Chem. Pharm. Bull. 31(5)1632—1640(1983)

## Syntheses of Acetylated Trisaccharides, $Man\alpha 1 \rightarrow 3Man\beta 1 \rightarrow 4GlcNAc$ and $Man\alpha 1 \rightarrow 2Man\beta 1 \rightarrow 4GlcNAc$ , relating to Mannosidosis

Yoshio Itoh and Setsuzo Tejima\*

Faculty of Pharmaceutical Sciences, Nagoya City University, Tanabe-dori, Mizuho-ku, Nagoya 467, Japan

(Received November 4, 1982)

The title trisaccharides (22 and 30) were synthesized by stepwise condensation of suitably protected monosaccharide units.

3-O-Allyl-2-O-benzoyl-4,6-di-O-benzyl- $\alpha$ -D-glucopyranosyl bromide (9) and 2-acetamido-1,6-anhydro-3-O-benzyl-2-deoxy- $\beta$ -D-glucopyranose (10) were coupled by a modified Koenigs-Knorr glycosidation to give the protected  $Glc\beta l \rightarrow 4GlcNAc$  (12) in 63.7% yield. After removal of the benzoyl group of 12, the C-2' hydroxyl group was isomerized to the D-manno configuration by a sequence consisting of oxidation to ulose and stereoselective borohydride reduction to give the protected  $Man\beta l \rightarrow 4GlcNAc$  (15). Benzylation of 15, followed by deallylation, gave the  $Man\beta l \rightarrow 4GlcNAc$  derivative (18) having an unprotected hydroxyl group at the C-3' position.  $\alpha$ -D-Mannosidation of 18 with acetobromomannose gave the protected  $Man\alpha l \rightarrow 3Man\beta l \rightarrow 4GlcNAc$  (20) in 47.1% yield. Deprotection of 20, followed by acetylation, yielded 22.

2-O-Acetyl-3,4,6-tri-O-benzyl- $\alpha$ -p-glucopyranosyl bromide (23) and 10 were coupled to give the protected  $\mathrm{Glc}\beta\mathrm{l}\to 4\mathrm{GlcNAc}$  (24) in 39.2% yield. Compound 30 was obtained from 24 via four steps using procedures analogous to those used to obtain 22 from 12.

**Keywords**—acetylated Manαl→3Man $\beta$ l→4GlcNAc; acetylated Manαl→2Man $\beta$ l→4GlcNAc; protected Glc $\beta$ l→4GlcNAc; DMSO-Ac<sub>2</sub>O oxidation; protected Man $\beta$ l→4GlcNAc; 1,6-anhydro sugar derivative; <sup>1</sup>H-NMR; <sup>13</sup>C-NMR

Mannosidosis has been shown to be an inherited lysosomal storage disease, in which oligosaccharides containing p-mannose (Man) and N-acetylglucosamine (GlcNAc) are the storage substances. Lundblad  $et\ al.^{1)}$  isolated a trisaccharide, Man $\alpha$ 1 $\rightarrow$ 3Man $\beta$ 1 $\rightarrow$ 4GlcNAc (1), from pooled urine of patients as the major storage material in mannosidosis, and suggested that the oligosaccharide is probably a degradation product derived from the inner core of glycoprotein chains. Recently, Jeanloz  $et\ al.^{2)}$  utilized 1 as a starting material for the synthesis of Man $\alpha$ 1 $\rightarrow$ 3Man $\beta$ 1 $\rightarrow$ 4GlcNAc $\beta$ 1 $\rightarrow$ 4GlcNAc  $\alpha$ -phosphate, the synthetic precursor of "lipid intermediate." In addition, 1 is also the common core structure in accumulating complex oligosaccharides isolated from the livers of patients suffering from a deficiency of  $\beta$ -p-galactosidase, 3) from  $G_{M1}$ -gangliosidosis, Type I,4) and fucosidosis.5)

In order to develop a reasonable approach toward the synthesis of sugar chains present in glycoproteins, in which a high-mannose or complex-type sugar chain is linked to protein by an N-glycosidic linkage, 1 and  $\text{Man}\alpha 1 \rightarrow 2\text{Man}\beta 1 \rightarrow 4\text{GlcNAc}$  (2), a structural isomer of 1, were chosen as targets for our synthetic studies on oligosaccharides of biological interest. These compounds may provide useful substrates for studies of the glycosidases involved in the biosynthesis and catabolism of glycoproteins.

In this paper, fully acetylated 1 and 2 (22 and 30) were synthesized by stepwise condensation of suitably protected monosaccharide units. The key point of this work is the synthesis of an amino disaccharide bearing a  $\beta$ -D-mannopyranosyl linkage at the C-4 position of GlcNAc (Man $\beta$ 1 $\rightarrow$ 4GlcNAc). This is because, despite several attempts, 6) stereoselective synthesis of  $\beta$ -D-mannopyranosides from D-mannopyranosyl halides has not yet been established, and the low reactivity of the hydroxyl group at the C-4 position of GlcNAc provides additional difficulties.

In order to overcome these barriers, the authors selected, as glucosyl donors, partially

etherated bromides bearing acyl groups at the C-2 position and, as a glucosyl acceptor, a 1,6-anhydro- $\beta$ -GlcNAc derivative having an unprotected hydroxyl group at the C-4 position and a benzyl group at the C-3 position. We subsequently converted the  $Glc\beta1\rightarrow 4GlcNAc$  thus obtained into a  $Man\beta1\rightarrow 4GlcNAc$  derivative by a sequence of oxidation and stereoselective reduction, resulting in epimerization at C-2'. Therefore, synthesis of the glucosyl donor for amino disaccharide synthesis is first described.

3-O-Allyl-1,2: 5,6-di-O-isopropylidene-α-D-glucofuranose<sup>7)</sup> prepared from 1,2: 5,6-di-O-isopropylidene-α-D-glucofuranose (3), was deisopropylidenated to give the 3-O-allyl ether. Without further purification, subsequent acetylation of the 3-O-allyl ether gave 1,2,4,6-tetra-O-acetyl-3-O-allyl-β-D-glucopyranose (4) as fine needles. In the proton nuclear magnetic resonance (¹H-NMR) spectrum, the anomeric proton (H-1) of 4 appeared as a doublet with a reasonable coupling constant for the assigned configuration. The corresponding α-bromide was prepared from 4 by treatment with hydrogen bromide in acetic acid. Without purification, the bromide was converted to the corresponding orthoester, 4,6-di-O-acetyl-3-O-allyl-1,2-O-(1-ethoxyethylidene)-α-D-glucopyranose (5), according to a slight modification of the procedure described for the synthesis of the fully acetylated analog. Benzylation of 5 with benzyl bromide and base, followed by removal of the ethoxyethylidene group by acid hydrolysis, gave 3-O-allyl-4,6-di-O-benzyl-D-glucopyranose (6).

