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Marine Terpenes and Terpenoids. V.¹⁾ Oxidation of Sarcophytol A, a Potent Anti-tumor-Promoter from the Soft Coral Sarcophyton glaucum

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Sarcophytol A (1a), a potent anti-tumor-promotor cembranoid from the soft coral Sarcophyton glaucum, was subjected to various oxidation reactions. m-Chloroperbenzoic acid oxidation of 1a gave two epoxides (2a, 2b) which were converted to the corresponding diols 6b, 11b, 12b, 12'b and triols 4b, 9b. Lithium aluminum deuteride reduction of 2a gave a C-7 monodeuterated diol; this was used as a model experiment for the tritiation of 1a. Chromic acid oxidation of 1a gave seco-cembranoids 13b—18, a dienone 19, and a dihydrofuran derivative 20a. Sarcophytol A acetate (1b) was less reactive in chromic acid oxidation and afforded a seco-aldehyde 22a and a dihydroxy derivative 23a in low yield. Hydrolysis of 22a followed by acid treatment afforded the furan 18. Hydrolysis of 23a gave a triol, which, on further oxidation, gave the aldehyde 16, the major product of the chromic acid oxidation of 1a.

Keywords—soft coral; *Sarcophyton glaucum*; cembranoid; seco-cembranoid; sarcophytol A; oxidation

Sarcophytol A (1a) is a simple monohydroxy cembratetraene which we previously isolated from *Sarcophyton glaucum*, a common soft coral found in the coral reefs of Indo-Pacific coastal waters.²⁾ Recently, Fujiki and co-workers found that sarcophytol A efficiently inhibits the activity of the powerful tumor promoter teleocidin,^{3a)} in a two-stage carcinogenesis experiment on the mouse dorsal skin using dimethylbenzanthracene as an initiator.^{3b,4)} The effects on the growth and average numbers of tumors were observed even with an equimolar amount of 1a with respect to teleocidin, and its potency is almost equal to those of known anti-tumor-promoter retinoic acid derivatives. Usually retinoids simultaneously cause severe hypervitaminosis A syndrome when applied to test animals,⁵⁾ which limits the prospects of clinical use of retinoids in the future. The potency of sarcophytol A is also unparalleled when compared with known natural anti-tumor-promoters such as pentagalloylglucose,⁶⁾ flavonoids,⁷⁾ or cembranoids from tobacco leaves,⁸⁾ since they invariably exert their activity only when amounts more than thousand times that of the promoters are applied.

There are several reasons that led us to examine the derivatization of sarcophytol A. In the first place its tritium labeling is indispensable for biochemical and physiological experiments. Secondly, unsaturated cembranoids are generally susceptible to oxidation and the structures and properties of the possible oxidative products should be taken into account. Also, the lipophilic nature of 1a sometimes made it difficult to dissolve at the required concentrations in aqueous media, so that more hydrophilic derivatives of 1a were desirable as possible substitutes for 1a. For these reasons we prepared several oxidized derivatives of 1a and also examined its stability to autoxidation.

S. glaucum is one of the most intensively studied soft corals as regards its chemical components but the major cembranoids varied according the localities where the coral was

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collected.⁹⁾ S. glaucum collected at Ishigaki Island, Okinawa, contains mainly **1a** (about a quarter of the total lipids) with its dihydroxy derivatives.^{2,13)} The geometry at the C-1 double bond and the absolute configuration at C-14 of **1a** remained unsettled. The proton nuclear magnetic resonance (1 H-NMR) spectrum of **1a** showed 1,2-Z geometry since there are significant nuclear Overhauser effects (NOEs) between the protons at C-14 (δ 4.99, dd, J= 10.0, 4.0 Hz) and at C-3 (δ 5.99, br d, J=11.5 Hz), and also between the protons at C-2 (δ 6.15, d, J=11.5 Hz) and C-16, 17, and C-18 (δ 1.74, d, J=1.0 Hz). The absolute configuration at C-14 was shown to be S by Horeau's method.^{10,11)}

Epoxidation of the acetate 1b gave two monoepoxides 2a and 2b. The lack of NOEs between the methyl group and the vicinal oxymethine proton of the epoxide rings indicated that both 2a and 2b are E-epoxides. Treatment of 2a with p-toluenesulfonic acid in benzene gave a mixture of allyl alcohols from which the major product 3a was isolated. Similar treatment of 2b gave an allyl alcohol 8a as the predominant product. The products from 2a were complex and a small amount of 8a was also obtained. The ¹H-NMR and ultraviolet (UV) spectra of 3a and 8a (Experimental) indicated that their conjugated diene moieties were unaffected by epoxidation and acid treatment. Since the coupling patterns of C-14 acetoxy methine protons in 3a and 8a were unchanged (3a, δ 5.82, dd, J=11.4, 3.3 Hz; 8a, δ 5.73, dd, J=10.3, 4.2 Hz), as compared with that of 1b (δ 6.03, dd, J=9.5, 4.5 Hz), both 2a and 2b are 7,8-epoxides. Formation of 8a from 2a was assumed to be due to the acid-catalyzed isomerization from 3a, or to the partial conversion of 2a to 2b by hydrolysis of the epoxide ring, giving 4a or 9a, followed by recyclization. The absolute configurations at C-7 of 3a and 8a were shown by Horeau's method to be R and S, respectively, and hence the configuration of the epoxide group of 2a is TR,8R and that of 2b is TS,8S.

