4232 Vol. 36 (1988)

Communications to the Editor

Chem. Pharm. Bull. 36(10)4232—4235(1988)

PULOSARIOSIDE, A NEW BITTER TRIMERIC-IRIDOID DIGLUCOSIDE,
FROM AN INDONESIAN JAMU, THE BARK OF
ALYXIA REINWARDTII BL. (APOCYNACEAE)

Isao Kitagawa,*,a Hirotaka Shibuya,a Nam In Baek,a Yoshihiro Yokokawa,a Aya Nitta,b Harry Wiriadinata,c and Masayuki Yoshikawa

Faculty of Pharmaceutical Sciences, Osaka University, a 1-6, Yamada-oka, Suita, Osaka 565, Japan, Faculty of Pharmaceutical Sciences, Kyoto University, b Yoshida, Shimoadachi-cho, Sakyo-ku, Kyoto 606, Japan and Puslitbang Biologi, Herbarium Bogoriense, Jalan Raya Juanda, 22-24, Bogor, Indonesia

A new bitter trimeric-iridoid diglucoside named pulosarioside ($\frac{1}{2}$) was isolated from an Indonesian folk-medicine (Jamu, "pulosari"), the air-dried bark of $Alyxia\ reinwardtii\ B\underline{L}$. (Apocynaceae), and the absolute configuration of $\frac{1}{2}$ has been determined on the basis of chemical and physicochemical evidence.

KEYWORDS — pulosarioside; trimeric-iridoid diglucoside; Indonesian Jamu; $Alyxia\ reinwardtii;$ Apocynaceae; (+)-pinoresinol glucoside; loganin

The air-dried bark of Alyxia reinwardtii BL. (Apocynaceae) (called "pulosari" in Indonesia) is one of the popular Indonesian folk medicines (Jamu). It has been used for various intestinal diseases and diarrhea. It is also known as a stomachic and an antispasmodic. During the course of our investigations in search of new biologically active constituents in naturally occurring drug materials in Indonesia, 1) we have chemically analyzed "pulosari." Along with the identification of known lup-20(30)-en-3 β -yl acetate, (+)-pinoresinol, 2) and (+)-pinoresinol β -D-glucopyranoside, 2) we have isolated a new bitter trimericitiodid diglucoside named pulosarioside (1) and have elucidated the structure on the basis of the following evidence. 3

The MeOH extract of the air-dried bark (obtained at Sukabumi, Java) was partitioned into a mixture of AcOEt and water. Chromatographic separation of the AcOEtsoluble portion using silica gel and reversed phase silica gel (Bondapak C_{18}) provided lup-20(30)-en-3 β -yl acetate (0.31% from the bark), (+)-pinoresinol (0.10%), and (+)-pinoresinol β -D-glucopyranoside (0.07%). The water-soluble portion was subjected to silica gel (eluting with CHCl $_3$ -MeOH-H $_2$ O=7:3:1, lower layer) and reversed phase silica gel (Bondapak C_{18} , MeOH-H $_2$ O=1:1) column chromatography to furnish pulosarioside ($_1$, 0.14%), mp 132 °C (EtOH), [α] $_0^{24}$ -94° (MeOH), C_{43} H $_6$ O $_{23}$ ·2H $_2$ O, 4) UV (MeOH): 230 nm ($_1$ 17600), IR (KBr): 3380 (br), 2913, 1695, 1625 cm⁻¹, SIMS: $\frac{m}{2}$ 967 (M+Na) $_1$, a mixture of its monoacetal (0.02%), and a diacetal (0.05%). 5)

Alkaline hydrolysis (3% NaOMe-MeOH, 25 °C, 20 min) of pulosarioside (1) and subsequent acetylation (Ac₂O-pyridine=1:2, 25 °C, 40 min) of the product yielded 2a, a white powder, $[\alpha]_D^{20}$ +129° (CHCl₃), $C_{14}H_{20}O_6$, IR (CHCl₃): 2960, 1736, 1236 cm⁻¹, 3a, a white powder, $[\alpha]_D^{20}$ -10° (CHCl₃), $C_{14}H_{20}O_6$, IR (CHCl₃): 2936, 1734. 1224 cm⁻¹, $\widetilde{5a}$, a white powder, $[\alpha]_D^{20}$ -133° (CHCl₃), $C_{25}H_{32}O_{14}$, IR (CHCl₃): 2938, 1751, 1704,

