

# Nuclear Magnetic Resonance of Semicarbazones and Thiosemicarbazones of an Aliphatic Aldehyde<sup>1)</sup>

HARUO OGURA, TSUNEO ITOH, TAKASHI OKAMOTO,  
and SATOSHI OMURA

*College of Pharmaceutical Sciences, Kitasato University<sup>2)</sup>*

(Received August 14, 1968)

In the previous report,<sup>3,4)</sup> it was described that leucomycin A<sub>3</sub>, a family of macrolide antibiotic, contains an aliphatic aldehyde group such as -CH<sub>2</sub>CHO, from the value of the ultraviolet absorption and nuclear magnetic resonance (NMR) spectra of the thiosemicarbazone of leucomycin A<sub>3</sub>. Evans and Gillam<sup>5)</sup> studied the ultraviolet absorption spectra of aldehydes, and those of their semicarbazones and thiosemicarbazones, and showed that the thiosemicarbazones of saturated aldehydes exhibit absorption bands near 270 mμ which are some 500 times more intense than those of the parent carbonyl compound. It would therefore seem necessary that simple type of thiosemicarbazones of saturated aldehydes have absorption at around 230 and 270 mμ. Table I shows the corresponding ultraviolet absorption data on the thiosemicarbazones of leucomycin A<sub>1</sub> and A<sub>3</sub>, magnamycin B,<sup>6)</sup> and spiramycin B,<sup>7)</sup> and it is clear from these data that the results closely parallel those of analogous saturated aldehydes.

On the other hand, our interest in the problems of NMR on the multiplicity and coupling constant of the aldehyde proton of leucomycin A<sub>3</sub><sup>3,4)</sup> have led us to examine the NMR spectra of the thiosemicarbazones and semicarbazones of aliphatic aldehydes. Because of a very small coupling constant of the aldehyde itself (1-3 cps),<sup>8)</sup> these derivatives are suitable not only for calculating the coupling constant by the enlargement of the coupling constant to 4-7 cps, but also give bands in the region of 7-8 ppm<sup>9)</sup> by the shielding effect of derivatives. Bhacca and Williams<sup>10)</sup> indicated the aldehyde proton resonance in the NMR spectrum of 20-formylprogn-4-en-3-one, and its chemical shift and 9.58 ppm and the coupling constant to C-20 methine hydrogen was 3 cps. Karabatsos and his co-workers<sup>8,9,11)</sup> studied the conformation and configuration of several aldehydes and ketones, and their oxime O-methyl ethers, and N-methylphenylhydrazones.

In the present investigation, the coupling constant of the aldehyde proton to hydrogens attached to the neighboring carbon atom was studied in the NMR spectra of saturated aliphatic aldehydes, and their thiosemicarbazones and semicarbazones. The obtained data are summarized in Tables I and II.

- 1) Abstract of Papers, the 24th Annual Meeting of the Pharmaceutical Society of Japan, Kyoto, April 1967, p. 517.
- 2) Location: Shiba Shirogane Sankochō, Minato-ku, Tokyo.
- 3) S. Omura, H. Ogura, and T. Hata, *Tetrahedron Letters*, 1967, 609, 1267.
- 4) S. Omura, M. Katagiri, H. Ogura, and T. Hata, *Chem. Pharm. Bull.* (Tokyo), 15, 1529 (1967).
- 5) L.K. Evans and A.E. Gillam, *J. Chem. Soc.*, 1943, 565.
- 6) F.A. Hochstein and K. Murai, *J. Am. Chem. Soc.*, 76, 5080 (1954).
- 7) R. Corbaz, L. Ettlinger, E. Gaumann, W. Keller-Schierlein, F. Kradolfer, E. Kyburz, L. Neipp, V. Prelog, A. Wettstein, and H. Zahner, *Helv. Chim. Acta*, 39, 304 (1956).
- 8) G.J. Karabatsos and N. Hsi, *J. Am. Chem. Soc.*, 87, 2864 (1965).
- 9) G.J. Karabatsos and N. Hsi, *Tetrahedron*, 23, 1079 (1967).
- 10) N.S. Bhacca and D.H. Williams, "Applications of NMR Spectroscopy in Organic Chemistry," Holden-Day, Inc., San Francisco, 1964, pp. 76-77.
- 11) G.J. Karabatsos, R.A. Taller, and F.M. Vane, *Tetrahedron Letters*, 1964, 1081; G.J. Karabatsos and R.A. Taller, *J. Am. Chem. Soc.*, 86, 4373 (1964).

Kuehne and Benson<sup>12)</sup> have recently reported that magnamycin A and B had shown a singlet aldehyde proton (9.50 ppm), while spiramycins had shown a doublet at 9.65 ppm. This observation initially suggested the structure of magnamycin would require an aldehyde function attached to a quaternary carbon atom, and spiramycin would require an aldehyde attached to a secondary carbon atom. In the case of leucomycins, the expected aldehyde proton triplet could not be observed and was indicated as a poor split in Table I. However, conversion of leucomycin A<sub>1</sub> and A<sub>3</sub>, and magnamycin B to their thiosemicarbazones resulted in the splitting appearing as the expected triplet at 7.00, 7.62, and 7.12 ppm ( $J=5.0-5.1$  cps), respectively.

