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Short-Step Synthesis of Plant Growth-Promoting Brassinosteroids

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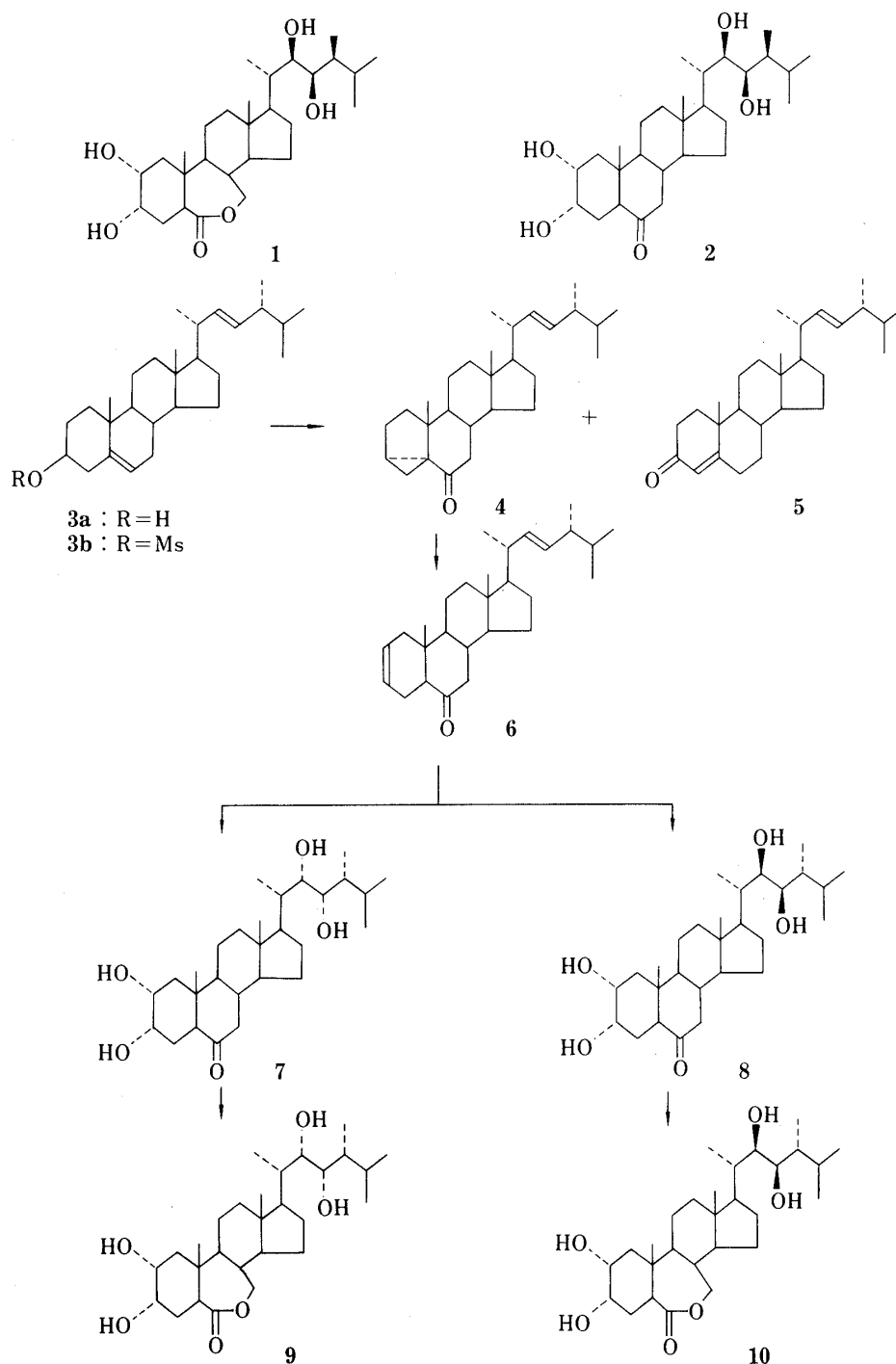
Brassinolide analogues, (22*R*, 23*R*, 24*R*)-2 α ,3 α ,22,23-tetrahydroxy-*B*-homo-7-oxa-5 α -ergostan-6-one (24-epibrassinolide) (**10**) and (22*S*, 23*S*, 24*R*)-2 α ,3 α ,22,23-tetrahydroxy-*B*-homo-7-oxa-5 α -ergostan-6-one (**9**), were synthesized from brassicasterol (**3a**) in five steps and with *ca.* 20% overall yield. The key steps are the direct formation of (22*E*, 24*R*)-3 α ,5-cyclo-5 α -ergost-22-en-6-one (**4**) from brassicasterol mesylate (**3b**), the acid-catalyzed rearrangement of **4** to (22*E*, 24*R*)-5 α -ergosta-2,22-dien-6-one (**6**), and the Baeyer–Villiger oxidation of the tetrahydroxy-5 α -ergostan-6-ones **7** and **8**.