Treatment of 2a with dilute perchloric acid in acetone gave a glycol 4a. The ring-opening occurred at C-8 since dehydration of the diacetate 4c with thionyl chloride in pyridine followed by hydrolysis gave 3b. Similarly, 2b gave the glycol 9a which on acetylation, dehydration, and hydrolysis afforded 8b. The major product was the terminal olefin (5 from 4c and 10 from 9c) in both cases. Lithium aluminum hydride reduction of 2a gave a diol 6b as the major product. In contrast, the reduction of 2b gave the 8,14-diol 11b and significant amounts of 7,14-diols 12b and 12'b, whose configurations at C-8 are unknown at present. These ten di- and trihydroxycembranoids (3b, 4b, 5, 6b, 8b, 9b, 10, 11b, 12b, 12'b) retain the 1,3-diene and 14-hydroxy moieties of 1a. However, their diverse specific rotations and the chemical shifts of the protons at C-2 and C-3, and C-14 to C-18 (Experimental) indicate that there are significant conformational variations among these compounds. Hydrolysis of one acetoxyl group sometimes caused large differences; for example, the triol monoacetate 9a ($[\alpha]_D + 118$) on hydrolysis gave the triol 9b, $[\alpha]_D - 105$.

For the tritiation of 1a, a model experiment was carried out using deuterium. Dehydration of the monoacetate 6a with phosphoryl chloride or thionyl chloride in pyridine gave sarcophytol A acetate (1b) as the major product, with smaller amounts of isomeric olefins. The by-products were easily removable on silver nitrate-impregnated silica gel chromatography. Pyrolysis of the diacetate of 6b led to similar results but in poor overall yield. Lithium aluminum deuteride reduction and acetylation of 2b gave 6c in which the C-7 pro-(S) hydrogen was replaced by deuterium. The introduced deuterium would not be simply antiperiplanar with respect to the C-8 hydroxyl group, unlike acyclic compounds, but the anti-elimination reaction was supposed to give an olefin in which the deuterium at C-7 was mostly lost. Indeed, treatment of 6c with phosphoryl chloride or thionyl chloride in pyridine and subsequent purification afforded 1b which retained only 25% or 37% of deuterium respectively, as judged from the intensity of C-7 olefinic proton signal $(\delta 4.98, \text{ br t}, J=7.0 \text{ Hz})$ in the $^1\text{H-NMR}$ spectra. However, treatment of 6c with thionyl chloride in refluxing benzene, which was proposed to be a syn-elimination reaction, 12 was quite efficient and the purified

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product 7 showed nearly complete loss of the signal at $\delta 4.98$. One of the olefinic methyl signals ($\delta 1.47$) of 7 was heightened as compared with that of 1b, showing it to be that of the C-19 protons. These three dehydration methods are however equally applicable for the tritiation of 1a to a sufficient specific activity for ordinary use.

Compound la was next subjected to several oxidative conditions. Manganese dioxide oxidation gave only a low yield of the dienone 19.13) After several attempts, chromic acid oxidation of 1a using Jones' reagent was studied in detail since it gave a variety of products, some of which are products of autoxidation of 1a. A solution of 1a in ether was stirred briefly with excess Jones' reagent and the product was separated to yield a minor acidic fraction, which gave 13b—15b after methylation, and a major neutral fraction, which gave 16—19 and 20a. Compounds 13b—15b were found to be the seco-acid derivatives formed by oxidative cleavage at C-1,14 (13b, 14b), and C-1,14 and C-3,4 (15b). Compound 13b was an α,β unsaturated ketone [UV, 222 nm (ε , 12000)] and its ¹H-NMR spectrum showed the signals of two olefinic methyls (δ 1.59, 3H, s; 1.68, 3H, br d, J=0.7 Hz), an isopropyl (δ 1.13, 6H, d, J=6.8 Hz), one tertiary methyl which is adjacent to oxygen (δ 1.34, 3H, s), two olefinic protons linked to isolated double bonds (δ 5.14 and 5.26, each 1H, brt, $J=7.0\,\mathrm{Hz}$), and olefinic protons of an α,β -conjugated ketone (δ 6.41, 1H, d, J=15.6 Hz; 6.88, 1H, d, J=15.6 Hz). Its infrared (IR) spectrum showed absorptions due to hydroxyl (3450 cm⁻¹), ester (1735 cm⁻¹), and α,β -unsaturated ketone (1690, 1668, 1625 cm⁻¹) moieties. All these data support the view that oxidative cleavage occurred at C-1,14 and the conjugated diene moiety in 1a was converted to a γ -hydroxy- α , β -unsaturated ketone in 13b. Compound 14b was a conjugated enol dihydrofuran derivative of 13a, and showed signals of the olefinic protons of a conjugated diene at δ 5.91 (d, J = 5.9 Hz) and 6.26 (d, J = 5.9 Hz) in the ¹H-NMR spectrum. The carbon-13 NMR (¹³C-NMR) spectrum showed characteristic signals of the enol moiety at δ 153.6 (C-1) and 98.5 (C-15) ppm. Compound 15b was the methylketone derivative (1 H-NMR δ : 2.14, 3H, s) formed by cleavage at two sites (C-1,14 and C-3,4).

