

## Communications to the Editor

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NEW QUASSINOID GLYCOSIDES, YADANZIOSIDES A - H, FROM *BRUCEA JAVANICA*

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Eight new antileukemic quassinoid glycosides, yadanziosides A - H (1 - 8) were isolated from seeds of *Brucea javanica* and their structures were determined by chemical transformation and spectral analysis.

KEYWORDS—bitter principle; antileukemic; quassinoid; glycoside; Simaroubaceae; *Brucea javanica*; <sup>1</sup>H-NMR; <sup>13</sup>C-NMR

As a continuation of studies on bitter principles of Simaroubaceous plants, glycosides in *Brucea javanica* (L.) MERR were investigated.<sup>1)</sup> This paper describes the structures of eight new quassinoid glycosides, some of which showed antileukemic activities against the murine P-388 lymphocytic leukemia.

The methanolic extract of defatted seeds of *B. javanica* was partitioned between dichloromethane and water. The organic layer, on evaporation, gave a residue, which was dissolved in hot methanol. After cooling, brusatol,<sup>2)</sup> which precipitated as crystals, was filtered. The filtrate was evaporated *in vacuo* and subjected to separation by silica-gel column chromatography. After elution with benzene-ethyl acetate, elution with ethyl acetate-methanol afforded fractions containing crude glycosides. The combined fractions were separated by silica-gel chromatography (elution: lower layer of chloroform-methanol-water), and then either by partition chromatography on silicic acid (elution: chloroform-ethanol), a Lobar column Lichroprep RP-8 (elution: methanol-water), or Toyopearl HW-40S (elution: methanol) to give eight new glycosides, yadanziosides A (1; 0.01%), B (2; 0.002%), C (3; 0.001%), D (4; 0.001%), E (5; 0.002%), F (6; 0.001%), G (7; 0.01%), and H (8; 0.001%) together with known brusatol,<sup>2)</sup> dehydrobrucein A,<sup>3)</sup> brucein D,<sup>4)</sup> brucein E,<sup>4)</sup> bruceoside A,<sup>5)</sup> bruceoside B,<sup>5)</sup> and yadanziolide A.<sup>1)</sup>

Yadanzioside B (2), mp 189-195°C; IR (KBr) 3425, 1735, and 1635 cm<sup>-1</sup>; UV (EtOH) 255 nm, gave a peak at *m/z* 707 due to [M+Na]<sup>+</sup> in SI-MS, from which the molecular formula, C<sub>32</sub>H<sub>44</sub>O<sub>16</sub>, was deduced, and was shown to be a hexoside from an appearance of a peak at *m/z* 522.2082 corresponding to [M-C<sub>6</sub>H<sub>10</sub>O<sub>5</sub>]<sup>+</sup> in the EI-high resolution mass spectrum. <sup>1</sup>H- and <sup>13</sup>C-NMR spectra suggest that yadanzioside B (2) must be a β-D-glucoside of brucein A (9).<sup>3,6)</sup> Glycosylation shifts (Δδ +18.1 ppm and Δδ +1.9 ppm) of the signals due to C-3 and C-4 respectively were observed in the <sup>13</sup>C-NMR, indicating that D-glucose is attached at C-3 of the aglycone. An anomeric proton was observed at δ 5.46 as a doublet (*J*=7.3 Hz) in the <sup>1</sup>H-NMR

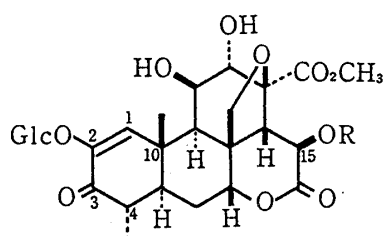
spectrum. On acid hydrolysis with 1M  $\text{H}_2\text{SO}_4$ -methanol, **2** gave brucein A (**9**), and hydrolysis with HCl-methanol afforded methyl D-glucoside, which was identified by GLC after trimethylsilylation. Thus the structure of yadanzioside B (**2**) was determined to be 3-O- $\beta$ -D-glucosyl brucein A.

