

## Communications to the Editor

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SWEET AND BITTER GLYCOSIDES OF THE CHINESE PLANT DRUG, BAI-YUN-SHEN  
(ROOTS OF SALVIA DIGITALOIDES)

Takashi Tanaka,<sup>a</sup> Osamu Tanaka,<sup>\*,a</sup> Zhong-Wen Lin,<sup>b</sup> Jun Zhou<sup>b</sup> and Hiroyuki Ageta<sup>c</sup>  
Institute of Pharmaceutical Sciences, Hiroshima University School of Medicine,<sup>a</sup> Kasumi, Minami-ku, Hiroshima, 734 Japan, Kunming Institute of Botany, Academia Sinica,<sup>b</sup> Kunming, Yunnan, China and Showa College of Pharmaceutical Sciences<sup>c</sup>, 5-1-8 Tsurumaki, Setagaya-ku, Tokyo 154, Japan

From a Chinese plant drug, Bai-Yun-Shen (roots of Salvia digitaloides), a new sweet glycoside named baiyunoside (**1**) was isolated. The structure of the acid-unstable aglycone of **1** named baiyunol (**2**) was elucidated by MS and NMR spectroscopy especially by comparison with the <sup>13</sup>C chemical shifts of β-onocerin (**3**). The structure of **1** was determined to be β-D-xylopyranosyl(1→2)-β-D-glucopyranoside of **2** by MS and NMR spectroscopy. Besides this sweet principle (**1**), several bitter iridoid glucosides were also isolated from this plant. Two of these glucosides were identified as shanzhiside methyl ester (**13**) and its 8-O-acetate (=acetylbarlerin, **10**), respectively. A new glucoside (**9**) was formulated as 6-O-syringyl-8-O-acetyl shanzhiside methyl ester by the correlation with **13**.

KEYWORDS — Bai-Yun-Shen; Salvia digitaloides Diels; Labiatae; sweet glycoside; baiyunoside; labdane-type diterpene; <sup>13</sup>C-NMR; β-onocerin; iridoid glucoside; acylated shanzhiside methyl ester

We have been engaged in the investigation of the sweet principles of Chinese plants, i.e., a sweet steviol-glycoside, rubusoside from Chinese Rubus chingii Hu (Rosaceae),<sup>1)</sup> dihydrochalcone-glycosides, phlorizin and trilobatin from Lithocarpus litseifolius (Hance) Rehd. (Fagaceae)<sup>2)</sup> and trilobatin from Vitis piasezkii Maxim. and V. saccharifera Makino (Vitaceae)<sup>3)</sup>. The present communication deals with the chemical study of sweet and bitter glycosides of the Chinese plant drug "Bai-Yun-Shen", roots of Salvia digitaloides Diels (Labiatae) which has been used for women's diseases.

An aqueous suspension of the methanolic extract of the dried roots collected in Li-jiang, Yunnan, China, was washed with ether and then extracted with 1-butanol saturated with water. The butanolic fraction was chromatographed to give a new sweet principle named baiyunoside (**1**), white powder, C<sub>31</sub>H<sub>48</sub>O<sub>11</sub>, EI-MS as its hexa-acetate: M+ m/z 890, [α]<sub>D</sub><sup>26</sup> +13.8° (c=2.50, MeOH), UV λ<sub>max</sub><sup>EtOH</sup> 213nm (logε=3.8), yield: 0.11%. This glycoside (**1**) is about 500-fold sweeter than sucrose and its sweetness lasts more than one h.

On hydrolysis with crude hesperidinase,<sup>4)</sup> an acid unstable aglycone, named baiyunol (**2**), was obtained from **1** as colorless needles mp 85.5-86°C (from n-hexane), C<sub>20</sub>H<sub>30</sub>O<sub>2</sub>, EI-MS: M+ 302, [α]<sub>D</sub><sup>21</sup> +64.0° (c=1.47, CHCl<sub>3</sub>). It was not sweet. The UV

absorption maximum at 212nm (in MeOH) ( $\log \epsilon = 3.9$ ), strong ions at  $m/z$  81 and 95 in its EI-MS,  $^1\text{H-NMR}$  signals at  $\delta$  7.35(1H, t,  $J=1.8\text{Hz}$ ), 7.23(1H, broadened dd) and 6.30(1H, broadened dd) in  $\text{CDCl}_3$  as well as  $^{13}\text{C}$  signals at  $\delta$  125.5 (or 126.7) (s, 13-C), 110.8(d, 14-C), 142.6(d, 15-C) and 138.4(d, 16-C) in  $\text{CDCl}_3$  revealed the presence of a  $\beta$ -substituted furan ring<sup>5</sup>) in **2**. Further, multiplicities of the  $^{13}\text{C-NMR}$  signals of **2** demonstrated the presence of one carbonyl carbon, one tetra-substituted double bond, four methyls, six methylenes, two methines and two quaternary carbons. Its proton signals in  $\text{CDCl}_3$  at  $\delta$  0.81(3H, s), 0.97(3H, s), 1.02(3H, s), 1.61(3H, s) and 3.25(1H, dd,  $J=6.0$  and  $10.8\text{Hz}$ ) revealed the presence

