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Studies on Monoterpene Glucosides and Related Natural Products. XLI.¹⁾ Chemical Conversion of Geniposide into 10-Hydroxyloganin²⁾

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Geniposide (9) was chemically converted into 10-hydroxyloganin (3a), which is presumed to be a precursor for secologanin (2). The stereoisomers 8a, 18a and 19a were also prepared. The absolute configurations of these four compounds were determined on the basis of chemical reactions and NMR measurements.

Keywords——iridoid glucosides; geniposide; 10-hydroxyloganin; chemical conversion; NOE experiments

Around 1966, several research groups showed that the non-tryptophan moiety of indole alkaloids is of monoterpene origin.^{4–7)} Subsequently, Battersby *et al.* demonstrated that both loganin (1) and secologanin (2) play an important role in the biosynthesis of indole alkaloids, *i.e.*, secologanin (2) formed by the cyclopentane ring cleavage of loganin (1) undergoes condensation with tryptamine, leading to the formation of indole alkaloids.^{8,9)} At present, secologanin (2) is known to be a key intermediate not only for indole alkaloids but also for ipecacuanha alkaloids¹⁰⁾ and secoiridoid glucosides.¹¹⁾ However, the mechanism of the formation of secologanin (2) from loganin (1) still remains obscure. For this process, Battersby proposed a mechanism involving the hydroxylation of loganin (1) to 10-hydroxyloganin (3a) followed by phosphorylation to the 10-phosphate and concerted cyclopentane ring cleavage of the last compound initiated by the elimination of the phosphoryloxy group.¹²⁾ Although no unequivocal evidence for this mechanism has so far been reported, it has been proved that the protons on C-7^{9,11)} and C-8¹³⁾ of loganin (1) are retained during the ring cleavage.

Meanwhile, Tietze succeeded in the total synthesis of 10-hydroxyloganin (3a)¹⁴⁾ and incidentally found that a base-catalyzed reaction of the tosylate (4) of 10-hydroxyloganin aglucone 1-O-methyl ether gave the oxetane-type compound (6), whereas the corresponding 7-epi derivative (5) furnished, through ring cleavage, secologanin aglucone 1-O-methyl ether (7).¹⁵⁾ However, the problem of whether or not 10-hydroxyloganin (3a) (or its 7-epimer (8a)) can actually be a biosynthetic precursor for secologanin (2) has not hitherto been discussed.

This paper deals with the chemical conversion of easily obtainable geniposide (9) into 10-hydroxyloganin (3a) which is indispensable for experiments to solve this important problem.

Hydroboration of an appropriate derivative of geniposide (9) would give a compound having the same plane structure as 10-hydroxyloganin (3a). Even if the configuration at C-7 and/or C-8 of the product differs from that of 3a, this could be overcome in view of the easy epimerization of 8-epi-7-dehydrologanin tetraacetate (10) with an 8 α -methyl group to 7-dehydrologanin tetraacetate (11) with an 8 α -methyl group as well as the smooth Walden inversion of the C-7 hydroxy group of 7-epi-loganin tetraacetate (12). For the protection of the hydroxy groups, benzylidene and benzyl groups would be suitable, because they survive hydroboration.

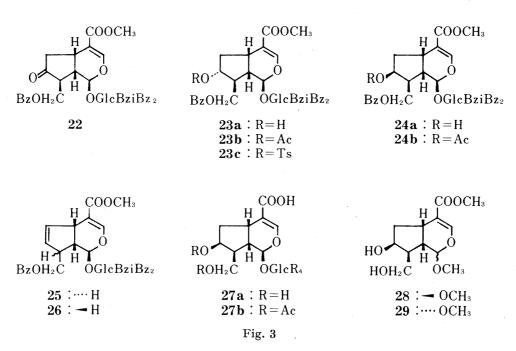
Geniposide (9) was treated with benzaldehyde and ZnCl₂ to yield 4',6'-O-benzylidene-geniposide (13) (mp 223—225°), which was in turn benzylated with NaH and benzyl bromide in benzene-DMSO in the presence of tetra-n-butylammonium iodide to provide 4',6'-O-benzylidene-2',3',10-tri-O-benzyl-geniposide (14) (mp 109—110°). This compound was then subjected to

Fig. 1

hydroboration to yield the mono-ol (15a) in 53% yield. The proton nuclear magnetic resonance (${}^{1}H$ NMR) spectrum of 15a lacked the signal of the vinyl group at δ 5.83 appearing in 14, but showed a signal of a hydroxy group at δ 2.10—2.32, which disappeared on addition of $D_{2}O$. Furthermore, the spectrum of the acetate (15b) of 15a showed a signal of an acetyl group at δ 1.99 and a double triplet assignable to a methine group bearing an acetoxy group at δ 5.07. These data clearly indicate that, as expected, the hydroxy group was introduced at the C-7 position. Since the hydroboration of compounds such as 1-methylcyclopentene is known to proceed through cis addition, i 15a is assumed to be a (7S,8R)-compound formed by the attack of i 15a from the less hindered convex face of the iridoid skeleton. The compound (15a) was then subjected to Jones oxidation to give the 7-dehydro derivative (16), which, without further purification, was reduced with i 15a in dioxane, furnishing 15a and another mono-ol (17a) (mp 110—111.5°) in a ratio of 7 to 13. The latter was thought to be the 7-epimer of 15a, i.e., the (7R, 8R)-compound. The inference regarding the stereochemistry of 15a and 17a

was verified in the following way. Benzylidene and benzyl protecting groups of 15a and 17a were removed through hydrogenolysis over Pd-C to afford the free glucosides 18a and 19a (mp 199—201°), which on acetylation gave the hexaacetates 18b (mp 139—140.5°) and 19b (mp 95—98°), respectively. The compound (18a) was heated in 3.5% aq. HCl to yield the aglucone (20a), $C_{11}H_{14}O_5$ (mp 74—75°). The composition of 20a as well as ¹H NMR data of its acetate (20b) (mp 65.5—66°) showing the presence of only one acetoxy group indicates that 20a is an anhydro derivative. Therefore, it was deduced that the configuration at C-8 of 18a and hence that of 15a are R. Likewise, 19a was converted into another anhydro derivative (21) (mp 81.5—82°), indicating the R-orientation at C-8 of 17a. On the other hand, the ¹H NMR signal (in acetone- d_6) of the C-7 proton in 20b appeared as a multiplet at δ 4.94, which collapsed to a double doublet (J=4.0 and 2.0 Hz) on irradiation of the C-8 proton at δ 2.97, whereas the corresponding signal of 21 appeared as a double triplet (J=11.0 and

6.5 Hz) at δ 5.10; these coupling constants suggest that the C-7 protons of 20b and 21 assume the α -quasi-equatorial and β -quasi-axial configurations, respectively. This was supported by NOE experiments with the tosylate (20c) (mp 91—92.5°) of 20a; ca.5% NOE was observed between the C-7 Ha (δ 4.75, m) and the C-10 Hb (δ 3.51, dd, J=10.0 and 3.5 Hz). This finding is indicative of the configuration at C-7 of 20c and hence that of 20a being S. From these findings, it became evident that, of the stereoisomers of 10-hydroxyloganin (3a), 18a and 19a, the former is the (7S, 8R)-compound (designated as allo-10-hydroxyloganin), whereas the latter is the (7R, 8R)-compound (designated as 7-epi-allo-10-hydroxyloganin). Since the R-configuration is assigned to the C-8 position of the 7-dehydro derivative (16), the next step required for the conversion of 16 into 3a should be epimerization at the C-8 position. This was achieved by PLC of 16 on silica gel with benzene-ether to afford the thermodynamically more favorable 4',6'-O-benzylidene-2',3',10-tri-O-benzyl-7-dehydro-10-hydroxyloganin (22) (mp 122.5—124°). The latter gave on reduction with NaBH₄ a pair of C-7 epimers 23a (mp 128.5—130.5°) and 24a in a ratio of 13 to 1. As 7-dehydrologanin tetraacetate (11) is known to afford exclusively 7-epi-loganin tetraacetate (12) with 7R-chirality upon NaBH₄ reduction, 16) the major product (23a) and also its acetate (23b) are presumed to have 7R,8S-configuration, whereas the minor one (24a) has 7S,8S-orientation. Since the yield of 24a in this reaction was so low, the conversion of 23a into 24a through Walden inversion was then attempted. Treatment of 23a with TsCl in pyridine yielded the tosylate (23c) (mp 100—102°), which was in turn refluxed with tetraethylammonium acetate in dry acetone to give the acetate (24b) in 79% yield, along with a minor product (25) formed by the elimination of p-toluenesulfonic acid. The tosylates 15c and 17c also gave on Walden inversion, besides the major products 17b and 15b, minor elimination products 14 and 26, respectively. The formation of the latter two products suggests trans stereochemistry for the elimination reaction. Since the elimination reaction of 23c is also considered to proceed in the same fashion, exclusive formation of 25 from 23c would amount to collateral evidence for the inference that the C-7 substituent and C-8 proton of 23c are in cis configuration, i.e., 23a assumes the 7R,8S-configuration.