1215 cm⁻¹, and vogeloside tetraacetate (6a), 6) approximately in 1:1:2:2 ratio. 7)

The unknown C-7' configuration of 6a has been determined as S on the basis of the ¹H NMR analysis 8) including the NOE experiments. 9) Furthermore, 5a has been shown to be the C-7' epimer of 6a by ¹³C (Table I) and ¹H NMR comparisons and NOE experiments. 11)

The 1 H NMR spectrum of 2 A showed, besides two acetoxyl signals (6 2.06, 2.08), signals at 6 4.21 (d, J=12.0 Hz), 4.32 (dd, J=3.5, 12.0 Hz) (1-H₂), 3.04 (ddd, J=6.0, 7.0, 13.5 Hz, 4 6 B-H), 2.98 (dddd, J=8.0, 10.5, 11.0, 13.5 Hz, 5 6 B-H), 1.36 (ddd, J=3.5, 10.5, 14.0 Hz), 1.98 (dd, J=8.0, 14.0 Hz) (6 B-H₂), 5.20 (dd, J=3.5, 3.5 Hz, 7 6 B-H), 2.07 (ddq, J=3.5, 7.0, 11.0 Hz, 8 6 B-H), 2.21 (ddd, J=3.5, 11.0, 11.0 Hz, 9 6 B-H), 1.02 (d, J=7.0 Hz, 8-CH₃), 4.06 (dd, J=7.0, 11.5 Hz), 4.48 (dd, J=6.0, 11.5 Hz) (11-H₂). The NOE's were observed between the following pairs of protons 10 : 10 B-H 6 B-H (4%), 10 B-H 6 B-H (11%), 10 B-H 6 B-H (3%), 10 B-H 6 B-H (8%), 10 B-H 6 B-CH₃ (4%), and 10 B-CH₃ 10 B-H (10%). Based on these physical data and detailed 10 C NMR examinations, the relative configuration of 10 A has been elucidated as shown. In addition, by the detailed 11 H and 12 C NMR examinations of 3 A including the NOE experiments of 1 H NMR, 12

To determine the absolute configurations of 2a and 3a, loganin (8) ¹³ was converted to 2a and 3a. Thus, loganin (8) was first hydrolyzed to give $8a^{13}$ by aq. 4% KOH treatment (reflux, 3 h). Enzymatic hydrolysis of 8a (β -glucosidase, H_2O , 41 °C, 64h, with stirring) followed by NaBH₄ reduction in aq. 95% EtOH (25 °C, 2 h) furnished two products: 2 (11% from 8), a white powder, $[\alpha]_D^{25}$ +117° (MeOH), $C_{10}H_{16}O_4$, IR (KBr): 3464 (br), 2894, 1719 cm⁻¹ and 3 (10% from 8), a white powder, $[\alpha]_D^{25}$ -35° (MeOH), $C_{10}H_{16}O_4$, IR (KBr): 3545 (br), 2879, 1721 cm⁻¹. Respective acetylation of 2 and 3 provided 2a and 3a. Thus the absolute configurations of two iridoid-lactones, which were obtained above by alkaline hydrolysis of pulosarioside (1), have been clarified.

The 1 H NMR spectrum (in d₅-pyridine-D₂O) of pulosarioside (1) showed signals due to one $sec.CH_3$ group (δ 1.06, d, J=7 Hz), one methoxycarbonyl group (δ 3.75), two aldehydic protons (δ 9.85, 9.87) and two olefinic protons (δ 7.71, 7.76, both br s, 3'-H, 3"-H). In addition to accumulated spectral properties of pulosarioside (1), the product ratio (vide~supra) of two iridoid-lactones (2a, 3a) and two secoiridoid-glucosides (5a, 6a) obtained by alkaline treatment of pulosarioside (1), have shown that 1 contains one mole of iridoid and two moles of secoiridoid-glucosides connecting through two ester-linkages in its molecule.

