

## Communications to the Editor

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SYNTHESIS OF DIASTEREOMERIC 24,25-DIHYDROXYVITAMIN D<sub>2</sub>  
AND SEPARATION OF ITS (24R)- AND (24S)-ISOMERS

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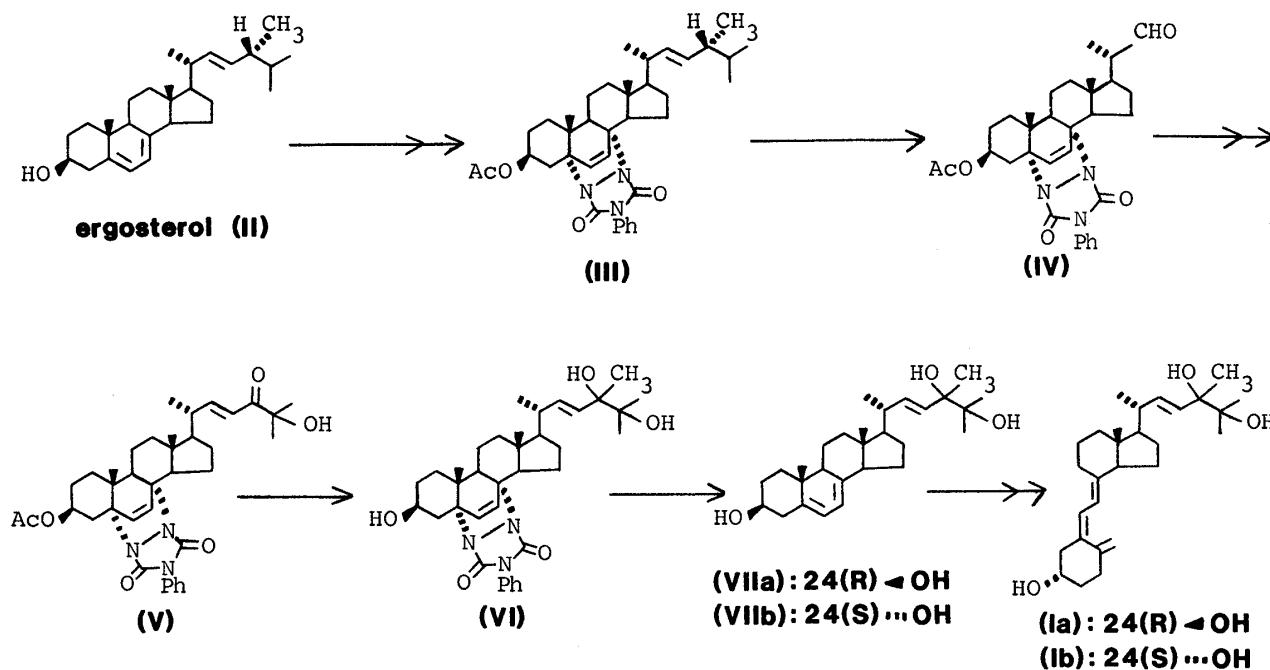
Diastereomeric 24,25-dihydroxyvitamin D<sub>2</sub> (I) was synthesized from ergosterol (II) through an efficient route and successfully separated into the (24R)- and (24S)-isomers by high-performance liquid chromatography (HPLC). The absolute configurations of the isomers were determined by co-chromatography with the authentic respective specimens.

KEYWORDS ——— 24,25-dihydroxyvitamin D<sub>2</sub>; 24R,25-dihydroxyvitamin D<sub>2</sub>; 24S,25-dihydroxyvitamin D<sub>2</sub>; ergosterol; high-performance liquid chromatography; vitamin D<sub>2</sub>; vitamin D