The compound (23a) was hydrogenated over Pd-C to remove the protecting groups, giving rise to 7-epi-10-hydroxyloganin (8a), which on acetylation afforded 7-epi-10-hydroxyloganin hexaacetate (8b) (mp 127—128°). The compound (24b) was subjected to Zemplén reaction to give 24a, which was hydrogenated over Pd-C to afford the desired 3a, C₁₇H₂₆O₁₁·1/2

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H₂O; $[\alpha]_{\rm p}^{\rm 27}$ –58.9° (lit. $[\alpha]_{\rm p}^{\rm 29}$ –56°); $\lambda_{\rm max}^{\rm MeOH}$ (log ε) 236 nm (4.05) (lit. $\lambda_{\rm max}^{\rm MeOH}$ (log ε) 236 nm (4.02)). It gave on acetylation the hexaacetate (3b), C₂₉H₃₈O₁₇·H₂O; $[\alpha]_{\rm p}^{\rm 28}$ –54.0° (lit. $[\alpha]_{\rm p}^{\rm 25}$ –58°); $\lambda_{\rm max}^{\rm EtOH}$ (log ε) 232 nm (4.04) (lit. $\lambda_{\rm max}^{\rm EtOH}$ (log ε) 232 nm (4.01)); $\nu_{\rm max}^{\rm EtOH}$ 1750, 1735, 1700 (sh), 1635 cm⁻¹; these data are in good accordance with those of the acetate reported by Tietze. The compound (3a) was hydrolyzed to 10-hydroxyloganic acid (27a) followed by acetylation to afford 10-hydroxyloganic acid hexaacetate (27b), C₂₉H₃₈O₁₇. mp 138.5—139.5° (lit. mp 158°): $[\alpha]_{\rm p}^{\rm 29}$ –52.1° (lit. $[\alpha]_{\rm p}^{\rm 20}$ –56°); $\lambda_{\rm max}^{\rm MeOH}$ (log ε) 229 nm (4.01) (lit. $\lambda_{\rm max}^{\rm MeOH}$ (log ε) 234 nm (3.97)); $\nu_{\rm max}^{\rm KBF}$ 3155, 1760, 1740, 1715, 1640.

The stereotructure of 3a was eventually established in the following way. The compound (3a) was hydrolyzed with β -glucosidase to give an aglucone, which was treated with Amberlite IR-120 (H⁺-form) in dry MeOH giving rise to a pair of aglucone methyl ethers 28 and 29 in a ratio of 9 to 2. The ¹H NMR signal of the C-1 proton of 28 appeared at δ 4.49 as a doublet (J=6.0 Hz), whereas the corresponding signal of 29 appeared at δ 4.92 as a doublet (J=2.5 Hz). Comparison of these chemical shifts and coupling constants led us to the presumption that the C-1 proton of 28 is α , whereas that of 29 is β -oriented. Furthermore, NOE experiments with 28 (in C_5D_5N) showed 6—9% NOE between the C-1 (δ 3.89, d, J=5.5 Hz) and C-8 (δ 1.26, m) protons as well as between the C-7 (δ 3.68, td, J=5.0 and 2.0 Hz) and C-8 protons, indicating the α -orientation of the protons on C-1, C-7 and C-8. Thus, it was demonstrated that 3a is no other than the desired 10-hydroxyloganin, whereas 8a is 7-epi-10-hydroxyloganin. Thus, we successfully prepared all the C-7 and C-8 stereoisomers of 10-hydroxyloganin.

In contrast to the overall yield of 0.04% for the total synthesis of 10-hydroxyloganin (3a) by Tietze, ¹⁴⁾ the above method starting from an easily available natural product, geniposide (9), gave 3a in 8.2% yield. Furthermore, our method is preferable to Tietze's for the preparation of labelled 3a in terms of easy preparation of optically pure 3a from geniposide (9) without resolution as well as the shortness of the synthetic process of 3a from the 7-dehydro compound (16) following NaB³H₄ reduction.

Experimental

General Procedures—Melting points were measured on a Yanagimoto micro-apparatus and are uncorrected. Optical rotations were taken with a Union PM 201 automatic digital polarimeter or a JASCO DIP-181 automatic digital polarimeter. Ultraviolet (UV) spectra were recorded on a Hitachi model 200-20 spectrophotometer, and infrared (IR) spectra on a Hitachi model 215 grating infrared spectrophotometer. NMR spectra were taken on a Varian A-60, a Varian HA-100 or a Hitachi R-22 spectrometer with TMS or DSS as the internal standard. Mass (MS) spectra were recorded on a JEOL JMS-01SG-2 spectrometer. Silica gel AR-100 (Mallinckrodt) was used for column chromatography and silica gel PF₂₅₄ for medium pressure column chromatography. Furthermore, silica gel 60 GF₂₅₄ was employed for thin-layer chromatography (TLC) and spots were visualized under UV light, or by treatment with I₂ vapor or anisaldehyde-H₂SO₄ reagent. Silica gel 60 GF₂₅₄ or PF₂₅₄ was used for preparative thin-layer chromatography (PLC), and the following solvent systems were employed: (I) benzene-Et₂O (8: 2); (II) benzene-Et₂O (7: 3); (III) CHCl₃-MeOH (9: 1); (IV) CHCl₃-MeOH (7: 3); (V) CHCl₃-MeOH (1: 1); (VI) Et₂O. Bands were detected under UV light. The ratios of solvents are expressed by volume.

Conversion of Geniposide (9) into 4′,6′-O-Benzylidene-geniposide (13)—Geniposide (9) (32.74 g) and ZnCl₂ (34 g) were added to benzaldehyde (160 ml) and the reaction mixture was stirred overnight at room temperature. The resulting solution was diluted with H₂O (200 ml) and extracted with AcOEt (200 ml × 3). The AcOEt layer was washed with satd. aq. NaCl and concentrated *in vacuo* to give an oily residue, which on crystallization from EtOH afforded crude crystalline 4′,6′-O-benzylidene-geniposide (13) (25.64 g). On the other hand, the mother liquor was concentrated *in vacuo* and the residue was chromatographed on silica gel (100 g); the column was eluted first with CHCl₃ (500 ml) to remove benzaldehyde, and then successively with 1% (200 ml), 2% (500 ml) and 3% MeOH-CHCl₃ (300 ml), collecting 100 ml fractions. On concentration, Fr. 8—10 also gave crude crystalline 4′,6′-O-benzylidene-geniposide (13) (5.99 g). The combined crystals (13) (total 31.63 g) were recrystallized from MeOH as colorless plates (26.26 g), mp 223—225°, [α]²⁹ = 17.3° (c=1.00, DMF). UV λ ^{MeOH}_{max} nm (log ε): 236 (4.03). IR ν ^{MBF}_{max} cm⁻¹: 3390, 1695, 1625, 1430, 760, 748, 695. NMR (CDCl₃) δ : 2.50—3.40 (2H, m, OH, disappeared on addition of D₂O), 3.71 (3H, s, COOCH₃), 4.22 (2H, br s, 10-H₂), 4.20—4.70 (1H, m, OH, disappeared on addition of D₂O), 4.84 (1H, d, J=7.0 Hz, 1′-H), 4.88 (1H, d, J=8.5 Hz, 1-H), 5.48 (1H, s, -CHPh), 5.78 (1H, m, 7-H), 7.17—7.63 (6H, m, 3-H and arom. H₅). Anal. Calcd for C₂₄H₂₈O₁₀: C, 60.50; H, 5.92. Found: C, 60.62; H, 5.89.

