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## Acridone Alkaloids. VI.<sup>1)</sup> The Constituents of *Citrus depressa*. Isolation and Structure Elucidation of New Acridone Alkaloids from *Citrus* genus<sup>2)</sup>

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Five new acridone alkaloids, citracridone-I (1a), -II (1b), citpressine-I (2a), -II (2b), and prenylcitpressine (3) along with a known acridone alkaloid, 5-hydroxynoracronycine (5), and three known coumarins, clausarin (6), suberosin (7a), and xanthyletin (8) were isolated from the root bark of Citrus depressa (Rutaceae) collected in Taiwan and characterized. This is the first report of isolation of acridone alkaloids from Citrus species. In the structure elucidations, the use of nuclear Overhauser effect (NOE) experiment on the O-methoxymethyl derivatives of phenolic acridone alkaloids was shown to be useful for the determination of the location of the phenolic hydroxyl groups.

**Keywords**——*Citrus depressa*; Rutaceae; acridone alkaloid; coumarin; citracridone-I; citracridone-II; citpressine-II; prenylcitpressine; <sup>1</sup>H-NMR

Citrus depressa (Rutaceae) is a small green tree, and the peel is popularly used as the source of Chen-pi, an important Chinese drug in Taiwan, which has been used for treating heartburn, abdominal pain, diarrhea, tussis, vomiting and inappetence, and as an expectorant.<sup>3)</sup> In a preliminary communication,<sup>2)</sup> we described the isolation and structures of five new acridone alkaloids contained in C. depressa. The full details of the structure elucidation are now presented in this paper.

The ethanolic extract of the root bark of *C. depressa*, which grows naturally in Taiwan, was treated by the procedure described in the experimental section, and five new acridone alkaloids named citracridone-I, -II, citpressine-I, -II, and prenylcitpressine along with a known acridone alkaloid, 5-hydroxynoracronycine (5),<sup>4)</sup> and three known coumarins, clausarin (6), suberosin (7a), and xanthyletin (8), were isolated and characterized.

Citracridone-I was obtained as orange plates from acetone, mp 275-278°C. Mass spectrometry and microanalysis established the molecular formula C<sub>20</sub>H<sub>19</sub>NO<sub>5</sub> for this alkaloid. The ultraviolet (UV) spectrum showed typical absorption associated with a 9-acridone nucleus<sup>5,6)</sup> (see Experimental), and a bathochromic shift was observed on addition of sodium methoxide (NaOCH<sub>3</sub>) or aluminium chloride (AlCl<sub>3</sub>), indicating the presence of phenolic hydroxyl groups.<sup>4)</sup> The proton nuclear magnetic resonance (1H-NMR) spectrum showed the presence of two one-proton singlets at  $\delta$  14.52 and 9.33 due to two hydroxyl protons, indicating that at least one of them was strongly hydrogen bonded. Hydrogen bonding of the 1-hydroxyl group in the 9-acridone nucleus is well known to occur.<sup>5)</sup> The <sup>1</sup>H-NMR data for citracridone-I are shown in Table I. The AB-type quartets at  $\delta$  6.63 and 5.61 (J=10 Hz) and a six-proton singlet at  $\delta$  1.53 reflect the presence of a dimethylpyran system attached to ring C since the lower signal of this AB quartet had a long range coupling (J=1 Hz) with the doublet at  $\delta$  6.23 (H-2). An NMR nuclear Overhauser effect (NOE) study gave a 9.5% enhancement of the signal at  $\delta$  6.63 on irradiation at the frequency corresponding to the N-methyl protons at  $\delta$  3.75, allowing the assignment of an angular orientation of the dimethylpyran ring. The <sup>13</sup>C-NMR spectrum of citracridone-I also supported these conclusions, and signals of an N-methyl carbon and the olefinic C-1' of the dimethylpyran ring appeared at  $\delta$  48.6 and 120.8, respectively.<sup>7,8)</sup> An additional AB-type signal (J=9 Hz) at  $\delta$  7.00 and 8.01 in the <sup>1</sup>H-NMR spectrum was assigned to mutually ortho-located protons; the lower field signal could be assigned to H-8 on the basis of the deshielding by the C-9 carbonyl moiety.