Keywords—brassinolide; brassinosteroid; plant growth hormone; brassicasterol; Baeyer–Villiger oxidation

Brassinolide (**1**),¹⁾ (22*R*,23*R*,24*S*)-2 α ,3 α ,22,23-tetrahydroxy-*B*-homo-7-oxa-5 α -ergostan-6-one, and castasterone (**2**),²⁾ (22*R*,23*R*,24*S*)-2 α ,3 α ,22,23-tetrahydroxy-5 α -ergostan-6-one, are naturally occurring plant growth-promoting steroids. Brassinolide (**1**) showed a wide variety of biological activities in a number of bioassay systems.³⁾ Among the brassinolide analogues already synthesized, (22*R*,23*R*,24*R*)-2 α ,3 α ,22,23-tetrahydroxy-*B*-homo-7-oxa-5 α -ergostan-6-one, 24-epibrassinolide (**10**), and (22*S*,23*S*,24*R*)-2 α ,3 α ,22,23-tetrahydroxy-*B*-homo-7-oxa-5 α -ergostan-6-one (**9**) are promising candidate compounds for applications in agriculture⁴⁾ not only because these steroidal lactones were found to be highly active (comparable to brassinolide (**1**)) in a number of bioassays,⁵⁾ but also because an efficient synthesis in gram quantities from the commercially available ergosterol, (22*E*,24*R*)-ergosta-5,7,22-trien-3 β -ol, has been developed by USDA scientists.⁶⁾ The high biological activity of brassinosteroids **9** and **10** prompted us to report our short-step synthesis of **9** and **10** from brassicasterol, (22*E*,24*R*)-ergosta-5,22-dien-3 β -ol (**3a**).

In order to introduce 2 α ,3 α - and 22,23-*vicinal* diol functions, (22*E*,24*R*)-5 α -ergosta-2,22-dien-6-one (**6**) had to be elaborated from ergosterol or brassicasterol *via* the key intermediate, (22*E*,24*R*)-3 α ,5-cyclo-5 α -ergost-22-en-6-one (**4**). Thompson *et al.*⁶⁾ have transformed ergosterol into the 2,22-dien-6-one **6** in eight steps. Anastasia *et al.*⁷⁾ have obtained brassicasterol (**3a**) and its Δ^7 isomer in a ratio of 3:2 from the 1,4-cycloadduct of ergosterol acetate and 4-phenyl-1,2,4-triazoline-3,5-dione by reduction with lithium and ethylamine. They transformed brassicasterol (**3a**) into the 2,22-dien-6-one **6** in five steps. The common transformations to obtain the 3,5-cyclo-6-one **4** are as follows; tosylation, solvolysis to the cyclopropyl-6 β -ol, and oxidation (and reduction of the C-7 (8) double bond).