It has been documented that vitamin D is metabolized to 25-hydroxyvitamin D (25-OH-D) in the liver and subsequently to 1 $\alpha$ ,25-dihydroxyvitamin D [1 $\alpha$ ,25-(OH)<sub>2</sub>-D] or 24R,25-dihydroxyvitamin D [24R,25-(OH)<sub>2</sub>-D] in the kidney according to respectively lower or higher plasma calcium levels than normal.<sup>1)</sup> There are two groups of vitamin D, namely D<sub>2</sub> and D<sub>3</sub>, which are different only in the structures of side chain but they are known to have practically the same biological activity for mammals including humans. Though a number of syntheses of various metabolites of vitamin D<sub>3</sub> have been reported, only few reports have appeared on the synthesis of vitamin D<sub>2</sub> metabolites due to difficulties in the synthesis by inserting a double bond into the 22- and a methyl group into the 24S-positions and also to the difficulty of securing synthetic compounds in vitamin D<sub>2</sub> series. Though the synthesis of 24,25-(OH)<sub>2</sub>-D<sub>2</sub> from stigmastanol was reported by Jones *et al.*,<sup>2,3)</sup> their synthetic route is apparently complicated, resulting in only a poor overall yield. Therefore, we have investigated the modification of the synthesis of vitamin D<sub>2</sub> metabolites and succeeded in the establishment of an improved synthesis of a vitamin D<sub>2</sub> metabolite, 24,25-(OH)<sub>2</sub>-D<sub>2</sub>, its separation into (24R)- and (24S)-isomers, and the confirmation of their absolute configuration.

As shown in Chart 1, ergosterol (II) was converted into the known 20-aldehyde (IV) *via* the route involving the protection of the 5,7-diene group and ozonolysis according to the procedure given by Barton *et al.*<sup>4)</sup> The 20-aldehyde (IV) was then converted to the enone (V) by the condensation with 3-methyl-3-(tetrahydropyran-2-yl-oxy)butan-2-one according to the procedure given by Eyley and Williams<sup>5)</sup> in 36% yield. Methylation of the enone (V) with methylolithium afforded the methylated 24,25-glycol (VI) as a mixture of diastereomers in 60% yield, which without separation was refluxed

with lithium aluminum hydride in tetrahydrofuran to afford the desired 24,25-dihydroxyvitamin D<sub>2</sub> [24,25-(OH)<sub>2</sub>-pro-D<sub>2</sub> (VII)] also as a mixture of diastereomers in 70% yield [MS: m/z, 428 (M<sup>+</sup>); <sup>1</sup>H-NMR (CDCl<sub>3</sub>): δ 0.64 (3H, s, 13-Me), 1.07 (3H, d, J= 6Hz, 20-Me), 1.20, 1.22 and 1.26 (each s, 24-Me and 25-Me<sub>2</sub>), 5.34~5.64(m, 6-, 7-, 22- and 23-H); UV λ<sub>max</sub><sup>ethanol</sup> : 272, 281 and 292 nm].



**Chart 1. Our Synthetic Course of 24,25-Dihydroxyvitamin D<sub>2</sub>**

Separation of two diastereomers (VIIa and VIIb) was performed by HPLC using a Zorbax SIL column with 2.5% isopropanol in n-hexane as a mobile phase. As shown on the profile of HPLC in Fig. 1, two peaks were clearly separated with nearly same peak heights; the first peak was confirmed as (24S)-24,25-(OH)<sub>2</sub>-pro-D<sub>2</sub> (VIIb) and the second peak as (24R)-isomer (VIIa) by converting the respective fractions into the corresponding 24,25-(OH)<sub>2</sub>-D<sub>2</sub> (Ib and Ia) upon ultraviolet (UV) irradiation followed by thermal isomerization. The UV irradiation was carried out by using a monochromatic ray at 295 nm obtained from a spectroirradiator (Japan Spectroscopic Co., CRM-FA type) and thermal isomerization was performed by refluxing the UV irradiated ethanolic solution for 2 h. Each isomerized product was applied to HPLC which clearly separated respective products as shown in Fig. 2, thus identifying the former peak as (24S)-24,25-(OH)<sub>2</sub>-D<sub>2</sub> (Ib) and the latter as (24R)-24,25-(OH)<sub>2</sub>-D<sub>2</sub> (Ia). [(Ib) MS: m/z, 428 (M<sup>+</sup>); <sup>1</sup>H-NMR (CDCl<sub>3</sub>): δ 0.50 (s, 13-Me), 0.98 (d, J=6Hz, 20-Me), 1.14, 1.16 and 1.20 (s, 24-Me and 25-Me<sub>2</sub>), 3.88 (m, 3-H), 4.76 and 4.99 (br s, 19-methylene), 5.50 (m, 22- and 23-H), 5.97 and 6.20 (d, J=11Hz, 6- and 7-H); UV λ<sub>max</sub><sup>ethanol</sup> : 265 nm and λ<sub>min</sub><sup>ethanol</sup> : 228 nm].