Conversion of 4′,6′-0-Benzylidene-geniposide (13) into 4′,6′-0-Benzylidene-2′,3′,10-tri-0-benzyl-geniposide (14)——Compound (13) (10 g) and n-Bu₄NI (126 mg) were added to a stirred suspension of NaH (4.54 g) (50%, washed with dry petr. ether to remove mineral oil) in a mixture of benzene (180 ml) and DMSO (32 ml). The mixture was stirred for 15 min, then benzyl bromide (9 ml) was added and the whole was stirred at room temperature for 25 hr, diluted with H₂O and extracted with AcOEt (100 ml × 3). The AcOEt extract was washed with H₂O, dried over MgSO₄ and concentrated in vacuo. The residue was chromatographed on silica gel (375 g); the column was eluted successively with benzene (3.4 l), 1% (1 l) and 2% Et₂O-benzene (3.2 l), collecting 200 ml fractions. On concentration, Fr. 28—38 afforded a crystalline substance (11.73 g), which was recrystallized from Et₂O-petr. ether to give 14 (9.85 g) as colorless needles, mp 109—110°, [α]_D²⁷ -20.3° (c=0.99, CHCl₃). UV λ _{max}^{MOOH} nm (log ε): 237 (4.00). IR ν _{max}^{MOOH} cm⁻¹: 1705, 1635, 1495, 1450, 745, 700. NMR (CDCl₃) δ : 3.64 (3H, s, COOCH₃), 4.18 (2H, br s, 10-H₂), 5.32 (1H, d, J=5.5 Hz, 1-H), 5.55 (1H, s, -CHPh), 5.83 (1H, m, 7-H), 7.00—7.68 (21H, m, 3-H and arom. H₂₀). Anal. Calcd for C₄₅H₄₆O₁₀: C, 72.37; H, 6.21. Found: C, 72.31; H, 6.12.

Hydroboration of 4',6'-O-Benzylidene-2',3',10-tri-O-benzyl-geniposide (14)——A solution of NaBH₄ (685 mg) in dry diglyme (4 ml) was added dropwise to a stirred solution of BF_3 -etherate (2 ml) in dry diglyme (4 ml). Diborane gas generated was introduced into a stirred solution of 14 (9.52 g) in dry THF (80 ml) (cooled with ice-H₂O) by applying a slight pressure of N₂. After completion of the NaBH₄ addition (30 min), the B₂H₆ generator was heated for 30 min at 80° to ensure the complete transfer of the diborane. The reaction mixture was allowed to warm to room temperature and was stirred for an additional 2 hr. After decomposition of the excess diborane by addition of H2O (10 ml) the resulting organoborane was oxidized by adding $3\,\mathrm{N}$ NaOH followed by $30\%~\mathrm{H_2O_2}$ (each $2.5~\mathrm{ml}$) and the stirring was continued at room temperature for 30min. The mixture was diluted with H2O and extracted with benzene (80 ml × 3). The benzene layer was washed with H₂O, dried over MgSO₄ and concentrated in vacuo. The residue (9.78 g) was chromatographed on silica gel (200 g); the column was eluted successively with benzene (0.3 l), 3% (1 l), 6% (1.5 l), 9% (1 l), 12% (0.6 l), 15% (1.2 l) and 20% Et₂O-benzene (1.4 l), collecting 100 ml fractions. The combined Fr. 44—45 were concentrated in vacuo to give a residue (1.52 g), which was rechromatographed on silica gel (50 g) in the same way as above to furnish 4',6'-O-benzylidene-2',3',10-tri-O-benzyl-allo-10-hydroxyloganin (15a) (1.01 g) as a white powder. The combined Fr. 46—62 afforded pure 15a (4.13 g). $[\alpha]_{D}^{27} - 78.8^{\circ}$ (c = 1.05, CHCl₃). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3410, 1705, 1635, 1495, 1450, 750, 740, 700. NMR (CDCl₃) δ : 2.05 (2H, t, J = 7.0Hz, 6-H₂), 2.10—2.32 (1H, m, OH, disappeared on addition of D₂O), 3.18 (1H, m, 5-H), 3.63 (3H, s, COOCH₃), 4.11 (1H, q, J = 7.0 Hz, 7-H), 4.29 (1H, dd, J = 10 and 4.5 Hz, 10-H), 19 5.38 (1H, d, J = 4.0 Hz, 1-H), 5.55 (1H, s, $-C\underline{H}Ph$), 7.13—7.62 (21H, m, 3-H and arom. H_{20}). Anal. Calcd for $C_{45}H_{48}O_{11}$: C, 70.67; H, 6.33. Found: C, 70.43; H, 6.49.

Jones Oxidation of 4',6'-O-Benzylidene-2',3',10-tri-O-benzyl-allo-10-hydroxyloganin (15a)—Jones reagent was added dropwise to a solution of 15a (633 mg) in acetone (20 ml) with stirring under ice-cooling until the solution showed a persistent pale orange color. Stirring was continued for a further 30 min, then the excess reagent was reduced with MeOH, and the mixture was diluted with H_2O (100 ml) and extracted with CHCl₃ (100 ml × 3). The CHCl₃ layer was washed with H_2O , dried over MgSO₄ and concentrated in vacuo to give 4',6'-O-benzylidene-2',3',10-tri-O-benzyl-7-dehydro-allo-10-hydroxyloganin (16) (622 mg) as a white powder. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1735, 1700, 1635, 1495, 1450, 750, 740, 700. NMR (CDCl₃) δ : 3.67 (3H, s, COOCH₃), 5.51 (1H, d, J=5.5 Hz, 1-H), 5.53 (1H, s, -CHPh), 7.12—7.68 (21H, m, 3-H and arom. H_{20}).

NaBH₄ Reduction of 4′,6′-O-Benzylidene-2′,3′,10-tri-O-benzyl-7-dehydro-allo-10-hydroxyloganin (16)—A solution of NaBH₄ (82.5 mg) in H₂O (0.5 ml) was added to a stirred solution of 16 (705 mg) in dioxane (20 ml) and the reaction mixture was stirred at room temperature for 1 hr. After decomposition of the excess NaBH₄ by addition of acetone under ice-cooling, the mixture was diluted with H₂O (50 ml) and extracted with benzene (100 ml × 3). The benzene layer was washed with H₂O, dried over MgSO₄ and concentrated in vacuo. The residue (678 mg) was chromatographed on silica gel (40 g); the column was eluted successively with 5% (100 ml), 8% (100 ml), 10% (500 ml), 12% (400 ml) and 15% Et₂O-benzene (300 ml), collecting 50 ml fractions. The combined Fr. 12—17 were concentrated in vacuo and the residue (346 mg) was recrystallized from EtOH to yield 4′,6′-O-benzylidene-2′,3′,10-tri-O-benzyl-7-epi-allo-10-hydroxyloganin (17a) (262 mg) as colorless needles, mp 110—111.5°, [α]²⁶_p -85.0° (c=0.99, CHCl₃). IR ν ^{msr}_{msx} cm⁻¹: 3500, 1685, 1635, 1495, 1450, 745, 695. NMR (CDCl₃) δ : 2.07 (1H, s, OH, disappeared on addition of D₂O), 3.68 (3H, s, COO-CH₃), 5.57 (1H, s, -CHPh), 5.59 (1H, d, J=7.0 Hz, 1-H), 7.15—7.65 (21H, m, 3-H and arom. H₂₀). Anal. Calcd for C₄₅H₄₈O₁₁: C, 70.67; H, 6.33. Found: C, 70.46; H, 6.60. The combined Fr. 19—27 gave a residue (164 mg), which was purified by PLC (solvent system I, 3 developments) to give 15a (141 mg).

