## Communications to the Editor

Chem. Pharm. Bull. 32(10)4229—4232(1984)

CHARACTERIZATION OF A MINOR COMPOUND, WHICH ACCOMPANIES THE USUAL  $22\alpha(\underline{R})-\underline{0},25\beta(\underline{S})$ -SPIROSTANOL GLYCOSIDE, AS A NOVEL TYPE OF  $22\beta(\underline{S})-\underline{0},25\alpha(\underline{S})$  ANALOG

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The structure of one of the minor compounds, which coexists only with a  $25\beta(\underline{S})$ -spirostanol glycoside (a steroid saponin) such as I, II, XI and XV, was determined to be the corresponding  $22\beta(\underline{S})-\underline{0}$ , $25\alpha(\underline{S})$  analog Ia, IIa, XIa and XVa, respectively.

Since it was believed that all the natural spirostanol glycosides have the  $22\alpha$   $(\underline{R})$ - $\underline{0}$  configuration, these compounds are worthy of note as the first glycosides of  $22\beta(\underline{S})$ -0-spirostanol so far isolated.

KEYWORDS —— steroid saponin;  $25\beta(\underline{S})$ -spirostanol glycoside; rhodeasapogenin; sarsasapogenin; neotokorogenin;  $22\beta(\underline{S})$ - $\underline{0}$ , $25\alpha(\underline{S})$ -spirostanol glycoside; IR; NMR; molecular rotation difference

Two glycosides  $R_3$ ,  $1-0-\alpha$ -L-rhamnopyranosyl-(1-2)- $\beta$ -D-xylopyranoside (rha-xyl) (I), and  $R_8$ ,  $3-0-\beta$ -D-glucopyranoside (glc) (II), of  $5\beta$ , $25\beta$ ( $\underline{S}$ )-spirostane- $1\beta$ , $3\beta$ -diol (rhodeasapogenin) (III), which had been isolated from the underground parts of <u>Rhodea japonica</u> and regarded to be homogeneous, were found by high performance liquid chromatography (HPLC) to consist of four glycosides Ia $^{\sim}$ d and IIa $^{\sim}$ d. They were successfully separated in preparative scale, and the major component Id and IId were identified as pure I and II, while the aglycones of Ic and IIc were identified as  $25\alpha$ (R)-epimer (isorhodeasapogenin) (IV) and those of Ib and IIb as 25(27)-dehydro derivative (convallamarogenin) (V) of III. Ib $^{\sim}$ d and IIb $^{\sim}$ d have the same sugar moieties. 2)

This paper deals with characterization of the fourth minor compounds Ia and IIa, and with examination of timosaponin A-III $^3$ ) and neotokoronin, both of which have been known also as  $5\beta,25\beta(\underline{S})$ -spirostanol glycosides.