Yadanzioside A (**1**), mp 200-204°C; IR (KBr) 3400, 1740, 1675, and 1625  $\text{cm}^{-1}$ ; UV (EtOH) 256 nm, was shown by SI-MS to be a glucoside with the same molecular formula as that of **2**. Yadanzioside A (**1**) was hydrolyzed with  $\beta$ -glucosidase at 37°C to give brucein A (**9**). Since a doublet signal due to  $\text{C}_{(4)}\text{-CH}_3$  of the aglycone part of **1** was observed at  $\delta$  1.13 ( $J=6$  Hz), the brucein A moiety isomerizes into a 3-keto-1-ene structure and forms the glycoside linkage through an oxygen atom on C-2. The structure of yadanzioside A (**1**) was formulated as 2-O- $\beta$ -D-glucosyl brucein A.

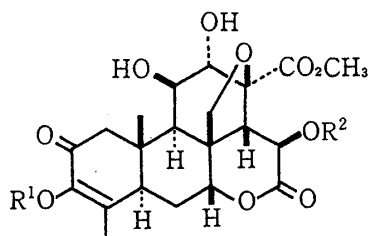
Yadanzioside C (**3**), mp 204-209°C; IR (KBr) 3445, 1740, 1680, and 1640  $\text{cm}^{-1}$ ; UV (EtOH) 221 and 254 nm, gave a peak at  $m/z$  749 due to  $[\text{M}+\text{Na}]^+$  in SI-MS, which corresponds to the molecular formula,  $\text{C}_{34}\text{H}_{46}\text{O}_{17}$ . On acid hydrolysis with 1M  $\text{H}_2\text{SO}_4$ -methanol and with HCl-methanol, **3** afforded brucein C (**10**)<sup>3,6,7</sup> and methyl D-glucoside, respectively. Since a doublet signal due to  $\text{C}_{(4)}\text{-CH}_3$  was observed at  $\delta$  1.19 ( $J=6.7$  Hz) in the  $^1\text{H}$ -NMR of **3**, the structure of yadanzioside C (**3**) was determined to be 2-O- $\beta$ -D-glucosyl brucein C.

Yadanzioside F (**6**), mp 202-207°C; IR (KBr) 3450, 1740, 1680, and 1630  $\text{cm}^{-1}$ ; UV (EtOH) 256 nm, exhibited a peak at  $m/z$  643 due to  $[\text{M}+\text{H}]^+$  in SI-MS, indicating the molecular formula  $\text{C}_{29}\text{H}_{38}\text{O}_{16}$ , and was hydrolyzed with  $\beta$ -glucosidase to give brucein B (**11**).<sup>3,6</sup> Since the  $\text{C}_{(4)}\text{-CH}_3$  of **6** was observed as a doublet signal at  $\delta$  1.15 ( $J=5$  Hz), yadanzioside F (**6**) was formulated as 2-O- $\beta$ -D-glucosyl brucein B.

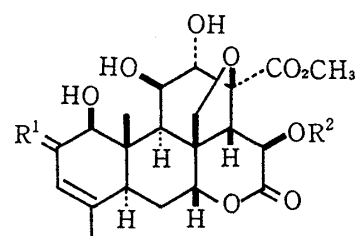
Yadanzioside G (**7**), mp 180-185°C; IR (KBr) 3450, 1740, 1685, and 1645  $\text{cm}^{-1}$ ; UV (EtOH) 222 and 253 nm, afforded a peak at  $m/z$  791 due to  $[\text{M}+\text{Na}]^+$  in SI-MS, corresponding to the molecular formula,  $\text{C}_{36}\text{H}_{48}\text{O}_{18}$ , and was hydrolyzed with  $\beta$ -



- 1:  $\text{R}=\text{CO}$    
 3:  $\text{R}=\text{CO}$    
 6:  $\text{R}=\text{COCH}_3$   
 7:  $\text{R}=\text{CO}$



- 2:  $\text{R}^1=\text{Glc}$ ;  $\text{R}^2=\text{CO}$    
 9:  $\text{R}^1=\text{H}$ ;  $\text{R}^2=\text{CO}$    
 10:  $\text{R}^1=\text{H}$ ;  $\text{R}^2=\text{CO}$    
 11:  $\text{R}^1=\text{H}$ ;  $\text{R}^2=\text{COCH}_3$   
 12:  $\text{R}^1=\text{H}$ ;  $\text{R}^2=\text{CO}$