The neutral fraction was composed of seco-cembranoid (16-18) and cembranoid (19, 20a) derivatives. The major product was the ketoaldehyde 16 and its ¹H-NMR signals were essentially the same (Experimental) as those of 13b except that the signal of the methoxyl group was replaced with that of an aldehyde (δ 9.96, 1H, t, J=2.5 Hz). Compound 17 was a 1:1 mixture of α,β -unsaturated Z- and E-aldehyde [1H-NMR δ : 9.99 and 9.92, each d, J=8.1 Hz]. Alkaline treatment of 16 gave 17 and a 1:1 mixture of conjugated dihydrofuran (21). It is interesting to note that compounds 13a-17 are related to retinoids both in their gross molecular weight and partial structure involving C-6 to C-14, and C-19 and C-20. Compound 18 was a seco-cembranoid having a furan ring formed by cleavage at C-3,4, followed by cyclization. Its ¹H-NMR spectrum showed the signals of a 2,3-disubstituted furan ring at δ 6.25 (1H, d, J = 2.0 Hz) and 7.23 (1H, d, J = 2.0 Hz), and the signal of a methylketo group at δ 2.13. Compound 19 was obtained in low yield and was identical with the dienone obtained by manganese dioxide oxidation of 1a.13) Compound 20a was also a cembranoid derivative of 1a having one extra oxygen atom. The IR spectrum showed the presence of a hydroxyl group which was not acetylated under the usual conditions (Ac₂O-pyridine). The UV absorption of the diene was lost and the ¹H-NMR showed the signals of the protons on a 2,3,5trisubstituted dihydrofuran ring (δ 4.70, 1H, dq, J=5.5, 1.0 Hz, 3-H; δ 4.86, 1H, br t, J= 5.5 Hz, 14-H; δ 5.41, 1H, q, J = 1.0 Hz, 2-H). These assignments were based on the presence of coupling of 14-H with one of the 13-H (δ 2.46, dd, J=13.6, 5.5 Hz) and also with 3-H, the latter being characteristic of such dihydrofuran ring systems, as found in deoxosarcophine or sarcophytonin A.^{2,14)} The acetate **20b**, prepared by acetic anhydride-triethylaminedimethylaminopyridine medium, showed little change in the signals of these protons (Experimental).

Sarcophytol A acetate (1b) was resistant to Jones' reagent and after prolonged stirring

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(22 h), it afforded small amounts of 22a and 23a (Chart 3). Compound 22a was an α,β -unsaturated aldehyde formed by the oxidative cleavage of the C-3,4 bond [UV: 234 nm (ϵ , 16000). ¹H-NMR δ : 2.04, 3H, s, Ac; 2.14, 3H, s, 18-H; 5.90, 1H, d, J=7.8 Hz, 2-H; 5.98, 1H, dd, J=7.8, 5.6 Hz, 14-H; 10.19, 1H, d, J=7.8 Hz, 3-H]. Compound 23a was the 1,4-dihydroxy derivative with an E-double bond at C-2,3 (¹H-NMR δ : 5.74, 1H, d, J=16.1 Hz; 6.11, 1H, d, J=16.1 Hz). The structures of 22a and 23a were confirmed by correlating to 18 and 16, respectively. Alkaline hydrolysis of 22a gave an alcohol which on brief treatment with dilute acid was converted to 18 in 85% yield. Alkaline hydrolysis of 23a gave a triol 23b, which, on brief treatment with Jones' reagent, afforded 16 in 60% yield. These results suggest that the primary product of the chromic acid oxidation of 1a is probably the 3,4-epoxide 24, and 20a is an its internal cyclization product. Conversion of 24 to the glycol 25, or to the glycol 23b with double bond migration, and subsequent oxidation would afford compounds 13a, 15a, 16, and 22b. Probably the strongly acidic Jones' reagent catalyzed the cyclization of 13a and 22b into

Chart 3

Chart 4

14a and 18, respectively (Chart 4).

Examination of the complex autoxidation products of 1a showed that compound 20a was one of the major components. Compound 18 was also a minor component but the presence of other synthetic compounds, described above, were not confirmed. When 1a was kept in air at room temperature, it afforded mainly two more polar compounds, supposed to be 24 and a derivative. They are quite labile and after separation soon changed to mixtures, of which compound 20a was the major product. Similar but much slower conversion was observed when 1a was kept in dilute solution in benzene or hexane—ethyl acetate (8:2). Facile decomposition of 1a was observed when 1a was kept in chloroform. A yellow color developed gradually and decomposition was complete in a week. Probably the trace of hydrochloric acid in chloroform accelerated the decomposition. In contrast, 1a was stable in ethanol solution and showed virtually no change on standing at room temperature for 7 d.