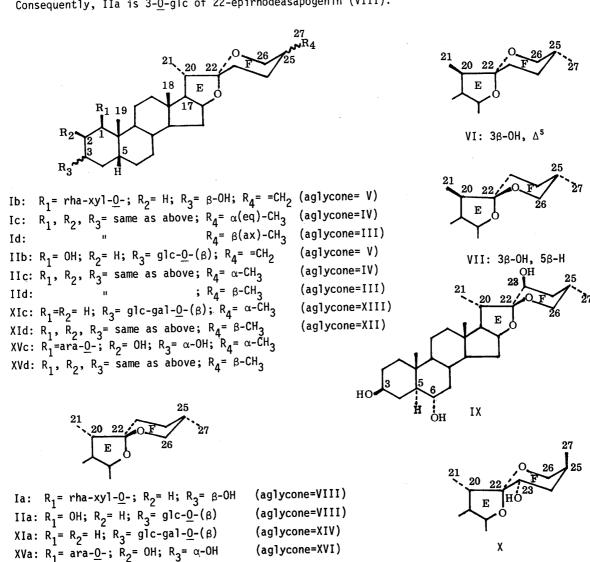
Ia, prisms  $^{5)}$  (Me $_2$ CO-MeOH-water), mp 275-277°C, $^{6)}$  [ $\alpha$ ] $_0$  -51.4°, $^{7)}$  showed the [M+H] $^+$  ion at  $\underline{m/z}$  711 and [M] $^+$  at 710 in the field desorption(FD) - MS. $^{8)}$  Taking the elemental analytical data into account, Ia has the same molecular formula,  $C_{38}H_{62}O_{12}\cdot H_2O$ , as Ic,d. In the  $^{13}C$ -NMR $^{9)}$  spectrum of Ia, only the signals assignable to C-20  $\sim$  C-27 and C-21  $\sim$  C-24 as well as C-26 of the aglycone were different from the respective signals of Id and Ic (Table I). Ia showed an IR spectrum  $^{11)}$  somewhat unlike the spectra of Ic and Id in the characteristic absorptions of the spiroketal side chain,  $^{12}$  but the relative intensity, 916 < 985 cm $^{-1}$ , was more like that, 919 < 900, in Ic rather than the 919 > 894 in Id (Table II), suggesting a probable presence of the  $25\alpha$ (eq)-CH $_3$  group. When Ia was hydrolyzed with 2N-H $_2$ SO $_4$  in 50% EtOH with refluxing for 1.5 h, rhamnose and xylose together with III were yielded. But on treatment with 1N-HC1 in 50% EtOH at 0°C for 2.5 h, Ia was transformed nealy quantitatively into Id. Thus, it was assumed that the aglycone of Ia might be different in stereo-structure from those of Ic and Id with respect to the E  $\sim$  F ring moiety. The possibility of a  $\beta$ -configuration of the 2O-CH $_3$  group could be excluded by comparing the  $^1$ H-NMR $^{9)}$  (Table III)  $^{13}$  and  $^1$ C-NMR data (Table I) of Ia with those of Ic, Id, cyclopseudo-diosgenin (VI) and -sarsasapogenin (VII).

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Therefore, the aglycone of Ia is regarded as the  $22\beta-\underline{0}$ , $25\alpha(eq)-CH_3$  isomer of III. The configuration of the  $25\alpha(eq)$ -CH $_3$  group, suggested by the IR spectrum, was endorsed by the similarity of the signals of 26-H $_2$  and 27-H $_3$  in the  $^1$ H-NMR spectrum and C-25 and C-27 in the  $^1$  $^3$ C-NMR spectrum with those of Ic (Tables I and III), and the unusual  $22\beta-\underline{0}$ -configuration was corroborated by the large molecular rotation difference (+379°) between Ia and Id.  $^{17)}$  Ia is now considered to be  $1-\underline{0}$ -rha-xyl of  $5\beta$ ,  $22\beta(\underline{S})-\underline{0}$ ,  $25\alpha(\underline{S})$ -spirostane-1 $\beta$ ,  $3\beta$ -diol (VIII), conventionally named 22-epirhodeasapogenin.

IIa, needles (dil.MeOH), mp 257-259°C, [ $\alpha$ ]<sub>D</sub> -31,2°, has the same molecular formula,  $^{\rm C}_{33}{}^{\rm H}_{54}{}^{\rm O}_{\rm O}$ (FD-MS:  $[M+H]^+$ , m/z 595; elemental analytical data), as IIc,d. The <sup>13</sup>C-NMR and IR spectral data Tables I and II) suggested, as in the case of Ia, that IIa has the  $25\alpha(eq)$ -CH $_3$  group, but that the stereo-structure of the E  $\sim$  F-ring moiety is different from either IIc or IId. In the  $^1\text{H-NMR}$  spectrum of IIa in comparison with that of Ia (Table III), the signal of 19-H $_3$  was shifted to an upper field by 0.06 ppm, but others, 18-, 21- and 27-H $_3$  as well as 26-H $_2$ , showed the  $\delta$ -values and the coupling patterns identical to those of Ia. This implys that IIa has the same aglycone as Ia, but that the sugar moiety is combined with the hydroxyl group at C-3 and not C-1. The fact that acid treatment of IIa at  $0^{\circ}\text{C}$  in the same way as for Ia afforded IId, and the molecular rotation difference (+198°) between IIa and IId supported the  $22\beta-\underline{0}$  configuration of the aglycone of IIa.

Consequently, IIa is  $3-\underline{0}$ -glc of 22-epirhodeasapogenin (VIII).



