been observed that L⁴HNa is highly soluble in water, and a very high dosage (50 mg kg⁻¹ in aqueous solution) does not cause the death of any mice. All the organotin complexes studied in the experiment are highly toxic to mice. Complex 14 is less toxic than 15 and 16. Although the exact cause of toxicity was not studied, it could be due to free radical generation from the organotins to the living system.

T/C values (using the NCI protocol for screening new anticancer drugs)¹⁷ showed that neither L⁴HNa nor the organotin complexes possess any antitumour properties. However, some diorganotin complexes of cognate ligands displayed cytotoxicity when tested *in vitro* against MCF-7, EVSA-T, WiDr, IGROV, M19 MEL, A498 and H226 human tumour cell lines.¹³ Further studies with the organotin complexes were not undertaken owing to their toxicity; however, studies with L⁴HNa are warranted owing to its water solubility and non-toxic nature.

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10

30.76

02

6.15

11.2

35

04

12.3

02

6.15

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 4 ± 0.77

 3.1 ± 0.6

 19.1 ± 4.6

59

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^a T/C is the ratio of the MST of treated mice to MST of control mice.