Experimental

Melting points were determined on a Kofler hot stage apparatus and are uncorrected. Optical rotations were determined on a JASCO DIP-4 digital polarimeter. ¹H-NMR spectra were determined in CDCl₃ solution with tetramethylsilane as an internal standard, on a JNM GX-270 spectrometer at 270 MHz unless otherwise specified. ¹³C-NMR spectra were determined in CDCl₃ solution on a JNM FX-90Q spectrometer at 22.5 MHz. Mass spectra (MS) were determined on a JEOL JMS D300 spectrometer. IR spectra were taken on a JASCO A 102 spectrometer. UV spectra were determined on a Hitachi EPS-3T spectrometer. Chromatography was carried out on a column of silica gel by the flash chromatography method. ¹⁵⁾

Configuration at C-14 of 1a—Compound 1a (288 mg, 1 mmol) was esterified with 620 mg (2 mmol) of α -phenylbutyric anhydride in pyridine for 1 h. Thin-layer chromatography (TLC) of the mixture showed about 80% of 1a was esterified. The mixture was worked up according the standard method.¹⁰⁾ The resultant benzene solution (5 ml) showed rotation of -0.28° and, accordingly, 14S configuration.

Epoxidation of 1b—Compound 1b (11.5 g) in 120 ml of CHCl₃ was treated dropwise with a solution of *m*-chloroperbenzoic acid (9.24 g) in 120 ml of CHCl₃ at 0 °C over a period of 1 h. The mixture was washed with 10% Na₂SO₃ solution (200 ml), H₂O, and saturated NaCl solution, then the solvent was evaporated off. Column chromatography of the residue in portions over silica gel with ethyl acetate—hexane (1:9 and 1:1) afforded a mixture (0.6 g) which consisted of unreacted 1b and unidentified compounds, 2a (4.4 g), 2b (2.9 g), and a mixture of more polar compounds which did not show any strong UV absorption. 2a, oil, [α]_D +150 ° (c=1.02, CHCl₃). ¹H-NMR δ: 1.06, 1.16 (each 3H, d, J=7.0 Hz, 16,17-H), 1.30 (3H, s, 19-H), 1.52 (3H, s, 20-H), 1.79 (3H, s, 18-H), 2.05 (3H, s), 2.54 (1H, epot, J=7 Hz, 15-H), 2.79 (1H, t, J=6.0 Hz, 7-H), 5.19 (1H, br t, J=6.0 Hz, 11-H), 5.92 (1H, dd, J=9.5, 4.0 Hz, 14-H), 6.08 (1H, br d, J=11.5 Hz, 3-H), 6.23 (1H, d, J=11.5 Hz, 2-H). UV $\lambda_{\text{max}}^{\text{EIOH}}$ nm (ε): 251 (15000). IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 1735, 1650, 1605. MS m/z: 346 (M⁺), 331, 286, 243, 137, 109. High-resolution MS [Found (Calcd)] m/z: C₂₂H₃₄O₃ (M⁺), 346.25063 (346.25073). 2b, oil, [α]_D +110 ° (c=1.18, CHCl₃). ¹H-NMR δ: 1.05, and 1.08 (each 3H, d, J=7.0 Hz, 16,17-H), 1.18 (3H, s, 19-H), 1.63 (3H, s, 20-H), 1.71 (3H, s, 18-H), 2.06 (3H, s), 2.49 (1H, sept, J=7.0 Hz, 15-H), 2.76 (1H, dd, J=8.5, 3.5 Hz, 7-H), 5.03 (1H, tq, J=7.0, 1.5 Hz, 11-H), 6.00 (1H, dd, J=7.8, 6.2 Hz, 14-H), 6.18 (1H, d, J=11.0 Hz, 2-H), 6.24 (1H, br d, J=11.0 Hz, 3-H). UV $\lambda_{\text{max}}^{\text{EIOH}}$ nm (ε): 250 (17400). IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 1735, 1650, 1610. MS m/z: 346 (M⁺), 331, 286, 243, 137, 109. High-resolution MS [Found (Calcd)] m/z: C₂₂H₃₄O₃ (M⁺), 346.24956 (346.25073).

Conversion of 2a to 3a and 3b—A mixture of 2a (200 mg) and p-toluenesulfonic acid (17 mg) in benzene (40 ml) was stirred at 55° C for 15 min then washed with H_2O , saturated NaHCO₃ solution, H_2O , and saturated NaCl