Benzoylation of 6 gave 3-O-allyl-1,2-di-O-benzoyl-4,6-di-O-benzyl- $\beta$ -D-glucopyranose (7) as a syrup. In order to obtain crystalline di-O-acylates, anisoylation of 6 was carried out, but the resultant di-O-anisoyl ester (8) was also a syrup, and so this route was not further pursued. Treatment of 7 with hydrogen bromide in acetic acid gave 3-O-allyl-2-O-benzoyl-4,6-di-O-benzyl- $\alpha$ -D-glucopyranosyl bromide (9), which was subjected to glucosidation.

$$R^{4}O$$

$$R^{1}O$$

$$R^{3}O$$

$$R^{2}$$

$$R^{1}=OAc, R^{2}=H, R^{3}=R^{4}=Ac$$

$$6: R^{1}, R^{2}=H, OH, R^{3}=H, R^{4}=Bn$$

$$7: R^{1}=OBz, R^{2}=H, R^{3}=Bz, R^{4}=Bn$$

$$8: R^{1}=OAnis, R^{2}=H, R^{3}=Anis, R^{4}=Bn$$

$$9: R^{1}=H, R^{2}=Br, R^{3}=Bz, R^{4}=Bn$$

$$11: R^{1}, R^{2}=H, OH, R^{3}=Bz, R^{4}=Bn$$

$$Ac=acetyl, All=allyl, Anis=anisoyl Bn=benzyl, Bz=benzoyl$$

$$OAc$$

$$AcO$$

$$AllO$$

Chart 1

Condensation of one molar equivalent of 2-acetamido-1,6-anhydro-3-O-benzyl-2-deoxy- $\beta$ -D-glucopyranose (10)<sup>9)</sup> with about three molar equivalents of 9 in benzene-nitromethane in the presence of mercuric cyanide and Drierite gave 2-acetamido-1,6-anhydro-3-O-benzyl-2-deoxy-4-O-(3-O-allyl-2-O-benzyl-4,6-di-O-benzyl- $\beta$ -D-glucopyranosyl)- $\beta$ -D-glucopyranose (12) in a yield of 63.7% with a small amount of 3-O-allyl-2-O-benzyl-4,6-di-O-benzyl-D-glucopyranosyl)

pyranose (11). Treatment of 12 with alkali caused selective debenzoylation to yield the disaccharide derivative (13) bearing only one unprotected hydroxyl group at the C-2 position of the p-glucose moiety. Acetylation of 13 gave the mono-O-acetate (14). The  $\beta$ -p-configuration of the newly introduced glucosidic linkage was confirmed by <sup>1</sup>H-NMR and carbon-13 nuclear magnetic resonance (<sup>13</sup>C-NMR) spectroscopies.

The unprotected hydroxyl group in 13 was then isomerized to D-manno configuration by reference to the method for isomerization of the amino disaccharide derivative. Thus, oxidation of 13 with dimethyl sulfoxide-acetic anhydride (DMSO-Ac<sub>2</sub>O) and, without purification, subsequent stereospecific reduction of the ulose with sodium borohydride yielded the  $\beta$ -D-mannopyranosyl disaccharide (15), which was acetylated to give the acetate (16).

Comparison of 13 with 15 showed differences in optical rotations (13,  $[\alpha]_D^{20}$  -41.4°; 15, -48.5°) and different mobilities on thin-layer chromatography (TLC). In the <sup>1</sup>H-NMR spectra of 13 and 15, the one-proton singlet due to the hydroxyl at C-2' was observed at 2.96 and 2.38 ppm, respectively. In the <sup>13</sup>C-NMR spectra of 13 and 15, the resonances of C-1' were observed at 102.1 and 99.3 ppm with <sup>1</sup>J values of 154.4 and 155.6 Hz, respectively. Therefore, the occurrence of isomerization from p-gluco to p-manno was confirmed.<sup>11</sup>

The unprotected hydroxyl group at C-2' of 15 was benzylated to give the corresponding benzyl ether (17). In order to remove the allyl group at the C-3' position, 17 was treated with tris(triphenylphosphine)rhodium chloride, 12) and the resultant 1-propenyl ether was removed with acid 13) to yield the deallylated product (18) in 43.2% yield. In the 1H-NMR spectrum, the resonance of the hydroxyl proton at C-3' was newly observed at 2.38 ppm as a one-proton singlet. Removal of the allyl group could be also effected by reaction of 17 with 10% palladium on charcoal. This method was recently recommended as a one-step deallylation procedure by Ogawa and Matsui. 14) However, the yield of 18 was not improved as much as expected.

Protected trisaccharide synthesis was then carried out by a conventional Koenigs–Knorr condensation as described for the preparation of the protected disaccharide (12). Namely, 2,3,4,6-tetra-O-acetyl-α-D-mannopyranosyl bromide (19) was coupled with 18. After a preliminary chromatographic purification, which did not separate the trisaccharide derivative (20) from 19, the crude 20 was de-O-acetylated, and the product was separated by preparative TLC (PTLC). Re-acetylation gave 20 in 47.1% yield from 18. It was characterized by elementary analysis, and infrared spectra (IR), <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectroscopies.

Hydrogenolytic removal of the benzyl groups of 20, followed by acetylation, gave the fully acetylated 1,6-anhydro- $\beta$ -trisaccharide (21) in 66.9% yield. The structure was characterized as described for 20. Finally, the 1,6-anhydro- $\beta$ -linkage of 21 was acetolyzed at 0°C with a mixture of boron trifluoride etherate and acetic anhydride to give the fully acetylated Man $\alpha$ 1 $\rightarrow$ 3Man $\beta$ 1 $\rightarrow$ 4GlcNAc (22) as an anomeric mixture in 70.1% yield. The product was separated as a white powder having  $[\alpha]_D^{20} + 20^\circ$ . Jeanloz et al.<sup>2)</sup> reported the monohydrate of 22 to be an amorphous solid having mp 106—109°C and  $[\alpha]_D^{20}$  0°.

The fully acetylated  $\text{Man}\alpha 1 \rightarrow 2\text{Man}\beta 1 \rightarrow 4\text{GlcNAc}$  (30), which is the second title sugar and a structural isomer of 22, was synthesized *via* analogous procedures from 9 and 10.

The first step, condensation of 10 and a glucosyl donor, 2-O-acetyl-3,4,6-tri-O-benzyl- $\alpha$ -D-glucopyranosyl bromide (23),<sup>15)</sup> was carried out by a modified Koenigs-Knorr reaction. After column chromatographic purification, the amino disaccharide derivative (24) was isolated in 39.2% yield. <sup>13</sup>C-NMR spectroscopy confirmed the  $\beta$ -D-gluco configuration of the newly introduced glucosidic linkage. Treatment of 24 with alkali gave the amino disaccharide derivative (25) bearing only one unprotected hydroxyl group at the C-2' position.

The second step, isomerization of the  $\beta$ -D-glucopyranosyl moiety of 25 to  $\beta$ -D-manno configuration, was carried out by a sequence of oxidation with DMSO-Ac<sub>2</sub>O and subsequent stereoselective reduction with sodium borohydride to give the  $\beta$ -D-mannosyl amino disaccharide (26). Acetylation of 26 yielded the mono-O-acetyl mannosyl amino disaccharide (27). The inversion from D-gluco to D-manno was confirmed by comparison of the Rf values on TLC and

No. 5

<sup>13</sup>C-NMR spectral data for 25 and 26, or 24 and 27.

The third step is synthesis of the fully protected trisaccharide derivative (28). Condensation of acetobromomannose (19) with 26 was carried out by a modified Koenigs-Knorr reaction. The crude trisaccharide derivative was purified by a sequence of de-O-acetylation, chromatographic separation, re-acetylation, and PTLC. The yield was 40.7%. The  $\alpha$ -D-configuration of the newly introduced mannosidic linkage was confirmed by <sup>13</sup>C-NMR spectroscopy.

Chart 3

1636 Vol. 31 (1983)

The final step is conversion of the protecting groups of 28 to acetyl groups. Hydrogenolytic debenzylation followed by acetylation gave the fully acetylated 1,6-anhydro- $\beta$ -trisaccharide (29). The 1,6-anhydro- $\beta$ -linkage was then acetolyzed to give the fully acetylated Man $\alpha$ 1 $\rightarrow$ 2Man $\beta$ 1 $\rightarrow$ 4GlcNAc (30), which was separated as a white powder in 77.4% yield from 29. The product was characterized as the  $\alpha$ -acetate by <sup>13</sup>C-NMR spectroscopy.

The present work further confirms<sup>9,16)</sup> that 1,6-anhydro- $\beta$ -derivatives of monosaccharides and oligosaccharides are extremely versatile starting materials or key intermediates for syntheses of complex oligosaccharides.