- 4:  $\text{R}^1=\alpha\text{-OGlc}, \beta\text{-H}$ ;  $\text{R}^2=\text{COCH}_3$   
 5:  $\text{R}^1=\alpha\text{-OGlc}, \beta\text{-H}$ ;  $\text{R}^2=\text{CO}$    
 8:  $\text{R}^1=\alpha\text{-OGlc}, \beta\text{-H}$ ;  $\text{R}^2=\text{CO}$    
 13:  $\text{R}^1=\alpha\text{-OH}, \beta\text{-H}$ ;  $\text{R}^2=\text{COCH}_3$   
 14:  $\text{R}^1=\text{O}$ ;  $\text{R}^2=\text{COCH}_3$   
 15:  $\text{R}^1=\alpha\text{-OH}, \beta\text{-H}$ ;  $\text{R}^2=\text{CO}$    
 16:  $\text{R}^1=\text{O}$ ;  $\text{R}^2=\text{CO}$    
 17:  $\text{R}^1=\alpha\text{-OH}, \beta\text{-H}$ ;  $\text{R}^2=\text{CO}$    
 18:  $\text{R}^1=\text{O}$ ;  $\text{R}^2=\text{CO}$

glucosidase to give bruceantinol (12).<sup>3,7)</sup> In the  $^1\text{H-NMR}$  spectrum of 7, a doublet signal due to  $\text{C}_{(4)}\text{-CH}_3$  was observed at  $\delta$  1.17 ( $J=6$  Hz). The structure of yadanzioside G (7) was formulated as 2-O- $\beta$ -D-glucosyl bruceantinol.

Yadanzioside D (4), mp 207-212°C, exhibited the IR-absorption bands due to hydroxyl ( $3430\text{ cm}^{-1}$ ) and  $\delta$ -lactone ( $1745\text{ cm}^{-1}$ ), but no UV absorption maximum characteristic of a conjugated carbonyl group. In the EI-MS of 4, a peak at  $m/z$  480 due to an aglycone appeared and the  $^1\text{H-NMR}$  spectrum suggests the presence of an acetoxyl group. On hydrolysis with  $\beta$ -glucosidase, 4 yielded the aglycone (13),  $\text{C}_{23}\text{H}_{30}\text{O}_{11}$ ,  $^1\text{H-NMR}$   $\delta$ : 1.52, 1.55, 2.09, 3.78, 4.01, and 6.76; MS  $m/z$ : 482, 464, 446, 440, 422, and 404. The aglycone (13) was oxidized with  $\text{MnO}_2$  to give isobrucein B (14).<sup>8,9)</sup> Since a doublet signal due to  $\text{C}_{(1)}\text{-H}$  appeared at  $\delta$  4.01 ( $J_{1,2}=8$  Hz) in the  $^1\text{H-NMR}$  of 13 and a glycosylation shift ( $\Delta\delta$  +11.0 ppm) due to C-2 was observed in the  $^{13}\text{C-NMR}$  spectrum, the configuration of  $\text{C}_{(2)}\text{-OH}$  of 13 was established to be  $\alpha$ -equatorial and the glycoside linkage occurred through the C-2 oxygen atom. The structure of yadanzioside D (4) was determined to be the  $\beta$ -D-glucoside of the allylic alcohol (13) derived from isobrucein B (14).

Yadanzioside E (5), mp 190-195°C; IR (KBr)  $3450$ ,  $1740$ , and  $1645\text{ cm}^{-1}$ , possesses the molecular formula,  $\text{C}_{32}\text{H}_{44}\text{O}_{16}$ , deduced from a peak at  $m/z$  707 due to  $[\text{M}+\text{Na}]^+$  in SI-MS and was hydrolyzed with  $\beta$ -glucosidase to afford an aglycone (15),  $^1\text{H-NMR}$   $\delta$ : 1.54, 1.66, 2.14, 3.74, 4.02, 5.08, and 6.75. This was treated with  $\text{MnO}_2$  to yield a keto alcohol (16),  $^1\text{H-NMR}$   $\delta$ : 1.19, 1.94, 1.96, 2.19, 3.78, and 6.23; MS  $m/z$ : 520, 502, 484, 437, 420, 402, 388, 374, 345, and 315, the structure of which could be established by  $^1\text{H-NMR}$  and MS spectra. The structure of yadanzioside E (5) was thus determined to be the  $\beta$ -D-glucoside of 15.