Catalytic Hydrogenation of 4',6'-O-Benzylidene-2',3',10-tri-O-benzyl-allo-10-hydroxyloganin (15a)—A solution of 15a (432 mg) in MeOH (30 ml) was hydrogenated over Pd-C (prepared from 5% PdCl₂-HCl solution (3.0 ml) and activated charcoal (DARCO G-60, 600 mg)). After completion of the H₂ uptake, the catalyst was filtered off and the filtrate was concentrated in vacuo to give a residue (228 mg), which was subjected to PLC (solvent system IV, Rf 0.24) to furnish allo-10-hydroxyloganin (18a) (167 mg) as a white powder, $[\alpha]_D^{29}$ -76.4° (c=0.89, MeOH). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ε): 237 (4.03). IR ν_{\max}^{ERS} cm⁻¹: 3300, 1680, 1625. NMR (D₂O) δ : 3.75 (3H, s, COOCH₃), 5.58 (1H, d, J=4.5 Hz, 1-H), 7.49 (1H, s, 3-H). Anal. Calcd for C₁₇H₂₆O₁₁· 1/4H₂O: C, 49.69; H, 6.50. Found: C, 49.59; H, 6.42.

Acetylation of Allo-10-hydroxyloganin (18a) ——Compound (18a) (52.3 mg) was acetylated with Ac₂O and pyridine (each 0.5 ml) in the usual way and the product (94.2 mg) was recrystallized from EtOH to give allo-10-hydroxyloganin hexaacetate (18b) (50.2 mg) as colorless needles, mp 139—140.5°, [α]₅ = -65.7° (c=0.82, CHCl₃). UV $\lambda_{\max}^{\text{BIOR}}$ nm (log ε): 233 (4.06). IR ν_{\max}^{KBF} cm⁻¹: 1740, 1700, 1630. NMR (CDCl₃) δ : 1.94, 2.01, 2.03, 2.05, 2.10 (each s, $6 \times \text{OCOCH}_3$), 3.72 (3H, s, COOCH₃), 5.36 (1H, d, J=3.5 Hz, 1-H), 7.37 (1H, br s, 3-H). Anal. Calcd for C₂₉H₃₈O₁₇: C, 52.89; H, 5.82. Found: C, 52.88; H, 5.71.

Catalytic Hydrogenation of 4',6'-O-Benzylidene-2',3',10-tri-O-benzyl-7-epi-allo-10-hydroxyloganin (17a) — A solution of 17a (301 mg) in MeOH (12 ml) was hydrogenated over Pd-C catalyst (prepared from 5% PdCl₂-HCl solution (1.5 ml) and activated charcoal (DARCO G-60, 500 mg)). After completion of the H₂ uptake, the catalyst was filtered off and the filtrate was concentrated *in vacuo* to give a residue (171 mg), which, on recrystallization from EtOH-AcOEt, afforded 7-epi-allo-10-hydroxyloganin (19a) (73.7 mg) as colorless needles, mp 199—201°, [α]²⁷_D -95.4° (c=1.00, MeOH). UV λ ^{MeOH}_{max} nm (log ε): 238 (4.08). IR ν ^{KBr}_{max} cm⁻¹: 3450—3200, 1670, 1625. NMR (D₂O) δ : 3.75 (3H, s, COOCH₃), 4.42 (1H, m, 7-H), 4.87 (1H, d, J=7.0 Hz, 1'-H), 5.64 (1H, d, J=7.5 Hz, 1-H), 7.54 (1H, s, 3-H). *Anal.* Calcd for C₁₇H₂₆O₁₁: C, 50.24; H, 6.45. Found: C, 50.16; H, 6.38.

Acetylation of 7-Epi-allo-10-hydroxyloganin (19a) ——Compound (19a) (51.2 mg) was acetylated with Ac₂O and pyridine (each 0.5 ml) and the product (75.4 mg) was recrystallized from Et₂O-petr. ether to furnish 7-epi-allo-10-hydroxyloganin hexaacetate (19b) (31.4 mg) as colorless needles, mp 95—98°, $[\alpha]_{5}^{27}$ —84.7° (c=0.98, CHCl₃). UV $\lambda_{\max}^{\text{MoOH}}$ nm (log ε): 234 (4.08). IR ν_{\max}^{KBr} cm⁻¹: 1740, 1695, 1635. NMR (CDCl₃) δ : 1.99, 2.01, 2.03, 2.09 (each s, $6 \times \text{OCOCH}_{3}$), 3.72 (3H, s, COOCH₃), 5.62 (1H, d, J=5.0 Hz, 1-H), 7.43 (1H, br s, 3-H). Anal. Calcd for C₂₉H₃₈O₁₇: C, 52.89; H, 5.82. Found: C, 52.80; H, 5.80.

Acid Treatment of Allo-10-hydroxyloganin (18a) — A solution of 18a (265 mg) in 3.5% aq. HCl (6 ml) was heated at 90° for 60 min. After cooling, the reaction mixture was diluted with $\rm H_2O$ (40 ml) and extracted with AcOEt (50 ml × 3). The AcOEt layer was washed with $\rm H_2O$, dried over MgSO₄ and evaporated to dryness in vacuo. The residue (59.5 mg) was subjected to PLC (solvent system VI, 4 developments) to give an oily residue (48.1 mg), which was recrystallized from $\rm Et_2O$ -petr. ether to afford the anhydride (20a) (12.4 mg) as colorless plates, mp 74—75°, [α] $_{\rm D}^{25}$ +88.3° (c=0.45, CHCl₃). IR $v_{\rm max}^{\rm eHCl_3}$ cm⁻¹: 3430, 1695, 1640. NMR (CDCl₃) δ : 1.52 (1H, ddd, J=13.0, 9.5 and 4.0 Hz, 6-H_{quasi-ax}), 2.58 (1H, s, OH, disappeared on addition of D₂O), 3.59 (1H, dd, J=9.5 and 4.0 Hz, 10-H), 3.74 (3H, s, COOCH₃), 3.98 (1H, dd, J=9.5 and 8.0 Hz, 10-H), 4.16 (1H, m, 7-H), 5.42 (1H, d, J=4.0 Hz, 1-H), 7.55 (1H, s, 3-H). Anal. Calcd for C₁₁H₁₄O₅: C, 58.40; H, 6.24. Found: C, 58.34; H, 6.25.

Acetylation of the Anhydride (20a) — The anhydride (20a) (19.8 mg) was acetylated with Ac₂O and pyridine (each 0.5 ml) and the product (19.9 mg) was purified by PLC (solvent system II, Rf 0.28) to give the acetate (20b) (13.4 mg), which was recrystallized from Et₂O-petr. ether to furnish colorless needles (6.3 mg), mp 65.5—66°, $[\alpha]_D^{27}$ +50.1° (c=0.52, CHCl₃), UV $\lambda_{\max}^{\text{Etoff}}$ nm (log ε): 231 (4.03). IR ν_{\max}^{KBr} cm⁻¹: 1730, 1690, 1640. NMR (CDCl₃) δ : 1.58 (1H, ddd, J=14.0, 10.5 and 4.0 Hz, 6-H_{quasi-ax}), 2.04 (3H, s, OCOCH₃), 2.40 (1H, ddt, J=14.0, 7.5 and 2.0 Hz, 6-H_{quasi-eq}), 2.76 (1H, td, J=9.5 and 5.0 Hz, 9-H), 2.75—3.15 (1H, m, 5-H), 3.14 (1H, m, 8-H), 3.67 (1H, dd, J=10.0 and 4.5 Hz, 10-H), 3.74 (3H, s, COOCH₃), 3.98 (1H, dd, J=10.0 and 8.5 Hz, 10-H), 5.01 (1H, m, 7-H), 5.40 (1H, d, J=5.0 Hz, 1-H), 7.54 (1H, d, J=1.0 Hz, 3-H). Anal. Calcd for C₁₃H₁₆O₅: C, 58.20; H, 6.01. Found: C, 58.18; H, 6.16.