## Experimental

Solutions were concentrated with a Büchi-Shibata Rotavapor EL-130 below 45°C under a vacuum. Melting points were determined with a Yanagimoto MP-S2 micro melting point apparatus, and are uncorrected. Optical rotations were measured with a Union Giken PM-201 automatic digital polarimeter in a 0.5 dm tube. <sup>1</sup>H- and <sup>13</sup>C-NMR spectra were recorded at 100 and 25 MHz, with JEOL JNM-MH-100 and -FX-100 spectrometers, respectively. Tetramethylsilane (TMS) was used as an internal standard. Chemical shifts are given in ppm from TMS. IR spectra were recorded with a JASCO IRA-2 or an A-102 spectrometer. TLC, PTLC, and preparative–layer chromatography (PLC) were performed on pre-coated plates of Silica Gel 60 F<sub>254</sub> 0.25 mm thick, 0.5 mm thick, and PLC plates 2 mm thick (E. Merck), respectively. The following solvent combinations were used for TLC: (A), CHCl<sub>3</sub>-acetone (6: 1); (B), CHCl<sub>3</sub>-acetone (3: 1); (C), hexane-ether (1: 1). Detection was effected by ultraviolet (UV) irradiation at 254 nm or with a spray reagent (A), anisaldehyde–H<sub>2</sub>SO<sub>4</sub>–EtOH at 125°C;<sup>171</sup> (B), 1% KMnO<sub>4</sub> in 2% Na<sub>2</sub>CO<sub>3</sub> solution. Column chromatography was performed on Silica Gel 60 (70—230 mesh, E. Merck). All solvent compositions are given as v/v.

1,2,4,6-Tetra-O-acetyl-3-O-allyl- $\beta$ -D-glucopyranose (4)—3-O-Allyl-1,2: 5,6-di-O-isopropylidene- $\alpha$ -D-glucofuranose, prepared from 1,2: 5,6-di-O-isopropylidene- $\alpha$ -D-glucofuranose (3, 12 g, 46.1 mmol) by the method of Corbett and McKay, in 1.2% (w/v) aq.  $H_2SO_4$  (180 ml) was heated to reflux for 3 h. The solution was neutralized with BaCO<sub>3</sub>, filtered, and the filtrate was extracted with CHCl<sub>3</sub> to remove by-products. The aqueous phase was concentrated to dryness, and the residue was acetylated by heating for 1.5 h with Ac<sub>2</sub>O (100 ml) and anhyd. AcONa (5 g). The mixture was poured into ice- $H_2O$  and extracted with CHCl<sub>3</sub>. The combined extracts were successively washed with  $H_2O$ , ice-cold aq. NaHCO<sub>3</sub>, and  $H_2O$ , then dried (MgSO<sub>4</sub>), and concentrated to a syrup, which was crystallized from EtOH-hexane as fine needles (10.06 g, 56.2% based on 3), mp 119—120°C, [ $\alpha$ ]<sup>22</sup> +4.7° (c=1.3, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 2.08, 2.09, 2.10, 2.11 (12H, each s, OAc×4), 5.62—6.08 (1H, m, CH<sub>2</sub>=CHCH<sub>2</sub>-), 5.72 (1H, d,  $J_{1,2}$ =8 Hz, H-1). IR  $\nu$ <sup>max</sup><sub>max</sub> cm<sup>-1</sup>: 1735 (OAc). TLC: Rf 0.78 (solvent A). Anal. Calcd for C<sub>17</sub>H<sub>24</sub>O<sub>10</sub>: C, 52.58; H, 6.23. Found: C, 52.27; H, 6.19.

4,6-Di-O-acetyl-3-O-allyl-1,2-O-(1-ethoxyethylidene)- $\alpha$ -D-glucopyranose (5)—A stirred solution of 4 (3 g, 7.72 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (20 ml) was treated dropwise with 30% (w/v) HBr-AcOH (12 ml) at 0°C. After being stirred for 1 h at 0°C, the mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, successively washed with H<sub>2</sub>O, ice-cold aq. NaHCO<sub>3</sub>, and H<sub>2</sub>O, then dried (MgSO<sub>4</sub>), and concentrated to yield a syrupy bromide.

A mixture of the bromide, EtOH (1.5 ml), 2,6-lutidine (1.8 ml), and nitromethane (20 ml) was stirred at 40°C for 16 h. After being diluted with  $CH_2Cl_2$ , the solution was successively washed with  $H_2O$ , ice-cold 1 n  $H_2SO_4$  and aq. NaHCO<sub>3</sub>, and  $H_2O$ , then dried (MgSO<sub>4</sub>), and concentrated to a syrup (2.56 g, 88.6%). For analysis, the syrup was chromatographed on a column with  $CHCl_3$ -acetone (5: 1) to yield pure 5,  $[\alpha]_2^{20}$ 

 $+42.8^{\circ}$  (c=0.32, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.20 (3H, t, J=7 Hz, CH<sub>3</sub>CH<sub>2</sub>O-), 1.70 (3H, s, CC), CH<sub>3</sub> (2.09 (6H, s, OAc×2), 3.60 (2H, q, J=7 Hz, CH<sub>3</sub>CH<sub>2</sub>O-), 5.63—6.20 (1H, m, CH<sub>2</sub>=CHCH<sub>2</sub>-), 5.74 (1H, d,  $J_{1,2}=5$  Hz, H-1). TLC: Rf 0.44 (solvent A). Anal. Calcd for  $C_{17}H_{26}O_{9}$ : C, 54.54; H, 7.00. Found: C, 54.17; H, 6.88.

3-O-Allyl-4,6-di-O-benzyl-p-glucopyranose (6)—Benzyl bromide (20 ml) and powdered KOH (20 g) were added to a solution of 5 (10.8 g, 29 mmol) in 1,4-dioxane (100 ml). The mixture was stirred at 70°C for 4 h, cooled, and diluted with CHCl<sub>3</sub>. After filtration, the filtrate was successively washed with  $H_2O$ , ice-cold 10%  $H_2SO_4$ , aq. NaHCO<sub>3</sub> and  $H_2O$ , and then concentrated to a syrup.

The resultant benzyl ether in 1,4-dioxane-1 m  $H_2SO_4$  [4: 1 (v/v), 150 ml] was heated under reflux for 4 h to hydrolyze it. After neutralization with solid NaHCO<sub>3</sub>, the mixture was concentrated to a syrup, which was dissolved in CHCl<sub>3</sub>. The solution was washed with  $H_2O$ , dried (MgSO<sub>4</sub>), and concentrated to a syrup, which was column-chromatographed with hexane-AcOEt (1: 1) to yield 6 as a white solid (4.67 g, 40.2%). For analysis, the solid was crystallized from ether-pentane as white silky needles, mp 84—85°C, [ $\alpha$ ]<sup>20</sup> +72.5° (c=0.8, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 5.80—6.20 (1H, m, CH<sub>2</sub>=CHCH<sub>2</sub>-), 7.31 (10H, s, PhCH<sub>2</sub>×2), 3.19—5.50 [17H, 15H (unresolved ring protons) and 2H (exchangeable with D<sub>2</sub>O)], IR  $\nu$ <sup>RBP</sup><sub>max</sub> cm<sup>-1</sup>: 3330 (OH). TLC:

Rf 0.60 (solvent B). Anal. Calcd for C<sub>23</sub>H<sub>28</sub>O<sub>6</sub>: C, 68.98; H, 7.05. Found: C, 69.01; H, 7.19.