The stereochemistry of the 17-side chain of the spirostane nucleus was extensively investigated,  $^{10,12\sim18)}$  but the studies were based on the fact that only two stereoisomers due to an asymmetric carbon atom in the E  $\sim$  F-ring moiety had so far been found in nature. It has been believed that both the configurations at C-20 and -22 are R  $(20\alpha$ -CH<sub>3</sub> and  $22\alpha$ -O), and that the two natural isomers have either the  $25(R)(\alpha(eq)$ -CH<sub>3</sub>) (more stable, iso-type) or the  $25(S)(\beta(ax)$ -CH<sub>3</sub>) (normal- or neotype) configuration.

Lately, Chakravarty et al<sup>19</sup>) obtained, by Smith degradation of the crude glycoside of <u>Solanum hispidum</u>, a novel type of spirostanol, hispigenin (IX), which has an unusual  $22\beta-\underline{0}$ - configuration, along with the usual  $(22\alpha-\underline{0})$  isomer,  $5\alpha,25\beta(\underline{S})$ -spirostane- $3\beta,6\alpha,23\alpha$ -triol (paniculogenin) (X).

Ia and IIa are noteworthy as the first naturally occurring  $22\beta(\underline{S})-\underline{0}$ -spirostanol glycoside ever isolated.

In contrast to the findings that Ia, IIa and IX accompany their  $22\alpha(\underline{R})-\underline{0},25\beta(\underline{S})$  analogs, the glycosides of  $22\alpha(\underline{R})-\underline{0},25\alpha(\underline{R})$ -spirostanols such as diosgenin, gitogenin and digitogenin are, as reported previously, coexistent only with their  $25(\underline{S})$ -epimers. In order to verify the generality of co-occurrence of the glycosides of  $22\alpha(\underline{R})-\underline{0},25\beta(\underline{S})$ -spirostanols with those of the corresponding 22-epimers, two additional  $25\beta(\underline{S})$ -spirostanol glycosides which had been regarded as homogeneous were examined.

Timosaponin A-III (XI),  $^3$ ) 3- $\underline{0}$ - $\beta$ -D-glucopyranosyl-(1-2)- $\beta$ -D-galactopyranoside (glc-gal) of 5 $\beta$ , 25 $\beta$ ( $\underline{S}$ )-spirostan-3 $\beta$ -ol (sarsasapogenin) (XII), prisms (70% MeOH), mp > 300°C,  $[\alpha]_D$  -41.6°, was shown by HPLC<sup>2</sup>) to consist of three compounds. They were separated to give XIa,c,d. XId (major), needles (MeOH-CHCl<sub>3</sub>), mp > 300°C,  $[\alpha]_D$  -47.5°, and XIc, needles (MeOH-CHCl<sub>3</sub>), mp > 300°C,  $[\alpha]_D$  -35.3°, were respectively identified as pure XI and its 25 $\alpha$ ( $\underline{R}$ )-epimer (3- $\underline{0}$ -glc-gal of smilagenin (XIII)).

XIa, prisms (MeOH-CHCl $_3$ ), mp 295-300°C, [ $\alpha$ ] $_D$  -0.1°, has the same molecular formula,  $C_{39}H_{64}O_{13}$  (FD-MS: [M+H] $^+$ , m/z 741; [M+Na] $^+$ , 763. elemental analytical data), and its IR (Table II), <sup>13</sup>C- and <sup>1</sup>H-NMR data (Tables I and III) indicated that XIa is presumably the 22-epimer of XId. The large molecular rotation difference (+338°) between XIa and XId substantiated the above presumption.

Thus, XIa is defined as  $3-\underline{0}$ -glc-gal of  $5\beta$ , $22\beta(\underline{S})-\underline{0}$ , $25\alpha(\underline{S})$ -spirostan-3 $\beta$ -ol (22-episarsasapogenin) (XIV).

