This assumption is supported by the fact that the observed coupling constant of the proton at carbon-3 is larger than that of other protons in ESR spectrum of the free radical produced from I,7 and the final step, deoxygenation, would take place similarly by a free radical course as in the case of deoxygenation of 4-nitroquinoline 1-oxide. The formation of 3,3'-biquinoline compounds and deoxygenation of N-oxide were also recognized in the reaction of 4-nitroquinoline 1-oxide and 4-hydroxyquinoline 1-oxide, indicating that a free radical would be present as an intermediate. Further studies are in progress on the chemistry of 4-hydroxyamino-quinoline 1-oxide and 4-nitroquinoline 1-oxide, in view of the free radical reactivity in relation to their carcinogenic activity.

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Isolation and Structures of New Pregnane Derivatives from Adonis amurensis Regel et RADD

In a previous communication,¹⁾ we reported the isolation and the structure of a non-cardiac aglycone, adonilide, from *Adonis amurensis* Regel et Radd. In this communication, we wish to describe the isolation and the structures of three novel non-cardiac aglycones: fukujusone (I), ester A (V) and ester B (VI).

Fukujusone (I), mp $224-227^{\circ}$, $[\alpha]_{\text{p}}+100.0^{\circ}$ (c=1.0, CHCl₃) has a formula $C_{21}H_{32}O_{4}$ (Anal. Calcd. for $C_{21}H_{32}O_{4}$: C, 72.38; H, 9.26. Found: C, 72.61, H, 9.07. molecular peak: m/e 348). The infrared (IR) spectrum ($p_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 3550, 3529, 1680) demonstrates the presence of hydroxyl groups and a carbonyl function. The NMR spectrum shows the following signals: τ (in CDCl₃) 8.80 (3H, singlet, CH₃), 8.78 (3H, singlet, CH₃) 7.72 (3H, singlet, -COCH₃), 7.10 (1H, multiplet), 6.50 (1H, broad multiplet, CH –OH), 4.60 (1H, multiplet, vinylic proton). Acetylation of I with acetic anhydride in pyridine afforded a monoacetate (II), mp 196—198°, $[\alpha]_{\text{p}}+61.5^{\circ}$ (c=1.39, CHCl₃), (Anal. Calcd. for $C_{23}H_{34}O_{5}$: C, 70.74; H, 8.78. Found: C, 70.98; H, 8.95). The IR spectrum ($p_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 35600, 3400, 1720, 1680) shows the presence of hydroxyl groups which are not acetylable under this condition. The nuclear magnetic resonance (NMR) spectrum of II has the following signals: τ (in CDCl₃) 8.80 (3H, singlet, CH₃), 8.72 (3H, singlet, CH₃), 7.90 (3H, singlet, -COCH₃), 7.70 (3H, singlet, -COCH₃), 7.10 (1H, multiplet, -CH-CO-), 5.30 (1H, broad multiplet, CH -OAc), 4.50 (1H, multiplet, vinylic proton). From the above data, we concluded that fukujusone (I) has a secondary hydroxyl group, a methyl ketone, two angular methyl groups, and a vinylic proton. Considering the structural simil-

⁷⁾ N. Kataoka, A. Imamura, Y. Kawazoe, and G. Chihara, Bull. Chem. Soc. Japan, 40, 62 (1967).

⁸⁾ T. Kosuge, H. Adachi, M. Yokota, and T. Nakao, Yakugaku Zasshi, 85, 66 (1965).

¹⁾ Y. Shimizu, Y. Sato and H. Mitsuhashi, Chem. Pharm. Bull. (Tokyo), 15, 2005 (1967).

arities expected from the biogenetic view point, it was assumed that fukujusone (I) also comprises a partial structure of 3β -hydroxy-5-pregnen-20-one, as do the other isolated compounds. This was supported by the splitting pattern of signals at τ 6.50 and 4.60 characteristic of 5-en- 3β -ol, and the mass spectrum data (vide infra). The absence of other signals in the τ 4—6 region of the NMR spectrum suggests that the remaining two oxygens exist as tertiary hydroxyl groups. The presence of 17-hydroxyl group was excluded, because the multiplet at τ 7.10 was considered to be the C-17-methine proton, as exemplified later by the formation of the 17-iso-derivative. Acetylfukujusone (II) consumed one mole of lead tetraacetate, indicating the tertiary alcohols exist as an α -glycol. And the fact that both angular methyl groups are considerably deshielded made us assign one of the alcohols to 8β -position.²⁾ Accordingly, the other one should be located at either C-9 or C-14 position. The 8β , 14β -glycol is known to form a cyclic sulphite with SOCl₂ rather than to undergo dehydration.³⁾ Thus treatment of (II) with SOCl₂ in pyridine gave a five-membered cyclic sulphite, (III), mp 195—198°. IR $v_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 1213—1215 $\binom{O}{O}$ S=O). The remaining problem is the stereochemistry at C-17, and it was established by the following evidence.