896

In addition, the absence of NOE between the H-7 signal at  $\delta$  7.00 and the methoxyl signal at  $\delta$  3.91 suggested the location of a methoxyl and a hydroxyl group at C-5 and C-6, respectively.

To confirm these assignments, a deuterium exchange reaction of the *ortho* (and *para*)-located proton to the phenolic hydroxyl group was attempted. However, all attempts using general methods<sup>10</sup> were unsuccessful. Then, an NOE experiment (Chart 2) was carried out for the *O*-methoxymethyl derivative of citracridone-I prepared by treatment of citracridone-I with chloromethylmethyl ether and sodium hydroxide in the presence of phase-transfer catalyst.<sup>11</sup> Irradiation of the methylene protons of the methoxymethyl ether moiety at  $\delta$  5.30 gave a 14.9% enhancement of H-7 at  $\delta$  7.09. Irradiation of the *N*-methyl signal at  $\delta$  3.70 produced a 12.3% enhancement of the signal at  $\delta$  6.52 due to H-1'. These data were consistent with the structure 1a for citracridone-I.

NOE measurement for the methoxymethyl derivatives of phenolic alkaloids is considered to be a useful method for the determination of the location of the phenolic hydroxyl groups.

Chart 2

Next, methylation of 1a with diazomethane furnished a mono-O-methyl ether, which still showed a hydrogen-bonded hydroxyl proton at  $\delta$  14.35 in the <sup>1</sup>H-NMR spectrum. Thus, the structure of this product was assigned as 1b. Citracridone-II, mp 161—163°C,  $C_{21}H_{21}NO_5$  (M+: m/z 367) was shown to be identical with this O-methyl ether by infrared (IR), UV, <sup>1</sup>H-NMR and mass spectral comparisons, and mixed mp determination.

Citpressine-I, mp 183—185°C, and citpressine-II, mp 168—170°C, have the molecular formulae  $C_{16}H_{15}NO_5$  (M+: m/z 301) and  $C_{17}H_{17}NO_5$  (M+: m/z 315), respectively, and showed UV spectra typical of a 9-acridone nucleus.<sup>5,6)</sup> The <sup>1</sup>H-NMR spectra (Table I) of these alkaloids showed similar signal patterns, except for an additional methoxyl signal in that of citpressine-II. Treatment of citpressine-I with diazomethane gave a mono-O-methyl ether, which was identical

1'-H

2'-H

3'-(CH<sub>3</sub>),

with citpressine-II. The presence of a hydroxyl group being still hydrogen bonded in citpressine-II was indicated by the IR (3430 cm<sup>-1</sup>) and <sup>1</sup>H-NMR spectra (1H singlet at  $\delta$  14.71). In the aromatic proton region of the <sup>1</sup>H-NMR spectrum of citpressine-I, meta-coupled proton signals at  $\delta$  6.25 and 6.51 (each 1H, d, J=2 Hz), and ortho-coupled proton signals at  $\delta$  8.10 and 6.99 (each 1H, d, J=9 Hz) were observed. The lower ortho-coupled proton signal at  $\delta$  8.10 is characteristic of H-8 in a 9-acridone system. An NOE experiment (Chart 2) showed that the signal at  $\delta$  6.51, one of the meta-coupled protons, exhibited 14.8% and 20% enhancements on irradiation of the methoxyl signal at  $\delta$  3.97, and of the N-methyl signal at  $\delta$  4.10, respectively. At the same time, irradiation of the methoxyl at  $\delta$  3.97 gave a 22% enhancement of the other meta-coupled proton signal at  $\delta$  6.25. However, on irradiation of the methoxyl at  $\delta$  3.83, no NOE was observed at any proton signal. These data led to the structures 2a and 2b for citpressine-I and -II, respectively.

