

Pharmaceutical Institute,
Tohoku University,
Aoba-yama, Sendai, Japan

TOMIHISA OHTA
HIROSHI HIKINO

Received August 11, 1980

[Chem. Pharm. Bull.]
29(1) 282—285 (1981)]

Structures of Four New Triterpenoidal Oligoglycosides, Bivittoside A, B, C, and D, from the Sea Cucumber *Bohadschia bivittata* MITSUKURI

On the basis of chemical and physicochemical evidence, the structures of four triterpenoidal oligoglycosides, bivittoside A, B, C, and D from the sea cucumber *Bohadschia bivittata* MITSUKURI, have been elucidated as **6**, **8**, **9**, and **10**, respectively. A new homoannular dienic sapogenol was isolated as the acetate and the highly strained structure (**4a**) has been elucidated.

Keywords—sea cucumber; *Bohadschia bivittata*; lanostane-type triterpenoid; oligoglycoside; bivittoside; strained homoannular diene; *Turbo cornutus* glycosidase

During the course of systematic studies on the biologically active constituents of echinoderm,¹⁾ we have recently isolated four new triterpenoidal oligoglycosides, named bivittoside A (**6**), B (**8**), C (**9**), and D (**10**), from the sea cucumber *Bohadschia bivittata* MITSUKURI collected in Okinawa Prefecture in July. This communication deals with the evidence being consistent with the proposed structures.²⁾

The MeOH extract of the Cuvierian tubes of *B. bivittata* afforded bivittoside A, B, C, and D, after solvent-fractionation and chromatographic separation, in 2, 2, 2, and 8% yields (respectively from the MeOH ext.).

Bivittoside A (**6**), $C_{41}H_{66}O_{12} \cdot H_2O$,³⁾ mp 267—268°, $[\alpha]_D +9^\circ$ (pyr.), UV (MeOH): transparent above 210 nm, shows the infrared (IR) absorption bands [3400 (br), 1070 (br) cm^{-1}] characteristic to glycoside and the band due to γ -lactone (1750 cm^{-1}). The circular dichroism (CD) spectrum (MeOH) of bivittoside A demonstrates chirality of the γ -lactone moiety by a negative maximum: $[\theta]_{222} -7800$. On acidic hydrolysis, bivittoside A furnished the dienic artifact sapogenol seichellogenin (**1**),⁴⁾ a dihydroxy-triterpene lactone (**2**), $C_{30}H_{48}O_4$, mp 205—207°, $[\alpha]_D -21^\circ$ ($CHCl_3$), IR (KBr): 3350 (br), 1753 (br) cm^{-1} , and one mole each of D-xylose and D-quinovose, and another minor sapogenol (*vide post*). The structure of **2** having the 9(11)-en-12 β -ol moiety has been corroborated by the proton nuclear magnetic resonance (1H -NMR) signals observed at δ 4.39 (1H, m, $W_{h/2}=12$ Hz, 12 α -H) and δ 5.12 (1H, br.s, $W_{h/2}=6$ Hz, 11-H) and the CD spectrum (MeOH): $[\theta]_{204} +32000$ (pos. max.) [9(11)-ene],^{5b)} and also by the ready conversion of **2** giving **1** on acidic treatment. The unknown configuration at C-20 of seichellogenin (**1**) has been now defined S as based on the 1H -NMR analysis utilizing the pyridine-induced shift (Table).^{1,5)}

Methylation⁶⁾ of bivittoside A gave the fully methylated hexa-O-methyl derivative (**6a**) [two β -anomeric proton signals at δ 4.33 and 4.62 (1H both, d, $J=7$ Hz)], which, on methanolysis, liberated methyl pyranosides of 2,3,4-tri-O-methylquinovose and 3,4-di-O-methylxylose. Presence of the 9(11)-en-12 α -ol moiety in bivittoside A has been substantiated by the 1H -NMR signals observed at δ 4.52 (1H, d, $J=4$ Hz, 12 β -H) and δ 5.70 (1H, d, $J=4$ Hz, 11-H)^{5b)} and the CD spectrum (MeOH): $[\theta]_{212} +8100$ and by oxidation with CrO_3 -pyridine-*n*-BuOH-aq. H_2SO_4 ⁷⁾ providing the 12-keto derivative (**7**), $C_{41}H_{64}O_{12} \cdot 2H_2O$, mp 263—264°, $[\alpha]_D +12^\circ$