I: R=H, 17β II: R=Ac, 17β IV: R=H, 17α Being treated with a solution of 5% methanolic potassium hydroxide, (I) isomerized predominantly to isofukujusone (IV), mp 212—217°, $[\alpha]_{\text{\tiny D}}$ —31.0° (c=1.08, CHCl₃) (Anal. Calcd. for C₂₁H₃₂O₄: C, 72.38; H, 9.26. Found: C, 72.48; H, 9.52. molecular peak at: m/e 348); $\nu_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 3600, 3590, 3580, 1680: τ (in CDCl₃) 8.84 (3H, singlet, CH₃), 8.64 (3H, singlet, CH₃), 7.86 (3H, singlet, -COCH₃), 6.72 (1H, multiplet, -CHCO-), 6.50 (1H, broad multiplet, CH-OH), 4.68 (1H, multiplet, vinylic

$$CH_3$$
 CO
 Me
 CH_3
 CO
 Me
 CO
 Me
 CH_3
 CO
 Me
 CO

 $\begin{array}{llll} m/e & 330(\mathrm{M-H_2O}), & 315(\mathrm{M-H_2O-CH_3}), & 312(\mathrm{M-2H_2O}), & 297(\mathrm{M-2H_2O-CH_3}), \\ 287(\mathrm{M-H_2O-CH_3CO}), & 279(\mathrm{M-3H_2O-CH_3}), & 269(\mathrm{M-2H_2O-CH_3CO}), & 251(\mathrm{M-3H_2O-CH_3CO}), & 192(\mathrm{B-H_2O}), & 177(\mathrm{B-H_2O-CH_3}), & 149(\mathrm{B-H_2O-CH_3CO}), \\ 134(\mathrm{B-H_2O-CH_3-CH_3CO}), & 123(\mathrm{B-2H_2O-CH_3-CH_3CO}), & 105(\mathrm{A-CH_3}). \end{array}$

Chart 1. Mass Fragmentation of Fukujusone (I)

²⁾ a) K. Tori and E. Kondo, Tetrahedron Letters, 1963, 645; b) Y. Shimizu and H. Mitsuhashi, Tetrahedron, 24, 4143 (1968).

A. von Wartburg and J. Renz, Helv. Chim. Acta, 42, 1639 (1959); A. von Wartburg and J. Renz, ibid.,
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proton). The optical rotatory dispersion (ORD) curve of IV shows a negative Cotton effect (trough $[\phi]_{309}$ —2800°, peak $[\phi]_{268}$ +4240°, in MeOH), while the ORD curve of I possesses a maximum peak at 304 m μ ($[\phi]_{304}$ +3170°) with the trough covered under the strong positive background. This relation between the ORD sign and equilibration is that of 14 β -pregnan-20-one with C/D-cis ring juncture, where the 17 α -orientation is more stable.⁴)

The mass spectrum data of I and IV are very similar and fully compatible with the structures. The base peaks, m/e 120, m/e 210 and the other prominent peaks could be accounted for by the following scheme (Chart 1).

Thus, we believe that fukujusone has the structure of 3β ,8 β ,14 β -trihydroxy pregn-5-en-20-one. It is of some biogenetic interest that fukujusone corresponds to the 12-desoxy derivative of isolineolon which was also found in the same plant.

Ester A (V), mp 250—254°, [α]_D +46° (c=1.26, CHCl₃) has a formula C₂₇H₃₅O₆N (Anal. Calcd. for C₂₇H₃₅O₆N: C, 69.06; H, 7.51, N, 2.98. Found: C, 69.53; H, 7.87, N, 3.24). The ultraviolet (UV) spectrum of V has an absorption maximum at $\lambda_{\text{max}}^{\text{ECOH}}$ 264 m μ (log ε 3.60). From the IR spectrum ($\nu_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 3600, 3500, 3459, 1680, 1720, 1595), the presence of hydroxyl, carbonyl, ester, and aromatic groups was anticipated. The NMR spectrum exhibits the following signals: τ (in CDCl₃) 8.75 (3H, singlet, CH₃), 8.60 (3H, singlet, CH₃), 7.76 (3H, singlet, CH₃), 6.76 (1H, multiplet, -CHCO-), 6.50 (1H, broad multiplet, CH-OH), 5.00 (1H, quartet, CH-O-CO-), 4.60 (1H, multiplet, vinylic proton), 1.60—2.50 (4H, aromatic proton).