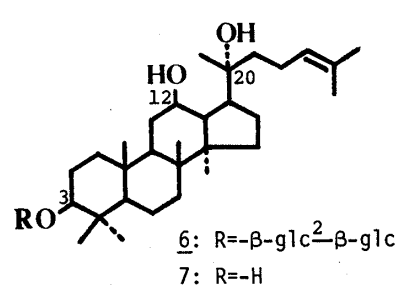
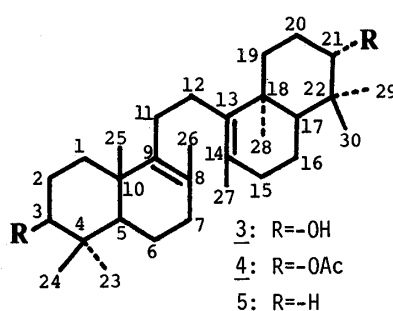
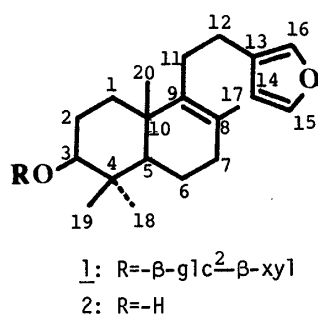


Table I.  $^{13}\text{C-NMR}$  Spectra of Aglycone Moieties (in Pyridine- $d_5$  at 25.15 MHz)

|     | 1      | 2      | 3     |        | 1      | 2      | 3    |
|-----|--------|--------|-------|--------|--------|--------|------|
| C-1 | 35.3   | 35.5   | 36.3  | C-11   | 29.0   | 29.2   | 28.7 |
| 2   | 27.2   | 29.2   | 29.1  | 12     | 26.1   | 26.1   |      |
| 3   | 89.3   | 78.1   | 78.0  | 13     | 126.1† | 126.1† |      |
| 4   | 38.7*  | 39.1*  | 39.2  | 14     | 111.5  | 111.4  |      |
| 5   | 51.6   | 51.4   | 51.4  | 15     | 143.3  | 143.3  |      |
| 6   | 18.9   | 19.5   | 19.4  | 16     | 139.0  | 139.1  |      |
| 7   | 34.0   | 34.1   | 34.6  | 17(26) | 19.0   | 19.2   | 20.7 |
| 8   | 126.5† | 126.6† | 126.5 | 18(23) | 28.0   | 28.7   | 28.8 |
| 9   | 139.9  | 140.1  | 141.1 | 19(24) | 16.4   | 16.5   | 16.4 |
| 10  | 39.6*  | 39.5*  | 39.5  | 20(25) | 20.2   | 20.3   | 21.0 |

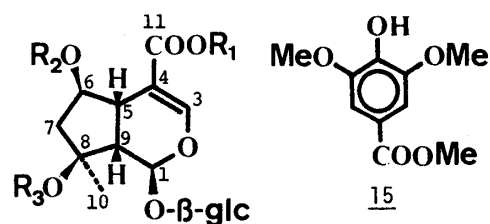
( ); For 3. \*,†; These signals may be reversed.

Table II.  $^{13}\text{C-NMR}$  Spectra of Sugar Moieties (in Pyridine- $d_5$  at 25.15 MHz)

|       | 1 (inner) | 6           |
|-------|-----------|-------------|
| glc-1 | 105.0     | glc-1 105.1 |
| 2     | 83.8      | 2 83.5      |
| 3     | 78.3*     | 3 78.2*     |
| 4     | 71.4†     | 4 71.7      |
| 5     | 78.1*     | 5 78.0*     |
| 6     | 62.6      | 6 62.7†     |
| xyl-1 | 106.8     |             |
| 2     | 76.5      |             |
| 3     | 77.9      |             |
| 4     | 71.0†     |             |
| 5     | 67.4      |             |

\*,†; These signals may be reversed.

glc; -D-glucopyranosyl, xyl; -D-xylopyranosyl.



