

UDC 581.134 : 547.566.1 : 582.893

104. Ko Kaneko : Biogenetic Studies of Natural Products. IV.<sup>1)</sup>  
Biosynthesis of Anethole by *Foeniculum vulgare*. (1).

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Anethole is contained in several Umbelliferae species and fennel oil contains 50~60% of anethole. For the biogenesis of anethole, Robinson has suggested the C<sub>6</sub>—C<sub>3</sub> type of condensation.<sup>2)</sup>

Nord, *et al.* described the biochemical pathway of lignin from phenylpyruvic acid by sugar-cane plant.<sup>3)</sup> Neish, *et al.* studied flavone biosynthesis from phenylpropane compounds, such as phenylalanine, cinnamic acid, caffeic acid, ferulic acid, and sinapic acid, by buckwheat and salvia plants. They concluded that the phenylpropane skeleton was incorporated into the B-ring and C-2, C-3, and C-4 of the flavone, without rearrangement of the side chain.<sup>4-6)</sup>

On the other hand, Shibata, *et al.*<sup>7,8)</sup> established the pathway from phenylalanine to ephedrine via 2-phenylethylamine and  $\omega$ -aminoacetophenone by Ephedra plant.

The biosynthetic study of anethole in comparison with flavone, lignin, and ephedrine is of interest. One of the possible ways of anethole biosynthesis is that acetyl-coenzyme-A (C<sub>2</sub> fragment) is transformed into C<sub>7</sub> fragment such as benzaldehyde, etc., but the result of feeding experiments showed that acetic acid[carboxyl-<sup>14</sup>C] was not incorporated into anethole.<sup>9)</sup> The results indicated that C<sub>7</sub>—C<sub>2</sub> type of condensation seems improbable.

In the present work, the origin of three carbons of the side chain and methoxycarbon of anethole was examined by the use of *dl*-phenylalanine[2-<sup>14</sup>C] and *dl*-methionine[methyl-<sup>14</sup>C].

### Experimental

**Cultivation of Plants**—*Foeniculum vulgare* MILLER was cultivated in the field for 3 months and it reached about 180 cm. in height, contained 0.15~0.25% of fennel oil per fresh weight, and had a few younger flowers on the top. Two kg. of the plant was harvested by severing the stem just above the ground level. The lower leaves and stems were removed, the middle and higher leaves and side stems were cut carefully, and 1 kg. of these leaves and side stems was placed in a 1-L. beaker which contained 500 cc. of half strength of Hoagland solution. This was covered with a large glass bell jar and CO<sub>2</sub>-free air was supplied, as shown in Fig. 1.

The hydroponic cultivation lasted for 5 days, during which the jar was cooled by tap water at 15~20° and the plant was illuminated by Hitachi-FL 30-W lamp during the day time. The weight of plant increased to 1150 g. during the experiment.

**Labeled Compounds**—*dl*-Phenylalanine[2-<sup>14</sup>C] and *dl*-methionine[methyl-<sup>14</sup>C] were procured from Daiichi Chem. Co., Tokyo.

**Identification of Anethole by Chromatostrip of Silica Gel**—The purity of anethole was detected by color change on the chromatostrip of silica gel with H<sub>2</sub>SO<sub>4</sub>. The silica gel chromatostrip was prepared in the following way: 30 g. of silica gel was mixed with 12 g. of 0.5% alginate acid solu-

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2) R. Robinson : "The Structural Relations of Natural Products," 1st Ed., 33(1955), Oxford University Press.

3) S. N. Acerbo, W. Walter, F. F. Nord : J. Am. Chem. Soc., 80, 1990(1958).

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5) A. S. Brown, A. C. Neish : *Ibid.*, 34, 769(1956).

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7) S. Shibata, I. Iiaseki, M. Yamazaki : This Bulletin, 5, 594(1957).

8) *Idem* : *Ibid.*, 7, 449(1959).

9) H. Mitsuhashi, K. Kaneko : Unpublished data (1958).