The identity of respective products (Ia and Ib) was established by co-chromatography with authentic samples of (24R)- and (24S)-24,25-(OH)<sub>2</sub>-D<sub>2</sub> kindly donated by Dr. G. Jones, as shown in Fig. 2, thus unambiguously confirming not only their structures but their absolute configurations.

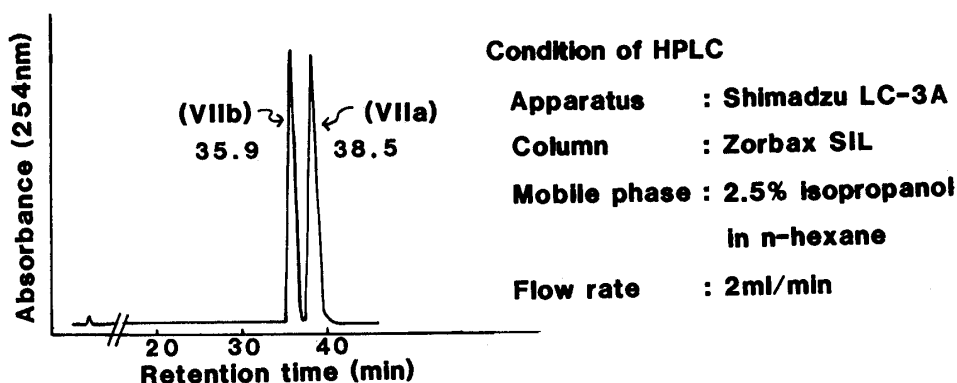


Fig. 1. Profile of HPLC on (VIIa) and (VIIb)

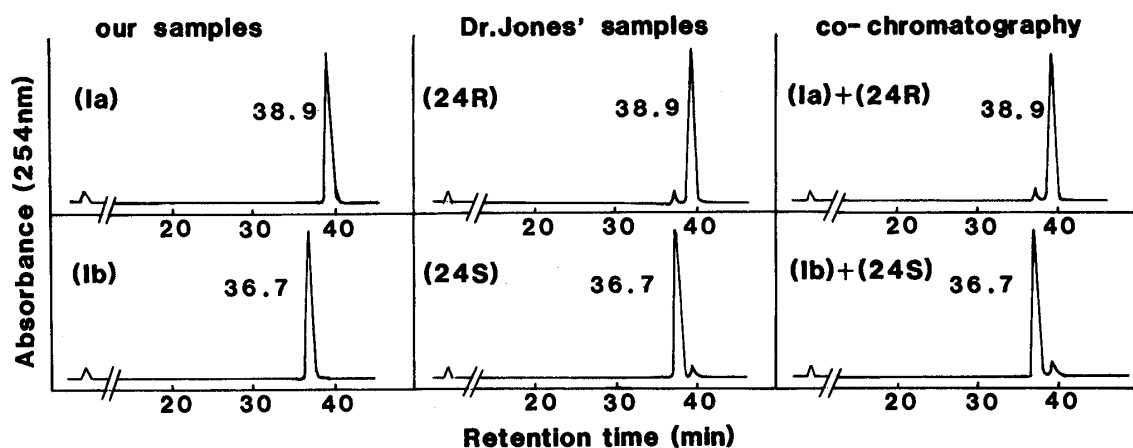


Fig. 2. Co-chromatography of Our Synthetic and Isolated Compounds (Ia and Ib) with the Authentic Isomers Kindly Donated by Dr. Jones. (The conditions of HPLC were the same as described in Fig.1.)

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