Silylation of 1 (t-butyldimethylsilyl chloride-imidazole in DMF, 35 °C, 72 h) and subsequent alkaline treatment of the product (2% NaOMe-MeOH, 25 °C, 1 h) provided two products: 2b, a white powder, $\left[\alpha\right]_{D}^{22}$ +6.0° (CHCl3), $C_{16}H_{30}O_{4}Si$, IR (CHCl3): 3680, 3425, 2929, 2856, 1754, 1604 cm⁻¹ and 7 (a mixture of C-7' epimers), IR (CHCl3): 2914, 1721 cm⁻¹, ^{1}H NMR: δ 3.67, 3.70 (totally 3H, both s, 7'-oCH3), 7.49 (br s, 3'-H). Oxidation of 2b with pyridinium chlorochromate (PCC) in CH2Cl2 gave 4, a white powder, $\left[\alpha\right]_{D}^{22}$ +16° (CHCl3), $C_{16}H_{28}O_{4}Si$, UV (MeOH): 238 nm (ε 1000), IR (CHCl3): 2927, 2857, 1741, 1080 cm⁻¹. The ^{1}H NMR spectrum of 4 showed, besides signals due to one t-butyldimethylsilyl group (δ 0.07, 6H, s; δ 0.89, 9H, s), signals at δ 4.31 (d, J=12.0 Hz), 4.49 (dd, J=3.5, 12.0 Hz) (1-H2), 3.04 (ddd, J=5.0, 7.0, 10.0 Hz, 4-H), 3.24 (dddd, J=3.0, 7.0, 10.0, 10.0 Hz, 5-H), 2.15 (ddd, J=2.0, 10.0, 18.0 Hz), 2.35 (dd, J=10.0, 18.0 Hz) (6-H2), 2.47 (ddd, J=2.0, 6.5,

Table I. ¹³C NMR Data

	1 ^{a)} ∼	la ^{b)}	_{2a} b)	_{3a} b)		<u>l</u> a)	la ^{b)}	5a ^{b)}	6a ^{b)}
1 3 4 5 6 7 8	64.3 62.9 46.1 41.0 36.0 76.3 37.5 44.3	64.5 64.5 46.0 41.2 36.1 76.7 37.9 44.2	67.4 172.4 41.6 35.2 35.4 77.9 40.5 43.3	69.2 172.4 43.2 36.9 39.0 78.9 41.0 43.5	1', 1" 3', 3" 4', 4" 5', 5" 6', 6" 7', 7" 8', 8" 9', 9"	96.6 152.7 108.9, 109.1 26.9, 27.3 44.1, 44.4 200.6 133.9 49.3	96.9 151.0 110.2 26.3 43.7 200.4, 200.5 133.2 49.4	95.4 151.5 105.3 21.9 29.1 101.4 131.2 41.7	96.2 151.5 104.7 24.2 30.4 103.5 130.8 42.1
10 11	14.5 174.8	14.6 173.7	12.6 62.3	13.3 62.3	10',10" 11',11"	119.6 166.3, 166.5	120.6 166.3, 166.6	121.1 164.3	121.4 164.3

a), b) Measured at 125 MHz a) in d_5 -pyridine or b) in CDCl $_3$.

6.5 Hz, 8-H), 2.38 (ddd, J=3.0, 3.5, 6.5 Hz, 9-H), 1.14 (d, J=6.5 Hz, 8 β -CH₃), 3.64 (dd, J=10.0, 10.0 Hz), 4.09 (dd, J=5.0, 10.0 Hz) (11-H₂). The NOE's were observed between the following pairs of protons: 4β -H & 5 β -H (11%), 5β -H & 4 β -H (4%), 5β -H & 9 β -H (9%), 9β -H & 5 β -H (17%), 9β -H & 8 β -CH₃ (3%), and 8β -CH₃ & 9 β -H (12%). Based on these findings together with 13 C NMR analysis, the stereostructures of 2b and 4 with 4(R) configurations have been elucidated. In addition, it has been shown that 1-OH and 7-OH groups in the iridoid moiety are respectively esterified with the carboxyl residues of two secoiridoid-glucoside moieties in pulosarioside (1).

