Ethylation of methylarsenic(III) compounds by sodium tetraethylborate

Vincent Clamagirand, Iain L Marr and James L Wardell
Department of Chemistry, University of Aberdeen, Meston Walk, Old Aberdeen AB9 2UE, UK

The compounds MeAsBr₂ and Me₂AsBr at concentrations of $(1-5)\times 10^{-3}\,\mathrm{M}$ in acetone solution are ethylated in high yield by NaBEt₄ to MeEt₂As and Me₂EtAs, as shown by ¹H NMR spectroscopy. The extents of ethylation of MeAs²⁺ and Me₂As⁺ (expressed as ions, by convention) in aqueous acid solutions [at concentrations of $(5-20)\times 10^{-6}\,\mathrm{M}$] were investigated using cold trap/AA and GC AA procedures. The species Me₂As⁺ was ethylated (to give Me₂EtAs) in good yield (88%); in contrast, MeAs²⁺ produced the volatile trialkylarsine, MeEt₂As, in poor yield (30%). No volatile trialkylarsine could be obtained on treating inorganic arsenic(III) (As³⁺) solutions with NaBEt₄.

Keywords: Organoarsenic, sodium tetraethylborate, gas chromatography-atomic absorption spectrometry, cold trap-atomic absorption spectrometry

INTRODUCTION

Various types of arsenic compounds have been found in environmental samples. 1-3 These range from arsenic(III) to arsenic(V) compounds and from inorganic to organic derivatives. The organoarsenic species include the simple methylarsenic species MeAs(O)(OH)₂, Me₂As(O)OH, Me₃As(O), Me₄As⁺], arsenocholine, (Me₃As⁺CH₂CH₂OH), arsenobetaine, (Me₃As⁺CH₂CO₂⁻), and arsenoribofuranosides.

As the toxicity of arsenic species varies widely, it is important to determine the amounts of individual compounds rather than just the total arsenic concentrations. So far, no general analytical method has been found which enables all arsenic species to be assayed. Several procedures have been devised which allow determination of some of the arsenic species, e.g. hydride generation using sodium tetrahydroborate (NaBH₄) for inorganic arsenic, Me₂As(O)OH and MeAs(O)(OH)₂ (organic arsenic moiety only);4-9 HPLC inducticoupled plasma (ICP) MS velv Me₃As⁺CH₂CO₂ and Me₂As(O)OH; polarography for inorganic arsenic, Me₂As(O)OH and MeAs(O)(OH)2;10 LC or ion chromatography/ QF AÀ/ÍCP ÁE or ICP MS,¹¹ reduction by HSCH₂CO₂Me, followed by GC HPLC/FAAS, flame atomic fluorescence (FAFS) ICP AES for $Me_2As(O)(OH)_2$ MeAs(O)(OH)2;12 HPLC/thermochemical hydrogenation/AAS for $Me_3As^+CH_2CO_2^-$, Me₃As⁺CH₂CH₂OH and Me₄As⁺; ¹³ and ion-pair and ion-exchange HPLC/FAAS for inorganic arsenic, Me₂As(O)OH, MeAs(O)(OH)₂ and $Me_3As^+CH_2CO_2^{-1}$

Sodium tetraethylborate (NaBEt₄) has been successfully used in speciation studies of tin, ¹⁵⁻¹⁷ mercury¹⁸⁻²⁰ and lead;²¹ reactions lead to the formation of volatile peralkylated-metal derivatives. Sodium tetraethylborate has the major advantage over other alkylating reagents, e.g. Grignard reagents, in allowing use in aqueous media.

This study was designed to investigate the potential of NaBEt₄ in the speciation of organoar-senic compounds.

Note: we express the arsenic species as ions, as is conventional for this work; the compounds are likely to be present mainly as the covalent species added).