3-O-Allyl-1,2-di-O-benzoyl-4,6-di-O-benzyl-β-D-glucopyranose (7)——Benzoyl chloride (0.4 ml) was added to a chilled solution of 6 (206 mg,  $5.14 \times 10^{-4}$  mol) in dry pyridine (5 ml), and the mixture was stirred for 18 h at room temperature. After dropwise addition of  $H_2O$  (1 ml) to decompose excess benzoyl chloride, the mixture was concentrated to a syrup. This was dissolved in  $CH_2Cl_2$ , washed with ice-cold dil. HCl,  $H_2O$ , ice-cold aq. NaHCO<sub>3</sub> solution, and  $H_2O$ , then dried (MgSO<sub>4</sub>), and concentrated to a syrup. On column chromatography with hexane-ether (3: 1), 7 (242.7 mg, 77.6%) was obtained as a syrup,  $[\alpha]_D^{10} + 143^\circ$  (c = 0.66, CHCl<sub>3</sub>).  $^{1}$ H-NMR (CDCl<sub>3</sub>): 5.87 (1H, d,  $J_{1,2} = 8$  Hz, H-1), 7.00 - 7.62 (16H, m, PhCH<sub>2</sub>×2 and meta and para to C=O of benzoyl×2), 7.03, 7.04 (4H, each d, J = 8 Hz, aromatic protons ortho to C=O of benzoyl×2). IR  $\nu_{max}^{nest}$  cm<sup>-1</sup>: 1725 (OBz). TLC: Rf 0.64 (solvent C). Anal. Calcd for  $C_{37}H_{36}O_8$ : C, 73.01; H, 5.96. Found: C, 72.81; H, 5.89.

3-O-Allyl-1,2-di-O-anisoyl-4,6-di-O-benzyl- $\beta$ -D-glucopyranose (8)—Anisoyl chloride (0.7 g) was added to a solution of 6 (212 mg,  $5.29 \times 10^{-4}$  mol) in dry pyridine (5 ml). After being stirred for 42 h at room temperature, the mixture was treated as described for the procedure employing 7. Two column chromatographies with CH<sub>2</sub>Cl<sub>2</sub> afforded 8 (283 mg, 80%) as a pale yellow syrup,  $[\alpha]_D^{2n} + 53.1^{\circ}$  (c=1.3, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 3.77 (6H, s, OCH<sub>3</sub>×2), 5.96 (1H, d,  $J_{1,2}=8$  Hz, H-1), 6.84, 6.86 (4H, each d, J=9 Hz, aromatic protons ortho to OCH<sub>3</sub> of anisoyl×2), 7.28, 7.31 (10H, each s, PhCH<sub>2</sub>×2), 7.96 (4H, d, J=9 Hz, aromatic protons ortho to C=O of anisoyl). IR  $\nu_{\max}^{\text{mest}}$  cm<sup>-1</sup>: 1722 (C=O). TLC: Rf 0.31 (solvent C). Anal. Calcd for C<sub>39</sub>H<sub>40</sub>O<sub>10</sub>: C, 70.05; H, 6.03. Found: C, 70.06; H, 5.88.

3-O-Allyl-2-O-benzoyl-4,6-di-O-benzyl- $\alpha$ -D-glucopyranosyl Bromide (9)—A mixture of 7 (3.5 g, 5.75 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (70 ml) with 30% (w/v) HBr-AcOH (18 ml) was stirred at 0°C for 40 min. After dilution with CH<sub>2</sub>Cl<sub>2</sub>, the mixture was washed with H<sub>2</sub>O, ice-cold aq. NaHCO<sub>3</sub> solution, and H<sub>2</sub>O, then dried (MgSO<sub>4</sub>), and filtered. The filtrate was concentrated to dryness by repeated co-distillation with dry toluene to yield 9 (2.94 g, 90.1%), which was immediately used.

2-Acetamido-1,6-anhydro-3-O-benzyl-2-deoxy-4-O-(3-O-allyl-2-O-benzyl-4,6-di-O-benzyl-β-D-gluco pyranosyl)-β-D-glucopyranose (12)—A solution of 9 (2.94 g, 5.18 mmol) in benzene-nitromethane (1:1, 10 ml) was added to a suspension of  $10^9$ ) (542 mg, 1.79 mmol), Hg(CN)<sub>2</sub> (2 g), and Drierite (0.6 g) in the same solvent (4 ml). The mixture was stirred for 25 h at room temperature, then filtered, and the filtrate was diluted with CHCl<sub>3</sub>. This solution was successively washed with H<sub>2</sub>O, satd. KI and NaHCO<sub>3</sub> solutions, and H<sub>2</sub>O, then dried (MgSO<sub>4</sub>), and concentrated to a syrup, which was column-chromatographed with hexane-ether (1:4). From the faster moving fractions having Rf 0.76 (solvent A), a trace of 3-O-allyl-2-O-benzoyl-4,6-di-O-benzyl-D-glucopyranose (11) was isolated after removal of the solvent. Compound 11 was crystallized from CHCl<sub>3</sub>-hexane as white needles, mp 139—140°C, [α]<sub>D</sub><sup>16</sup> +123.7° (c=0.35, CHCl<sub>3</sub>). Anal. Calcd for C<sub>30</sub>H<sub>32</sub>O<sub>7</sub>: C, 71.41; H, 6.39. Found: C, 71.64; H, 6.40.

After 11 had emerged, 12 was eluted with the same solvent, and isolated as a foamy solid (903 mg, 63.7%),  $[\alpha]_D^{22} - 32.9^\circ$  (c = 0.47, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 2.12 (3H, s, NAc), 6.44 (1H, d,  $J_{\rm NH,2} = 10$  Hz, NH), 7.06—8.22 (20H, m, aromatic protons). IR  $\nu_{\rm max}^{\rm RST}$  cm<sup>-1</sup>: 3360 (NH), 1716 (OBz), 1669 (amide I), 1509 (amide II). TLC: Rf 0.71 (solvent A). Anal. Calcd for  $C_{45}H_{49}NO_{11} \cdot 1/2H_2O$ : C, 68.51; H, 6.39; N, 1.78. Found: C, 68.27; H, 6.44; N, 1.89.

2-Acetamido-1,6-anhydro-3-O-benzyl-2-deoxy-4-O-(3-O-allyl-4,6-di-O-benzyl-β-D-glucopyranosyl)-β-D-glucopyranose (13)——A 0.5 N methanolic solution of MeONa (10 ml) was added to a solution of 12 (2.07 g, 2.62 mmol) in dry MeOH (50 ml). After being stirred overnight at room temperature, the mixture was decationated with Amberlite IR-120 (H<sup>+</sup>) resin, filtered, and concentrated to a syrup, which was chromatographed on a column with CHCl<sub>3</sub>-MeOH (100: 1). Removal of the solvent from the major fractions provided 13 as a foamy solid (1.4 g, 76%),  $[\alpha]_D^{22}$  -41.4° (c=0.46, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.98 (3H, s, NAc), 2.92 (1H, s, OH), 6.30 (1H, d,  $J_{NH,2}$ =8 Hz, NH), 7.26, 7.29 (15H, each s, aromatic protons). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 102.1 ( ${}^1J_{C-1'-H-1'}$ =154.4 Hz, C-1'), 100.5 ( ${}^1J_{C-1-H-1}$ =173.3 Hz, C-1). IR  $v_{max}^{KBT}$  cm<sup>-1</sup>: 3380 (OH, NH), 1650 (amide I), 1522 (amide II). TLC: Rf 0.46 (solvent B). Anal. Calcd for  $C_{38}H_{45}NO_{10} \cdot 1.5H_2O$ : C, 64.94; H, 6.88; N, 1.99. Found: C, 65.05; H, 6.67; N, 2.13.

2-Acetamido-1,6-anhydro-3-O-benzyl-2-deoxy-4-O-(2-O-acetyl-3-O-allyl-4,6-di-O-benzyl-β-D-glucopyranosyl)-β-D-glucopyranose (14)——A mixture of 13 (36.9 mg,  $5.25 \times 10^{-5}$  mol) in Ac<sub>2</sub>O (0.5 ml) and pyridine (1 ml) was stirred overnight at room temperature, then concentrated to a syrup. This was column-chromatographed with toluene-acetone (4: 1) to isolate 14 (37.5 mg, 99.5%) as a syrup,  $[\alpha]_D^{24}$  – 101.5° (c=0.13, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 2.09, 2.16 (6H, each s, OAc, NAc), 6.33 (1H, d,  $J_{\rm NH,2}$ =10 Hz, NH), 7.12—7.48 (15H, m, aromatic protons). IR  $\nu_{\rm max}^{\rm nest}$  cm<sup>-1</sup>: 3375 (NH), 1738 (OAc). TLC: Rf 0.63 (solvent A). Anal. Calcd for C<sub>40</sub>H<sub>47</sub>NO<sub>11</sub>: C, 66.93; H, 6.60; N, 1.95. Found: C, 66.83; H, 6.88; N, 2.11.