solution, then the solvent was evaporated off. Repeated column chromatography of the residue over silica gel with ethyl acetate-hexane (1:3) afforded 3a (32 mg) and 8a (10 mg). 3a, oil. ¹H-NMR δ : 1.07 and 1.16 (each 3H, d, J= 7.0 Hz, 16,17-H), 1.62 (3H, t, J = 1.0 Hz), 1.68 (3H, s), 1.72 (3H, s), 2.02 (3H, s), 2.55 (1H, sept, J = 7.0 Hz, 15-H), 2.87 (1H, ddd, J = 13.7, 11.5, 8.5 Hz, 10-H), 3.85 (1H, dd, J = 11.0, 4.8 Hz, 7-H), 5.01 (1H, ddd, J = 11.5, 4.0, 1.0 Hz), 5.33(1H, t, J = 8.4 Hz), 5.76 (1H, br d, J = 11.7 Hz, 3-H), 5.82 (1H, dd, J = 11.4, 3.3 Hz, 14-H), 6.18 (1H, d, J = 11.7 Hz, 2-Hz)H). ¹³C-NMR δ: C-1 (141.9), C-2, C-3 (121.6, 123.9), C-4 (135.2), C-5, C-6 (26.7, 29.4), C-7 (77.9), C-8, C-12 (130.8, 132.3), C-9,11 (127.9, 125.3), C-10 (36.5), C-13 (42.3), C-14 (73.3), C-15 (28.0), C-16, C-17 (24.4, 25.4), C-18 (14.3), C-18 (14.3), C-19 (14.3), C 19 (9.4), C-20 (18.1), OAc (21.4, 170.3). Assignments were based on the structural similarity and dissimilarity with 1b. UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (ϵ): 254 (16500). IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 3400, 1730. MS m/z: 346 (M⁺), 286, 243, 225, 203, 137, 109. Compound 8a was identical with the compound obtained from 2b in several TLC systems. Compound 3a (10 mg) was treated with α-phenylbutyric anhydride as shown for 1a. The resultant excess acid in 1 ml of benzene showed the rotation of $+0.05^{\circ}$ and hence 7R configuration of 3a. 10) The blank test showed rotation of less than $\pm 0.01^{\circ}$. Hydrolysis of 3a by 2.5% KOH in MeOH followed by the usual work-up gave 3b. Oil, $[\alpha]_D + 100^{\circ} (c = 0.65, CHCl_3)$. ¹H-NMR δ : 1.12 and 1.15 (each 3H, d, J = 7.0 Hz, 16,17-H), 1.66 and 1.63 (each 3H, s, 19,20-H), 1.72 (3H, s, 18-H), 2.62 (1H, sept, J =7.0 Hz, 15-H), 2.85 (1H, ddd, J = 13.6, 11.4, 8.4 Hz, 10-H), 3.86 (1H, dd, J = 11.0, 5.0 Hz, 7-H), 4.82 (1H, dd, J = 11.0, 3.2 Hz, 14-H), 5.01 (1H, br dd, J = 11.5, 3.5 Hz), 5.26 (1H, br t, J = 8.4 Hz), 5.70 (1H, br d, J = 11.7 Hz, 3-H), 6.14 (1H, J = 11.7 Hz, J =d, J = 11.7 Hz, 2-H). UV $\lambda_{\text{max}}^{\text{EiOH}}$ nm (ϵ): 254 (23000). MS (m/z): 304 (M⁺), 286, 271, 261, 243, 235, 217, 137.

Conversion of 2b to 8a and 8b——A solution of 2b (310 mg) in benzene (40 ml) was treated with p-toluenesulfonic acid (23 mg) at 55 °C for 15 min and then the mixture was worked up as above. Silica gel column chromatography of the mixture with ethyl acetate-hexane (1:3) gave 210 mg of 8a as an oil. ¹H-NMR δ : 1.05 and 1.15 (each 3H, d, J= 6.8 Hz, 16,17-H), 1.48 (3H, s), 1.64 (3H, t, J=1.0 Hz), 1.72 (3H, br s, 18-H), 2.04 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.04 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.04 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.04 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.04 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.04 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.04 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.94 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.94 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.94 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.94 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.94 (3H, s), 2.90 (1H, dt, J=13.7, 9.6 Hz), 1.72 (3H, br s, 18-H), 2.94 (3H, s), 2.94 (3H, s10-H), 3.95 (1H, brt, J = 7.3 Hz, 7-H), 5.2—5.5 (2H, m), 5.73 (1H, dd, J = 10.3, 4.2 Hz, 14-H), 5.89 (1H, brd, J = 10.3, 4.2 Hz, 14-H, 14-H, 14-Hz, 1 11.7 Hz, 3-H), 6.16 (1H, d, J = 11.7 Hz, 2-H). ¹³C-NMR δ : C-1 (141.8), C-2,3 (121.8, 123.1), C-4 (134.9), C-5,6 (27.3, 29.8), C-7 (80.0), C-8 (136.5), C-9,11 (125.3, 125.8), C-10 (36.6), C-12 (130.0), C-13 (42.1), C-14 (74.3), C-15 (28.8), C-10 (136.5), C-10 (136.5), C-10 (136.5), C-10 (136.5), C-10 (136.5), C-10 (136.6), C-12 (130.0), C-13 (136.5), C-14 (74.3), C-15 (136.5), C-10 (136.5), C-10 (136.5), C-10 (136.6), C-12 (130.0), C-13 (136.5), C-14 (136.5), C-15 (13 16,17 (23.8, 25.2), C-18 (15.1), C-19 (10.5), C-20 (19.0). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (ϵ): 254 (15500). IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 3400, 1735. MS m/z: 346 (M⁺), 328, 286, 271, 243, 225, 137, 109. Compound 8a (10 mg) was treated with α -phenylbutyric anhydride as described for 1a. The resultant excess acid in 1 ml of benzene showed the rotation of -0.07° , and hence 7S configuration of 8a.¹⁰⁾ The blank test showed rotation of less than ± 0.01 °. Hydrolysis of 8a by 2.5% KOH in MeOH followed by usual work-up gave **8b**, $[\alpha]_D - 15^{\circ} (c = 0.95, CHCl_3)$. ¹H-NMR δ : 1.11 and 1.15 (each 3H, d, J = 7.0 Hz, 16,17-H), 1.49, 1.64, 1.73 (each 3H, s), 2.53 (1H, sept, $J = 7.0 \,\mathrm{Hz}$, 15-H), 2.89 (1H, dt, J = 13.5, 9.5 Hz, 10-H), 3.96 (1H, dd, J=9.5, 4.0 Hz, 7-H), 4.76 (1H, br dd, J=10.0, 3.5 Hz, 14-H), 5.24 (1H, br dd, J=9.0, 6.5 Hz), 5.36 (1H, br dd, J = 9.5, 7.5 Hz), 5.88 (1H, br d, J = 11.5 Hz, 3-H), 6.12 (1H, d, J = 11.5 Hz, 2-H). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (ϵ): 254 (18000). MS m/z: 304 (M⁺), 286, 271, 261, 243, 235, 217, 137.