10: R<sub>1</sub> = Me, R<sub>2</sub> = H, R<sub>3</sub> = Ac

13: R<sub>1</sub> = Me, R<sub>2</sub> = R<sub>3</sub> = H

14: R<sub>1</sub> = R<sub>2</sub> = R<sub>3</sub> = H

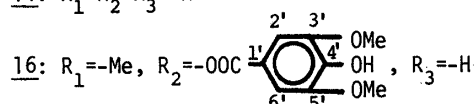


Table III.  $^{13}\text{C-NMR}$  Spectra of Iridoids (in Pyridine- $d_5$  at 25.15 MHz)

|     | 9      | 16     | 10    | 13    |             | 9      | 16     | 10    | 13   |
|-----|--------|--------|-------|-------|-------------|--------|--------|-------|------|
| C-1 | 95.1   | 94.9   | 95.8  | 94.9  | 11-COOMe    | 51.1   | 51.3   | 51.1  | 52.0 |
| 3   | 153.6  | 152.6  | 152.9 | 152.3 | OCOMe       | 170.5  |        | 170.9 |      |
| 4   | 107.7  | 109.3  | 108.9 | 110.6 | OCOMe       | 21.9*  |        | 22.0  |      |
| 5   | 39.2   | 38.2   | 42.0  | 41.5  |             |        |        |       |      |
| 6   | 78.1   | 78.3   | 74.7  | 76.6  | 1'          | 120.4  | 120.9  |       |      |
| 7   | 44.7   | 48.1   | 47.4  | 48.8  | 2',6'       | 108.4  | 108.4  |       |      |
| 8   | 88.3   | 77.7   | 88.6  | 78.3  | 3',5'       | 148.5  | 148.6  |       |      |
| 9   | 49.7   | 51.3   | 49.6  | 51.1  | 4'          | 143.0  | 142.7  |       |      |
| 10  | 21.5*  | 25.7   | 22.0  | 24.9  | $\phi$ -OMe | 56.4   | 56.4   |       |      |
| 11  | 166.6† | 167.1† | 167.5 | 168.0 | $\phi$ -COO | 165.9† | 166.4† |       |      |

\*,†; These signals may be reversed.

of three methyl groups on quarternary carbons, one allylic methyl group and one equatorial secondary hydroxyl group, respectively. This evidence suggests that **2** is a monohydroxy-labdane-type diterpene having a  $\beta$ -substituted furan ring. One of the present authors, Ageta, has investigated the  $^{13}\text{C}$ -NMR spectra of onocerane-type triterpenes in connection with his chemical studies of fern triterpenes, assigning the carbon signals of  $\beta$ -onocerin (**3**)<sup>6)</sup> derived from  $\alpha$ -onocerin<sup>7)</sup> by comparing them with those of its acetate (**4**) and  $\beta$ -onoceradiene (**5**)<sup>8)</sup> with consideration for the substitution and acetylation effects. As shown in Table I, it was found that the carbon signals of **2** other than those associated with its  $\beta$ -substituted furan moiety appeared at almost the same positions as those of **3** except for the slight differences in the chemical shift of the allylic methyl signal (C-17 for **2** and C-26 and -27 for **3**), leading to the formulation of **2** as illustrated in Chart 1. The  $^1\text{H}$ -NMR of **2** was consistent with this formulation. The chirality of the 3-equatorial-hydroxyl group of **2** was established as S-configuration by the modified Horeau's method.<sup>9)</sup> Accordingly, the absolute configuration of the skeleton of **2** should be the normal type.

On mineral acid hydrolysis, **1** yielded glucose and xylose. The EI-MS of the acetate of **1** exhibited fragment ions at  $m/z$  547 ((glucose-xylose) $\text{Ac}_6$ ) and 259 ((xylose) $\text{Ac}_3$ ). The coupling constants of two anomeric proton signals of **1** ( $\delta$  4.93 and 5.25 (1H d each,  $J=6.0\text{Hz}$ ) in  $\text{C}_5\text{D}_5\text{N}$ ) indicated the  $\beta$ -configuration of both glycosyl linkages. Inspection of the  $^{13}\text{C}$ -NMR spectrum of **1** in comparison with those of known glycosides revealed that signals due to the sugar moiety of **1** consisted of those due to a terminal  $\beta$ -xylopyranosyl unit and a 2-O-glycosylated  $\beta$ -glucopyranoside unit, the latter of which appeared at almost the same positions as those of the 2-O-substituted  $\beta$ -D-glucopyranosyl unit of the 3-O- $\beta$ -sophorosyl moiety of the prosapogenin (**6**) of Ginseng saponins<sup>10)</sup> as shown in Table II. It was revealed that the glycosylation shifts are significantly related to the combination of absolute configurations of an aglycone alcohol<sup>11)</sup> and a sugar moiety. On going from **2** to **1**, the glycosylation shift values of C-2, -3 and -4 as well as that of the anomeric carbon signal of the inner glucosyl unit were found to be almost identical with those for **6** from 20(R)-protopanaxadiol (**7**).<sup>11)</sup> This evidence led to the formulation of **1** including absolute configuration of the sugar moiety as  $\beta$ -D-xylopyranosyl(1 $\rightarrow$ 2)- $\beta$ -D-glucopyranoside of **2**.