Furthermore, enzymatic hydrolysis of $\hat{1}$ with lipase (Triacylglycerol lipase from Porcine pancreas, Type II Crude, Sigma) in a phosphate buffer (pH 5.5) (38 °C, 72 h) followed by treatment with ionic resin (Dowex 50w x 8, H⁺ form) provided $\hat{2}$. On alkaline treatment (2% NaOMe-MeOH, 20 °C, 10 h), $\hat{2}$ was epimerized to $\hat{3}$, while $\hat{3}$ was not affected by a similar alkaline treatment. Consequently, the C-4 configuration in $\hat{1}$ has been confirmed as \hat{R} which is preserved in $\hat{2}$. Finally, the above-mentioned findings and the $\hat{1}^3$ C NMR examinations in detail of $\hat{1}$, $\hat{1}$, $\hat{2}$, $\hat{3}$, $\hat{5}$, and $\hat{6}$, have substantiated the absolute configuration of pulosarioside (1) as shown.

It is interestingly pointed out here that pulosarioside (1) is very bitter comparable to swertiamarin which is the bitter principle of $Swertia\ japonica\ Makino\ 16$)

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and this property may be important for the use of "pulosari" as a stomachic.

The authors are grateful to the Ministry of Education, Science, and Culture of Japan for Grant-in-Aid for Overseas Scientific Survey (No. 60041044, 61043040).

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4) The molecular composition of the compound given with the chemical formula was determined by elemental analysis or high resolution mass spectrometry.

- 5) The aq. 50% acetone extract of the air-dried bark ("pulosari") contained pulosarioside (1) but lacked these monoacetal and diacetal derivatives as shown by TLC and HPLC, so that these acetals were considered secondarily formed by TLC and HPLC, so that these acetals were considered secondarily formed during the MeOH extraction. Monoacetal mixture, IR (KBr): 3494 (br), 2920, 1696, 1625, 1275, 1062 cm⁻¹, ¹H NMR (d5-pyridine-D₂O, δ): 1.09 (d, J=6.5 Hz, 8-CH₃), 3.35, 3.58 (both s, OCH₃x₂), 3.74 (s, COOCH₃), 9.86 (s, CHO), ¹³C NMR (d5-pyridine, $\delta_{\rm C}$): 14.6 (10-C), 51.4, 52.0, 53.0 (OCH₃x₃), 103.1 (¹/₂C), 103.3 (¹/₂C), 200.6 (7', 7"-C), SIMS: m/z 1013 (M+Na)⁺. Diacetal, mp 104-105 °C (EtOH), $\alpha_{\rm C}$ C (MeOH), $\alpha_{\rm C}$ C (MeOH), $\alpha_{\rm C}$ C (HoH), $\alpha_{\rm C}$ C (EtOH), $\alpha_{\rm C}$ C (MeOH), $\alpha_{\rm C}$ C (MeOH), $\alpha_{\rm C}$ C (HoH), $\alpha_{$
- 7) TLC examination of the reaction mixture showed that only iridoid-lactone 2 was liberated at the initial stage of alkaline treatment and 3 was formed gradually at the time proceeded.

8) The $^{
m 1}$ H NMR spectra were measured at 500 MHz in CDCl $_{
m 3}$ unless otherwise specified.

fied.