Glycolation of 2a and 2b—(a) A solution of 2a (250 mg) in 4 ml of acetone was treated for 5 min at room temperature with 0.6 ml of HClO₄ solution made from 0.15 ml of 70% perchloric acid and 5 ml of H₂O. The mixture was diluted with Et₂O and washed with H₂O and saturated NaCl solution, then the solvent was evaporated off. Column chromatography of the residue with Et₂O–CHCl₃ (15:85) gave 4a (200 mg) as an oil, $[\alpha]_D$ + 160° (c=0.81, CHCl₃). ¹H-NMR δ: 1.07 and 1.19 (each 3H, d, J=7.0 Hz, 16,17-H), 1.13 (3H, s, 19-H), 1.55 (3H, s, 20-H), 1.77 (3H, s, 18-H), 2.03 (3H, s), 2.57 (1H, sept, J=7.0 Hz, 15-H), 3.54 (1H, br t, J=7.0 Hz, 7-H), 5.56 (1H, br dd, J=9.8, 3.0 Hz, 11-H), 5.73 (1H, dd, J=9.8, 2.8 Hz, 14-H), 6.07 (1H, br d, J=11.2 Hz, 3-H), 6.27 (1H, d, J=11.2 Hz, 2-H). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (ε): 251 (16000). IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 3500, 1720. Hydrolysis of 4a with 2.5% KOH in MeOH gave 4b (180 mg). Oil, $[\alpha]_D$ + 140° (c=1.0, CHCl₃). ¹H-NMR δ: 1.13, 1.18 (each 3H, d, J=7.0 Hz, 16,17-H), 1.12 (3H, s, 19-H), 1.56 (3H, s, 20-H), 1.77 (3H, s, 18-H), 2.64 (1H, sept, J=7.0 Hz, 15-H), 3.49 (1H, m, 7-H), 4.74 (1H, br d, J=9.2 Hz, 14-H), 5.47 (1H, br dd, J=9.5, 4.0 Hz, 11-H), 6.02 (1H, br d, J=11.2 Hz, 3-H), 6.24 (1H, d, J=11.2 Hz, 2-H). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (ε): 251 (15500). IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 3350. MS m/z: 322 (M⁺), 304, 289, 137, 109. High-resolution MS [Found (Calcd)] m/z: $C_{20}H_{34}O_3$ (M⁺), 322.25206 (322.25076).

(b) Compound **2b** (200 mg) was treated in the same way as shown in (a) and gave 173 mg of **9a**. Oil, $[\alpha]_D + 118^{\circ}$ (c = 0.80, CHCl₃). ¹H-NMR δ : 1.09 and 1.18 (each 3H, d, J = 7.0 Hz, 16,17-H), 1.13 (3H, s, 19-H), 1.58 (3H, s, 20-H), 1.70 (3H, s, 18-H), 2.07 (3H, s), 2.45 (1H, sept, J = 7.0 Hz, 15-H), 3.45 (1H, dd, J = 10.3, 1.8 Hz, 7-H), 5.41 (1H, brt, J = 7.0 Hz, 11-H), 5.64 (1H, dd, J = 7.7, 4.8 Hz, 14-H), 6.19 (2H, s, 2,3-H). UV $\lambda_{\text{max}}^{\text{E10H}}$ nm (ϵ): 251 (15000). IR $\nu_{\text{max}}^{\text{neat}}$ cm ⁻¹: 3500, 1720. Hydrolysis of **9a** as in (a) gave the triol **9b** quantitatively. Oil, $[\alpha]_D - 105^{\circ}$ (c = 1.24, CHCl₃). ¹H-NMR δ : 1.10, 1.14 (each 3H, d, J = 7.0 Hz, 16,17-H), 1.09 (3H, s, 19-H), 1.57 (3H, s, 20-H), 1.71 (3H, s, 18-H), 3.60 (1H, dd, J = 10.0, 3.0 Hz, 7-H), 4.48 (1H, m, 14-H), 5.43 (1H, br dd, J = 9.0, 4.5 Hz, 11-H), 6.16 (1H, d, J = 11.0 Hz, 2-H), 6.45 (1H, br d, J = 11.0 Hz, 3-H). UV $\lambda_{\text{max}}^{\text{E10H}}$ nm (ϵ): 250 (16000). MS m/z: 322 (M⁺), 304, 289, 286, 261, 243, 235, 222, 137, 109.