Brassicasterol (**3a**), present in rapeseed oil at 5–19% of the free sterol fraction⁸⁾ and now commercially available, is the most suitable starting material for the synthesis of brassinolide analogues **9** and **10**. In our case the preparation of the 3,5-cyclo-6-one **4** was achieved by treatment of the brassicasterol mesylate (**3b**) with sodium acetate⁹⁾ or potassium acetate in dimethyl sulphoxide at 90–100 °C in *ca.* 50% yield. The by-product (*ca.* 10%) was (22*E*,24*R*)-ergosta-4,22-dien-3-one (**5**). The use of triethylamine instead of sodium acetate failed to yield the cyclopropyl ketone **4**. The major product in this case was the dienone **5**. Acid-catalyzed



isomerization of the 3,5-cyclo-ketone **4** to (22*E*,24*R*)-5 α -ergosta-2,22-dien-6-one (**6**) was effected by heating with *p*-toluenesulphonic acid and sulpholane¹⁰⁾ in 65% yield. The 2,22-dien-6-one **6** in *tert*-BuOH-THF-H₂O (10:3:1) was treated with a catalytic amount of osmium tetroxide and 6.5 molar equivalents of *N*-methylmorpholine *N*-oxide¹¹⁾ at room temperature for 3 days to give a separable mixture of the tetraols **7** and **8**. Flash chromatography on silica gel provided the less polar (22*S*,23*S*,24*R*)-2 α ,3 α ,22,23-tetrahydroxy-5 α -ergostan-6-one (**7**, 50%), mp 184–185 °C, (lit.⁷⁾ mp 184–185 °C) and the more polar (22*R*,23*R*,24*R*)-2 α ,3 α ,22,23-tetrahydroxy-5 α -ergostan-6-one (**8**, 30%), mp 241–242 °C (lit.⁷⁾ mp 241–242 °C). Baeyer-Villiger oxidation¹²⁾ of **7** and **8** with trifluoroperacetic acid followed by recrystallization of the product provided (22*S*,23*S*,24*R*)-2 α ,3 α ,22,23-

tetrahydroxy-*B*-homo-7-oxa-5 α -ergostan-6-one (**9**), mp 193—195 °C (lit.⁷) mp 193—195 °C), and (22*R*,23*R*,24*R*)-2 α ,3 α ,22,23-tetrahydroxy-*B*-homo-7-oxa-5 α -ergostan-6-one (**10**), mp 256—257 °C (lit.⁷) mp 256—257 °C), respectively, in *ca.* 80% yield. The overall yield of the five-step synthesis of both **9** and **10** was *ca.* 20%.

Experimental

Melting points were determined with a hot-stage microscope and are uncorrected. Infrared (IR) spectra were recorded on a Hitachi 260-10 spectrometer. Proton nuclear magnetic resonance (¹H-NMR) spectra were taken with a Hitachi R-24 (60 MHz) spectrometer with tetramethylsilane as an internal standard. Mass spectra (MS) were taken with a Shimadzu LKB-9000S or a Shimadzu GC-MS 6020 mass spectrometer. Column chromatography was done on Kieselgel 60 F₂₅₄ (70—230 mesh, E. Merck). Analytical thin layer chromatography (TLC) was carried out on precoated Kieselgel 60 F₂₅₄ (0.25 mm thickness, E. Merck). The usual work-up refers to dilution with water, extraction with the organic solvent indicated in parentheses, washing of the extract to neutrality, drying over MgSO₄, filtration, and removal of the solvent by evaporation under a vacuum. The following abbreviations are used for ¹H-NMR data; s, singlet; d, doublet; dd, double doublet; m, multiplet.

(22*E*,24*R*)-3 α ,5-Cyclo-5 α -ergost-22-en-6-one (**4**)—Brassicasterol (**3a**) (4.1 g, 10.3 mmol) was treated with methanesulphonyl chloride (3 ml) and pyridine (20 ml) at room temperature for 3 h, then ice-water was added. The whole was extracted with ethyl acetate. The usual work-up gave the mesylate **3b** (4.9 g). This was treated with sodium acetate (5.0 g, 60.98 mmol) and dimethyl sulphoxide (160 ml) at 90—100 °C for 5 h. The usual work-up (ether) gave a crude product, which was applied to a column of silica gel (50 g). Elution with benzene provided (22*E*,24*R*)-3 α ,5-cyclo-5 α -ergost-22-en-6-one (**4**) (2.1 g, 51%), mp 109—111 °C (lit.⁷) mp 110—111 °C (from aqueous acetone). IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 1710. ¹H-NMR (CDCl₃) δ : 0.72 (3H, s, 18-H₃), 0.96 (3H, s, 19-H₃), 5.20 (2H, m, 22-H and 23-H). *Anal.* Calcd for C₂₈H₄₄O: C, 84.78; H, 11.18. Found: C, 84.66; H, 11.30.