(pyr.), UV (MeOH): 256 nm ($\epsilon=10000$). The latter conversion simultaneously shows that the oligosaccharide moiety in bivittoside A attaches to 3β -OH of the sapogenol. Based on the above-mentioned evidence, the structure of bivittoside A has been elucidated as **6**. The C-12 configuration of **6** has been inverted during the acidic hydrolysis to furnish **2**.

Isolation of the above-mentioned minor sapogenol was effected after acetylation. The monoacetate (**4a**), $C_{32}H_{48}O_4$, mp 172–173°, $[\alpha]_D -299^\circ$ ($CHCl_3$), shows two olefinic proton signals at δ 5.85 and 6.09 (1H each, ABq, $J=9$ Hz) in the 1H -NMR spectrum. It gave seychellogenin acetate (**1a**) on acidic treatment, thus presence of the 8,11-diene moiety being suggested. The UV spectrum of the acetate shows the homoannular diene absorption maximum with unusual large red-shift: λ_{max} 302 nm ($\epsilon=2500$). The similar characteristic has been observed in the CD spectrum (MeOH): $[\theta]_{302} -89000$ (neg. max.) (homoannular diene $\pi\rightarrow\pi^*$) and $[\theta]_{239} +40000$ (pos. max.) (γ -lactone). However, $LiAlH_4$ reduction of the acetate gave a triol (**5**), $C_{30}H_{50}O_3$, amorphous, $[\alpha]_D -5^\circ$ ($CHCl_3$), 1H -NMR (δ): 6.04 (2H, s, 11-H, 12-H), 3.56, 3.86 (1H each, ABq, $J=11$ Hz, 18- H_2). The UV and CD spectra of the