RESULTS

The NMR experiments showed that Me₂AsBr or MeAsBr₂ in acetone as well as in aqueous acetone [at concentrations of ca (1–5)×10⁻³ M were ethylated by NaBEt₄ to Me₂EtAs or MeEt₂As in high yield. The NMR reactions were best monitored using the methylarsenic peaks. Only one ethyl group of each NaBEt₄ molecule was active. In reactions between MeAsBr₂ and more than one equivalent of NaBEt₄, the intermediate species, MeEtAsBr, was detected. Values of the chemical shift, δ^1 H(Me) for the various methylarsenic species are listed in Table 1.

Ethylation of arsenic(III) species was next attempted at lower concentration in purely aqueous media at different pH values. Aqueous

Table 1 Values of $\delta^{1}H$ (Me) for methylarsenic species in CD₃COCD₃ solution

Compound	δ¹H (Me)	
Me ₂ AsBr	1.84	
MeAsBr ₂	2.70	
Me ₂ EtAs	0.96	
MeEt ₂ As	0.90	
MeEtAsBr	1.80	

^{*}Relative to Me4Si.

solutions of As3+, MeAs2+ or Me2As+ as bromides at different pH values were treated with aqueous NaBEt₄. The volatile arsenic species were determined by AA after stripping the solutions, using a stream of nitrogen, into a cold trap. As shown in the results given in Table 2, no volatile species were formed from As3+ at any pH. Both MeAs²⁺ and Me₂As⁺ are ethylated to different extents, to MeEt, As and Me, EtAs respectively, depending on the pH value. At any of the pH regions investigated, little or no further alkylation was detected after 400 s from the start of the reaction, further addition of NaBEt₄ giving only a slight increase in the overall response. The shape of the responses (in particular the tailing) suggested that the ethylations were not immediate reactions. The better guide to the total amount of volatile trialkylarsine products was considered to be the total area of response in AA rather than the peak heights. From the results in Table 1, solutions at pH 1 give the best overall response and so all alkylations were subsequently carried out at pH 1; however, it should be noted that NaBEt₄ is sensitive to acids and that some decomposition occurs in these acid solutions.

Table 2 Determination of arsenic by derivatization using $NaBEt_4$ -cold trap- AA^a

рН	Arsenic reagent						
	As ³⁺		MeAs ²⁺		Me ₂ As ⁺		
	Peak height ^b	Peak area ^b	Peak height ^b	Peak area ^b	Peak height ^b	Peak area ^b	
0	0	0			0.509	25.6	
1	0	0	0.132	30.8	0.437	72.4	
2	0	0	0.134	23.1	0.546	73.1	
4.5	0	0	0.045	5.2	0.045	71.0	

^a Each solution contained the same mass of arsenic (80 μg).

^b Relative values.

Table 3 shows that there is a linear correlation of the response obtained with the amount of arsenic species originally present. The response from Me₂As⁺ was consistently greater than that from MeAs²⁺ (by a factor of 2.8). Use of the ethylation/simple cold trap/AA combination did not result in any significant separation of the peaks arising from MeEt₂As and Me₂EtAs from mixed derivatizations. It is likely that the boiling point of MeEt₂As is too similar to that of Me₂EtAs (84 °C) for it to be separated simply by sequential release from the cold trap on warming. This should be contrasted with the separation of AsH₃ (b.p. -55 °C), MeAsH₂ (b.p. 2 °C) and Me₂AsH (b.p. 36 °C) in the hydride generation/ cold trap/AA approach; relative response times on removing the cold trap here were 27 s, 44 s and 58 s.]

Use of the ethylation/GC/AA approach does however allow separation of Me₂EtAs and MeEt₂As. Derivatization of Me₂As⁺ or MeAs²⁺ in aqueous hydrochloric acid (HCl) solution by NaBEt₄, followed by extraction into pentane, GC separation and AA analysis gave the following retention times (min): pentane/Me₂EtAs/MeEt₂As = 1.04/2.26/3.79.

