bly counteracted by administration of the moulting hormone (Table II). This suggests that DAEA causes lack of moulting hormones due to an additional block in the metabolic pathway from cholesterol to moulting hormones. Studies designed to further our understanding of the effects of DAEA on sterol metabolism in *Bombyx* are now in progress.

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The Stereochemistry of Enzymatic Aromatization of 19-Norsteroids

The bioconversion of C_{19} steroids into estrogens requires two general steps: (1) hydroxylation and subsequent loss of the C-19 methyl group, and (2) removal of hydrogens from both C-1 and C-2 resulting in a double-bond in ring A.¹⁾ The 19-norsteroids also serve as estrogen precursors in the placental and ovarian tissues²⁾ and hence are useful model compounds for the study of the aromatization reaction. In addition the metabolism of the 19-norsteroids is of particular interest in itself, since they are widely used for the contraceptive and chemotherapeutic purposes. Recently the β -cis nature of C-1,2 dehydrogenation in the enzymatic aromatization was established with C_{19} steroids.³⁾ Concerning the 19-norsteroids, however, the evidence only for the elimination of the 1β -hydrogen in this process has been demonstrated.⁴⁾ We now wish to report on the stereochemistry of hydrogen loss from C-2 during the placental aromatization of estr-4-ene-3,17-dione.

An initial project was focused to the synthesis of the substrates stereospecifically labeled with tritium in the 2β and 2α positions of estrenedione. The preparation of the desired substrates was carried out along the way which had already been developed for epimeric 2-deuterioestr-4-ene-3,17-diones.⁵⁾ Reduction of 2β ,3 β -epoxyestrane-5 β ,17 β -diol (II) derivable from estr-2-ene-5 β ,17 β -diol (I) with [³H]LiAlH₄ yielded the [2α -³H]3 β ,5 β ,17 β -triol (III). Usual acetylation and dehydration with thionyl chloride in pyridine gave [2α -³H]estr-4-ene-

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 3β ,17 β -diol diacetate (IV). Hydrolysis with alkali and subsequent oxidation with chromium trioxide-pyridine complex afforded one of the desired substrates, $[2\alpha^{-3}H]$ estr-4-ene-3,17-dione (V). Hydroboration of I with tritiated diborane, which was generated from lithium aluminum tritiide and boron trifluoride, followed by oxidation gave the $[2\beta^{-3}H]3\beta$,5 β ,17 β -triol (VI). Transformation of VI into the other substrate, $[2\beta^{-3}H]$ estr-4-ene-3,17-dione (VII), was performed in the same manner as described above for the 2α -labeled substrate. These tritiated compounds were mixed with an appropriate amount of $[4^{-14}C]$ estr-4-ene-3,17-dione and repeatedly recrystallized to constant isotope ratio, respectively.

Tracental Afonatization		
	$^3\mathrm{H}/^{14}\mathrm{C}^a$)	% 3H los
[2a-3H]Estr-4-ene-3,17-dione (V)	37.8	
Estrone	30.1	20
1β-Hydroxyestr-4-ene-3,17-dione	30.3	
Recovered substrate	35.6	
$[2\beta$ -3H]Estr-4-ene-3,17-dione (VII)	37.6	
Estrone	8.0	7 9
1β -Hydroxyestr-4-ene-3,17-dione	29.2	
Recovered substrate	35.0	

TABLE I. Tritium Loss of Products and Substrates in Placental Aromatization

The labeled substrates were incubated with human placental preparation according to the method of Ryan.⁶⁾ The incubation mixture consisted of $50 \,\mu g$ of the steroid substrate, placental $10000 \times g$ supernatant equivalent to $16 \, g$ wet weight of tissue and NADPH-generating system suspended in $11 \, \text{ml}$ of $0.05 \, \text{m}$ phosphate buffer, pH 7.0. After incubation at 37° for 1 hr, the mixture was extracted with ethyl acetate and the extract was separated by thin-layer chromatography on silica gel. The products and unchanged substrate thus obtained were diluted with carrier and then recrystallized to constant specific activity, respectively.

The results of the incubations are collected in Table I. Estrone formed from VII with the label in the 2β -position exhibited a 79% loss of tritium. In contrast the 2α -labeled substrate (V) lost only 20% tritium. These figures unambiguously tell us that the hydrogen loss from C-2 in the aromatization process is stereoselectively β and in consequence C-1, 2 hydrogen elimination is β -cis. The 1β -hydroxylated products obtained from both incubations showed almost the same tritium content with a slight decrease. The retention of tritium in the recovered substrates implies that the hydrogen removal from C-2 is not a reversible process.

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a) expressed as ratio of disintegration per minute (dpm)

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