Dehydration of 4c and 9c—(a) Compound 4c was prepared from 4a in a usual way (Ac₂O-pyridine). A solution of 4c (74.3 mg) in 0.5 ml of pyridine was treated with 40 μ l of thionyl chloride at room temperature for 1 h. After addition of H₂O, the mixture was extracted with Et₂O. The extract was washed with H₂O, 5% HCl solution, saturated NaHCO₃ solution, H₂O, and saturated NaCl solution, then the solvent was evaporated off. The residue was hydrolyzed with 2.5% KOH in MeOH (30 min at room temperature) and the mixture was worked up as usual. Silica

gel column chromatography of the residue with ethyl acetate-hexane (1:4) gave **3b** (20 mg) and **5** (20 mg). **5**, $[\alpha]_D + 200^{\circ}$ (c = 1.50, CHCl₃). ¹H-NMR δ : 1.11 and 1.14 (each 3H, d, J = 7.0 Hz, 16,17-H), 1.53 (3H, s, 20-H), 1.77 (3H, s, 18-H), 2.62 (1H, sept, J = 7.0 Hz, 15-H), 3.87 (1H, br t, J = 6.0 Hz, 7-H), 4.87 (1H, dd, J = 10.0, 4.0 Hz, 14-H), 4.99 and 5.06 (each 1H, s, 19-H), 5.19 (1H, br t, J = 6.0 Hz, 11-H), 5.89 (1H, br d, J = 11.3 Hz, 3-H), 6.19 (1H, d, J = 11.3 Hz, 2-H). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (ε): 250 (22000). IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 3350, 1005, 910. MS m/z: 304 (M⁺), 286, 271, 261, 243, 235, 137, 109. Compound **3b** ([α]_D + 95°) was identical with that obtained from **2a** as judged from UV, ¹H-NMR and mass spectra, and in several TLC systems.

(b) Compound **9c** was prepared in a usual way (Ac₂O-pyridine) from 200 mg of **9a**. It was dissolved in 2 ml of pyridine and 150 μ l of thionyl chloride was added with cooling. After standing at room temperature for 1 h, the mixture was worked up as in (a). Silica gel column chromatography of the hydrolysis product as shown in (a) gave **8b** (40 mg) and **10** (105 mg). Compound **8b**, $[\alpha]_D - 16.1^\circ$ (c = 0.62, CHCl₃), was identical with that obtained from **2b** as judged from UV, ¹H-NMR, and mass spectra, and in several TLC systems. **10**, oil, $[\alpha]_D + 150^\circ$ (c = 0.71, CHCl₃). ¹H-NMR δ : 1.13 and 1.15 (each 3H, d, J = 7.0, 16,17-H), 1.56 (3H, s, 20-H), 1.70 (3H, s, 18-H), 2.55 (1H, sept, J = 7.0 Hz, 15-H), 4.04 (1H, br t, J = 6.5 Hz, 7-H), 4.78 (1H, dd, J = 9.0, 4.0 Hz, 14-H), 4.92 and 5.04 (each 1H, s, 19-H), 5.32 (1H, t, J = 7.0 Hz, 11-H), 6.07 (1H, br d, J = 10.5 Hz, 3-H), 6.13 (1H, d, J = 10.5 Hz, 2-H). UV λ_{max}^{EtOH} nm (ϵ): 250 (15000). MS m/z: 304 (M⁺), 286, 271, 261, 243, 235, 217, 137, 109.