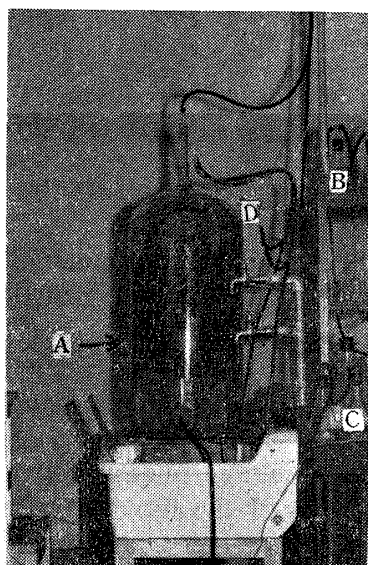


Fig. 1. Experimental Arrangement

A is the jar used for cultivation and it is supplied with CO<sub>2</sub>-free air from B trap, C trap is collection trap for CO<sub>2</sub> released from the plant. Jar A is cooled by tap water from D line.

TABLE I. Components of Hydroponic Culture Solution

	Concentration ( $\times 10^{-3}$ moles)	
	Expt. 1	Expt. 2
KNO <sub>3</sub>	2.5	2.5
Ca(NO <sub>3</sub> ) <sub>2</sub> ·4H <sub>2</sub> O	2.5	2.5
MgSO <sub>4</sub> ·7H <sub>2</sub> O	1.0	1.0
KH <sub>2</sub> PO <sub>4</sub>	0.5	0.5
Supplement	trace	trace
<i>dl</i> -Phenylalanine	6.0 (50 $\mu$ c)	6.0 (Not labeled)
<i>dl</i> -Methionine	6.0 (Not labeled)	6.0 (50 $\mu$ c)

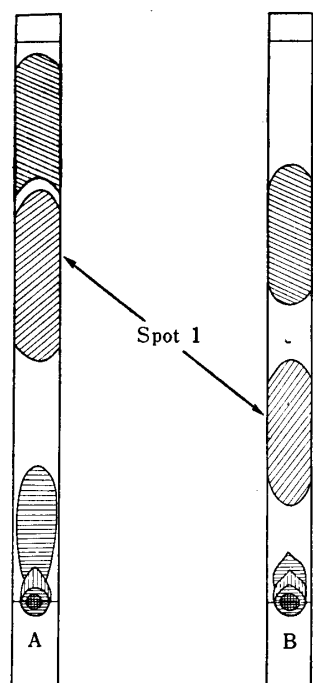


Fig. 2. Chromatostrip of Fennel Oil

A : Developed by hexane containing 15% AcOEt.

B : Developed by hexane.

Spot 1 is anethole fraction.

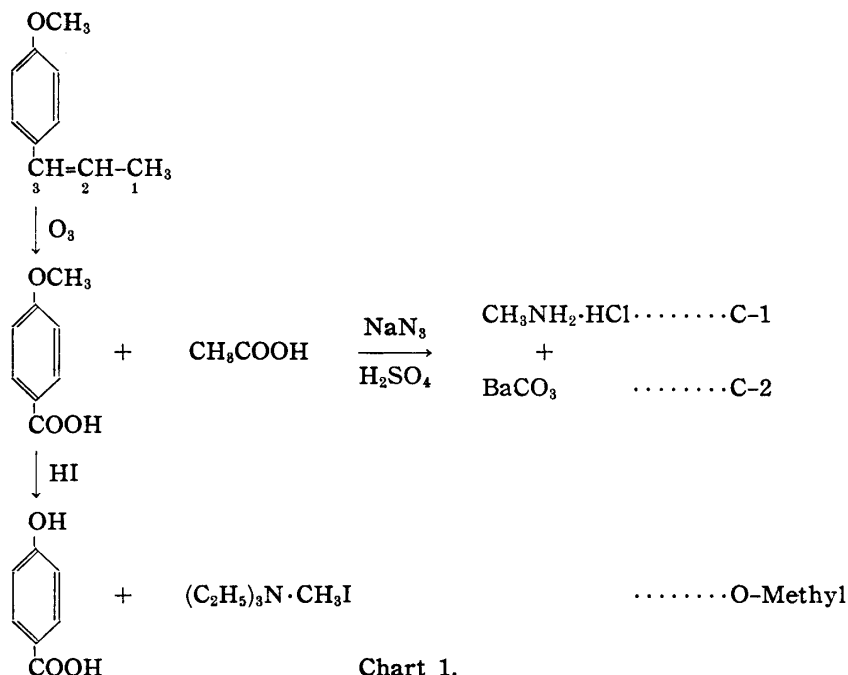
tion and 80 cc. of H<sub>2</sub>O, the mixture was extended on 1.5×3.0 cm. of glass plate, dried in an air bath at 80~90° for 1 hr., and kept in a desiccator (CaCl<sub>2</sub>) overnight.