2-Acetamido-1,6-anhydro-3-O-benzyl-2-deoxy-4-O-(3-O-allyl-4,6-di-O-benzyl- $\beta$ -D-mannopyranosyl)- $\beta$ -D-glucopyranose (15)—A solution of 13 (233.7 mg,  $3.33\times10^{-4}$  mol) in DMSO-Ac<sub>2</sub>O (2:1, v/v, 6 ml) was stirred for 48 h at room temperature. After dilution with CHCl<sub>3</sub>, the whole was washed with H<sub>2</sub>O, dried (MgSO<sub>4</sub>), and concentrated to a syrup by repeated co-distillation with toluene.

A mixture of this syrup and NaBH<sub>4</sub> (200 mg) in CH<sub>2</sub>Cl<sub>2</sub>-MeOH (1:1, 6 ml) was stirred for 4 h at room temperature, and then diluted with CHCl<sub>3</sub>. The whole was successively washed with H<sub>2</sub>O, ice-cold 10%

citric acid and satd. NaHCO<sub>3</sub> solutions, and H<sub>2</sub>O, dried (MgSO<sub>4</sub>), and concentrated to a syrup. This was chromatographed on a column with CHCl<sub>3</sub>–MeOH (100: 1) to provide 15 (137.1 mg, 58.6%) as a foamy solid, [ $\alpha$ ]<sup>19</sup>  $-48.5^{\circ}$  (c=0.41, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.99 (3H, s, NAc), 2.38 (1H, s, OH), 5.48 (1H, s, H-1), 6.46 (1H, d,  $J_{\rm NH,2}=10$  Hz, NH), 7.36, 7.41 (15H, each s, aromatic protons). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 100.6 (<sup>1</sup> $J_{\rm C_{-1-H-1}}=175.2$  Hz, C-1), 99.3 (<sup>1</sup> $J_{\rm C_{-1'-H-1'}}=155.6$  Hz, C-1'). IR  $v_{\rm max}^{\rm kBr}$  cm<sup>-1</sup>: 3380 (NH, OH), 1652 (amide I), 1520 (amide II). TLC: Rf 0.31 (solvent B). Anal. Calcd for C<sub>38</sub>H<sub>45</sub>NO<sub>10</sub>·1.5H<sub>2</sub>O: C, 64.94; H, 6.88; N, 1.99. Found: C, 64.89; H, 6.95; N, 2.24.

2-Acetamido -1,6-anhydro -3-O- benzyl-2-deoxy-4-O-(2-O-acetyl-3-O-allyl-4,6-di-O-benzyl- $\beta$ -D-mannopyranosyl)- $\beta$ -D-glucopyranose (16)—Acetylation of 15 (30.4 mg,  $4.33 \times 10^{-5}$  mol) with Ac<sub>2</sub>O (0.5 ml) and pyridine (1 ml) as described for 14 provided 16 (31 mg, 99.5%) as a syrup, [α]<sub>D</sub><sup>17</sup> -100° (c=0.12, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 2.07, 2.17 (6H, each s, OAc, NAc), 6.18 (1H, d,  $J_{NH,2}$ =10 Hz, NH), 7.26, 7.31 (15H, each s, aromatic protons). IR  $\nu_{max}^{neat}$  cm<sup>-1</sup>: 3370 (NH), 1730 (OAc). TLC: Rf 0.49 (solvent A). Anal. Calcd for  $C_{40}H_{47}$ NO<sub>11</sub>: C, 66.93; H, 6.60; N, 1.95. Found: C, 66.80; H, 6.43; N, 1.75.

2-Acetamido-1,6-anhydro-3-O-benzyl-2-deoxy-4-O-(3-O-allyl-2,4,6-tri-O-benzyl-β-D-mannopyranosyl)-β-D-glucopyranose (17)——Benzyl bromide (2 ml) was added to a mixture of 15 (0.64 g, 9.11 × 10<sup>-4</sup> mol), powdered BaO (1.7 g), and Ba(OH)<sub>2</sub>·8H<sub>2</sub>O (0.7 g), suspended in dry DMF (30 ml). The mixture was stirred at 50°C for 48 h, cooled, diluted with CHCl<sub>3</sub>, and filtered. The filtrate was successively washed with ice-cold dil. HCl, H<sub>2</sub>O, ice-cold NaHCO<sub>3</sub> solution, and H<sub>2</sub>O, dried (MgSO<sub>4</sub>), and concentrated to a syrup, which was column-chromatographed with hexane-ether (1: 4). Fractions having Rf 0.58 (solvent A) were further purified by PLC with toluene-acetone (4: 1). A zone having Rf 0.38 was excluded from the plates and extracted with CHCl<sub>3</sub>-MeOH (9: 1) to isolate 17 (0.48 g, 69.5%) as a syrup, [α]<sup>13</sup><sub>D</sub> -87° (c=0.94, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.61 (3H, s, NAc), 6.05 (1H, d,  $J_{NH,2}$ =9 Hz, NH), 7.22 (20H, s, aromatic protons). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 100.6 (C-1, C-1'). IR  $\nu_{max}^{nest}$  cm<sup>-1</sup>: 3380 (NH), 1665 (amide I), 1505 (amide II). TLC: Rf 0.58 (solvent A).

2-Acetamido-1,6-anhydro-3-*O*-benzyl-2-deoxy-4-*O*-(2,4,6-tri-*O*-benzyl-β-D-mannopyranosyl)-β-D-glucopyranose (18)——A) Deallylation with Pd-catalyst: A mixture of 17 (56.4 mg,  $7.36 \times 10^{-5}$  mol) and 10% Pd-on-charcoal (60 mg) suspended in EtOH-AcOH-H<sub>2</sub>O (2:1:1, 5 ml) was stirred at 75°C for 5 h. then filtered. The filtrate was concentrated to dryness by repeated co-distillation with EtOH and toluene. The residue was purified by PTLC with solvent B. The band having Rf 0.63 was excluded from the plates and extracted with CHCl<sub>3</sub>-MeOH (9:1) to isolate 18 (19.1 mg, 35.7%) as a foamy solid,  $[\alpha]_D^{21} - 85.9^\circ$  (c = 0.17, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.70 (3H, s, NAc), 2.38 (1H, s, OH), 6.07 (1H, d,  $J_{NH,2} = 10$  Hz, NH), 7.33 (20H, s, aromatic protons). <sup>18</sup>C-NMR (CDCl<sub>3</sub>): 100.7 ( $^1J_{C-1'-H-1'} = 158.1$  Hz, C-1'), 100.6 ( $^1J_{C-1-H-1} = 171.5$ Hz, C-1). IR  $r_{msr}^{msr}$  cm<sup>-1</sup>: 3400 (NH, OH), 1664 (amide I), 1515 (amide II). TLC: Rf 0.63 (solvent B). *Anal.* Calcd for  $C_{42}H_{47}NO_{10}$ : C, 69.50; H, 6.53; N, 1.93. Found: C, 69.61; H, 6.68; N, 1.96.