Yadanzioside H (8), mp 180-185°C; IR (KBr)  $3450$ ,  $1750$ , and  $1640\text{ cm}^{-1}$ , has the molecular formula,  $\text{C}_{32}\text{H}_{46}\text{O}_{16}$ , deduced from a peak at  $m/z$  709 due to  $[\text{M}+\text{Na}]^+$  in SI-MS and was hydrolyzed with  $\beta$ -glucosidase to give an aglycone (17),  $^1\text{H-NMR}$   $\delta$ : 0.94, 0.97, 1.52, 1.53, 3.80, 3.98, and 6.82, which on oxidation with  $\text{MnO}_2$  afforded isobrucein A (18).<sup>10)</sup> The position of the glycoside linkage was determined to be C-2 from the glycosylation shift ( $\Delta\delta$  +11.0 ppm). Yadanzioside H (8), on hydrogenation (10% Pd-C/EtOH), gave a dihydro derivative, which was identical with a

Table I.  $^1\text{H-NMR}$  Spectra (90MHz,  $\text{C}_5\text{D}_5\text{N}$ )<sup>a)</sup> of Yadanziosides A-H (1-8)

|                             | <u>1</u>             | <u>2</u> <sup>b)</sup>   | <u>3</u> <sup>b)</sup> | <u>4</u>  | <u>5</u>       | <u>6</u>  | <u>7</u>       | <u>8</u>             |
|-----------------------------|----------------------|--------------------------|------------------------|-----------|----------------|-----------|----------------|----------------------|
| 1-H                         | 7.22s                | 3.25d(16)                | 7.29s                  | c)        | c)             | 7.22s     | 7.25s          | c)                   |
| 2-H                         | -                    | -                        | -                      | 4.45m     | 4.42m          | -         | -              | 4.42m                |
| 3-H                         | -                    | -                        | -                      | 5.72brs   | 5.69brs        | -         | -              | 5.69brs              |
| 15-H                        | 6.84d(13)            | 6.9br                    | 6.8br                  | 6.78d(12) | 6.76d(13)      | 6.76d(13) | 6.81d(13)      | 6.82d(13)            |
| $\text{C}_{(4)}\text{-Me}$  | 1.13d(6)             | 2.04d(1.2)               | 1.19d(6.7)             | 1.43brs   | 1.43brs        | 1.15d(5)  | 1.17d(6)       | 1.42brs              |
| $\text{C}_{(10)}\text{-Me}$ | 1.61s                | 1.71s                    | 1.63s                  | 1.43brs   | 1.43brs        | 1.63s     | 1.64s          | 1.42brs              |
| $\text{CO}_2\text{Me}$      | 3.84s                | 3.84s                    | 3.73s                  | 3.77s     | 3.72s          | 3.81s     | 3.89s          | 3.79s                |
| 2'-H                        | c)                   | c)                       | 6.77d(1.2)             | -         | 5.82brs        | -         | 6.06s          | c)                   |
| 3'-Me                       | 0.96d(6)<br>0.99d(6) | 0.95d(6.7)<br>0.98d(6.7) | 2.40brs                | -         | 1.67s<br>2.14s | -         | 2.26s          | 0.94d(6)<br>0.97d(6) |
| 4'-Me                       | -                    | -                        | 1.41s<br>1.42s         | -         | -              | -         | 1.42s<br>1.46s | -                    |
| -OAc                        | -                    | -                        | -                      | 2.09s     | -              | 2.10s     | 1.95s          | -                    |
| Anomeric<br>-H              | c)                   | 5.46d(7.3)               | 5.37d(7.0)             | 4.93d(8)  | 4.94d(6)       | c)        | c)             | c)                   |

a) Coupling constants in Hz in parentheses. b) Measured at 400 MHz. c) Not measured.