Further elution with benzene-ethyl acetate (50:1) gave (22*E*,24*R*)-ergosta-4,22-dien-3-one (**5**) (0.49 g, 12%), mp 150—151 °C (from methanol). ¹H-NMR (CDCl₃) δ : 0.74 (3H, s, 18-H₃), 1.15 (3H, s, 19-H₃), 5.20 (2H, m, 22-H and 23-H), 6.14 (1H, s, 4-H). *Anal.* Calcd for C₂₈H₄₄O: C, 84.78; H, 11.18. Found: C, 84.70; H, 11.21.

(22*E*,24*R*)-5 α -Ergosta-2,22-dien-6-one (**6**)—The cyclopropyl ketone **4** (2.1 g, 5.28 mmol) was treated with *p*-toluenesulphonic acid (100 mg) and sulpholane (16 ml) at 160 °C for 1.5 h. The usual work-up (ether) gave a crude product, which was applied to a column of silica gel (50 g). Elution with hexane-benzene (1:5) provided (22*E*,24*R*)-5 α -ergosta-2,22-dien-6-one (**6**) (1.3 g, 65%), mp 123—124 °C (lit.⁷) mp 123—124 °C (from methanol). IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 1705. ¹H-NMR (CDCl₃) δ : 0.68 (3H, s, 18-H₃), 0.70 (3H, s, 19-H₃), 5.20 (2H, m, 22-H and 23-H), 5.60 (2H, m, 2-H and 3-H). *Anal.* Calcd for C₂₈H₄₄O: C, 84.78; H, 11.18. Found: C, 84.72; H, 11.27.

(22*S*,23*S*,24*R*)-2 α ,3 α ,22,23-Tetrahydroxy-5 α -ergostan-6-one (**7**) and (22*R*,23*R*,24*R*)-2 α ,3 α ,22,23-Tetrahydroxy-5 α -ergostan-6-one (**8**)—The diene **6** (1.3 g, 3.43 mmol) in *tert*-BuOH-THF-H₂O (10:3:1, 20 ml) was treated with osmium tetroxide (20 mg) and *N*-methylmorpholine *N*-oxide (3.0 g) at room temperature for 3 d, then sat. NaHSO₃ solution (20 ml) was added. The mixture was stirred at room temperature for 1 h. The usual work-up (dichloromethane) gave two separable products (1.5 g), which were purified by flash chromatography. Then 500 mg of the products was applied to a column of silica gel (3.5 cm i.d. \times 15 cm, Kieselgel 60, 230—400 mesh, E. Merck). Elution with chloroform-methanol (15:1) provided the less polar (22*S*,23*S*,24*R*)-2 α ,3 α ,22,23-tetrahydroxy-5 α -ergostan-6-one (**7**) (253 mg), mp 184—185 °C (lit.⁷) mp 184—185 °C (from ethyl acetate). EI-MS *m/z*: 446 (*M*⁺ - 18), 394, 393 (*M*⁺ - 71, C₂₃-C₂₄ fission), 364 (*M*⁺ - 101, C₂₂-C₂₃ fission + H, base peak), 363, 345, 327, 287, 263, 245, 175, 173, 155, 147, 107, 101, 95, 43. Emitter CI-MS (isobutane) *m/z*: 465 (*M*⁺ + 1, base peak), 447, 429. *Anal.* Calcd for C₂₈H₄₈O₅: C, 72.41; H, 10.43. Found: C, 72.28; H, 10.50. Tetraacetate of **7**; ¹H-NMR (CDCl₃) δ : 0.67 (3H, s, 18-H₃), 1.93 (3H, s, acetyl), 2.03 (9H, s, three acetyls), 4.50—5.50 (4H, m, 2-H, 3-H, 22-H, and 23-H).