TABLE I. 1H-NMR Spectra Data for Acridone Alkaloids from Citrus depressa

	1a	1b	2a
1-OH	14.52 (1H, s) <sup>a)</sup>	14.35 (1H, s) <sup>a)</sup>	14.85 (1H, s) <sup>a)</sup>
2-H	6.23 (1H, d; 1)	6.26 (1H, s)	6.25 (1H, d; 2)
3-OCH <sub>3</sub>		, , ,	3.97 (3H, s)
4-H			6.51 (1H, d; 2)
N-CH <sub>3</sub>	3.75 (3H, s)	3.74 (3H, s)	4.10 (3H, s)
5-OR	3.91 (3H, s)	3.90 (3H, s)	3.83 (3H, s)
6-OR	$9.33 (1H, s)^{a}$	4.01 (3H, s)	$2.83 (1H, s)^{a}$
7-H	7.00 (1H, d; 9)	7.01 (1H, d; 9)	6.99 (1H, d; 9)
8-H	8.01 (1H, d; 9)	8.13 (1H, d; 9)	8.10 (1H, d; 9)
1'-H	6.63 (1H, dd; 1, 10)	6.62 (1H, d; 10)	(- ,,,
2'-H	5.61 (1H, d; 10)	5.58 (1H, d; 10)	
3'-(CH <sub>3</sub> ) <sub>2</sub>	1.53 (6H, s)	1.51 (6H, s)	
	2b	3	5
1-OH	14.71 (1H, s) <sup>a)</sup>	14.31 (1H, s) <sup>a)</sup>	14.57 (1H, s) <sup>a)</sup>
2-H	6.22 (1H, d; 2)	6.30 (1H, s)	6.18 (1H, s)
3-OCH <sub>3</sub>	3.96 (3H, s)	$3.06 (1H, s)^{a}$	` , ,
4-H	6.46 (1H, d; 2)		
N-CH <sub>3</sub>	4.04 (3H, s)	3.64 (3H, s)	3.87 (3H, s)
5-OR	3.84 (3H, s)	3.91 (3H, s)	$2.90 (1H, s)^{a}$
6-OR	4.06 (3H, s)	9.32 (1H, br)	7.40 (1H, dd; 2, 7)
7-H	7.16 (1H, d; 9)	6.94 (1H, d; 9)	7.25 (1H, t; 7)
8-H	8.12 (1H, d; 9)	7.92 (1H, d; 9)	7.88 (1H, dd; 2, 7)
4 / TT		0 =0 (0.77	

Taken in acetone- $d_6$ , except for 1a (CDCl<sub>3</sub>+DMSO- $d_6$ ) and 1b (CDCl<sub>3</sub>). Values are in ppm. Multiplicities are indicated by the usual symbols: s, singlet; d, doublet; t, triplet; m, multiplet; dd, double doublet. Figures in parentheses are coupling constants in Hz.  $\alpha$ ) These signals disappeared on addition of D<sub>2</sub>O.

3.58 (2H, m)

5.36 (1H, m)

1.71 (3H, s)

1.81 (3H, s)

Prenylcitpressine was isolated as yellow needles from ether, mp  $160-162^{\circ}$ C,  $C_{20}H_{21}NO_{5}$  (M+: m/z 355). The UV and IR spectra (see Experimental) also showed absorptions characteristic of 9-acridones.<sup>5,6)</sup> The <sup>1</sup>H-NMR studies revealed the presence of a strongly intramolecularly hydrogen bonded proton at  $\delta$  14.31 assignable to the C-1 hydroxyl group of an acridone molecule. In addition, two hydroxyl protons at  $\delta$  9.32 and 3.06, which disappeared with D<sub>2</sub>O, were observed. Furthermore, the presence of an N-methyl, a methoxyl and a prenyl group in