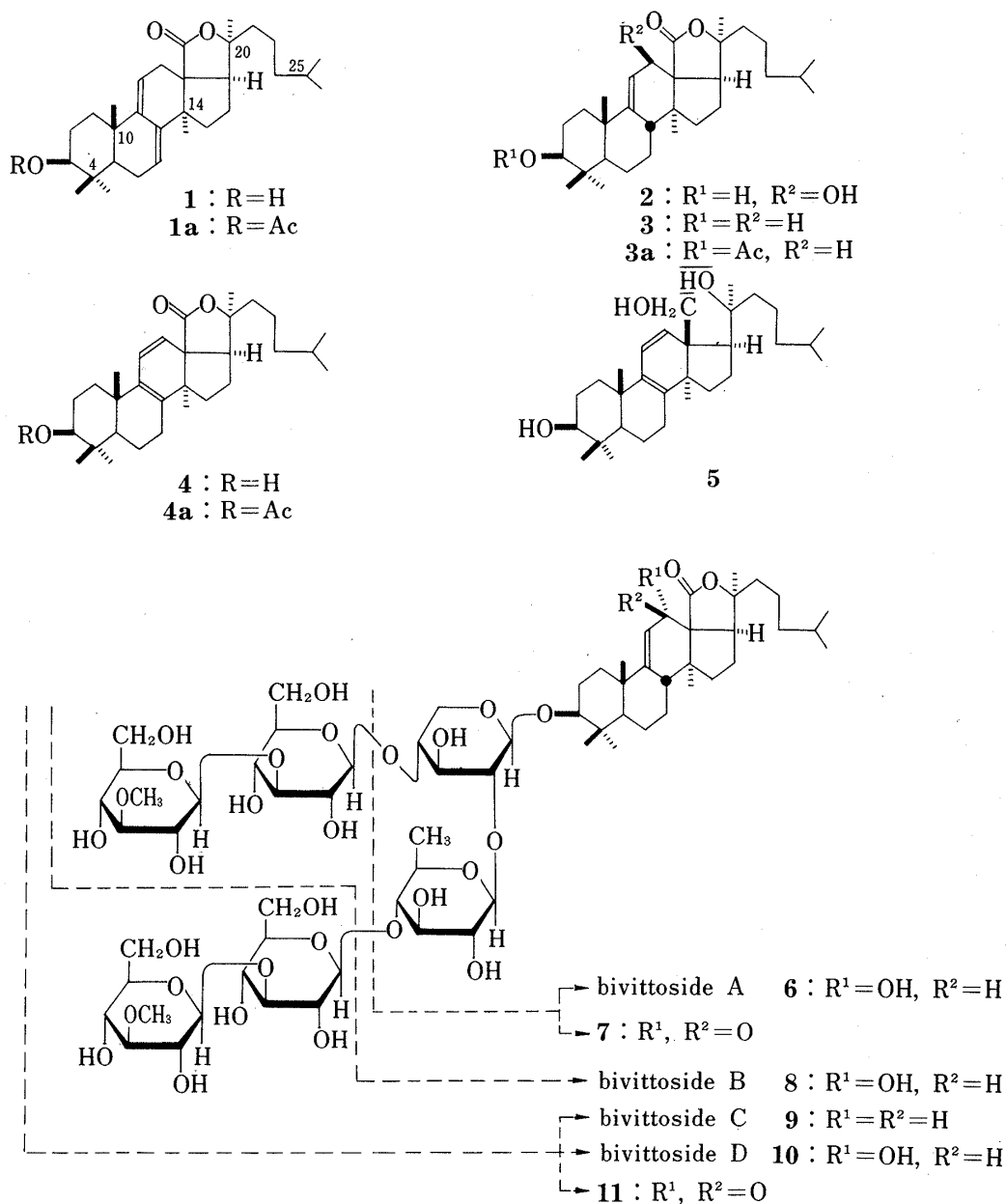


Chart 1

TABLE I. ^1H -NMR Data (90 MHz)

	Solvent	4-Me ₂	10-Me	14-Me	20-Me	25-Me ₂
1	CDCl_3	0.90, 1.00	1.10	1.00	1.38	0.88
	$d_5\text{-pyr.}$	1.11, ^{a)} 1.21	1.35	1.07 ^{a)}	1.35	0.88
2	CDCl_3	0.85, 1.00	1.19	0.91	1.58	0.88
	$d_5\text{-pyr.}$	1.03, ^{a)} 1.20	1.34	0.98 ^{a)}	1.89	0.88

^{a)} The assignments are interexchangeable.

triol show the homoannular diene maximum at the normal wave length: λ_{max} 275 nm ($\epsilon=2500$), $[\theta]_{275} -25000$ (neg. max.), thus the structure **4a** being evidenced. It has been assumed that the sapogenol (**4**) with the 8,11-diene moiety may be an intermediary compound in the process providing seychellogenin (**1**) from bivittoside A (**6**). The reason for the unusual large red-shift of the UV absorption maximum of **4a** is yet unclear, although the severe strain on the homoannular diene chromophore may be ascribable as one of the reasons.

Enzymic hydrolysis of bivittoside B (**8**), $\text{C}_{54}\text{H}_{88}\text{O}_{22} \cdot 3\text{H}_2\text{O}$, mp 270—273°, $[\alpha]_{\text{D}} +6^\circ$ (pyr.), with crude hesperidinase afforded bivittoside A (**6**).⁸⁾ On methanolysis, the dodeca-O-methyl derivative (**8a**) [four β -anomeric proton signals at δ 4.32 (d, $J=8$ Hz), 4.43 (d, $J=7$ Hz), 4.70 (d, $J=8$ Hz), and 4.72 (d, $J=8$ Hz)] liberated methyl pyranosides of 2,3,4,6-tetra-O-methylglucose, 2,4,6-tri-O-methylglucose, 2,3,4-tri-O-methylquinovose, and 3-O-methylxylose, thus the structure **8** being substantiated.

