

[Chem. Pharm. Bull.  
36(10)4157—4161(1988)]

# <sup>1</sup>H- and <sup>13</sup>C-Nuclear Magnetic Resonance Spectra of Rifampicin and 3-[(Dimethylhydrazono)methyl]rifamycin SV

MASAHITO TAGUCHI,\*<sup>a</sup> YUKINOBU YAMANE,<sup>b</sup> NORIO AIKAWA,<sup>a</sup>  
and GORO TSUKAMOTO<sup>a</sup>

Pharmaceuticals Research Center<sup>a</sup> and Development Research Laboratories,<sup>b</sup>  
Kanebo, Ltd., Tomobuchi-cho, Miyakojima-ku, Osaka 534, Japan

(Received March 22, 1988)

Assignments of labile protons (NH, OH-1, OH-4 and OH-8) and naphthalene carbon atoms in the <sup>1</sup>H- and <sup>13</sup>C-nuclear magnetic resonance spectra of rifampicin (1) and 3-[(dimethylhydrazono)methyl]rifamycin SV (2) in CDCl<sub>3</sub> were made based on long-range selective decoupling experiments. The published assignments of the labile protons and two naphthalene carbon atoms (C-5 and C-9) of 1 should be revised.

**Keywords**—rifamycin; <sup>1</sup>H-NMR; <sup>13</sup>C-NMR; rifampicin

Rifamycins are important natural products, and signal assignments in the <sup>13</sup>C-nuclear magnetic resonance (<sup>13</sup>C-NMR) spectra of naphthoquinone-type rifamycins have been discussed in detail.<sup>1)</sup> Assignments of signals in the <sup>13</sup>C-NMR spectra for 1,4-naphthalenediol-type rifamycins including rifampicin<sup>2)</sup> (1), a most valuable drug for the treatment of tuberculosis, have also been published,<sup>3)</sup> but the basis of the assignments was not described. Recently, we reinvestigated the structure of the deep-blue compound, which was formed in the reaction of rifamycin S with 1,3,5-tri-*tert*-butylhexahydro-1,3,5-triazine,<sup>4)</sup> and in connection with this work, it became necessary to confirm the published <sup>13</sup>C-NMR assignments of the naphthalene carbon atoms (C-1—C-10) of 1,4-naphthalenediol-type rifamycins. This paper describes the results of long-range selective proton decoupling (LSPD) experiments on 1 and 3-[(dimethylhydrazono)methyl]rifamycin SV (2), and points out that the published assignments<sup>3b,c)</sup> of C-5 and C-9 of 1,4-naphthalenediol-type rifamycins should be interchanged. Signal assignments of labile protons (NH, OH-1, OH-4 and OH-8) in the <sup>1</sup>H-nuclear magnetic resonance (<sup>1</sup>H-NMR) spectra of 1 and 2 are also described.

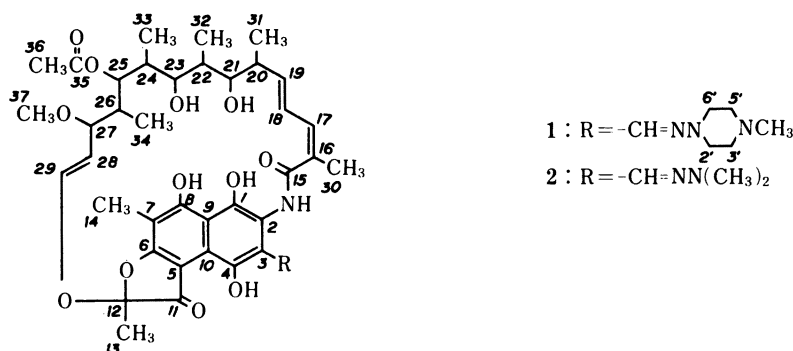


Chart 1

### Results and Discussion

Chemical shifts and assignments in the  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra of **1** and **2** in  $\text{CDCl}_3$  are summarized in Tables I and II. Assignments of protons and carbon atoms of **1** and **2**, except for four labile protons in the chromophore (NH, OH-1, OH-4 and OH-8) and all quaternary carbon atoms, were made based on H-H and C-H correlation spectroscopy, and we confirmed the published assignments<sup>3a,b,5a)</sup> of these protons and carbon atoms of **1**. In order to assign labile protons in the chromophore and all quaternary carbon atoms, proton non-decoupling spectra and LSPD spectra were recorded. The data are summarized in Tables

TABLE I.  $^1\text{H}$ -NMR Chemical Shifts of Rifampicin (**1**) and 3-[(Dimethylhydrazono)methyl]rifamycin SV (**2**) in  $\text{CDCl}_3$  (Concn. ca.  $10^{-1}\text{ M}$ ) at 300 MHz