6.81 (1H, d; 10)

5.73 (1H, d; 10)

1.51 (6H, s)

this alkaloid was suggested by the <sup>1</sup>H-NMR spectrum (Table I), and aromatic protons appeared as an *ortho*-coupled AB-type quartet and a sharp one-proton singlet. Cyclization of this alkaloid with formic acid afforded yellow needles, mp 247—250°C. The IR, <sup>1</sup>H-NMR, UV, and mass spectra of this product were superimposable on those of 4 obtained by catalytic hydrogenation of 1a. On the basis of these results, prenylcitpressine can be represented by the formula 3.

A known acridone alkaloid, 5-hydroxynoracronycine  $(5)^{4}$  was also isolated from the same plant, and identified by comparison with an authentic sample. Among the non-alkaloidal components, clausarin (6), 12) suberosin (7a), 13,14) and xanthyletin  $(8)^{13,15}$  were isolated and characterized.

Previous chemical investigation of the alkaloidal constituents of several species of the genus *Citrus* have revealed only the presence of simple alkylamines, quinolones and furoquinolines.<sup>16)</sup> This is the first report of the presence of acridone alkaloids in *Citrus* genus.

## Experimental

All melting points were measured on a micro melting point hot stage apparatus (Yanagimoto).  $^{1}$ H-and  $^{13}$ C-NMR spectra were recorded on PS-100 (JEOL) and FX-100 (JEOL) spectrometers, respectively in acetone- $d_8$  except where otherwise stated. Chemical shifts are given in ppm ( $\delta$ ) with tetramethylsilane (TMS) as an internal reference. Mass spectra (MS) were taken with an M-52 spectrometer (Hitachi) with a direct inlet system. UV spectra were determined in MeOH and infrared (IR) spectra were recorded in KBr tablets unless otherwise noted. Silica Gel GF<sub>254</sub> (Merck) and Silica gel 60 (70—230 mesh ASTM) (Merck) were used for thin layer chromatography (TLC) and column chromatography, respectively. The abbreviations used are as follows: s, singlet; d, doublet; dd, double doublet; t, triplet; q, quartet; m, multiplet; br, broad; sh, shoulder.

Isolation of Acridone Alkaloids and Coumarins from Citrus depressa—The root bark of Citrus depressa HAYATA (250 g) collected on Orchid Island, Taiwan, was extracted with EtOH. The EtOH extract was partitioned between CHCl<sub>3</sub> and H<sub>2</sub>O. The CHCl<sub>3</sub> layer was separated, dried, and concentrated to give a brown syrup (9.0 g). This was subjected to silica gel column chromatography. Elution with acetone-benzene (1:9) yielded five fractions (each 500 ml). Fraction 2 was rechromatographed on a silica gel column. Elution with hexane–EtOAc (4:1) provided successively 6, 7a, 8,  $\beta$ -sitosterol, 1b, 2b, 5, and 1a. Fraction 3 was also subjected to silica gel column chromatography and elution with CHCl<sub>3</sub>-acetone (9:1) gave successively 1a, 2a, and 3.

Citracridone-I (1a)——Orange plates (380 mg) from acetone. mp 275—278°C. UV  $\lambda_{\text{max}}$  nm (log  $\varepsilon$ ): 209 (4.28), 270 (4.67), 282 (sh, 4.63), 295 (sh, 4.58), 340 (4.05), 400 (3.57). UV  $\lambda_{\text{max}}$  (+AlCl<sub>3</sub>) nm: 209, 215 (sh), 272, 290, 307, 363, 452. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 3380, 1620, 1590, 1565. MS m/z (%): 353 (M+, 33), 338 (100), 323 (35), 322 (32), 308 (9), 294 (15), 280 (20), 176 (15), 169 (15), 161 (32). Anal. Calcd for C<sub>20</sub>H<sub>19</sub>NO<sub>5</sub>: C, 67.98; H, 5.42; N, 3.96. Found: C, 67.94; H, 5.42; N, 3.78.