Tosylation of the Anhydride (20a)—p-TsCl (410 mg) was added to a solution of 20a (45.8 mg) in pyridine (2 ml). After standing overnight at room temperature, the reaction mixture was poured into ice-H₂O and extracted with CHCl₃ (30 ml × 3). The CHCl₃ layer was washed successively with 1 n HCl, satd. aq. NaHCO₃, and H₂O, dried over MgSO₄ and concentrated *in vacuo*. The residue (74.5 mg) was purified by PLC (solvent system II, Rf 0.36) followed by recrystallization from Et₂O to give the tosylate (20c) (64.2 mg) as colorless plates, mp 91—92.5°. IR $\nu_{\rm max}^{\rm CHCl_3}$ cm⁻¹: 1700, 1645, 1360, 1175. NMR (CDCl₃) δ : 1.47 (1H, ddd, J=14.0, 10.0 and 4.0 Hz, 6-H_{quasi-ax}), 2.37 (1H, ddt, J=14.0, 7.5 and 2.0 Hz, 6-H_{quasi-eq}), 2.45 (3H, s, arom. CH₃), 3.51 (1H, dd, J=10.0 and 3.5 Hz, 10-H), 3.69 (3H, s, COOCH₃), 3.90 (1H, dd, J=10.0 and 8.5 Hz, 10-H), 4.75 (1H, m, 7-H), 5.37 (1H, d, J=5.0 Hz, 1-H), 7.50 (1H, s, 3-H), 7.23—7.97 (4H, A'₂B'₂ type arom. H₄). High resolution MS, Calcd for C₁₈H₂₀O₅S (M⁺): 380.0930. Found: 380.0931.

Conversion of 7-Epi-allo-10-hydroxylogain (19a) into the Corresponding Aglucone Acetate (21)——A solution of 19a (274.4 mg) in 3.5% aq. HCl (6 ml) was heated at 90° for 80 min. After cooling, the mixture was diluted with H_2O (30 ml) and extracted with AcOEt (30 ml×5). The AcOEt layer was washed with H_2O , dried over MgSO₄ and concentrated in vacuo. The oily residue (71.7 mg) was acetylated with Ac₂O and pyridine (each 1.0 ml) and the product (63.1 mg) was subjected to PLC (solvent system II, 2 developments), yielding a colorless syrup (19.7 mg), which was recrystallized from Et_2O -petr. ether to give the aglucone acetate (21) (9.0 mg) as colorless needles, mp 81.5—82°, $[\alpha]_D^{26}$ +91.6° (c=0.23, CHCl₃). UV $\lambda_{\max}^{\text{BIOT}}$ nm (log ε), 231 (4.01). IR ν_{\max}^{KBr} cm⁻¹: 1725, 1695, 1640. NMR (CDCl₃) δ : 1.00—1.80 (1H, m, 6-H), 2.03 (3H, s, OCOCH₃), 2.33—3.33 (4H, m, 5-, 6-, 8- and 9-H₄), 3.72 (3H, s, COOCH₃), 3.73 (1H, dd, J=10.0 and 8.0 Hz, 10-H), 4.08 (1H, dd, J=10.0 and 4.5 Hz, 10-H), 5.10 (1H, dt, J=11.0 and 6.5 Hz, 7-H), 5.41 (1H, d, J=5.0 Hz, 1-H), 7.52 (1H, s, 3-H). Anal. Calcd for $C_{13}H_{16}O_5$: C, 58.20; H, 6.01. Found: C, 57.92; H, 5.86.

Epimerization of 4',6'-O-Benzylidene-2',3',10-tri-O-benzyl-7-dehydro-allo-10-hydroxyloganin (16)----

Compound (16) (116 mg) was subjected to PLC (1 plate, solvent system I). A band around Rf 0.37 was scraped off and extracted with CHCl₃ (100 ml). Concentration of the CHCl₃ extract gave a residue (82.7 mg), which was recrystallized from Et₂O-petr. ether to afford 4′,6′-O-benzylidene-2′,3′,10-tri-O-benzyl-7-dehydro-10-hydroxyloganin (22) (73.6 mg) as colorless needles, mp 122.5—124°, $[\alpha]_D^{27}$ —134.3° (c=1.02, CHCl₃). IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 1735, 1700, 1635, 1495, 1450, 750, 700. NMR (CDCl₃) δ : 3.58 (3H, s, COOCH₃), 5.58 (1H, s, -CHPh), 5.68 (1H, d, J=2.5 Hz, 1-H), 7.13—7.72 (21H, m, 3-H and arom. H₂₀). Anal. Calcd for C₄₅H₄₆O₁₁: C, 70.85; H, 6.08. Found: C, 70.93; H, 5.87.

No. 4

NaBH₄ Reduction of 4',6'-O-Benzylidene-2',3',10-tri-O-benzyl-7-dehydro-10-hydroxyloganin (22)-A solution of NaBH₄ (87.1 mg) in H₂O (0.5 ml) was added to a stirred solution of 22 (794 mg) in dioxane (20 ml), and the reaction mixture was stirred for a further 1 hr at room temperature. The excess NaBH4 was decomposed by addition of acetone under ice-cooling, and the mixture was diluted with H2O (50 ml) and extracted with benzene (50 ml×4). The benzene layer was washed with H₂O, dried over MgSO₄ and concentrated in vacuo. The residue (754 mg) was reprecipitated from benzene-petr. ether, yielding 4',6'-O-benzylidene-2',3',10-tri-O-benzyl-7-epi-10-hydroxyloganin (23a) (579 mg), which was purified by recrystallization from Et₂O-petr. ether to give colorless needles, mp 128.5—130.5°, $[\alpha]_D^{20}$ -70.5° $(c=0.51, \text{CHCl}_3)$. IR $\nu_{\text{max}}^{\text{KBr}} \text{ cm}^{-1}$: 3200, 1700, 1635, 1495, 1450, 755, 738, 700. NMR (CDCl₃) δ : 1.54 (1H, dt, J = 13.0 and 7.0 Hz, 6-H), 1.89— 2.20 (1H, m, OH, disappeared on addition of D_2O), 2.48 (1H, dt, J=13.0 and 6.5 Hz, 6-H), 2.91 (1H, m, 5-H), 3.61 (3H, s, COOCH₃), 4.05 (1H, m, 7-H), 4.30 (1H, dd, J = 10.5 and 5.0 Hz, 10-H), 5.30 (1H, d, J = 10.5 and 5.0 Hz, 10-Hz, 10-H 4.5 Hz, 1-H), 5.53 (1H, s, $-C\underline{H}Ph$), 7.10—7.58 (21H, m, 3-H and arom. H_{20}). Anal. Calcd for $C_{45}H_{48}O_{11}$: C, 70.67; H, 6.33. Found: C, 70.64; H, 6.62. The mother liquor was concentrated in vacuo and the residue (176 mg) was chromatographed on silica gel (10 g); the column was eluted successively with 5% (50 ml), 8% (50 ml), 10% (100 ml) and 12% Et₂O-benzene (100 ml), collecting 25 ml fractions. The combined Fr. 8-9 were concentrated in vacuo to give 4',6'-O-benzylidene-2',3',10-tri-O-benzyl-10-hydroxyloganin (24a) (47.8 mg) as a white powder, whereas the combined Fr. 10-11 afforded, after reprecipitation from benzenepetr. ether, 23a (51.1 mg). 24a, $[\alpha]_D^{18}$ -69.2° (c=1.02, CHCl₃). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3430, 1695, 1625, 1490, 1445, 745, 735, 695. NMR (CDCl₃) δ: 2.37—2.67 (1H, m, OH, disappeared on addition of D₂O), 3.61 (3H, s, COOCH₃), 4.32 (1H, m, 7-H), 5.23 (1H, d, 1-H), 5.55 (1H, s, -CHPh), 7.08—7.63 (21H, m, 3-H and arom. H_{20}). Anal. Calcd for $C_{45}H_{48}O_{11}$: C, 70.67; H, 6.33. Found: C, 70.78; H, 6.27.