No volatile trialkylarsines remained in aqueous acid solution after the nitrogen purging. However, some non-volatile methylarsenic derivatives must still have been in the aqueous solution, due to the incomplete ethylations [viz. unreacted Me₂As⁺ and MeAs²⁺, as well as the partially ethylated MeEtAs⁺]. The amounts of arsenic(III) species remaining in the aqueous solution after the NaBEt₄ treatments of MeAs²⁺ and Me₂As⁺ were determined by subjecting the nitrogen-purged solution to the hydride generation procedure (hvdride generation/cold trapping/AA). It was established for the Me₂As⁺ reactions that 12% of the arsenic remained in

Table 3 Response curves for determination of arsenic by ethylation-cold trapping-AA

Mass of arsenic (ng)	Peak area		
	MeAs ²⁺	Me ₂ As ⁺	
0	_		
80	0.028	0.084	
160	0.059	0.163	
320	0.112	0.305	

^a Peak appeared at times between 160 and 180 s after removing the cold trap.

solution; whereas for reactions of MeAs²⁺, 30% of the original arsenic content remained.

CONCLUSIONS

The results indicate that NaBEt₄ has a limited use in determining organoarsenic compounds directly in aqueous acidic solutions at concentrations found in environmental samples. Only Me₂As⁺ was effectively ethylated (to ca 88%) under the conditions employed; in comparison only ca 30% of MeAs²⁺ was diethylated.

Much higher levels of ethylation were detected by NMR spectroscopy using higher concentrations in acetone or aqueous acetone solutions.

EXPERIMENTAL

Reagents

MeAsBr₂ and Me₂AsBr were Johnson Matthey GmbH and Alfa products.

An arsenic(III) stock solution (As³+) containing 50 µg As cm⁻³ was prepared by dissolving 21.7 mg of sodium arsenite in 250 cm³ of 0.10 M HCl in water. Appropriate dilutions were made using 0.10 M HCl. Monomethylarsenic or dimethylarsenic stock solutions containing 40 µg As cm⁻³ were prepared by dissolving 33.3 mg of MeAsBr₂ or 24.8 mg of Me₂AsBr in 250 cm³ water. Appropriate dilutions of the stock solutions were made using water. The stock solutions of As³+, MeAs²+ and MeAs⁺ were stable at 6°C for three weeks, three weeks and three days, respectively.

A 2% (w/v) solution of NaBH₄ (from Aldrich Chem. Co. Ltd) in 0.5% (w/v) NaOH solution in water was prepared daily. NaBEt₄ (from Stem Chemical Inc.) was stored under a nitrogen atmosphere and 0.4% (w/v) NaBEt₄ in water was prepared daily.

Apparatus

Hydride generation-cold trap-atomic absorption spectrometry system

This system comprised a discrete hydride generator and a cold trap with detection by AA. Solutions of the arsenic species (1 cm³) and aqueous 0.10 M HCl (10 cm³) were added to the

generator. Volatile arsenic species were produced in the generator after injection of 2 cm³ of the stock NaBH₄ solution and carried into a nitrogen flow. The gas stream was dried by passage through sodium hydroxide pellets and the arsenic species were trapped in a U-tube filled with broken Fenske helices and submerged in liquid nitrogen, at -190 °C. The arsines were collected in the cold trap for 100 s (a nitrogen carrier gas flow rate of 60 cm³ min⁻¹ was used); after removal of the liquid nitrogen, the trap was allowed to warm to room temperature, which resulted in the sequential elution of arsines in the order of their volatility into an air-acetylene-flame-heated quartz T-tube [ca 900 °C] in the lightpath of a Varian Spectra A.A-10 atomic absorption spectrometer (using the 193.7 mm wavelength). A hollow-cathode lamp was chosen as the arsenic line source, with a lamp current of 10 mA. A H₂-D₂ continuum source background correction system was used. The slit width was 0.5 mm. Relative times (s) found for $AsH_3/MeAsH_2/Me_2AsH$ were $27 \pm 1/44 \pm 1/58 \pm$ (°C) boiling points AsH₃/MeAsH₂/Me₂AsH were -55/2/36, respectively.

Ethylation-cold trap-AA

This system was similar to that used for the hydride generation-cold trap-AA system, with the following changes: (1) 2 cm³ of the NaBEt₄ solution was used instead of 2 cm³ of NaBH₄; and (2) the volatile alkylarsines were collected in the cold trap for 400 s.