B) Deallylation with Rhodium Complex: A mixture of 17 (252.3 mg,  $3.29 \times 10^{-4}$  mol), tris(triphenylphosphine) rhodium chloride (22 mg), and 1,4-diazabicyclo[2.2.2]octane (33 mg) dissolved in EtOH-benzene-H<sub>2</sub>O (7: 3: 1, 8 ml) was boiled under reflux for 4 h under stirring, then concentrated to a syrup. This was dissolved in 80% (v/v) aq. AcOH (13 ml). The solution was stirred at 80°C for 2 h, cooled, and concentrated to a syrup by repeated co-distillation with toluene. The resultant syrup was purified by PTLC as described in method A) to isolate the deallylation product (102.8 mg, 43.2%), which was indistinguishable from 18 by TLC, IR and NMR comparisons.

O-(2,3,4,6-Tetra-O-acetyl- $\alpha$ -p-mannopyranosyl)-(1 $\rightarrow$ 3)-O-(2,4,6-tri-O-benzyl- $\beta$ -p-mannopyranosyl)-(1 $\rightarrow$ 4) 2-acetamido-1,6-anhydro-3-O-benzyl-2-deoxy- $\beta$ -p-glucopyranose (20)——A solution of 19 (250mg, 6.08 × 10<sup>-4</sup> mol) in dry benzene (1.5 ml) was added to a suspension of 18 (43.3 mg, 5.97 × 10<sup>-5</sup> mol), Hg(CN)<sub>2</sub> (230 mg), and Drierite (80 mg) in dry nitromethane (1.5 ml). After being stirred at 50°C for 3 d, the mixture was diluted with CHCl<sub>3</sub>, and filtered, and the filtrate was treated as described in the case of 12 to separate the coupling product as a syrup. The resultant syrup in dry MeOH (4 ml) was de-O-acetylated with a 0.5 N methanolic solution of MeONa (0.2 ml), and the crude deacetylated product was purified by PTLC with CHCl<sub>3</sub>-acetone (1: 2). The band having Rf 0.17 was removed from the plates, extracted with CHCl<sub>3</sub>-MeOH (1: 1), and concentrated to a syrup.

The syrup was re-acetylated with Ac<sub>2</sub>O (1 ml) and pyridine (2 ml) for 24 h at room temperature, then concentrated to dryness. The residue was purified by PTLC with CHCl<sub>3</sub>-acetone (6: 1). From the band having Rf 0.44, 20 (29.7 mg, 47.1%) was isolated as a foamy solid,  $[\alpha]_{1}^{21} - 27.8^{\circ}$  (c = 0.22, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.68 (3H, s, NAc), 2.02, 2.04, 2.07, 2.08 (12H, each s, OAc×4), 6.09 (1H, d,  $J_{NH,2}=9$  Hz, NH), 7.31 (20H, s, aromatic protons). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 101.0 ( ${}^{1}J_{C-1'-H-1'}=156.3$  Hz, C-1'), 100.7 ( ${}^{1}J_{C-1-H-1}=170.9$  Hz, C-1), 99.6 ( ${}^{1}J_{C-1''-H-1''}=175.8$  Hz, C-1"). IR  $\nu_{max}^{kBr}$  cm<sup>-1</sup>: 3410 (NH), 1750 (OAc), 1674 (amide I), 1511 (amide II). TLC: Rf 0.44 (solvent A). Anal. Calcd for C<sub>56</sub>H<sub>65</sub>NO<sub>19</sub>: C, 63.69; H, 6.20; N, 1.33. Found: C, 63.50; H, 6.35; N, 1.35.

O-(2,3,4,6-Tetra-O-acetyl- $\alpha$ -D-mannopyranosyl)- $(1\rightarrow 3)$ -O-(2,4,6-tri-O-acetyl- $\beta$ -D-mannopyranosyl)- $(1\rightarrow 4)$ -2-acetamido-3-O-acetyl-1,6-anhydro-2-deoxy- $\beta$ -D-glucopyranose (21)—A solution of 20 (42.8 mg,  $4.05\times 10^{-5}$  mol) in glacial AcOH (3 ml) was mixed with 10% Pd-on-charcoal (40 mg), and hydrogenated at room temperature under atmospheric pressure for 48 h. The catalyst was filtered off, and the filtrate was concentrated to give a glassy mass, which was acetylated with Ac<sub>2</sub>O (1 ml) and pyridine (2 ml) for 24 h at room

temperature. The mixture was concentrated to dryness, and the residue was purified by PTLC with CHCl<sub>3</sub>-acetone (3: 1). From the band having Rf 0.25, 21 was isolated as a foamy solid. For analysis, the product was precipitated from CHCl<sub>3</sub>-hexane as a white powder (23.4 mg, 66.9%),  $[\alpha]_D^{21} - 10^\circ$  (c = 0.1, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.99, 2.02, 2.06, 2.10, 2.14, 2.27 (27H, each s, OAc × 8, NAc), 6.14 (1H, d,  $J_{NH,2} = 9$  Hz, NH). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 100.4 ( ${}^1J_{C-1-H-1} = 175.8$  Hz, C-1), 99.2 ( ${}^1J_{C-1''-H-1''} = 175.8$  Hz, C-1"), 96.5 ( ${}^1J_{C-1'-H-1'} = 158.7$  Hz, C-1'). IR  $\nu_{max}^{KBT}$  cm<sup>-1</sup>: 3410 (NH), 1746 (OAc), 1676 (amide I), 1516 (amide II). TLC: Rf 0.25 (solvent B). Anal. Calcd for  $C_{36}H_{49}NO_{23} \cdot 1/2H_2O$ : C, 49.54; H, 5.77; N, 1.60. Found: C, 49.35; H, 5.65; N, 1.69.

O-(2, 3, 4, 6-Tetra-O-acetyl- $\alpha$ -p-mannopyranosyl)-(1 $\rightarrow$ 3)-O-(2, 4, 6-tri-O-acetyl- $\beta$ -p-mannopyranosyl)-(1 $\rightarrow$ 4)-2-acetamido-1,3,6-tri-O-acetyl-2-deoxy-p-glucopyranose (22)—A solution of 21 (26 mg, 2.98 × 10<sup>-5</sup> mol) in an ice-cold acetolysis reagent [boron trifluoride etherate-Ac<sub>2</sub>O (1: 25, v/v) 0.65 ml] was stirred for 2 h at 0°C. A piece of ice was added, and the mixture was stirred overnight at room temperature to decompose excess acetolysis reagent. The solution was then diluted with CHCl<sub>3</sub>, and neutralized with solid NaHCO<sub>3</sub>. The separated organic layer was washed with H<sub>2</sub>O, dried (MgSO<sub>4</sub>), and concentrated to dryness. The resultant syrup was treated with CHCl<sub>3</sub>-hexane to yield 22 (20.2 mg, 70.1%) as a white powder, [ $\alpha$ ]<sup>22</sup><sub>p</sub> +20° (c=0.14, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.93, 1.98, 2.05, 2.09, 2.11, 2.19 (33H, each s, OAc × 10, NAc), 6.12 (d, J<sub>1.2</sub>=4 Hz, H-1). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 97.9 (C-1,  $\beta$ ), 90.6 (C-1,  $\alpha$ ). IR  $\nu$ <sup>KBT</sup><sub>max</sub> cm<sup>-1</sup>: 3380 (NH), 1746 (OAc), 1684 (amide I), 1526 (amide II). TLC: Rf 0.29 and 0.22 (anomeric mixture, solvent B). Anal. Calcd for C<sub>40</sub>H<sub>55</sub>NO<sub>26</sub>. 1/2H<sub>2</sub>O: C, 49.28; H, 5.79; N, 1.44. Found: C, 49.25; H, 5.82; N, 1.44. lit.<sup>2)</sup> amorphous (monohydrate), mp 106—109°C, [ $\alpha$ ]<sup>20</sup><sub>p</sub> 0° (c=2.3, 5: 1 CHCl<sub>3</sub>-MeOH), <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 6.1 (d, 8 Hz, H-1 $\beta$ ).