Acetylation of 4',6'-O-Benzylidene-2',3',10-tri-O-benzyl-allo-10-hydroxyloganin (15a) ——Compound (15a) (53.9 mg) was acetylated with Ac₂O and pyridine (each 0.5 ml) and the product (53.4 mg) was purified by PLC (solvent system I, Rf 0.37) to give 15b (48.4 mg) as a white powder. $[\alpha]_{5}^{29}$ —80.7° (c=0.81, CHCl₃). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1730, 1700, 1635, 1495, 1450, 750, 740, 700. NMR (CDCl₃) δ : 1.99 (3H, s, OCOCH₃), 3.15 (1H, m, 5-H), 3.61 (3H, s, COOCH₃), 4.27 (1H, dd, J=10.0 and 4.5 Hz, 10-H), 4.89 (1H, d, J=8.0 Hz, 1'-H), 5.07 (1H, q, J=5.0 Hz, 7-H), 5.54 (1H, s, -CHPh), 5.59 (1H, d, J=4.5 Hz, 1-H), 7.11—7.67 (21H, m, 3-H and arom. H₂₀). Anal. Calcd for C₄₇H₅₀O₁₂: C, 69.96; H, 6.25. Found: C, 69.92; H, 6.51.

Acetylation of 4′,6′-O-Benzylidene-2′,3′,10-tri-O-benzyl-7-epi-allo-10-hydroxyloganin (17a) ——Compound (17a) (83.5 mg) was acetylated with Ac₂O and pyridine (each 0.8 ml) and the product (93.4 mg) was purified by PLC (solvent system I, Rf 0.30), giving 17b (86.3 mg) as a white powder. [α]²⁸ -86.5° (c= 0.60, CHCl₃). IR ν ^{RBr}_{max} cm⁻¹: 1720, 1705, 1640, 1500, 1460, 755, 700. NMR (CDCl₃) δ : 1.87 (3H, s, OCOCH₃), 2.33 (1H, q, J=7.0 Hz, 9-H), 3.68 (3H, s, COOCH₃), 4.98 (1H, d, J=8.0 Hz, 1′-H), 5.47 (1H, d, J=8.0 Hz, 1-H), 5.56 (1H, s, -CHPh), 7.17—7.56 (21H, m, 3-H and arom. H₂₀). Anal. Calcd for C₄₇H₅₀O₁₂: C, 69.96; H, 6.25. Found: C, 70.05; H, 6.48.

Acetylation of 4′,6′-O-Benzylidene-2′,3′,10-tri-O-benzyl-7-epi-10-hydroxyloganin (23a) ——Compound (23a) (53.8 mg) was acetylated with Ac₂O and pyridine (each 0.5 ml) and the product (58.8 mg) was purified by PLC (solvent system I, Rf 0.36), giving the acetate (23b) (55.3 mg) as a white powder, $[\alpha]_D^{29}$ —98.5° (c=0.75, CHCl₃). IR ν_{\max}^{KBr} cm⁻¹: 1725, 1705, 1635, 1495, 1450, 750, 698. NMR (CDCl₃) δ : 1.93 (3H, s, OCOCH₃), 2.48 (1H, dd, J=15.0 and 7.0 Hz, 6-H), 3.03 (1H, m, 5-H), 3.62 (3H, s, COOCH₃), 4.28 (1H, dd, J=10.0 and 4.5 Hz, 10-H), 5.52 (1H, d, J=3.5 Hz, 1-H), 5.56 (1H, s, -CHPh), 7.11—7.64 (21H, m, 3-H and arom. H₂₀). Anal. Calcd for C₄₇H₅₀O₁₂: C, 69.96; H, 6.25. Found: C, 69.86; H, 6.31.

Tosylation of 4',6'-O-Benzylidene-2',3',10-tri-O-benzyl-allo-10-hydroxyloganin (15a)—A solution of 15a (122 mg) and p-TsCl (108 mg) in pyridine (1 ml) was allowed to stand overnight at room temperature. The reaction mixture was worked up in the usual way. The crude product (141 mg) was subjected to PLC (solvent system I, Rf 0.48), giving the tosylate (15c) (119 mg) as a white powder. $[\alpha]_D^{27} - 84.2^{\circ}$ (c = 0.99, CHCl₃). IR ν_{\max}^{KBr} cm⁻¹: 1700, 1635, 1495, 1450, 1360, 1175, 745, 735, 695. NMR (CDCl₃) δ : 2.40 (3H, s, arom. CH₃), 3.59 (3H, s, COOCH₃), 5.54 (1H, s, -CHPh), 7.00-7.90 (25H, 3-H and arom. H₂₄). Anal. Calcd for $C_{52}H_{54}O_{13}S$: C, 67.96; H, 5.92; S, 3.49. Found: C, 67.81; H, 6.15; S, 3.38.

Walden Inversion of the Tosylate (15c)——Et₄NOAc (2.00 g) was added to a solution of the tosylate (15c) (860 mg) in dry acetone (50 ml) and the mixture was heated under reflux for 25 hr. Removal of the solvent by evaporation gave a residue, which was redissolved in $CHCl_3$ (150 ml). The $CHCl_3$ layer was washed successively with satd. aq. NaCl and H_2O , dried over $MgSO_4$ and concentrated *in vacuo*. The residue was subjected to medium pressure column chromatography on silica gel (60 g); the column was eluted first with benzene (100 ml) and then successively with 5% (200 ml), 7% (200 ml) and 10% Et_2O —benzene (300 ml), collecting 10 g fractions. The combined Fr. 30—33 were concentrated *in vacuo* to give a residue (46.5 mg),

which was recrystallized from Et₂O-petr. ether to yield the olefin (26) (25.6 mg) as colorless needles, mp 118—119°. The combined Fr. 47—63 afforded a powdery substance (639 mg), which was identical with the above-mentioned 17b (TLC, IR and NMR comparisons). 26, $[\alpha]_D^{e7}$ –145.5° (c=0.55, CHCl₃). IR ν_{\max}^{KBF} cm⁻¹: 1710, 1640, 1500, 1455, 755, 740, 700. NMR (CDCl₃) δ : 2.83 (1H, td, J=8.5 and 2.0 Hz, 9-H), 3.21 (1H, m, 5-H), 3.63 (3H, s, COOCH₃), 4.30 (1H, dd, J=10.0 and 4.5 Hz, 10-H), 5.56 (1H, s, -CHPh), 5.68 (1H, dt, J=6.0 and 2.0 Hz, 6-H or 7-H), 5.89 (1H, d, J=2.0 Hz, 1-H), 6.12 (1H, m, 7-H or 6-H), 7.05—7.58 (21H, m, 3-H and arom. H₂₀). Anal. Calcd for C₄₅H₄₆O₁₀: C, 72.37; H, 6.21. Found: C, 72.41; H, 6.32.