Gas chromatography coupled with AA

A Perkin-Elmer E17 gas chromatograph was interfaced with a Perkin-Elmer 360 double-beam flame AA.²² A Megabore DB-S capillary column was used. To the nitrogen flow used as carrier gas in the GC was added a supplementary nitrogen flow at the end of the column, in order to transport the separated compounds with greater efficiency. These were carried to the atomization cell, which consisted of a flame-heated quartz T-tube to which was added hydrogen and air flow for better atomization. The conditions used were as follows:

Injector temperature: 225 °C

Oven temperature: 60 °C

Interface temperature: 180 °C

H₂ flow rate: 100 cm³ min⁻¹

Air flow rate: 50 cm³ min⁻¹

Original solution	Peak area (h)	Peak area ^a	Retention time (min) ^b	Assignment
Pentane	_		1.04	Pentane
Pentane + MeAs ²⁺			1.04	Pentane
	26 451	1	3.79	MeEt ₂ AS
Pentane + Me ₂ As ⁺	_	_	1.02	Pentane
	77 600	2.9	2.26	Me ₂ EtAs

Table 4 GC-AA data for ethylation of MeAs²⁺ and Me₂As⁺ (containing 80 µg of arsenic) by NaBEt₄

Nitrogen flow rate: 1-5 psg for capillary GC and 125 cm³ min⁻¹ for additional flow to interface at the quartz cell.

Other instruments

For the NMR work a Bruker 250 MHz instrument was used.

Methods

Effect of pH on ethylation of As⁺, MeAs²⁺ and Me₂As⁺ by aqueous NaBEt₄

To a solution of the arsenic compound [containing 80 µg As in 20 cm³ of acetate/HCl-buffered solution at pH 0, 1, 2 or 4.5 was added 2 cm³ of 0.4% (w/v) NaBEt₄ solution in water. The volatile organoarsenic species were analysed by the cold trap-AA technique. Results are given in Table 2. As a result remaining determinations involving ethylation-cold trapping-AA reactions were performed at a pH of 1, for different concentrations of MeAs²⁺ and Me₂As⁺ (Table 3).

Determination of organoarsenic species remaining after ethylation of MeAs²⁺ and Me₂As⁺ by NaBEt₄

To a solution of MeAs²⁺ or Me₂As⁺ (containing 80 µg As) in 20 cm³ of a 0.10 M HCl solution was

Table 5 Solutions used in ¹H NMR studies

Methylarser	M.DE.			
Compd	(mg)	(µmol)	(mg)	(µmol)
Me ₂ AsBr	10.1	54.6	8.20	54.6
Me ₂ AsBr	10.1	54.6	16.40	109.2
Me ₂ AsBr	10.1	54.6	4.10	27.3
MeAsBr ₂	13.0	52.0	7.81	52.0
MeAsBr ₂	13.0	52.0	15.62	104.0
MeAsBr ₂	13.0	52.0	23.43	156.0
	Compd Me ₂ AsBr Me ₂ AsBr Me ₂ AsBr Me ₂ AsBr MeAsBr ₂ MeAsBr ₂	Compd (mg) Me ₂ AsBr 10.1 Me ₂ AsBr 10.1 Me ₂ AsBr 10.1 Me ₃ AsBr 10.1 MeAsBr ₂ 13.0 MeAsBr ₂ 13.0	Me ₂ AsBr 10.1 54.6 Me ₂ AsBr 10.1 54.6 Me ₂ AsBr 10.1 54.6 MeAsBr ₂ 13.0 52.0 MeAsBr ₂ 13.0 52.0	Compd (mg) (μmol) (mg) Me ₂ AsBr 10.1 54.6 8.20 Me ₂ AsBr 10.1 54.6 16.40 Me ₂ AsBr 10.1 54.6 4.10 MeAsBr ₂ 13.0 52.0 7.81 MeAsBr ₂ 13.0 52.0 15.62

added 2 cm³ of the 0.4% solution of NaBEt₄ in water. This was purged for 400 s and then 100 µl of the remaining solution was subjected to the hydride generation technique. The remaining quantities generated by NaBH4 were found to be 47 ng As from DMA In 100 µl (i.e. 10 µg overall) and 110 ng As from MeAs²⁺ (i.e. 24 µg overall).