Hydrolysis of (V) with 5% methanolic potassium hydroxide gave an amino acid, which was separated by ion-exchange resin and identified as nicotinic acid. The neutral fraction was a mixture of two substances, lineolon (VII) and isolineolon (VIII), which were identified with the samples isolated from *Cynanchum caudatum*. Therefore, ester A (V) should be formulated as 12-nicotinoyllineolon or -isolineolon. The location of the ester linkage was deduced by the splitting pattern of the hydrogen adjacent to the esterified hydroxyl group (quartet, J=5 and 11 cps). The authors believe that the ester A is the first example of a plant steroid conjugated with an amino acid.

$$CH_3$$
 RO
 CO
 OH
 OH
 $V: R = CO-, 17\beta$
 $VI: R = Phenyl-CO-, 17\beta$
 $VII: R = H, 17\alpha$
 $VIII: R = H, 17\beta$

The other ester aglycone, ester B (VI), mp $245-250^{\circ}$, $[\alpha]_{\text{p}}+45^{\circ}$ (c=0.82, CHCl₃) has a formula, $C_{28}H_{36}O_{6}\cdot 1/2H_{2}O$ (Anal. Calcd. for $C_{28}H_{36}O_{6}\cdot 1/2H_{2}O$: C, 70.44 H, 7.80. Found: C, 71.06 H, 7.80). The UV spectrum of VI has an absorption maximum at $\lambda_{\text{max}}^{\text{EioH}}=233 \text{ m}\mu$ (log ε 4.10), 278 m μ (log ε 3.13). The IR spectrum ($\nu_{\text{max}}^{\text{Nijol}}$ cm⁻¹: 3620, 3600, 3500, 1720, 1690, 1590, 1270) shows the presence of hydroxyl, carbonyl, ester and aromatic groups. The hydrolysis of VI with 5% methanolic potassium hydroxide gave an acidic substance, which was identified with benzoic acid. The neutral fraction consisted of lineolon (VII) and isolineolon (VIII). Thus VI is a monobenzoyl ester of lineolon or isolineolon. However, ester B (VI) is different from the substance G reported by Abisch, et al.5 but identical with the sample isolated from Cynanchum boerhavifolium, and its structure will be published in a different paper.6 In addition, free lineolon (VII) and isolineolon (VIII) were isolated from the polar fraction of the aglycone mixture.

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Occurrence of "NIH Shift" during Hydroxylation of Aromatic Steroid

It has recently been discovered that during enzymatic hydroxylation of aromatic substrates the substituent (²H, ³H, Cl, Br, *etc.*) displaced by the entering hydroxyl group migrates to an adjacent position. These phenomena called "NIH shift" have been demonstrated with enzymes derived from animal and plant sources.^{1,2)} We have previously reported that hydroxylation does take place at C-2 and C-3 when 3-deoxyestrone (I) is orally given to rabbit.³⁾ Therefore it seemed to be of considerable interest to us to elucidate whether the aromatic steroid would similarly undergo "NIH shift" or not. We now wish to report the *in vivo* hydroxylation of the specifically deuterated 3-deoxyestrone.

I: $R_1=R_2=H$, $R_3=O$ II: $R_1=OH$, $R_2=H$, $R_3= \stackrel{OH}{H}$

 $II: R_1=1$ -phenyl-5-tetrazolyloxy-, $R_2=H$, $R_3=\langle {}_{H}^{OH}$

 \mathbb{N} : $R_1=D$, $R_2=H$, $R_3=O$ \mathbb{V} : $R_1=H$, $R_2=OH$, $R_3=O$

 $VI: R_1=H, R_2=1$ -phenyl-5-tetrazolyloxy-, $R_3=O$

 $M: R_1=H, R_2=D, R_3=O$

First, 2-deuteriosteroid was prepared as a substrate from 2-hydroxy-3-deoxyestradiol (II) in three steps. Condensation of II with 1-phenyl-5-chlorotetrazole in the presence of potassium carbonate⁴⁾ gave the 2-(1-phenyl-5-tetrazolyl) ether (III), mp 140—141°, as colorless needles (from aq. acetone). Catalytic reduction over palladium—on–barium carbonate under a stream of deuterium gas followed by oxidation with Jones reagent furnished the desired 2-deuterio-3-deoxyestrone (IV), mp 141—142°, as colorless needles (from ether). Likewise 3-deuterio-3-deoxyestrone (VII) was also synthesized starting from estrone (V) by way of 3-(1-phenyl-5-tetrazolyl) ether (VI), mp 204—206° (from aq. acetone). The distribution and quantity of the isotope in these selectively labelled steroids were determined by means of nuclear magnetic resonance and mass spectrometries.

A single dose of suspension of 2-deuterio-3-deoxyestrone (IV) (475 mg) in Tween 80 was orally given to a male rabbit weighing about 2.3 kg. The urine was collected for the following

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