Compound	NH	OH-1	OH-4	OH-8	H-13	H-14	H-17	H-18	H-19	H-20	H-21
<b>1</b>	$\delta$ 13.19	12.8—13.8	11.98	12.8—13.8	1.80	2.21	6.39	6.57	5.93	2.37	3.77
<b>2</b>	$\delta$ 13.18	13.72	11.77	13.13	1.83	2.14	6.38	6.46	5.91	2.34	3.71

Compound	OH-21	H-22	H-23	OH-23	H-24	H-25	H-26	H-27	H-28	H-29	H-30
<b>1</b>	3.50	1.70	3.00	3.71	1.52	4.95	1.33	3.47	5.10	6.20	2.08
<b>2</b>	3.45	1.67	3.00	3.67	1.47	4.91	1.28	3.42	5.05	6.20	2.11

Compound	H-31	H-32	H-33	H-34	H-36	H-37	CH=N	H-2',6'	H-3',5'	NCH <sub>3</sub>	N(CH <sub>3</sub> ) <sub>2</sub>
<b>1</b>	0.88	1.01	0.59	-0.31	2.05	3.04	8.25	3.13	2.56	2.33	—
<b>2</b>	0.80	0.98	0.57	-0.39	2.03	3.01	7.71	—	—	—	2.90

TABLE II.  $^{13}\text{C}$ -NMR Chemical Shifts of Rifampicin (**1**) and 3-[(Dimethylhydrazono)methyl]rifamycin SV (**2**) in  $\text{CDCl}_3$  (Concn. ca.  $10^{-1}\text{ M}$ ) at 75.47 MHz

Compound	C-1	C-2	C-3	C-4	C-5	C-6	C-7	C-8	C-9	C-10	C-11
<b>1</b>	138.56	120.43	110.88	147.92	104.53	174.47	106.05	169.19	112.82	117.90	195.42
<b>2</b>	138.17	119.77	111.30	146.63	104.39	173.92	104.99	168.76	111.66	117.46	194.73

Compound	C-12	C-13	C-14	C-15	C-16	C-17	C-18	C-19	C-20	C-21	C-22
<b>1</b>	108.74	21.40	7.50	169.67	129.37	135.05	123.21	142.64	38.54	70.62	33.36
<b>2</b>	108.65	21.42	7.30	169.47	129.47	134.34	122.99	142.05	38.28	70.37	33.10

Compound	C-23	C-24	C-25	C-26	C-27	C-28	C-29	C-30	C-31	C-32	C-33
<b>1</b>	76.89	37.49	74.41	39.48	76.74	118.64	142.62	20.66	17.80	10.84	8.42
<b>2</b>	76.57	37.28	74.18	39.38	76.57	118.59	142.63	20.54	16.52	10.52	8.24

Compound	C-34	C-35	C-36	C-37	CH=N	C-2',6'	C-3',5'	NCH <sub>3</sub>	N(CH <sub>3</sub> ) <sub>2</sub>
<b>1</b>	8.85	172.00	20.67	57.04	134.41	50.25	53.93	45.87	—
<b>2</b>	8.65	171.69	20.46	56.71	129.80	—	—	—	41.71

TABLE III.  $^{13}\text{C}$ -NMR Chemical Shifts and Multiplicities of Quaternary Carbon Atoms in a Proton Non-decoupling Spectrum of Rifampicin (**1**) in  $\text{CDCl}_3$  and Change of Multiplicities by LSPD

Chemical shifts	Multiplicities ( <i>J</i> , Hz)	Assignments	Change of multiplicities ( <i>J</i> , Hz) by LSPD			
			$\delta$ 2.21 (H-14)	$\delta$ 8.25 (CH = N)	$\delta$ 11.98 (OH-4)	$\delta$ 13.19 (NH)
195.42	q (2)	C-11	—	—	—	—
174.47	q (4)	C-6	s	—	—	—
172.00	dq (4, 7)	C-35	—	—	—	—
169.67	dqd (1, 4, 12)	C-15	—	—	—	qd (4, 12)
169.19	q (4)	C-8	s	—	—	—
147.92	t (2)	C-4	—	d (2)	d (2)	—
138.56	dd (1, 5)	C-1	—	d (5)	—	d (1)
129.37	dq (1, 7)	C-16	—	—	—	—
120.43	d (6)	C-2	—	s	—	—
117.90	d (1)	C-10	—	—	s	—
112.82	s	C-9	a)	—	—	—
110.88	dt (3, 6)	C-3	—	dd (3, 6)	dd (3, 6)	t (6)
108.74	quintet (4.5)	C-12	—	—	—	—
106.05	q (6)	C-7	s	—	—	—
104.53	s	C-5	—	—	—	—

a) The peak height was clearly enhanced.