Determination of arsenic by ethylation-GC-AA MeAs²⁺ or Me₂As⁺ (80 μ g As) in 20 cm³ of a 0.1 M HCl solution was derivatized with 2 cm³ of 0.4% NaBEt₄ solution in water. After 7 min, the solution was extracted into pentane (5.0 cm³); aliquots (2 µl) of the pentane solution were injected into the GC-AA (oven temperature 60°C). Results are given in Table 4.

NMR study of reaction between methylarsenic(III) compounds and NaBEt4 in CD₃COCD₃ and D₂O/CD₃COCD₃ $(D_2O/CD_3COCD_3=1:9)$

To a solution of a methylarsenic bromide (Me₂AsBr or MeAsBr₂) in CD₃COCD₃ solution (1 cm³) was added a known quantity of NaBEt₄ under nitrogen. The reactions were monitored by ¹H NMR spectroscopy; the extent of the ethylation reactions was determined from integration of the singlet absorption for the methyl groups in the relevant methylarsenic species. Values of $\delta^1 H$ (Me) for the methylarsenic reagents, products and intermediates are given in Table 1. The solutions used are listed in Table 5.

REFERENCES

- 1. Shibata, Y and Morita, M Appl. Organomet. Chem.,
- 2. Francesconi, K A, Edmonds, J S and Stick, R V J. Chem. Soc., Perkin Trans. 1, 1992, 1349
- 3. Francesconi, K A, Edmonds, J S, Stick, R V, Skelton, B W and White, A H J. Chem. Soc., Perkin Trans. 1, 1991, 2707

^a Relative values. ^b The cold trap was removed at time = 0.

- Chana, B S and Smith, N J Anal. Chim. Acta, 1987, 197: 177
- Braman, R S, Johnson, D L, Foreback, C C, Ammons, J M and Bricker, J L Anal. Chem., 1977, 49: 621
- Howard, A G and Comber, D W Mikrochim. Acta, 1992, 109: 27
- Michel, P, Averty, B and Colandini, V Mikrochim. Acta, 1992, 109: 35
- 8. Rubio, R, Padro, A, Alberti, J and Rauret, G Mikrochim. Acta, 1992, 109: 39
- Aizpunz Fernandez, R, Valdes-Hevia, C, Tempramo, Y, Fernandez de la Campa, M R, Sanz-Medel, A and Neil, P Talanta, 1992, 39: 1517
- Greschonig, H and Irgolic, K J Appl. Organomet. Chem., 1992, 6: 565
- Morin, P, Amran, M B, Favier, S, Heimburger, R and Leroy, M Fresenius J. Anal. Chem., 1991, 339: 504
- Ebdon, L, Hill, S, Walton, A P and Ward, R W Analyst (London), 1983, 113: 1159

- Blais, J S, Momplaisir, G M and Marshall, W D Anal. Chem., 1990, 62: 1161
- Larsen, E H and Hansen, S H Mikrochim. Acta, 1992, 109: 47
- Ashby, J R, Clark, S and Craig, P J J. Anal. At. Spectrom., 1988, 3: 735
- Ashby, J R and Craig, P J Appl. Organomet. Chem., 1991, 5: 173
- Ashby, J R and Craig, P J Sci. Total Environ., 1989, 78:
 219
- Rapsomanikis, S and Craig, P J Anal. Chem. Acta, 1991, 248: 563
- Craig, P J, Mennie, D, Ostah, N, Donald, O F X and Martin, F Analyst (London), 1992, 117: 823
- 20. Bloom, N Can. J. Fish. Aquat. Sci., 1989, 46: 1131
- Rapsomanikis, S, Donald, O F X and Weber, J H Anal. Chem., 1986, 58: 35
- 22. Raes, A MSc Thesis, University of Aberdeen, 1991