Tosylation of 4',6'-O-Benzylidene-2',3',10-tri-O-benzyl-7-epi-allo-10-hydroxyloganin (17a) ——A solution of 17a (50.1 mg) and p-TsCl (38.2 mg) in pyridine (0.5 ml) was allowed to stand for 2 days. After further addition of p-TsCl (37.1 mg), the reaction mixture was allowed to stand for another 1 day. The conventional work-up of the mixture gave a crude product (60.8 mg), which was subjected to PLC (solvent system I). A band at around Rf 0.38 furnished 17c (41.1 mg) as a white powder, $[\alpha]_{b}^{20}$ —70.1° (c=0.41, CHCl₃). IR ν_{max}^{RBT} cm⁻¹: 1700, 1635, 1495, 1450, 1360, 1175, 750, 745, 700. NMR (CDCl₃) δ : 2.36 (3H, s, arom. CH₃), 3.66 (3H, s, COOCH₃), 5.47 (1H, d, J=7.5 Hz, 1-H), 5.59 (1H, s, -CHPh), 7.12—7.87 (25H, m, 3-H and arom. H₂₄). Anal. Calcd for $C_{52}H_{54}O_{13}S$: C, 67.96; H, 5.92; S, 3.49. Found: C, 67.67; H, 5.85; S, 3.49. From a band at around Rf 0.13, 17a (10.9 mg) was recovered.

Walden Inversion of the Tosylate (17c)—A solution of 17c (125 mg) and Et₄NOAc (360 mg) in dry acetone (15 ml) was refluxed for 20 hr. After removal of the solvent, the oily residue was diluted with CHCl₃ (50 ml), washed with satd. aq. NaCl, dried over MgSO₄ and concentrated *in vacuo*. The residue was subjected to PLC (solvent system I). A band at around Rf 0.37 gave a powdery substance (21.9 mg), which was identical with an authentic sample of 15b (TLC, IR and NMR comparisons). The band at around Rf 0.47 afforded a crystalline compound (73.9 mg), which was identical with an authentic specimen of 14 (TLC, IR and NMR comparisons).

Tosylation of 4′,6′-O-Benzylidene-2′,3′,10-tri-O-benzyl-7-epi-10-hydroxyloganin (23a) ——Compound (23a) (235 mg) was tosylated with p-TsCl (158 mg) and pyridine (2 ml) in the usual way and the product (270 mg) was purified by PLC (solvent system I, Rf 0.45) followed by recrystallization from EtOH to give crystals of 23c (237 mg), mp 100—102°, $[\alpha]_D^{23}$ —83.7° (c=1.03, CHCl₃). IR ν_{\max}^{KBr} cm⁻¹: 1700, 1630, 1490, 1450, 1360, 1175, 750, 700. NMR (CDCl₃) δ : 2.37 (3H, s, arom. CH₃), 3.59 (3H, s, COOCH₃), 5.36 (1H, d, J=4.0 Hz, 1-H), 5.55 (1H, s, -CHPh), 7.08—7.90 (25H, m, 3-H and arom. H₂₄). Anal. Calcd for C₅₂H₅₄O₁₃S: C, 67.96; H, 5.92; S, 3.49. Found: C, 67.90; H, 6.21; S, 3.63.

Walden Inversion of the Tosylate (23c)—A solution of 23c (867 mg) and Et₄NOAc (1.90 g) in dry acetone (50 ml) was refluxed for 24 hr. The conventional work-up of the mixture afforded a crude residue, which was subjected to medium pressure column chromatography on silica gel (60 g); the column was eluted successively with benzene (100 ml), 5% (200 ml), 7% (300 ml) and 10% Et₂O-benzene (300 ml), collecting 10 g fractions. The combined Fr. 30—36 and 44—55 furnished 25 (77.9 mg) and 24b (598 mg) as white powders, respectively. 24b, $[\alpha]_D^{28}$ -74.9° (c=0.53, CHCl₃). IR ν_{\max}^{KBr} cm⁻¹: 1730, 1700, 1635, 1495, 1450, 750, 740, 700. NMR (CDCl₃) δ: 1.96 (3H, s, OCOCH₃), 3.60 (3H, s, COOCH₃), 4.27 (1H, dd, J=10.0 and 4.5 Hz, 10-H), 5.33 (1H, m, 7-H), 5.53 (1H, d, J=3.0 Hz, 1-H), 5.59 (1H, s, -CHPh), 7.08—7.75 (21H, m, 3-H and arom. H₂₀). Anal. Calcd for C₄₇H₅₀O₁₂: C, 69.96; H, 6.25. Found: C, 70.26; H, 6.46. 25, $[\alpha]_D^{28}$ -41.1° (c=0.29, CHCl₃). IR ν_{\max}^{KBr} cm⁻¹: 1705, 1640, 1500, 1450, 750, 735, 698. NMR (CDCl₃) δ: 2.47 (1H, ddd, J=8.0, 7.0 and 4.0 Hz, 9-H), 3.04 (1H, m, 5-H), 3.63 (3H, s, COOCH₃), 4.26 (1H, dd, J=10.0 and 5.0 Hz, 10-H), 5.46 (1H, d, J=4.0 Hz, 1-H), 5.56 (1H, s, -CHPh), 5.60 (1H, dt, J=5.5 and 2.0 Hz, 6-H or 7-H), 6.06 (1H, dt, J=5.5 and 2.2 Hz, 7-H or 6-H), 7.06—7.57 (21H, m, 3-H and arom. H₂₀). Anal. Calcd for C₄₅H₄₆O₁₀·1/2H₂O: C, 71.51; H, 6.27. Found: C, 71.47; H, 6.42.

Catalytic Hydrogenation of 4',6'-O-Benzylidene-2',3',10-tri-O-benzyl-7-epi-10-hydroxyloganin (23a)—A solution of 23a (307 mg) in MeOH (12 ml) was hydrogenated over Pd-C (prepared from 5% PdCl₂-HCl solution (1.5 ml) and activated charcoal (DARCO G-60, 500 mg)). After completion of the H₂ uptake, the catalyst was filtered off and the filtrate was concentrated in vacuo. The residue (172 mg) was purified by PLC (solvent system IV, Rf 0.24) to give 7-epi-10-hydroxyloganin (8a) (132 mg) as a white powder. [α] $_{\rm b}^{\rm N}$ -93.4° (c=0.92, MeOH). UV $\lambda_{\rm max}^{\rm MeoH}$ nm (log ε): 236 (4.03). IR $\nu_{\rm max}^{\rm KBT}$ cm⁻¹: 3330, 1680, 1625. NMR (D₂O) δ : 3.75 (3H, s, COOCH₃), 5.42 (1H, d, J=6.0 Hz, 1-H), 7.53 (1H, d, J=1.0 Hz, 3-H). Anal. Calcd for C₁₇H₂₆-O₁₁·1/2H₂O: C, 49.15; H, 6.55. Found: C, 49.11; H, 6.53.

Acetylation of 7-Epi-10-hydroxyloganin (8a) — 7-Epi-10-hydroxyloganin (8a) (96.7 mg) was acetylated with Ac₂O and pyridine (each 1 ml) and the product (156 mg) was recrystallized from EtOH to give 7-epi-10-hydroxyloganin hexaacetate (8b) (107 mg) as colorless needles, mp 127—128°, [α]_D²⁵ — 94.9° (c=0.52, CHCl₃), UV $\lambda_{\max}^{\text{BIOH}}$ nm (log ε): 232 (4.04). IR ν_{\max}^{RBT} cm⁻¹: 1755, 1735, 1705, 1635. NMR (CDCl₃) δ : 1.90, 1.94, 1.98, 2.01, 2.04, 2.07 (each s, 6×OCOCH₃), 3.71 (3H, s, COOCH₃), 7.32 (1H, d, J=2.0 Hz, 3-H). Anal. Calcd for C₂₉H₃₈O₁₇: C, 52.89; H, 5.82. Found: C, 52.87; H, 5.90.