Table II.  $^{13}\text{C}$ -NMR Spectra (22.5 MHz,  $\text{C}_5\text{D}_5\text{N}$ ) of Yadanzioids A-H (1-8)

|   | $\delta$                                                                                                                                                                                                                                                             |
|---|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| 1 | 12.5q, 17.9q, 22.4q, 22.5q, 25.8d, 30.0t, 39.5s, 40.6d, 41.3d, 43.3t, 43.8d, 46.6s, 50.4q, 52.3d, 62.3t, 68.4d, 71.3d, 73.4d, 73.7t, 74.5d, 75.9d, 78.3d, 78.7d, 82.6s, 83.5d, 102.0d, 129.6d, 148.8s, 168.1s, 171.1s, 171.6s, 194.6s                                |
| 2 | 15.3q, 15.8q, 22.4q, 22.5q, 25.9d, 29.3t, 40.8s, 42.2d, 43.3d, 43.3t, 46.1s, 50.6d, 51.1t, 52.4q, 62.8t, 68.4d, 71.6d, 73.1d, 73.6t, 75.7d, 76.0d, 78.4d, 78.6d, 82.7s, 83.5d, 104.9d, 146.2s, 147.9s, 168.1s, 171.2s, 171.6s, 193.6s                                |
| 3 | 12.6q, 15.6q, 18.0q, 28.9q, 28.9q, 30.0t, 39.7s, 40.5d, 41.4d, 43.8d, 46.7s, 50.3d, 52.4q, 62.4t, 68.4d, 71.3d, 73.2s, 73.6d, 73.8t, 74.7d, 76.1d, 78.6d, 79.0d, 82.6s, 83.5d, 102.0d, 112.9d, 129.3d, 149.2s, 166.5s, 168.2s, 168.3s, 171.2s, 194.5s                |
| 4 | 12.1q, 20.6q, 20.7q, 28.3t, 43.0d, 43.1d, 44.4s, 46.7s, 50.6d, 52.2q, 62.8t, 68.8d, 71.7d, 74.1t, 75.5d, 75.6d, 76.1d, 78.6d, 78.7d, 81.2d, 82.5s, 84.3d, 84.9d, 107.0d, 124.3d, 135.8s, 168.0s, 169.7s, 171.5s                                                      |
| 5 | 12.1q, 20.2q, 20.8q, 27.0q, 28.3t, 42.9d, 43.1d, 44.4s, 46.7s, 50.6d, 52.2q, 62.7t, 68.2d, 71.6d, 74.1t, 75.6d, 75.7d, 76.0d, 78.5d, 78.6d, 81.2d, 82.5s, 84.2d, 84.8d, 106.9d, 116.0d, 124.3d, 135.9s, 158.2s, 165.3s, 168.2s, 171.4s                               |
| 6 | 12.6q, 18.0q, 20.6q, 30.0t, 39.6s, 40.6d, 41.4d, 43.9d, 46.6s, 50.5q, 52.3d, 62.4t, 68.8d, 71.3d, 73.4d, 73.7t, 74.7d, 76.1d, 78.5d, 78.8d, 82.7s, 83.6d, 102.0d, 129.5d, 148.8s, 168.0s, 169.7s, 171.3s, 194.5s                                                     |
| 7 | 12.5q, 14.5q, 18.0q, 21.4q, 25.8q, 26.4q, 30.1t, 39.6s, 40.4d, 41.4d, 43.9d, 46.6s, 50.2d, 52.6q, 62.4t, 68.7d, 71.4d, 73.4d, 73.7t, 74.6d, 76.0d, 78.4d, 78.8d, 82.3s, 82.6s, 83.5d, 102.0d, 113.7d, 129.7d, 148.8s, 163.3s, 165.7s, 168.0s, 169.5s, 171.7s, 194.7s |
| 8 | 12.1q, 20.7q, 22.4q, 25.5q, 25.8d, 28.3t, 43.1d, 43.1d, 43.3t, 44.4s, 46.7s, 50.8d, 52.2q, 62.8t, 68.4d, 71.6d, 74.1t, 75.5d, 75.7d, 76.1d, 78.6d, 78.7d, 81.2d, 82.5s, 84.3d, 84.9d, 107.0d, 124.3d, 135.8s, 168.2s, 171.4s, 171.6s                                 |

tetrahydro derivative of yadanzioid E (5). Thus the structure of yadanzioid H (8) was formulated as the  $\beta$ -D-glucoside of the allylic alcohol (17) derived from isobrucein A (18).

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