2-Acetamido-1,6-anhydro-3-O-benzyl-2-deoxy-4-O-(2-O-acetyl-3,4,6-tri-O-benzyl-β-D-glucopyranosyl)-β-D-glucopyranose (24)——A solution of 23<sup>15</sup> (2.69 g, 4.84 mmol) in dry benzene (8 ml) was added to a suspension of 10° (514.9 mg, 1.7 mmol), Hg(CN)<sub>2</sub> (2 g), and Drierite (1 g) in nitromethane (8 ml). After being stirred at 55°C for 24 h, the mixture was treated as described for 12, yielding crude 24, which was column-chromatographed with hexane-ether (1: 4). Removal of the solvent from the fractions having Rf 0.62 (solvent A) provided pure 24 (511.2 mg, 39.2%),  $[\alpha]_D^{2i}$  –59.7° (c=0.58, CHCl<sub>3</sub>), as a foamy solid. <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 2.01, 2.05 (6H, each s, OAc, NAc), 5.23 (1H, s, H-1), 6.28 (1H, d,  $J_{NH,2}$ =10 Hz, NH), 7.25 (20H, s, aromatic protons). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 101.2 ( $^1J_{C-1-H-1}$ =174.6 Hz, C-1), 98.9 ( $^1J_{C-1'-H-1'}$ =158.7 Hz, C-1'). IR  $\nu_{max}^{RBT}$  cm<sup>-1</sup>: 3360 (NH), 1729 (OAc), 1667 (amide I), 1505 (amide II). TLC: Rf 0.62 (solvent A). Anal. Calcd for C<sub>44</sub>H<sub>49</sub>NO<sub>11</sub>: C, 68.82; H, 6.43; N, 1.82. Found: C, 68.61; H, 6.48; N, 1.82.

2-Acetamido-1,6-anhydro-3-*O*-benzyl-2-deoxy-4-*O*-(3,4,6-tri-*O*-benzyl- $\beta$ -D-glucopyranosyl)- $\beta$ -D-glucopyranose (25)—Deacetylation of 24 (148.7 mg, 1.94 × 10<sup>-4</sup> mol) in dry MeOH (4 ml) with a 0.5 N methanolic solution of MeONa (0.2 ml) was carried out as for 13 to yield crude 25, which was purified by PTLC with CHCl<sub>3</sub>-acetone (3: 1). The band having *Rf* 0.44 was removed from the plates and extracted with CHCl<sub>3</sub>-MeOH (3: 1). Removal of the solvent provided 25 as a foamy solid (133.5 mg, 94.8%), [α]<sub>D</sub><sup>23</sup> -41.9° (c = 0.27, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.89 (3H, s, NAc), 5.39 (1H, s, H-1), 6.36 (1H, d,  $J_{NH,2}$ =8 Hz, NH), 7.27 (20H, s, aromatic protons). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 102.1 ( $^{1}J_{C-1'-H-1'}$ =155.0 Hz, C-1'), 100.5 ( $^{1}J_{C-1-H-1}$ =172.3 Hz, C-1). IR  $\nu_{max}^{KBF}$  cm<sup>-1</sup>: 3300 (OH, NH), 1642 (amide I), 1505 (amide II). TLC: *Rf* 0.44 (solvent B). *Anal.* Calcd for C<sub>42</sub>H<sub>47</sub>NO<sub>10</sub>: C, 69.50; H, 6.53; N, 1.93. Found: C, 69.42; H, 6.49; N, 2.12.

2-Acetamido-1,6-anhydro-3-O-benzyl-2-deoxy-4-O-(3,4,6-tri-O-benzyl-β-D-mannopyranosyl)-β-D-glucopyranose (26)—Oxidation of 25 (378 mg,  $5.21 \times 10^{-4}$  mol) in DMSO-Ac<sub>2</sub>O (2: 1, v/v, 13.5 ml) to the corresponding ulose and subsequent reduction of the ulose to crude 26 with NaBH<sub>4</sub> (420 mg) was carried out as described for 15 (from 13). Pure 26 was obtained by column chromatography of the crude product with CHCl<sub>3</sub>-acetone (20: 1) as a foamy solid (227.3 mg, 58.7%), [α]<sup>18</sup><sub>D</sub> -54.2° (c=0.21, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.92 (3H, s, NAc), 2.82 (1H, br s, OH), 5.40 (1H, s, H-1), 6.40 (1H, d,  $J_{NH,2}=9$  Hz, NH), 7.25, 7.32 (20H, each s, aromatic protons). <sup>13</sup>C-NMR (CDCl<sub>3</sub>):  $100.6 (^1J_{C-1-H-1}=175.8$  Hz, C-1), 99.3 ( $^1J_{C-1'-H-1'}=156.3$  Hz, C-1'). IR  $\nu_{max}^{RBT}$  cm<sup>-1</sup>: 3380 (OH, NH), 1658 (amide I), 1530 (amide II). TLC: Rf 0.31 (solvent B). Anal. Calcd for  $C_{42}H_{47}NO_{10}\cdot H_2O$ : C, 67.82; H, 6.64; N, 1.88. Found: C, 67.84; H, 6.41; N, 2.16.

2-Acetamido-1,6-anhydro-3-O-benzyl-2-deoxy-4-O-(2-O-acetyl-3,4,6-tri-O-benzyl-β-D-mannopyranosyl)-β-D-glucopyranose (27)—Acetylation of 26 (30.5 mg,  $4.1 \times 10^{-6}$  mol) with Ac<sub>2</sub>O (0.5 ml) and pyridine (1 ml) was carried out as described for 14. After purification of the crude acetate by PTLC with CHCl<sub>3</sub>-acetone (6: 1, pure 27 was isolated as a syrup (27.2 mg, 85.4%), [α]<sub>D</sub><sup>20</sup> -56.5° (c=0.37, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 2.02, 2.15 (6H, each s, OAc, NAc), 5.36 (1H, s, H-1), 6.10 (1H, d,  $J_{\rm NH,2}$ =10 Hz, NH), 7.27, 7.29 (20H, each s, aromatic protons). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 101.1 ( ${}^{1}J_{\rm C-1-H-1}$ =173.3 Hz, C-1), 95.2 ( ${}^{1}J_{\rm C-1'-H-1'}$ =153.8 Hz, C-1'). IR  $\nu_{\rm max}^{\rm RBr}$  cm<sup>-1</sup>: 3390 (NH), 1738 (OAc), 1672 (amide I), 1509 (amide II). TLC: Rf 0.43 (solvent A). Anal. Calcd for C<sub>44</sub>H<sub>49</sub>NO<sub>11</sub>·1/2H<sub>2</sub>O: C, 68.03; H, 6.49; N, 1.80. Found: C, 68.07; H, 6.77; N, 1.91.

O-(2,3,4,6-Tetra-O-acetyl- $\alpha$ -D-mannopyranosyl)-(1 $\rightarrow$ 2)-O-(3,4,6-tri-O-benzyl- $\beta$ -D-mannopyranosyl)-(1 $\rightarrow$ 4)-2-acetamido-1,6-anhydro-3-O-benzyl-2-deoxy- $\beta$ -D-glucopyranose (28)—Glycosidation of 26 (113.7 mg, 1.53 × 10<sup>-4</sup> mol) with 19 (668.7 mg, 1.63 mmol) in benzene (3 ml) in the presence of a suspension of Hg(CN)<sub>2</sub> (600 mg) and Drierite (200 mg) in dry nitromethane (3 ml) was carried out as described for 20. Because the crude trisaccharide was contaminated with a side product having the same Rf value on TLC, it was de-O-acetylated with a 0.5 N methanolic solution of MeONa (0.5 ml) in dry MeOH (10 ml) as described for 13. From the band having Rf 0.17 on PTLC with CHCl<sub>3</sub>-acetone (1: 2), the pure de-O-acetylated trisac-

charide was isolated.