A small volume of crude anethole was spotted on this chromatostrip which was developed with hexane containing 15% AcOEt. When the developer was evaporated the chromatostrip was sprayed with conc. H<sub>2</sub>SO<sub>4</sub>, KMnO<sub>4</sub>, and KMnO<sub>4</sub> plus dil. HCl. The change of these colors is shown in Fig. 2.

The spot corresponding to anethole showed following color changes:  $\text{H}_2\text{SO}_4$ , reddish green  $\rightarrow$  orange  $\rightarrow$  yellow;  $\text{KMnO}_4$ , brown;  $\text{KMnO}_4$  plus dil.  $\text{HCl}$ , white  $\rightarrow$  yellow.  $R_f$  value of anethole was 0.74~0.76 (hexane containing 15%  $\text{AcOEt}$ ).

**Isolation and Purification of Anethole**—Plant materials administered with labeled compounds were carefully washed with tap water, minced into small pieces, and extracted by steam distillation. The distillate was extracted with  $\text{Et}_2\text{O}$  and fennel oil obtained was 2.00 g. in Expt. 1 and 2.50 g. in Expt. 2. The yield of pure anethole was 950 mg. in Expt. 1 and 1,130 mg. in Expt. 2. The purity was examined by above-mentioned glass-strip method. Pure anethole crystallized in the refrigerator. UV:  $\lambda_{\text{max}}^{\text{EtOH}}$  268  $\text{m}\mu$ .

**Degradation of Anethole and Determination of its Radioactivity**—The scheme of degradation is shown in Chart 1. One g. of anethole was dissolved in 20 cc. of  $\text{CHCl}_3$  and 5%  $\text{O}_3$  was passed through



this solution for 3~4 hr. under cooling. After  $\text{CHCl}_3$  was evaporated, residual yellow crystals were dissolved in 20 cc. of 5%  $\text{NaOH}$  solution on a boiling water bath, then oxidized by the dropwise addition of 10 cc. of  $\text{H}_2\text{O}_2$ , and heated for 2 hr. at  $80^\circ$ . The solution was acidified by the addition of 10 cc. of  $\text{H}_3\text{PO}_4$  (sp. gr., 1.7). The crude crystals of anisic acid were collected by filtration and recrystallized from water to 830 mg. of crystals, m.p.  $182^\circ$ . UV  $\lambda_{\text{max}}^{\text{EtOH}}$   $\text{m}\mu$ : 256, 210.

The filtrate was distilled and the distillate was titrated with 0.1N  $\text{NaOH}$  to determine the amount of generated  $\text{AcOH}$ . The crude  $\text{AcONa}$  was recrystallized from  $\text{EtOH}$ ; yield, 957 mg. (88%).

A mixture of 60 mg. of  $\text{AcONa}$  with 70 mg. of  $\text{NaN}_3$  and 1.5 cc. of conc.  $\text{H}_2\text{SO}_4$  was heated for 60 min. at  $70^\circ$ ,  $\text{CO}_2$  evolved was washed with 5%  $\text{KMnO}_4$ , and was absorbed in 0.2N  $\text{NaOH}$  solution ( $\text{CO}_2$ -free).  $\text{BaCl}_2$  was added to the alkali solution with exclusion of air and the resulting  $\text{BaCO}_3$  was collected. The radioactivity of this  $\text{BaCO}_3$  is due to that of C-2 of anethole.

5 cc. of 50%  $\text{NaOH}$  solution was added to the decarboxylated solution and the generated  $\text{MeNH}_2$  was trapped with 0.1N  $\text{HCl}$  solution.  $\text{MeNH}_2\cdot\text{HCl}$  solution was evaporated and the crystals were recrystallized from  $\text{EtOH}$ ; yield, 42 mg. (70%). The activity of  $\text{MeNH}_2\cdot\text{HCl}$  crystals corresponds to that of C-1 of anethole.