Further elution with the same solvent gave the more polar (22*R*,23*R*,24*R*)-2 α ,3 α ,22,23-tetrahydroxy-5 α -ergostan-6-one (**8**) (152 mg), mp 241—242 °C (lit.⁷) mp 241—242 °C (from ethyl acetate). EI-MS *m/z*: 446 (*M*⁺ - 18), 394, 393, 364, 363, 345, 327, 287, 263, 245, 175, 173, 155, 147, 107, 101, 95, 43. Emitter CI-MS (isobutane) *m/z*: 465 (*M*⁺ + 1), 447, 429. *Anal.* Calcd for C₂₈H₄₈O₅: C, 72.41; H, 10.43. Found: C, 72.40; H, 10.47. Tetraacetate of **8**; ¹H-NMR (CDCl₃) δ : 0.67 (3H, s, 18-H₃), 1.95 (3H, s, acetyl), 2.00 (6H, s, two acetyls), 2.05 (3H, s, acetyl), 4.65—5.45 (4H, m, 2-H, 3-H, 22-H, and 23-H). The purification procedure was repeated three times. The total amounts of **7** and **8** were 760 and 457 mg (total yield, 80%), respectively.

(22*R*,23*R*,24*R*)-2 α ,3 α ,22,23-Tetrahydroxy-*B*-homo-7-oxa-5 α -ergostan-6-one (**10**)—Trifluoroperacetic acid was prepared by adding trifluoroacetic anhydride (6.74 ml) to 30% aqueous H₂O₂ (1.0 g) in dichloromethane (7.4 ml) at 0 °C. Three molar equivalents of the prepared trifluoroperacetic acid solution was added to a solution of the ketone **8** (400 mg, 0.86 mmol) in dichloromethane (10 ml) at 0 °C. The mixture was stirred at 0 °C for 3 h, then sat. aqueous NaHSO₃ solution (10 ml) was added. The whole was extracted with dichloromethane. The usual work-up and recrystallization from ethyl acetate provided (22*R*,23*R*,24*R*)-2 α ,3 α ,22,23-tetrahydroxy-*B*-homo-7-oxa-5 α -ergostan-6-

one (**10**) (343 mg, 83%), mp 256—258 °C (lit.⁷⁾ mp 256—258 °C). EI-MS m/z : 465 ($M^+ - 15$), 462 ($M^+ - 18$), 447, 409, 380 ($M^+ - 101$, $C_{22}-C_{23}$ fission + H, base peak), 379, 361, 350, 343, 331, 325, 322, 313, 307, 303, 285, 177, 173, 155, 131, 101, 71, 43. Emitter CI-MS (isobutane) m/z : 481 ($M^+ + 1$, base peak), 463, 445. *Anal.* Calcd for $C_{28}H_{46}O_6$: C, 70.03; H, 9.85. Found: C, 70.01; H, 9.92. Tetraacetate of **10**; 1H -NMR ($CDCl_3$) δ : 0.69 (3H, s, 18- H_3), 1.98 (3H, s, acetyl), 2.02 (6H, s, two acetyls), 2.09 (3H, s, acetyl), 3.00 (1H, dd, $J = 13$ and 6 Hz, 5 α -H), 4.07 (2H, m, 7- H_2), 4.70—5.50 (4H, m, 2-H, 3-H, 22-H, and 23-H). The mother liquor contained some trifluoroacetate of **10** which was recovered by saponification with 5% KOH-MeOH and relactonization with conc. HCl.