The product was then re-acetylated with Ac<sub>2</sub>O (1 ml) and pyridine (2 ml), and the resultant acetate was purified by PTLC with CHCl<sub>3</sub>-acetone (6: 1). From the band having Rf 0.36, pure 28 was isolated as a foamy solid (65.8 mg, 40.7%), [ $\alpha$ ]<sup>19</sup>  $-29^{\circ}$  (c=0.2, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.99, 2.07, 2.09 (15H, all s, OAc×4, NAc), 6.16 (1H, d,  $J_{\rm NH,2}=10$  Hz, NH), 7.25, 7.29 (20H, each s, aromatic protons). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 100.9 ( $^{1}J_{\rm C_{1-C-H}}=153.8$  Hz, C-1'), 100.5 ( $^{1}J_{\rm C_{1-H-1}}=175.8$  Hz, C-1), 98.1 ( $^{1}J_{\rm C_{1-''-H-1''}}=173.3$  Hz, C-1"). IR  $\nu_{\rm max}^{\rm kBF}$  cm<sup>-1</sup>: 3410 (NH), 1750 (OAc), 1673 (amide I), 1511 (amide II). TLC: Rf 0.36 (solvent A). Anal. Calcd for C<sub>56</sub>H<sub>65</sub>NO<sub>19</sub>: C, 63.69; H, 6.20; N, 1.33. Found: C, 63.73; H, 6.23; N, 1.60.

O-(2,3,4,6-Tetra-O-acetyl- $\alpha$ -D-mannopyranosyl)-(1 $\rightarrow$ 2)-O-(3,4,6-tri-O-acetyl- $\beta$ -D-mannopyranosyl)-(1 $\rightarrow$ 4)-2-acetamido-3-O-acetyl-1,6-anhydro-2-deoxy- $\beta$ -D-glucopyranose (29)—Catalytic debenzylation of 28 (57.4 mg, 5.43 × 10<sup>-5</sup> mol) in glacial AcOH (3 ml) with 10% Pd-on-charcoal (50 mg) and subsequent acetylation of the resultant debenzylated product with Ac<sub>2</sub>O (1 ml) and pyridine (2 ml) were carried out as described for 14. The crude acetate was column-chromatographed with CHCl<sub>3</sub>-ether-MeOH (30: 5: 1). From the fractions having Rf 0.21 with CHCl<sub>3</sub>-acetone (3: 1), 29 was isolated as a syrup. For analysis, the syrup was treated with CH<sub>2</sub>Cl<sub>2</sub>-hexane to yield a white powder (35.3 mg, 75.3%), [ $\alpha$ ]<sup>21</sup><sub>D</sub> -39.8° (c=0.22, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 2.02, 2.05, 2.09, 2.16 (27H, each s, OAc×8, NAc), 6.12 (1H, d,  $J_{NH,2}$ =10 Hz, NH). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 100.6 ( ${}^{1}J_{C-1'-H-1'}$ =156.3 Hz, C-1'), 100.2 ( ${}^{1}J_{C-1-H-1}$ =178.2 Hz, C-1), 97.6 ( ${}^{1}J_{C-1''-H-1''}$ =170.9 Hz, C-1"). IR  $r_{max}^{max}$  cm<sup>-1</sup>: 3420 (NH), 1745 (OAc), 1679 (amide I), 1516 (amide II). TLC: Rf 0.21 (solvent B). Anal. Calcd for C<sub>36</sub>H<sub>49</sub>NO<sub>23</sub>: C, 50.06; H, 5.72; N, 1.62. Found: C, 49.89; H, 5.83; N, 1.58.

O-(2,3,4,6-Tetra-O-acetyl- $\alpha$ -D-mannopyranosyl)-(1 $\rightarrow$ 2)-O-(3,4,6-tri-O-acetyl- $\beta$ -D-mannopyranosyl)-(1 $\rightarrow$ 4)-2-acetamido-1,3,6-tri-O-acetyl-2-deoxy- $\alpha$ -D-glucopyranose (30)—Cleavage of the 1,6-anhydro-ring of 29 (28.3 mg,  $3.28 \times 10^{-5}$  mol) with acetolysis mixture (0.7 ml) was carried out as described for 22. The resultant syrup was treated with CH<sub>2</sub>Cl<sub>2</sub>-hexane to give 30 as a white powder (24.5 mg, 77.4%), [ $\alpha$ ]<sup>19</sup> +32.7° (c=0.11, CHCl<sub>3</sub>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.95, 2.02, 2.09, 2.11, 2.15, 2.19 (33H, each s, OAc × 10, NAc), 5.66 (1H, d,  $J_{NH,2}$ =9 Hz, NH), 6.11 (1H, d,  $J_{1,2}$ =4 Hz, H-1). <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 99.3 ( $^1J_{C-1'-H-1'}$ =161.1 Hz, C-1'), 98.9 ( $^1J_{C-1''-H-1''}$ =170.9 Hz, C-1"), 90.5 ( $^1J_{C-1H-1}$ =178.2 Hz, C-1). IR  $\nu_{\max}^{KB}$  cm<sup>-1</sup>: 3360 (NH), 1744 (OAc), 1682 (amide I), 1528 (amide II). TLC: Rf 0.20 (solvent B). Anal. Calcd for C<sub>40</sub>H<sub>55</sub>NO<sub>26</sub>: C, 49.74; H, 5.74; N, 1.45. Found: C, 49.88; H, 6.24; N, 1.43.

Acknowledgement We thank Miss S. Kato for the <sup>1</sup>H- and <sup>13</sup>C-NMR spectral measurements, and Misses S. Iwauchi and T. Naito for the microanalyses.

## References and Notes

- 1) N.E. Nordén, A. Lundblad, S. Svenson, P.-A. Öckerman, and S. Autio, J. Biol. Chem., 248, 6210 (1973).
- 2) C.D. Warren, R.W. Jeanloz, and G. Strecker, Carbohydr. Res., 92, 85 (1981).
- 3) N.M.K. Ng Ying Kin and L.S. Wolfe, Biochem. Biophys. Res. Comm., 66, 123 (1975).
- 4) L.S. Wolfe, R.G. Senior, and N.M.K. Ng Ying Kin, J. Biol. Chem., 249, 1828 (1974).
- 5) G.C. Tsay, G. Dawon, and S.-S. J. Sung, J. Biol. Chem., 251, 5852 (1976).
- 6) P.A.J. Gorin and A.S. Perlin, Can. J. Chem., 39, 2474 (1961); P.J. Garegg, T. Iversen, and T. Norberg, Carbohydr. Res., 73, 313 (1979); H. Paulsen and O. Lockhoff, Tetrahedron Lett., 1978, 4027; G. Wulff and J. Wichelhaus, Chem. Ber., 112, 2847 (1979).
- 7) W.M. Corbett and J.E. McKay, J. Chem. Soc., 1961, 2930.
- 8) N.K. Kochetkov, A.J. Khorin, and A.F. Bochkov, Tetrahedron, 23, 693 (1967).
- 9) Y. Itoh and S. Tejima, Chem. Pharm. Bull., 30, 3383 (1982).
- 10) M.A.E. Shaban and R.W. Jeanloz, Carbohydr. Res., 52, 115 (1976).
- 11) A.S. Perlin, Pure & Appl. Chem., 50, 1401 (1978).
- 12) C.D. Warren and R.W. Jeanloz, Carbohydr. Res., 53, 67 (1977).
- 13) J. Cunningham, R. Gigg, and C.D. Warren, Tetrahedron Lett., 1964, 1191.
- 14) T. Ogawa and M. Matsui, Carbohydr. Res., 62, C1 (1978).
- 15) D.R. Burfield and R.H. Smithers, J. Org. Chem., 43, 3966 (1978).
- T. Takamura, T. Chiba, and S. Tejima, Chem. Pharm. Bull., 29, 1027, 1076, 2270 (1981); S. Oguri and S. Tejima, ibid., 28, 3184, 3196 (1980), 29, 1629 (1981); Y. Itoh and S. Tejima, ibid., 31, 727 (1983).
- 17) E. Stahl and U. Kaltenbach, J. Chromatogr., 5, 351 (1961).