A mixture of 100 mg. of anisic acid and  $\text{HI}$  (sp. gr., 1.7) was heated at  $170\sim 180^\circ$  in a modified Pregl apparatus and  $\text{MeI}$  produced was absorbed in 5%  $\text{EtOH}$  solution of  $\text{Et}_3\text{N}$ . On standing overnight, about 144 mg. of white crystals of  $\text{Et}_3\text{N}\cdot\text{MeI}$  were obtained upon evaporation of the solution.<sup>10)</sup>

Upon cooling this demethylated solution, 74 mg. of *p*-hydroxybenzoic acid was obtained, which after recrystallization from water, melted at  $212^\circ$ . Amide, m.p.  $162^\circ$ .

**Measurement of Radioactivity**—Radioactive material was burned by the wet-combustion method and the resulting  $\text{BaCO}_3$  was prepared as a thin film on a copper plate. Radioactivity was measured by the flowcounter, Low-Background Proportional Counter PC-2B (Nuclear Measurement Co.).

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## Results and Discussion

It is evident that *Foeniculum* plants metabolize phenylalanine and methionine to anethole as shown in Table II, but, in Expt. 1 and 2, the ratio of radioactivity of anethole to fennel oil is 93/100 and 12/100, respectively. On silica gel chromatography of fennel oil (Expt. 2) strong radioactivity remained near top of the column.

When phenylalanine was metabolized to anethole by the *Foeniculum* plant,  $\alpha$ -carbon of phenylalanine was completely incorporated into the C-2 of anethole as shown in Expt. 1 in Table III.

TABLE II.

	Radioactivity (cpm $\times 10^6$ )	
	Expt. 1	Expt. 2
Culture medium		
Initial	322.400	335.910
Final	146.000	152.020
Net	176.400	182.900
Fennel oil	0.116	0.357
Anethole	0.108	0.043
Respiratory carbon dioxide	0.531	0.548

TABLE III.

	Radioactivity (cpm/mM)	
	Expt. 1 ( $\times 10^4$ )	Expt. 2 ( $\times 10^3$ )
Anethole	1.33	4.17
Anisic acid	0	4.04
<p><i>p</i>-Hydroxybenzoic acid</p>	—	0
Triethylmethylammonium iodide	—	3.88
Acetic acid	1.23	0
Methylamine hydrochloride	0	—
Carbon dioxide	1.30	—

In this plant, methyl group of methionine was transmethylated to the methoxyl group of anethole but not to the side chain of anethole. This result is summarized in Expt. 2 of Table III. If the transmethylation of methionine to the side chain of anethole occurs, the radioactivity would be distributed to the methyl-carbon of acetic acid produced from the ozone oxidation of anethole, but the activity of acetic acid was completely negative.

The radioactivity of respiratory carbon dioxide was five times higher than that of anethole during the metabolism of phenylalanine[2- $^{14}\text{C}$ ] in *Foeniculum* plant. It may be suggested that the phenylpropane skeleton is metabolized by the  $\beta$ -oxidation-like breakdown, while the products of  $\text{C}_7$  compounds are not resynthesized to anethole because acetic acid[carboxyl- $^{14}\text{C}$ ] is not incorporated into anethole as mentioned before.

In view of the experiments described above it can be suggested that the phenylpropane skeleton is incorporated into anethole without rearrangement of its side chain, though not rigidly demonstrated. Additional experiments are being conducted concerning this point.

The author wishes to thank Prof. H. Mitsuhashi for his interest and advices, and is indebted to Mr. N. Yoshida for the cultivation of plants. A part of the expenses of the present work was defrayed by the Grant-in-Aid for Institutional Research from the Ministry of Education which is gratefully acknowledged.

## Summary

It was found that *dl*-phenylalanine[2- $^{14}\text{C}$ ] and *dl*-methionine[methyl- $^{14}\text{C}$ ] are respectively incorporated into the  $\alpha$ -carbon of the side chain and methyl-carbon of methoxyl group in anethole molecule produced by *Foeniculum vulgare* MILLER, *in vivo*.

(Received November 13, 1959)