Table I. 13C-NMR Data 9,10)

					J 11/11( 1			
	C-20	C-21	C-22	C-23	C-24	C-25	C-26	C-27
Ia	42.1	17.0	110.6	28.3*	28.1*	30.7	69.6	17.3
Ιc	42.0	15.0	109.2	31.7	29.2	30.6	66.9	17.3
Id	42.5	14.9	109.7	26.3	26.3	27.4	65.1	16.3
IIa	42.1	17.0	110.3	28.2*	28.1*	30.6	69.4	17.3
ΙΙc	41.9	15.0	109.0	31,8	29.2	30.5	66.7	17.3
ΙΙd	42.4	14.8	109.5	26.1	26.1	27.4	64.9	16.2
<b>VI*</b> *	46.4	11.4	108.4	31.8	28.5	37.3	68.1	17.2
AII*	*45.9	9.9	106.8	34.1	29.1	29.9	67.5	17.1
ΧIa	42.2	17.0	110.5	28.2	28.2	30.9	69.6	17.4
XIc	42.0	15.1	109.2	31.8	29.3	30.6	66.9	17.4
ΧId	42.5	14.9	109.7	26.4	26.4	27.5	65.1	16.3
XVa	42.1	17.0	110.5	28.3*	28.4*	30.7	69.5	17.3
XVc	42.0	15.1	109.2	31.9	29.2	30.6	66.9	17.3
XVd	42.5	14.9	109.7	26.4	26.2	27.6	65.2	16.3

<sup>\*</sup> may be reversed in horizontal column. \*\* taken in CDCl<sub>3</sub>.

Table II. IR Data 11,12)

Ia	983	946	916 < 895	872	(cm <sup>-1</sup> )
Ιc	981	945	919 < 900	863	,
Ιd	986	945	919 > 894	850	
ΙΙa	987	946	917 < 895	832	
ΙΙc	980	945	918 < 900	864	
IId	988	945	920 > 898	850	
ΧIa	994	925	910 < 896	850	
XIc	981	949	919 < 899	861	
ΧId	986	948	919 > 898	850	

A glycoside, mp 277-278°C, obtained from the mother liquor of recrystallization (MeOH) of neotokoronin,  $5\beta$ , $25\beta(\underline{S})$ -spirostane- $1\beta$ , $2\beta$ , $3\alpha$ -triol  $1-\underline{0}-\alpha$ -L-arabinopyranoside (ara) (XV), $^4$ ) was also a mixture of three compounds XVa,c,d as shown by HPLC, $^2$ ) and they were isolated in pure state. XVd, needles (dil.MeOH), mp 284-286°C,  $[\alpha]_D$  -17.2°, and XVc, needles (dil.MeOH), mp 275-277°C,  $[\alpha]_D$  -13.0°,

<sup>1</sup>H-NMR Data 9,13) Table III.

	19-H <sub>3</sub>	18-H <sub>3</sub>	21-H <sub>3</sub>	27-Н <sub>3</sub>	26-H <sub>2</sub>
Ia	1.30 s	0.97 s	1.00 d (J=6 Hz)	0.69 m	3.67 m
Ic	1.33 s	0.85 s	1.13 d (J=6 Hz)	0.69 d (J=6 Hz)	3.57 m
Id	1.32 s	0.83 s	1.13 d (J=6 Hz)	1.07 d (J=6 Hz)	3.35 d (J=11
۷I**	1.01 s	0.95 s	1.13 d (J=8 Hz)	0.77 d (J=6 Hz)	3.45 m —
<b>**11V</b>	0.96 s	1.01 s	1.09 d $(\bar{J}=7 \text{ Hz})$	0.76 d $(\bar{J}=6 \text{ Hz})$	3.47 m
IIa	1.24 s	0.97 s	1.01 d $(\bar{J}=7 \text{ Hz})$	0.69 m	3.68 m
XIa	0.96 s	0.96 s	1.02 d $(\bar{J}=7 \text{ Hz})$	0.70 m	3.70 m

\*  $26\beta(eq)-H$ . \*\* taken in CDC1<sub>3</sub>.

were respectively identified as pure XV and tokoronin  $(25\alpha(\underline{R})).^{20}$  XVa, prisms (dil.MeOH), mp 287- $^{2}90^{\circ}\text{C}$ ,  $[\alpha]_{D}^{}$  +19.4°, has the same molecular formula,  $^{\text{C}}_{32}\text{H}_{52}^{}0_{9}$ , as XVc,d. In the same manner as for Ia, IIa and XIa ( $^{13}$ C-NMR data (Table I); molecular rotation difference (+214 $^{\circ}$ ) between XVa and XVd), it was determined to be 1-0-ara of 22-epitokorogenin  $(22\beta(\underline{S})-\underline{0},25\alpha(\underline{S}))$  (XVI).