TABLE IV.  $^{13}\text{C}$ -NMR Chemical Shifts and Multiplicities of Quaternary Carbon Atoms in a Proton Non-decoupling Spectrum of 3-[(Dimethylhydrazono)methyl]rifamycin SV (**2**) in  $\text{CDCl}_3$  and Change of Multiplicities by LSPD

Chemical shifts	Multiplicities ( <i>J</i> , Hz)	Assignments	Change of multiplicities ( <i>J</i> , Hz) by LSPD					
			$\delta$ 2.14 (H-14)	$\delta$ 7.71 (CH = N)	$\delta$ 11.77 (OH-4)	$\delta$ 13.13 (OH-8)	$\delta$ 13.18 (NH)	$\delta$ 13.72 (OH-1)
194.73	q (3)	C-11	—	—	—	—	—	—
173.92	dq (2, 4)	C-6	d (2)	—	—	q (4)	—	—
171.69	dq (4, 7)	C-35	—	—	—	—	—	—
169.47	qd (4, 11)	C-15	—	—	—	—	—	—
168.76	quintet (4)	C-8	d (4)	—	—	q (4)	—	—
146.63	t (2)	C-4	—	d (2)	d (2)	—	—	—
138.17	ddd (1, 3.5, 4.5)	C-1	—	dd (3.5, 4.5)	—	—	dd (1, 3.5)	dd (1, 4.5)
129.47	q (7.5)	C-16	—	—	—	—	—	—
119.77	d (6)	C-2	—	s	—	—	—	—
117.46	d (1)	C-10	—	—	s	—	—	—
111.66	t (4)	C-9	a)	—	—	d (4)	—	d (4)
111.30	dt (3.5, 6)	C-3	—	dd (3.5, 6)	dd (3.5, 6)	—	t (6)	—
108.65	quintet (4)	C-12	—	—	—	—	—	—
104.99	quintet (6)	C-7	d (6)	—	—	q (6)	—	—
104.39	s	C-5	—	—	—	—	—	—

a) The peak height was clearly enhanced.

III and IV. Assignments were made as follows.

**Assignments of C-11, C-12, C-15, C-16 and C-35**

Assignments of these five quaternary carbon atoms were made with reference to the published assignments<sup>3a,b)</sup> of **1**, and were confirmed by the multiplicities of the corresponding signals (Tables III and IV).

### Assignments of C-1, C-2, C-3, C-4, C-10 and Labile Protons in the Chromophore

**Assignments for 1**—LSPD of the hydrazonomethyl proton ( $\text{CH}=\text{N}$ ) affected four signals at 147.92, 138.56, 120.43 and 110.88, and hence these signals must be attributed to C-1, C-2, C-3, C-4 or C-10. The two lower signals at 147.92 and 138.56 are attributed to C-1 and C-4, because these carbons each bear an oxygen atom. The  $^1\text{H}$ -NMR spectrum of **1** showed two sharp singlets at  $\delta$  11.98 and 13.19 (each corresponding to one proton) and one very broad signal in the range of  $\delta$  12.8–13.8 (corresponding to two protons), due to four labile protons in the chromophore (NH, OH-1, OH-4 and OH-8). The singlet at  $\delta$  13.19 is assigned to NH, because LSPD of the  $\delta$  13.19 proton affected the signal at 169.67 (C-15). The LSPD of the same proton also affected the signal at 138.56, and this result allowed us to assign the signal at 138.56 to C-1. The signal at 147.92 is therefore assigned to C-4. LSPD of the  $\delta$  11.98 proton affected the signal at 147.92 (C-4), and hence the singlet at  $\delta$  11.98 is assigned to OH-4. The remaining broad signal in the range of  $\delta$  12.8–13.8 is therefore assigned to OH-1 and OH-8. These assignments of labile protons in the chromophore are in disagreement with the assignments in the literature.<sup>3a,b,5)</sup> The published assignments of these protons should be revised as above. The signal at 110.88 was affected by LSPD of the hydrazonomethyl proton ( $\text{CH}=\text{N}$ ), the amidic proton (NH), and the proton of the 4-hydroxyl group, and is assigned to C-3. The remaining signal at 120.43 is assigned to C-2, because the signal at 117.90 is assigned to C-10 which was affected by LSPD of the proton of the 4-hydroxyl group. These assignments of C-1, C-2, C-3, C-4 and C-10 are in agreement with the published assignments.<sup>3b)</sup>

**Assignments for 2**—Assignments of C-1, C-2, C-3, C-4 and C-10 of **2** were made with reference to the assignments described above for **1**. Differing from **1**, the  $^1\text{H}$ -NMR spectrum of **2** showed four sharp singlets due to four labile protons in the chromophore (NH, OH-1, OH-4 and OH-8), and assignments of these labile protons in the chromophore were made by LSPD experiments. The results are shown in Table IV.