Citracridone-II (1b)—Yellow needles (10 mg) from ether. mp 161—163°C. UV  $\lambda_{\text{max}}$  nm (log  $\varepsilon$ ): 209 (4.37), 268 (4.67), 283 (4.64), 341 (4.14), 405 (3.69). UV  $\lambda_{\text{max}}$  (+AlCl<sub>3</sub>) nm: 248 (sh), 272, 292, 305 (sh), 365, 417. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 1635, 1575, 1550. MS m/z (%): 367 (M<sup>+</sup>, 40), 352 (100), 336 (11), 322 (20).

Citpressine-I (2a)—Yellow needles (54 mg) from acetone, mp 183—185°C. UV  $\lambda_{\text{max}}$  nm (log  $\epsilon$ ): 220 (4.09), 265 (sh, 4.64), 271 (4.65), 294 (sh, 4.06), 332 (3.96), 384 (3.58). UV  $\lambda_{\text{max}}$  (+AlCl<sub>3</sub>) nm: 220, 273, 308 (sh), 352, 420. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 3430, 1620, 1580, 1560. MS m/z (%): 301 (M<sup>+</sup>, 100), 286 (65), 256 (14), 243

(18), 213 (9), 199 (6), 185 (12), 137 (44), 129 (27). Anal. Calcd for  $C_{16}H_{15}NO_5 \cdot 1/2H_2O$ : C, 61.93; H, 5.18; N, 4.51. Found: C, 61.86; H, 5.23; N, 4.30.

Citpressine-II (2b) — Yellow needles (153 mg) from ether, mp 168—170°C. UV  $\lambda_{\text{max}}$  nm (log  $\varepsilon$ ): 220 (4.14), 270 (4.79), 300 (sh, 4.15), 333 (4.12), 382 (3.74). UV  $\lambda_{\text{max}}$  (+AlCl<sub>3</sub>) nm: 230, 265 (sh), 277, 353, 428. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 1650, 1590. MS m/z (%): 315 (M+, 100), 300 (72), 285 (18), 257 (17). Anal. Calcd for C<sub>17</sub>H<sub>17</sub>-NO<sub>5</sub>: C, 64.75; H, 5.43; N, 4.44. Found: C, 64.81; H, 5.47; N, 4.27.

Prenylcit pressine (3)—Yellow plates (12 mg) from ether, mp 160—162°C. UV  $\lambda_{\rm max}$  nm (log \$\epsilon\$): 222 (4.29), 262 (4.51), 269 (4.49), 334 (4.18), 387 (3.76). UV  $\lambda_{\rm max}$  (+AlCl<sub>3</sub>) nm: 234, 250 (sh), 265 (sh), 279, 368, 433. IR  $\nu_{\rm max}$  (CHCl<sub>3</sub>) cm<sup>-1</sup>: 3500, 3230, 1620, 1590, 1555. MS m/z (%): 355 (M<sup>+</sup>, 54), 340 (100), 310 (52). Anal. Calcd for C<sub>20</sub>H<sub>21</sub>NO<sub>5</sub>·1/2H<sub>2</sub>O: C, 65.92; H, 6.09; N, 3.84. Found: C, 66.20; H, 6.53; N, 3.86.

5-Hydroxynoracronycine (5)—Red needles (92 mg) from acetone, mp 268—270°C. UV  $\lambda_{\text{max}}$  nm: 209, 236, 267, 284, 295 (sh), 323, 344 (sh), 418. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 3250, 1630, 1550. MS m/z: 323 (M<sup>+</sup>), 308 (100%), 293, 280, 264, 161, 154, 146, 132, 118. This was shown to be identical with an authentic sample of 5 by IR, <sup>1</sup>H-NMR, and MS comparisons, and mixed mp determination.