9) $6a: {}^{1}H$ NMR $(\delta): 5.37$ (d, J=1.5 Hz, $1'\alpha-H$), 7.59 (d, J=2.5 Hz, 3'-H), 3.22 (dddd, J=1.5, 2.5, 5.5, 13.5 Hz, $5'\beta-H$), 1.67 (ddd, J=3.0, 13.5, 13.5 Hz, $6'\alpha-H$), 1.82 (ddd, J=1.5, 5.0, 13.5 Hz, $6'\beta-H$), 5.29 (dd, J=3.0, 5.0 Hz, $7'\alpha-H$), 5.46 (ddd, J=9.5, 9.5, 17.0 Hz, 8'-H), 2.61 (ddd, J=1.5, 5.5, 9.5 Hz, $9'\beta-H$), 5.11 (dd, J=7.5, 17.0 Hz), 5.28 (dd, J=7.5, 9.5 Hz) ($10'-H_2$). NOE experiments: $5'\beta-H$ & $6'\beta-H$ (6^8), $5'\beta-H$ & $9'\beta-H$ (10^8), $6'\beta-H$ & $5'\beta-H$ (9^8), $6'\alpha-H$ & $7'\alpha-H$ (7^8), $7'\alpha-H$ & $6'\alpha-H$ (4^8), $7'\beta-OCH_3$ & $6'\beta-H$ (3^8). 10)

The magnitude of NOE (3^8) given in the parenthesis was obtained when the under-

10) The magnitude of NOE (%) given in the parenthesis was obtained when the underlined proton was irradiated.

- lined proton was irradiated.
 11) $5a: ^{1}H$ NMR $(\delta): 5.34$ (d, J=1.5 Hz, $1'\alpha-H)$, 7.56 (d, J=2.5 Hz, 3'-H), 2.82 (dddd, J=1.0, 2.5, 5.5, 9.5 Hz, $5'\beta-H)$, 1.54 (ddd, J=9.5, 10.0, 13.5 Hz, $6'\alpha-H)$, 1.94 (ddd, J=1.0, 2.5, 13.5 Hz, $6'\beta-H)$, 5.22 (dd, J=2.5, 10.0 Hz, $7'\beta-H)$, 5.43 (ddd, J=9.5, 10.0, 17.0 Hz, 8'-H), 2.67 (ddd, J=1.5, 5.5, 9.5 Hz, $9'\beta-H)$, 5.28 (dd, J=7.5, 10.0 Hz), 5.29 (dd, J=7.5, 17.0 Hz)($10'-H_2$). NOE experiments: $5'\beta-H$ & $6'\beta-H$ (4%), $6'\beta-H$ & $5'\beta-H$ & $8'\beta-H$ & $8'\beta-H$ & $7'\beta-H$ (5%), $7'\beta-H$ & $6'\beta-H$ (11%).
 12) $3a: ^{1}H$ NMR $(\delta): 3.99$ (dd, J=11.5, 11.5 Hz), 4.41 (dd, J=6.0, 11.5 Hz)($1-H_2$), 2.65 (ddd, J=5.0, 6.0, 11.0 Hz, $4\alpha-H$), 2.48 (ddd, J=7.5, 10.0, 11.0, 11.0 Hz, $5\beta-H$), 1.70 (ddd, J=4.0, 10.0, 14.0 Hz, $7\alpha-H$), 1.87 (ddq, J=4.0, 7.5, 14.0 Hz)($6-H_2$), 5.26 (ddd, J=1.0, 4.0, 4.0 Hz, $7\alpha-H$), 1.87 (ddq, J=4.0, 7.0, 9.5 Hz, $8\alpha-H$), 2.35 (dddd, J=6.0, 9.5, 11.0, 11.5 Hz, $9\beta-H$), 1.02 (d, J=7.0 Hz, $8\beta-CH_3$), 4.24 (dd, J=5.0, 11.5 Hz), 4.43 (dd, J=6.0, 11.5 Hz) ($1-H_2$). NOE experiments: $5\beta-H$ & 11-H (δ 4.24)(5%), $9\beta-H$ & 11-H (δ 4.43)(6%), $8\beta-CH_3$ & $9\beta-H$ (5%).
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 14) When the C-4 carbinyl group was protected with a t-butyldimethylsilyl group,
- 14) When the C-4 carbinyl group was protected with a t-butyldimethylsilyl group, the C-4 configuration seemed hardly epimerized on alkaline treatment.
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