Besides this sweet principle (**1**), six bitter iridoid-glucosides, **8-13**, were isolated from this plant in yields of 0.02, 0.19, 0.28, 0.11, 0.17 and 0.03%, respectively. Glucoside (**13**), white powder,  $[\alpha]_{\text{D}}^{22} -123.3^\circ$  ( $c=3.8$ , MeOH), UV  $\lambda_{\text{max}}^{\text{MeOH}}$  236nm ( $\log \epsilon=3.9$ ) was identified as shanzhiside methyl ester which was previously isolated from *Mussaenda parviflora* Miquel.<sup>12)</sup> Glucoside (**10**), white powder,  $[\alpha]_{\text{D}}^{26} -84.8^\circ$  ( $c=1.28$ , MeOH), UV  $\lambda_{\text{max}}^{\text{MeOH}}$  234nm ( $\log \epsilon=4.0$ ),  $^1\text{H}$ -NMR in  $\text{C}_5\text{D}_5\text{N}$ :  $\delta$  1.65(3H, s, 10- $\text{CH}_3$ ), 1.88(3H, s,  $\text{OCOCH}_3$ ), 3.60(3H, s,  $\text{COOCH}_3$ ), 5.32(1H, d,  $J=7.2\text{Hz}$ , anomeric H), 6.43(1H, d,  $J=1.0\text{Hz}$ , 1-H) and 7.68(1H, s, 3-H) afforded **13** on mild alkaline hydrolysis, being identical with 8-O-acetate of **13** named acetylbarlerin which was recently isolated from *Barleria prionitis* L..<sup>13)</sup>

On alkaline hydrolysis with 1N NaOH in methanol at room temperature, a new glucoside **9**, white powder,  $[\alpha]_{\text{D}}^{24} -71.0^\circ$  ( $c=1.01$ , MeOH), UV  $\lambda_{\text{max}}^{\text{MeOH}}$  221 and 276nm ( $\log \epsilon=4.2$  and 3.7) yielded shanzhiside (**14**) and methyl syringate (**15**). The  $^1\text{H}$ -NMR spectrum of **9** in  $\text{C}_5\text{D}_5\text{N}$  exhibited a signal at  $\delta$  1.93(3H, s,  $\text{OCOCH}_3$ ) along with

signals at  $\delta$  1.71(3H, s, 10-CH<sub>3</sub>), 3.60(3H, s, COOCH<sub>3</sub>), 3.83(6H, s, OCH<sub>3</sub>), 5.36(1H, d, J=7.2Hz, anomeric H), 6.31(1H, d, J=3.6Hz, 1-H), 7.69(2H, s, aromatic H) and 7.74(1H, d, J=1.2Hz, 3-H) indicating the presence of an acetoxyl group which was supported by its <sup>13</sup>C-NMR spectrum (Table III). Mild hydrolysis of **9** with 0.1N NaOH in methanol at room temperature resulted in deacetylation, yielding **16** (a syringate of **13**), white powder,  $[\alpha]_D^{22}$  -117.3° (c=1.05, MeOH), UV  $\lambda_{\max}^{\text{MeOH}}$  221 and 275nm (log  $\epsilon$  =4.3 and 3.9), <sup>1</sup>H-NMR in C<sub>5</sub>D<sub>5</sub>N:  $\delta$  1.43(3H, s, 10-CH<sub>3</sub>), 3.55(3H, s, COOCH<sub>3</sub>), 3.80(6H, s, OCH<sub>3</sub>), 5.37(1H, d, J=7.2Hz, anomeric H), 6.07(1H, d, J=4.5Hz, 1-H), 7.74(1H, s, 3-H) and 7.78(2H, s, aromatic H). As shown in Table III, on going from **13** to **16**, the signal due to 6-C was deshielded by 1.7 ppm and that due to 4-, 5- and 11-C were shielded, leading to the formulation of **16** as 6-O-syringate of **13**. On going from **13** to **9**, signals due to both 6- and 8-C were displaced downfield and those due to 4-, 5-, 7-, 9-, 10- and 11-C were shielded. It follows that **9** can be formulated as 6-O-syringyl-8-O-acetyl shanzhiside methyl esters. The structures elucidation of **8**, **11** and **12** are being studied.

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