Methylation of Citracridone-I (1a)—Compound 1a (25 mg) was treated with CH<sub>2</sub>N<sub>2</sub> in ether at room temp. overnight. Evaporation of the solvent left a solid, which was crystallized from ether to give yellow needles, mp 161—163°C. The IR, <sup>1</sup>H-NMR, UV, and mass spectra were superimposable on those of citracridone-II (1b).

Hydrogenation of Citracridone-I (1a) — A solution of 1a (30 mg) in EtOH (25 ml) was shaken under H<sub>2</sub> gas in the presence of PtO<sub>2</sub> (50 mg) at room temp. for 6 h. The solution was filtered and the filtrate was concentrated. The residue was crystallized from ether to give yellow needles (4) (30 mg), mp 248—250°C. UV  $\lambda_{\text{max}}$  nm: 225, 266, 270, 336, 382. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 3380, 1635, 1580, 1552. MS m/z (%): 355 (M+, 100), 340, 326, 310, 300, 288, 272, 256. <sup>1</sup>H-NMR δ: 1.45 (6H, s, 3'-(CH<sub>3</sub>)<sub>2</sub>), 1.86 (2H, t, J=7 Hz, 2'-H), 2.82 (1H, s, 6-OH), 2.95 (2H, t, J=7 Hz, 1'-H), 3.72 (3H, s, N-CH<sub>3</sub>), 3.94 (3H, s, 5-OCH<sub>3</sub>), 6.08 (1H, s, 2-H), 6.98 (1H, d, J=9 Hz, 7-H), 7.94 (1H, d, J=9 Hz, 8-H), 14.17 (1H, s, 1-OH). Anal. Calcd for C<sub>20</sub>H<sub>21</sub>NO<sub>5</sub>·1/2H<sub>2</sub>O: C, 65.92; H, 6.09; N, 3.84. Found: C, 66.13; H, 5.91; N, 3.73.

Methylation of Citpressine-I (2a)—Treatment of 2a (20 mg) with  $CH_2N_2$  in the usual way afforded yellow needles (20 mg), mp 168—170°C. The product was shown to be identical with citpressine-II (2b) by IR, and <sup>1</sup>H-NMR comparisons and mixed mp determination.

Cyclization of Prenylcitpressine (3)—A solution of 3 (7 mg) in HCOOH (85%, 1 ml) was heated at 80—90°C for 3 h, and then left at room temp. overnight. Water was added and the solution was extracted with CHCl<sub>3</sub>. The CHCl<sub>3</sub> layer was washed with aqueous NaHCO<sub>3</sub> and H<sub>2</sub>O, then dried and concentrated. The residue was chromatographed on silica gel and eluted with disopropyl ether. Fractions (each 5 ml) were collected and monitored by TLC. Fraction 18—23 left yellow residues after evaporation, and the residues were recrystallized from ether to afford yellow needles, mp 247—250°C. The IR, ¹H-NMR, UV and MS were superimposable on those of 4.

Methoxymethylation of Citracridone-I (1a) — A mixture of 1a (30 mg), CH<sub>2</sub>Cl<sub>2</sub> (10 ml), 1% NaOH aq. sol. (10 ml), and phase transfer catalyst (Adogen 464 from Aldrich, 2 mg) was stirred at room temp. for 30 min, and then chloromethylmethyl ether (1.0 ml) was added. After 1 h, the aqueous layer was separated and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The CH<sub>2</sub>Cl<sub>2</sub> layers were combined, dried with MgSO<sub>4</sub>, and evaporated to dryness. The residue was chromatographed on a silica gel column and elution with benzene—acetone (9: 1) afforded yellow plates, which were recrystallized from ether. mp 205—207°C. UV  $\lambda_{\text{max}}$  nm: 235, 269, 285, 295 (sh), 313 (sh), 342, 405. IR  $\nu_{\text{max}}$  (CHCl<sub>3</sub>) cm<sup>-1</sup>: 1622, 1580, 1550. MS m/z: 397 (M+), 382 (100%), 338, 322, 294. <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ: 1.51 (6H, s, 3'-(CH<sub>3</sub>)<sub>2</sub>), 3.52 (3H, s, OCH<sub>3</sub>), 3.70 (3H, s, N-CH<sub>3</sub>), 3.87 (3H, s, 5-OCH<sub>3</sub>), 5.30 (2H, s, O-CH<sub>2</sub>-O), 5.50 (1H, d, J=10 Hz, 2'-H), 6.16 (1H, s, 2-H), 6.52 (1H, d, J=10 Hz, 1'-H), 7.09 (1H, d, J=9 Hz, 7-H), 7.95 (1H, d, J=9 Hz, 8-H), 14.12 (1H, s, 1-OH).