It is very likely that, regardless of the structure of the A  $\sim$  B ring moiety of aglycone, the usual  $22\alpha(\underline{R})$ -0,25 $\beta(\underline{S})$ -spirostanol glycoside is generally accompanied by a minor amount of the corresponding glycosides of the 22-epimer  $(22\beta(\underline{S})-\underline{0},25\alpha(\underline{S}))$ . Absence of the glycoside of 25(27)-dehydro derivative in XI and XV is also noted.

ACKNOWLEDGEMENT This work was supported by a Grant-in-Aid for scientific research (Grant No. 56470119) from the Ministry of Education, Science and Culture of Japan, which is gratefully acknowledged.

## REFERENCES AND NOTES

- Present address: Faculty of Pharmaceutical Sciences, Setsunan University, 45-1, Nagaotoge-cho, Hirakata, Osaka 573, Japan. 1)

- K. Miyahara, K. Kudo and T. Kawasaki, Chem. Pharm. Bull., 31, 348 (1983).

  T. Kawasaki and T. Yamauchi, Chem. Pharm. Bull., 11, 1221 (1963).

  S. Kiyosawa, S. Huto, T. Nohara, K. Miyahara and T. Kawasaki, Abstracts of Papers, The 91st 4) Annual Meeting of Pharmaceutical Society of Japan, Fukuoka, April 1971, p.762.
- All crystals described in this paper were colorless.

  Melting points were determined on a micro melting point apparatus MP-S<sub>3</sub> (Yanagimoto) and are uncorrected. In all cases reported herein, melting was accompanied by decomposition.
- Optical rotations were taken with a JASCO DIP-SL automatic polarimeter (cell= 1 dm) at 18~28°C, 7) in a pyridine solution (c=  $0.2 \sim 1.4$ ).
- FD-MS spectra were recorded on a JEOL JMS-DX-300/JMA 3500.  $^{13}$ C- and  $^{1}$ H-NMR spectra were taken with a JEOL FX-100 (25 and 100 MHz, respectively) in a  $^{13}$ C-  $^{13}$ C- and  $^{1}$ H-NMR spectra were taken with a JEOL FX-100 (25 and 100 MHz, respectively) in a  $^{13}$ C-  $^{13}$ solution unless otherwise specified. Chemical shifts are given in  $\delta$  values (ppm) using TMS
- as an internal standard. cf. K. Tori, S. Seo, Y. Terui, J. Nishikawa and F. Yasuda, Tetrahedron Lett., <u>22</u>, 2405 (1981).
- IR spectra were recorded on a JASCO IR-G Spectrometer in KBr disk.
- 12)
- 13)
- IR spectra were recorded on a JASCO IR-G Spectrometer in KBr disk.

  M. E. Wall, C. R. Eddy, M. L. McClennan and M. E. Klumpp, Anal. Chem., 24, 1337 (1952).

  cf. K. Tori, J. Nishikawa, S. Seo, A. Uomori, F. Yasuda and K. Kushida, Steroids, 39, 73 (1982).

  L. F. Fieser and M. Fieser, "Steroids," Reinhold Pub. Co., New York, 1959, p.825.

  Prepared from diosgenin and sarsasapogenin, respectively, according to the reported methods.

  M. E. Wall, S. Serota and C. R. Eddy, J. Am. Chem. Soc., 77, 1230 (1955); M. E. Wall and H. A. Walens, ibid., 77, 5661 (1955); idem., ibid., 80, 1984 (1958); J. B. Ziegler, W. E. Rosen and A. C. Shabica, ibid., 77, 1223 (1955).

  M. E. Wass, Experientia, 11, 340 (1955).

  L. F. Fieser and M. Fieser, "Steroids," Reinhold Pub. Co., New York, 1959, p.817.

  A. K. Chakravarty, T. K. Dhar and S. C. Pakrashi, Tetrahedron Lett., 40, 3875 (1978).

  K. Miyahara and T. Kawasaki, Chem. Pharm. Bull., 17, 1369 (1969).

(Received August 27, 1984)