### Assignments of C-5, C-6, C-7, C-8 and C-9

**Assignments for 1**—Assignments of C-5, C-6, C-7, C-8 and C-9 were made with reference to assignments described below for **2**, and the assignments of C-5 and C-9 were found to be in disagreement with the published assignments.<sup>3b)</sup> The published assignments of C-5 and C-9 should be interchanged as shown in Tables II and III.

**Assignments for 2**—Among fifteen signals due to quaternary carbon atoms, the five signals at 173.92, 168.76, 111.66, 104.99 and 104.39 must be attributed to C-5, C-6, C-7, C-8, and C-9. The two signals at 173.92 and 168.76 are attributed to two carbons, C-6 and C-8, bearing an oxygen atom. The coupling constants between these carbons and the proton of the 8-hydroxyl group allowed us to assign the signal at 173.92 to C-6 and the signal at 168.76 to C-8, because the long-range coupling constant  $^2J$  is generally larger than  $^4J$ .<sup>6)</sup> The result of LSPD of H-14 allowed us to assign the signal at 104.99 to C-7. The signal at 111.66 was affected by LSPD of the protons of the 1-hydroxyl and the 8-hydroxyl groups, and hence this signal is assigned to C-9. The remaining signal at 104.39 must be attributed to C-5. These assignments mean that the published assignments<sup>3b,c)</sup> of C-5 and C-9 of 1,4-naphthalenediol-type rifamycins should be interchanged.

### Experimental

Compounds **1** and **2** were prepared according to the literature.<sup>7,8)</sup> The  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra were recorded on a Bruker AM-300 spectrometer in  $\text{CDCl}_3$  solution, and tetramethylsilane (TMS) was used as an internal reference. Typical experimental conditions are as follows. For  $^1\text{H}$ -NMR spectra: pulse width  $4\ \mu\text{s}$  ( $15\ \mu\text{s}$  for a  $90^\circ$  pulse); sweep width 6000 Hz at 300 MHz; relaxation delay 1 s; acquisition time 2.72 s; 32 k memory. For  $^{13}\text{C}$ -NMR spectra: pulse width  $3.6\ \mu\text{s}$  ( $7\ \mu\text{s}$  for a  $90^\circ$  pulse); sweep width 18500 Hz at 75.47 Hz; relaxation delay 1 s; acquisition time 1.769 s; 64 k memory. The results are summarized in Tables I–IV.

---

References

- 1) E. Martinelli, R. J. White, G. G. Gallo, and P. J. Beynon, *Tetrahedron*, **29**, 3441 (1973); H. Fuhrer, *Helv. Chim. Acta*, **56**, 2377 (1973); E. Martinelli, R. J. White, G. G. Gallo, and P. J. Beynon, *Tetrahedron Lett.*, **1974**, 1367.
- 2) N. Maggi, C. R. Pasqualucci, R. Ballotta, and P. Sensi, *Chemotherapia*, **11**, 285 (1966).
- 3) a) G. G. Gallo and P. Radaelli, "Analytical Profiles of Drug Substances," Vol. 5, ed. by K. Florey, Academic Press, New York, 1976, pp. 467—513; b) R. Cricchio, P. Antonini, G. C. Lancini, G. Tamborini, R. J. White, and E. Martinelli, *Tetrahedron*, **36**, 1415 (1980); c) P. Traxler, T. Schupp, and W. Wehrli, *J. Antibiot.*, **35**, 594 (1982); L. Marsili, M. Ballabio, G. Franceschi, G. Oronzo, and A. Vigevani, *ibid.*, **35**, 1621 (1982).
- 4) M. Taguchi, N. Aikawa, and G. Tsukamoto, *Bull. Chem. Soc. Jpn.*, **61**, 2431 (1988).
- 5) a) E. Martinelli, P. Gironi, and G. G. Gallo, *Farmaco Ed. Sci.*, **36**, 671 (1981); L. Cellai, S. Cerrini, A. Segre, M. Brufani, W. Fedeli, and A. Vaciago, *J. Org. Chem.*, **47**, 2652 (1982); b) M. F. Dampier, C.-W. Chen, and H. W. Whitlock, Jr., *J. Am. Chem. Soc.*, **98**, 7064 (1976).
- 6) J. W. Akitt, "NMR and Chemistry—An Introduction to Nuclear Magnetic Resonance Spectroscopy—," Chapman and Hall, Ltd., London, 1973 ["NMR Nyumon," translated by M. Hirota, Tokyo Kagaku Dozin, Ltd., Tokyo, 1975, p. 32].
- 7) S. p. A. Lepetit, *Neth. Appl.* 6509961 (1966) [*Chem. Abstr.*, **65**, 5462 (1966)].
- 8) L. Marsili and C. Pasqualucci, *Ger. Offen.* 2433105 (1975) [*Chem. Abstr.*, **82**, 156407s (1975)].