Suberosin (7a)—Colorless needles (30 mg) from MeOH, mp 88—90°C. UV  $\lambda_{\text{max}}$  nm: 210, 224, 255 (sh), 298 (sh), 333. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 1705, 1605. <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.72 (3H, s, CH<sub>3</sub>), 1.78 (3H, s, CH<sub>3</sub>), 3.32 (2H, d, J=8 Hz, H-1'), 3.92 (3H, s, OCH<sub>3</sub>), 5.33 (1H, m, H-2'), 6.25 (1H, d, J=10 Hz, H-3), 6.80 (1H, s, H-5), 7.20 (1H, s, H-8), 7.64 (1H, d, J=10 Hz, H-4). MS m/z: 244 (M+), 229 (100%), 201, 175. This was shown to be identical with an authentic sample prepared from demethylsuberosin (7b) by IR, <sup>1</sup>H-NMR, and MS comparisons, and mixed mp determination.

**Xanthyletin** (8)—Colorless plates (250 mg) from EtOH. mp 130—132°C. UV  $\lambda_{\text{max}}$  nm: 225, 265, 304 (sh), 348. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 1705, 1622. <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.47 (6H, s, 2×CH<sub>3</sub>), 5.72 (1H, d, J=10 Hz, H-7), 6.25 (1H, d, J=10 Hz, H-3), 6.37 (1H, d, J=10 Hz, H-6), 6.76 (1H, s, H-5), 7.07 (1H, s, H-10), 7.60 (1H, d, J=10 Hz, H-4). MS m/z: 228 (M<sup>+</sup>), 213 (100%), 200, 199, 185. This was shown to be identical with an authentic sample by IR, <sup>1</sup>H-NMR, and MS comparisons, and mixed mp determination.

Clausarin (6)—Colorless rod (30 mg) from acetone. mp 208—210°C. UV  $\lambda_{\text{max}}$  nm: 212, 229, 280, 337. IR  $\nu_{\text{max}}$  cm<sup>-1</sup>: 3150, 1715, 1665, 1640. <sup>1</sup>H-NMR  $\delta$ : 1.42 (6H, d, 2×CH<sub>3</sub>), 1.44 (6H, s, 2×CH<sub>3</sub>), 1.63 (6H, s, 2×CH<sub>3</sub>), 4.70 (1H, q, J=1.5 and 10 Hz, H-3′), 4.90 (1H, q, J=1.5 and 18 Hz, H-3′), 5.05 (1H, q, J=1.5 and 18 Hz, H-3″), 5.08 (1H, q, J=1.5 and 10 Hz, H-3″), 5.66 (1H, d, J=10 Hz, H-7), 6.18 (1H, q, J=10 and 18 Hz, H-2″), 6.28 (1H, q, J=10 and 18 Hz, H-2″), 6.72 (1H, d, J=10 Hz, H-6), 7.95 (1H, s, H-4), 8.37 (1H, br s, 5-OH). MS m/z: 380 (M<sup>+</sup>), 365 (100%), 337, 309, 297. This was shown to be identical

with an authentic sample by IR, 1H-NMR, and MS comparisons, and mixed mp determination.

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