The NMR spectrum of spiramycin B shows a doublet at 9.80 ppm ( $J=3.0$  cps). The thiosemicarbazone of spiramycin B shows the band at 7.28 ppm ( $J=5.4$  cps) in dioxan.

The thiosemicarbazone of chloral had already been shown to have the true hydrate structure from its infrared spectral studies.<sup>13)</sup> This was confirmed from the NMR spectra.

TABLE I. Ultraviolet Absorption and NMR Spectra of Aldehydes and Their Thiosemicarbazones

Compound Secondary aldehyde ( $R \cdot CH_2CHO$ ) $R$	mp (°C)	Thiosemicarbazone Ultraviolet absorption		Aldehyde in CCl <sub>4</sub>	NMR (ppm, cps) Thiosemicarbazone	
		$\lambda_{max}^{EtOH}$	$m\mu$ (log $\epsilon$ )		in Me <sub>2</sub> SO	in dioxan
H	148—149	241 (3.98), 269 (4.04)		9.66 q. 2.8 <sup>a)</sup>	7.49 q. 5.2	7.30 q. 5.4
CH <sub>3</sub>	155—156	229 (3.88), 270 (4.38)		9.70 t. 1.5	7.48 t. 4.2	7.28 t. 5.4
C <sub>2</sub> H <sub>5</sub>	109—110	230 (3.89), 270 (4.74)		9.69 t. 1.5	7.42 t. 5.1	7.27 t. 5.1
<i>n</i> -C <sub>3</sub> H <sub>7</sub>	66	230 (3.98), 271 (4.47)		9.66 t. 1.8	7.47 t. 5.1	7.28 t. 5.1
<i>iso</i> -C <sub>3</sub> H <sub>7</sub>	48	230 (3.99), 270 (4.44)		9.72 t. 1.8	7.47 t. 5.1	7.24 t. 4.5
<i>n</i> -C <sub>5</sub> H <sub>11</sub>	89—90	230 (3.93), 271 (4.40)		9.67 t. 1.5	7.46 t. 5.1	7.28 t. 5.1
<i>n</i> -C <sub>6</sub> H <sub>13</sub>	65—66	230 (3.98), 271 (4.37)		9.61 t. 1.5	7.45 t. 5.4	7.29 t. 5.1
<i>n</i> -C <sub>7</sub> H <sub>15</sub>	77—78.5	230 (4.01), 271 (4.45)		9.68 t. 1.5	7.46 t. 5.1	7.26 t. 5.4
Leucomycin A <sub>1</sub>	158—162	228 (4.48), 269 (4.33)				7.00 t. 5.1
Leucomycin A <sub>3</sub>	138—141	232 (4.54), 271 (4.37)		9.56 poor split		7.62 t. 5.0
Magnamycin B	135—138	235 (3.98), 273 (4.60)		9.58 poor split		7.12 t. 5.1
Tertiary aldehyde ( $\begin{smallmatrix} R \\ \diagup \\ R-CHCHO \end{smallmatrix}$ )						
CH <sub>3</sub>	86—87	230 (3.85), 270 (4.28)		9.54 d. 0.6	7.40 d. 5.4	7.26 d. 5.4
C <sub>2</sub> H <sub>5</sub>	92—93.5	230 (3.83), 270 (4.32)		9.46 d. 1.2	7.31 d. 6.6	7.20 d. 6.0
Spiramycin B	151—155	230 (4.55), 270 (4.35)		9.80 d. 3.0		7.28 d. 5.4
Quaternary aldehyde ( $\begin{smallmatrix} R \\ \diagup \\ R-CCHO \\ \diagdown \\ R' \end{smallmatrix}$ )						
CH <sub>3</sub>	113—114	228 (3.81), 270 (4.33)		9.37 s.	7.33 s.	7.13 s.
Cl	122—123 <sup>b)</sup> (decomp.)	245 (4.02)		8.96 s.	7.40 d. 5.4	notsoluble

a) in deuteriochloroform b) reported<sup>13)</sup> mp 104—105° (decomp.)