Bivittoside D (**10**), $\text{C}_{67}\text{H}_{110}\text{O}_{32} \cdot 3\text{H}_2\text{O}$, mp 219—221°, $[\alpha]_{\text{D}} -7^\circ$ (pyr.) is a hexaglycoside as shown by the ^1H -NMR spectrum of the octadeca-O-methyl derivative (**10a**) which shows six β -anomeric proton signals at δ 4.11 (1H, d, $J=7$ Hz), 4.31 (1H, d, $J=8$ Hz), 4.55 (1H, d, $J=7$ Hz), and 4.89 (3H, d, $J=7$ Hz). Enzymic hydrolysis of **10** with mixed glycosidase from *Turbo cornutus* afforded bivittoside B (**8**), while methanolysis of the fully methylated derivative gave methyl pyranosides of 2,3,4,6-tetra-O-methylglucose, 2,4,6-tri-O-methylglucose, 2,3-di-O-methylquinovose, and 3-O-methylxylose. Therefore, the structure **10** has been proposed to bivittoside D.

On acidic hydrolysis, bivittoside C (**9**), $\text{C}_{67}\text{H}_{110}\text{O}_{31} \cdot \text{H}_2\text{O}$, mp 216—218°, $[\alpha]_{\text{D}} -31^\circ$ (pyr.), furnished another sapogenol (**3**), $\text{C}_{30}\text{H}_{48}\text{O}_3$, mp 231—233°, $[\alpha]_{\text{D}} -16^\circ$ (CHCl_3), [monoacetate (**3a**), $\text{C}_{32}\text{H}_{50}\text{O}_4$, mp 225—226°]. The structure of bivittoside C (**9**) has been elucidated as based on the following evidence. Thus, acetylation followed by oxidation with *t*-butyl chromate and subsequent deacetylation of bivittoside C gave the 9(11)-en-12-one derivative (**11**), $\text{C}_{67}\text{H}_{108}\text{O}_{32} \cdot 2\text{H}_2\text{O}$, mp 218—220°, $[\alpha]_{\text{D}} -13^\circ$ (pyr.), UV (MeOH): 253 nm ($\epsilon=8300$), which was found to be identical with an oxidation product of bivittoside D (**10**) obtained by the method⁷⁾ as described above for oxidation of **6** giving **7**.

The antifungal activities of these bivittosides will be reported elsewhere.

Acknowledgement The authors are grateful to Foundation for the Promotion of Research on Medicinal Resources and to Ministry of Education, Science, and Culture of Japan (547116) for financial support.

References and Notes

- 1) I. Kitagawa, T. Inamoto, M. Fuchida, S. Okada, M. Kobayashi, T. Nishino, and Y. Kyogoku, *Chem. Pharm. Bull.*, **28**, 1651 (1980).
- 2) Presented at the 27th Annual Meeting of The Japanese Society of Pharmacognosy, held at Nagoya, Sept. 29—30, 1980, Abstract Paper, p. 21.
- 3) Compounds given with the chemical formulae gave the satisfactory analytical values.
- 4) Identified by comparison of the physicochemical properties with those reported in P. Roller, B. Tursch, and C. Djerassi, *J. Org. Chem.*, **35**, 2585 (1970).
- 5) a) P.V. Demarco, E. Farkas, D. Doddrell, N.L. Mylari, and E. Wenkert, *J. Am. Chem. Soc.*, **90**, 5480

- (1968); b) I. Kitagawa, T. Nishino, T. Matsuno, H. Akutsu, and Y. Kyogoku, *Tetrahedron Lett.*, **1978**, 985.
- 6) S. Hakomori, *J. Biochem.* (Tokyo), **55**, 205 (1964).
- 7) Y. Kobayashi, M. Ogawa, S. Amagaya, M. Asada, and Y. Ogihara, the 22nd Symposium on the Chemistry of Natural Products, Fukuoka, Oct. 23—26, 1979, Symposium Papers, p. 71.
- 8) The respective identifications of oligoglycosides obtained by enzymic hydrolysis hereafter were undertaken by mixed mp determination and comparison of TLC, $[\alpha]_D$, and ^{13}C -NMR spectra.

*Faculty of Pharmaceutical Sciences,
Osaka University,
133-1, Yamada-kami, Suita,
Osaka 565, Japan*

*Institute for Protein Research,
Osaka University,
5311, Yamada-kami, Suita,
Osaka 565, Japan*

ISAO KITAGAWA*
MOTOMASA KOBAYASHI
MANABU HORI

YOSHIMASA KYOGOKU

Received November 4, 1980