Zemplén Reaction of the Acetate (24b)—Methanolic NaOCH₃ (0.1 N, 0.8 ml) was added to a solution of 24b (236 mg) in dry MeOH (15 ml) and the mixture was refluxed for 30 min. After cooling, the mixture was diluted with H₂O (20 ml), neutralized with 1 N HCl and extracted with CHCl₃ (25 ml \times 3). The CHCl₃ layer was washed with H₂O, dried over MgSO₄ and concentrated *in vacuo*. The residue (217 mg) was purified by PLC (solvent system II, Rf 0.20) giving a white powder (203 mg). This compound was identical with an

authentic specimen of 24a (TLC, IR and NMR comparisons).

Catalytic Hydrogenation of 4',6'-O-Benzylidene-2',3',10-tri-O-benzyl-10-hydroxyloganin (24a) — A solution of 24a (294 mg) in MeOH (12 ml) was hydrogenated over Pd-C (prepared from 5% PdCl₂-HCl solution (1.2 ml) and activated charcoal (DARCO G-60, 300 mg)). After completion of the H₂ uptake, the catalyst was filtered off and the filtrate was concentrated in vacuo. Purification of the residue (167 mg) by PLC (solvent system IV, Rf 0.27) afforded 10-hydroxyloganin (3a) (138 mg) as a white powder, $[\alpha]_D^{27}$ -58.9° (c=0.58, MeOH). UV $\lambda_{\max}^{\text{meoH}}$ nm (log ε): 236 (4.05). IR ν_{\max}^{KBr} cm⁻¹: 3350, 1685, 1630. NMR (D₂O) δ : 3.76 (3H, s, COOCH₃), 4.38 (1H, m, 7-H), 4.81 (1H, d, J=8.0 Hz, 1'-H), 5.27 (1H, d, J=4.5 Hz, 1-H), 7.51 (1H, d, J=1.0 Hz, 3-H). Anal. Calcd for C₁₇H₂₆O₁₁·1/2H₂O: C, 49.15; H, 6.55. Found: C, 49.26; H, 6.45.

Acetylation of 10-Hydroxyloganin (3a) — 10-Hydroxyloganin (3a) (56.6 mg) was acetylated with Ac₂O and pyridine (each 0.5 ml) in the usual way and the product (83.1 mg) was purified by PLC (solvent system VI, Rf 0.28) giving 10-hydroxyloganin hexaacetate (3b) (71.3 mg) as a white powder, [α]²⁸ –54.0° (c=0.98, CHCl₃). UV $\lambda_{\max}^{\text{BIOH}}$ nm (log ε): 232 (4.04). IR $\nu_{\max}^{\text{CHCl}_3}$ cm⁻¹: 1750, 1735, 1700 (sh), 1635. NMR (CDCl₃) δ : 1.93, 2.01, 2.03, 2.09 (each s, $6 \times \text{OCOCH}_3$), 3.72 (3H, s, COOCH₃), 7.36 (1H, d, J=1.0 Hz, 3-H). Anal. Calcd for C₂₉H₃₈O₁₇·H₂O: C, 51.48; H, 5.96. Found: C, 51.49; H, 5.74.

Alkaline Hydrolysis of 10-Hydroxyloganin (3a) — 10-Hydroxyloganin (3a) (109.2 mg) was dissolved in 0.5 N aq. NaOH (2 ml) and the solution was stirred for 2.5 hr at room temperature. After neutralization of the solution with Amberlite IR-120 (H⁺-form), the resin was filtered off and the filtrate was concentrated in vacuo to afford a residue (105.5 mg), which was purified by PLC (solvent system V, Rf 0.36) to give 10-hydroxyloganic acid (27a) (90.3 mg) as a white powder, $[\alpha]_D^{ar}$ -62.7° (c=0.46, MeOH). UV λ_{max}^{MeOH} nm (log ε): 233 (4.03). IR ν_{max}^{RBr} cm⁻¹: 3330, 1670, 1625. NMR (D₂O) δ : 3.82 (2H, d, J=6.5 Hz, 10-H₂), 4.36 (1H, m, 7-H), 5.30 (1H, d, J=4.5 Hz, 1-H), 7.45 (1H, d, J=1.0 Hz, 3-H). Anal. Calcd for $C_{16}H_{24}O_{11} \cdot H_2O$: C, 46.83; H, 6.39. Found: C, 46.99; H, 6.30.

Acetylation of 10-Hydroxyloganic Acid (27a) ——10-Hydroxyloganic acid (27a) (43.9 mg) was acetylated with Ac₂O and pyridine (each 1 ml) and the product (66.3 mg) was recrystallized from Et₂O to give 10-hydroxyloganic acid hexaacetate (27b) (31.0 mg) as colorless needles, mp 138.5—139.5°, [α] $_{\rm max}^{\rm gc}$ —52.1° (c=0.44, CHCl₃). UV $\lambda_{\rm max}^{\rm moth}$ nm (log ε): 229 (4.01). IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 3155, 1760, 1740, 1715, 1640. NMR (CDCl₃) δ: 1.93, 1.98, 2.01, 2.02, 2.07 (each s, 6 × OCOCH₃), 3.03 (1H, m, 5-H), 3.73 (1H, m, 5'-H), 4.90—5.67 (1H, m, COOH, disappeared on addition of D₂O), 7.42 (1H, s, 3-H). Anal. Calcd for C₂₈H₃₆O₁₇: C, 52.17; H, 5.63. Found: C, 51.90; H, 5.52.

Conversion of 10-Hydroxyloganin (3a) into the Corresponding Aglucone Methyl Ethers 28 and 29-Glucosidase (prepared from almonds) (27 mg) was added to a solution of 3a (310 mg) in acetate buffer (pH 4.8) (45 ml). After standing at 39° for 15 hr, the reaction mixture was concentrated in vacuo. The residue was extracted with AcOEt (25 ml×4) and the combined AcOEt extracts were dried over MgSO₄ and evaporated to dryness. The residue was purified by PLC (solvent system III, Rf 0.33) to give a colorless syrup (130 mg), which was dissolved in dry MeOH (7 ml) and stirred with Amberlite IR-120 (H+-form) (980 mg) for 98 hr. The resin was filtered off and the filtrate was concentrated in vacuo to give an oily residue, which was subjected to PLC (solvent system VI, 4 developments). Of the two major bands, the more mobile one gave 29 (16.9 mg) as a colorless syrup, whereas the less mobile one afforded 28 (72.0 mg) as a colorless syrup. **28**, $[\alpha]_D^{29} - 47.8^{\circ}$ (c = 0.60, CHCl₃). IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3400, 1690, 1625. NMR (CDCl₃) δ : 1.54 (1H, ddd, J =14.0, 10.0 and 4.5 Hz, 6-H), 1.83—2.53 (3H, m, 6-H, 8-H and 9-H), 3.12 (1H, m, 5-H), 3.26 (2H, br s, OH, disappeared on addition of D_2O), 3.53 (3H, s, OCH₃), 3.71 (3H, s, COOCH₃), 4.49 (1H, d, J=6.0 Hz, 1-H), 7.46 (1H, d, J=1.0 Hz, 3-H). High resolution MS, Calcd for $C_{12}H_{18}O_6$ (M+): 258.1103. Found: 258.1104. 29, IR $v_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3400, 1695, 1625. NMR (CDCl₃) δ : 1.77—2.65 (4H, m, 6-H₂, 8-H and 9-H), 3.03 (1H, m, 5-H), 3.09 (2H, br s, OH, disappeared on addition of D_2O), 3.45 (3H, s, OCH_3), 3.72 (3H, s, $COOCH_3$), 3.88 $(2\mathrm{H,\ d},\ J=5.0\ \mathrm{Hz},\ 10\mathrm{-H_2}),\ 4.46\ (1\mathrm{H,\ m},\ 7\mathrm{-H}),\ 4.92\ (1\mathrm{H,\ d},\ J=2.5\ \mathrm{Hz},\ 1\mathrm{-H}),\ 7.59\ (1\mathrm{H,\ d},\ J=1.0\ \mathrm{Hz},\ 3\mathrm{-H}).$ High resolution MS, Calcd for $C_{12}H_{18}O_6$ (M+): 258.1103. Found: 258.1100.

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