TABLE II. Ultraviolet Absorption and NMR Spectra of Aldehydes and Their Semicarbazones

Compound	mp (°C)	Ultraviolet absorption		Aldehyde in CCl <sub>4</sub>	NMR (ppm, cps) Semicarbazone	
		$\lambda_{max}^{EtOH}$	$m\mu$ (log $\epsilon$ )		in Me <sub>2</sub> SO	in dioxan
CH <sub>3</sub> CHO	160—161	230 (4.08)		9.66 q. 2.8	7.11 q. 5.4	notsoluble
CH <sub>3</sub> (CH <sub>2</sub> ) <sub>4</sub> CHO	109—110	230 (4.07)		9.67 t. 1.5	7.13 t. 5.4	7.01 t. 5.4
(CH <sub>3</sub> ) <sub>2</sub> CHCHO	123—124	234 (4.09)		9.54 poor split	7.06 d. 4.8	6.97 d. 5.4
(CH <sub>3</sub> ) <sub>3</sub> CHO	194	228 (4.08)		9.37 s.	7.33 s.	7.13 s.

12) M.E. Kuehne and B.W. Benson, *J. Am. Chem. Soc.*, **87**, 4660 (1965).

13) P.K. Chang and T.L.V. Ulbricht, *J. Am. Chem. Soc.*, **80**, 976 (1958).

Chloral itself showed the expected aldehyde proton singlet at 8.96 ppm, but the chloral thiosemicarbazide showed the elusive splitting as a doublet at 7.40 ppm ( $J=5.4$  cps).

### Experimental<sup>14)</sup>

**Semicarbazones and Thiosemicarbazones**—Derivatives were prepared by the usual procedures.<sup>15)</sup> Their melting point, ultraviolet absorption, and NMR spectral data are summarized in Tables I and II.

- 14) All melting points are uncorrected. The NMR spectra were obtained with Hitachi H-60 spectrometer at 60 Mc and tetramethylsilane was used as an internal reference.  
 15) R.L. Shriner, R.C. Fuson, and D.Y. Curtin, "The Systematic Identification of Organic Compounds," 4th ed., John Wiley and Sons, Inc., New York, 1956, pp. 218—219.

[Chem. Pharm. Bull.,  
17(4) 846—848 (1969)]

UDC 547.92.07 : 547.834.2.07

## Studies on the Azasteroids and Related Compounds. IV.<sup>1)</sup> Reactions of 1,2,3,4,6a,7,8,9,10,10a-Decahydro-6H-benzo[c]quinolizin-6-one

ZEN-ICHI HORII, KOICHI MORIKAWA,<sup>2)</sup>  
and ICHIYA NINOMIYA<sup>2a)</sup>

Faculty of Pharmaceutical Sciences, Osaka University<sup>2)</sup>

(Received September 4, 1968)

In continuation of our synthetic studies on 14-azasteroids,<sup>1,3,4)</sup> which included preparation and some reactions of B/C *trans* fused decahydro-6H-benzo[c]quinolizin-6-one (I), we now describe angular methylation at C-4a position of I and attempt to prepare the benzo[c]quinolizine derivatives with functional group at C-4 position which corresponds to C-17 position in natural steroid structure, from the compound (I).

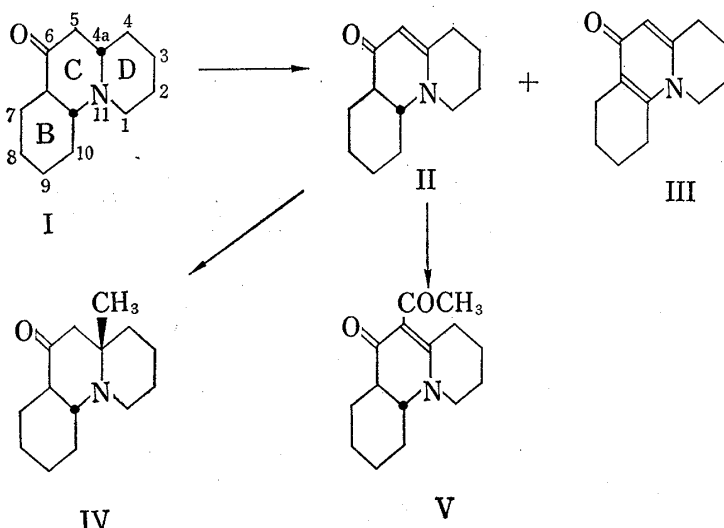


Chart 1

Dehydrogenation of the quinolizidinone (I) with mercuric acetate in 35% acetic acid solution afforded a mixture of decahydrobenzo[c]quinolizin-6-one (II) and the  $\gamma$ -pyridone (III). The former was methylated at the angular position

- 1) Part III: Z. Horii, K. Morikawa, C. Iwata, and I. Ninomiya, *Chem. Pharm. Bull.* (Tokyo), **16**, 1686 (1968).  
 2) Location: Toneyama, Toyonaka, Osaka; a) Present Address: Kobe Women's College of Pharmacy, Nakano, Motoyama, Higashinada, Kobe.  
 3) Z. Horii, K. Morikawa, and I. Ninomiya, *Chem. Pharm. Bull.* (Tokyo), **14**, 1399 (1966).  
 4) Z. Horii, K. Morikawa, and I. Ninomiya, *Chem. Pharm. Bull.* (Tokyo), **16**, 1472 (1968).