(22*S*,23*S*,24*R*)-2 α ,3 α ,22,23-Tetrahydroxy-*B*-homo-7-oxa-5 α -ergostan-6-one (**9**)—The 6-ketone **7** (500 mg, 1.08 mmol) was oxidized, as described for **10**, to give, after recrystallization from ethyl acetate, (22*S*,23*S*,24*R*)-2 α ,3 α ,22,23-tetrahydroxy-*B*-homo-7-oxa-5 α -ergostan-6-one (**9**) (415 mg, 80%), mp 193—195 °C (lit.⁷⁾ mp 193—195 °C). EI-MS m/z : 465 ($M^+ - 15$), 462 ($M^+ - 18$), 447, 409, 380, 379, 361, 350, 343, 331, 325, 322, 313, 307, 303, 285, 177, 173, 155, 131, 101, 71, 43. Emitter CI-MS (isobutane) m/z : 481 ($M^+ + 1$, base peak), 463, 445. *Anal.* Calcd for $C_{28}H_{48}O_6$: C, 70.03; H, 9.85. Found: C, 69.88; H, 9.87. Tetraacetate of **9**; 1H -NMR ($CDCl_3$) δ : 0.67 (3H, s, 18- H_3), 1.93 (3H, s, acetyl), 2.03 (9H, s, three acetyls), 3.00 (1H, dd, $J = 13$ and 6 Hz, 5 α -H), 4.07 (2H, m, 7- H_2), 4.50—5.50 (4H, m, 2-H, 3-H, 22-H, and 23-H). Some trifluoroacetate of **9** contained in the mother liquor was recovered as **9** by saponification and acidification as described above.

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References

- 1) M. D. Grove, G. F. Spencer, W. K. Rohwedder, N. B. Mandava, J. F. Worley, J. D. Warthen Jr., J. L. Flippen-Anderson, and J. C. Cook, Jr., *Nature* (London), **281**, 216 (1979).
- 2) T. Yokota, M. Arima, and N. Takahashi, *Tetrahedron Lett.*, **23**, 1275 (1982).
- 3) a) J. H. Yopp, N. B. Mandava, and J. M. Sasse, *Physiol. Plant.*, **53**, 445 (1981); b) N. B. Mandava, J. M. Sasse, and J. H. Yopp, *Physiol. Plant.*, **53**, 453 (1981); c) J. H. Yopp, N. B. Mandava, M. J. Thompson, and J. M. Sasse, 8th Proc. Plant Growth Reg. Soc. Am., 1981, p. 138; d) K. Wada, S. Marumo, N. Ikekawa, M. Morisaki, and K. Mori, *Plant and Cell Physiol.*, **22**, 323 (1981); e) L. E. Gregory and N. B. Mandava, *Physiol. Plant.*, **54**, 239 (1982).
- 4) T. H. Maugh II, *Science*, **212**, 33 (1981).
- 5) a) M. J. Thompson, N. B. Mandava, W. J. Meudt, W. R. Lusby, and D. W. Spaulding, *Steroids*, **38**, 567 (1981); b) M. J. Thompson, W. J. Meudt, N. B. Mandava, S. R. Dutky, W. R. Lusby, and D. W. Spaulding, *Steroids*, **39**, 89 (1982); c) S. Takatsuto, N. Yazawa, N. Ikekawa, T. Morishita, and H. Abe, *Phytochemistry*, **22**, 1393 (1983); d) S. Takatsuto, N. Yazawa, N. Ikekawa, T. Takematsu, Y. Takeuchi, and M. Koguchi, *Phytochemistry*, **22**, 2437 (1983).
- 6) M. J. Thompson, N. B. Mandava, J. L. Flippen-Anderson, J. F. Worley, S. R. Dutky, W. E. Robbins, and W. R. Lusby, *J. Org. Chem.*, **44**, 5002 (1979).
- 7) M. Anastasia, P. Ciuffreda, and A. Fiecchi, *J. Chem. Soc., Perkin Trans. 1*, **1983**, 379.
- 8) T. Itoh, T. Tamura and T. Matsumoto, *J. Am. Oil Chem. Soc.*, **50**, 122 (1973).
- 9) D. Libermann and R. Jacquier, *Bull. Soc. Chim. Fr.*, **1962**, 887.
- 10) a) D. H. R. Barton, P. G. Feakins, J. P. Poyser, and P. G. Sammes, *J. Chem. Soc., (C)*, **1970**, 1584; b) K. Mori, *Agric. Biol. Chem.*, **44**, 1211 (1980).
- 11) a) V. VanRheenen, R. C. Kelly, and D. Y. Cha, *Tetrahedron Lett.*, **1976**, 1973; b) L. Fieser and M. Fieser, "Steroids," Reinhold, New York, 1959, p. 274.
- 12) S. Takatsuto and N. Ikekawa, *Tetrahedron Lett.*, **24**, 917 (1983).