Lithium Aluminum Hydride Reduction of 2a and 2b——(a) LiAlH₄ (300 mg) was added in portions to a solution of 2a (1 g) in dry Et₂O (10 ml). After 3 h, excess reagent was decomposed with moist Et₂O and the Et₂O layer was washed with H₂O, 5% HCl, H₂O, and saturated NaCl solution, then the solvent was evaporated off. Column chromatography of the residue with ethyl acetate—hexane (1:9) gave 6b (300 mg). Oil, [α]_D – 30° (c=0.64, CHCl₃). ¹H-NMR δ: 1.12 and 1.13 (each 3H, d, J=7.0 Hz, 16,17-H), 1.14 (3H, s, 19-H), 1.58 (3H, s, 20-H), 1.69 (3H, s, 18-H), 4.60 (1H, m, 14-H), 5.35 (1H, br t, J=7.0 Hz, 11-H), 6.16 (1H, d, J=11.2 Hz, 2-H), 6.29 (1H, br d, J=11.2 Hz, 3-H). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (ε): 251 (17700). IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 3400. MS m/z: 306 (M⁺), 288, 273, 270, 255, 245, 137. High-resolution MS [Found (Calcd)] m/z: C₂₀H₃₄O₂ (M⁺), 306.25717 (306.25587). Acetylation of 6b under usual conditions (Ac₂O-pyridine) gave 6a. ¹H-NMR δ: 1.07 and 1.17 (each 3H, d, J=7.0 Hz, 16,17-H), 1.17 (3H, s, 19-H), 1.57 (3H, d, J=1.0 Hz, 20-H), 1.69 (3H, s, 18-H), 2.07 (3H, s), 2.48 (1H, sept, J=7.0 Hz, 15-H), 5.31 (1H, br t, J=7.0 Hz, 11-H), 5.75 (1H, dd, J=8.8, 4.0 Hz, 14-H), 6.07 (1H, br d, J=10.6 Hz, 3-H), 6.19 (1H, d, J=10.6 Hz, 2-H). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (ε): 251 (15000). IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 3500, 1730, 1650, 1605, 1240. MS m/z: 348 (M⁺), 288, 270, 255, 245, 227, 137.

(b) A solution of 2b (300 mg) in tetrahydrofuran (10 ml) was treated with lithium aluminum hydride (100 mg) at room temperature for 1 h. Excess reagent was decomposed by adding moist Et₂O and the mixture was worked up as in (a). Separation of the residue was difficult, so the whole residue was acetylated. Silica gel column chromatography of the acetate mixture with ethyl acetate—hexane (1:4) gave 12a, 12'a and 11a in that order, as oils. 11a, 1 H-NMR δ : 1.06 and 1.19 (each 3H, d, J = 7.0 Hz, 16,17rH), 1.12 (3H, s, 19-H), 1.53 (3H, s, 20-H), 1.69 (3H, s, 18-H), 2.03 (3H, s), 2.56 (1H, sept, J = 7.0 Hz, 15-H), 5.39 (1H, br t, J = 7.0 Hz, 11-H), 5.81 (1H, dd, J = 10.0, 3.7 Hz, 14-H), 6.07 (1H, br d, $J = 11.0 \text{ Hz}, 3-\text{H}), 6.22 (1\text{H}, d, J = 11.0 \text{ Hz}, 2-\text{H}). \text{ MS } m/z: 348 (M^+), 330, 288, 255, 245, 227, 137. 12a, ^1\text{H-NMR } \delta:$ 0.92 (3H, d, J = 6.6 Hz, 19-H), 1.04 and 1.18 (each 3H, d, J = 7.0 Hz, 16,17-H), 1.58 (3H, s, 20-H), 1.78 (3H, s, 18-H), 2.01 and 2.04 (each 3H, s), 2.57 (1H, sept, J = 7.0 Hz, 15-H), 4.68 (1H, dq, J = 10.5, 2.5 Hz, 7-H), 5.17 (1H, t, J =7.0 Hz, 11-H), 5.91 (1H, dd, J = 11.0, 3.3 Hz, 14-H), 6.07 (1H, br d, J = 11.4 Hz, 3-H), 6.24 (1H, d, J = 11.4 Hz, 2-H). MS m/z: 390 (M⁺), 330, 270, 255, 227, 137. **12'a**, ¹H-NMR δ : 0.82 (3H, d, J=6.6 Hz, 19-H), 1.07 and 1.16 (each 3H, d, J = 7.0 Hz, 16,17-H), 1.65 (3H, s, 20-H), 1.72 (3H, s, 18-H), 2.01 and 2.04 (each 3H, s), 2.48 (1H, sept, J = 7.0 Hz, 15-H), 4.78 (1H, dq, J = 9.8, 2.0 Hz, 7-H), 5.07 (1H, br t, J = 7.2 Hz, 11-H), 5.73 (1H, t, J = 6.5 Hz, 14-H), 6.17 (2H, s, 2,3-H). MS m/z: 390 (M⁺), 330, 270, 255, 227, 137. They were hydrolyzed with 2.5% KOH in MeOH, giving 11b, 12b and 12'b. 11b, mp 102—104°C, $[\alpha]_D$ +240° (c=0.67, CHCl₃). ¹H-NMR δ : 1.13 and 1.19 (each 3H, d, J=7.0 Hz, 16,17-H), 1.13 (3H, s, 19-H), 1.54 (3H, s, 20-H), 1.69 (3H, s, 18-H), 2.65 (1H, sept, J=7.0 Hz, 15-H), 4.82 (1H, dd, J=10.0), 1.13 (3H, s, 19-H), 9.8, 3.0 Hz, 14-H), 5.37 (1H, br dd, J = 9.0, 4.8 Hz, 11-H), 5.97 (1H, br d, J = 11.0 Hz, 3-H), 6.19 (1H, d, J = 11.0 Hz, 2-10 Hz, 3-H), 6.19 (1H, d, J = 11.0 Hz, 2-10 Hz, 3-H), 6.19 (1H, d, J = 11.0 Hz, 2-10 Hz, 3-H), 6.19 (1H, d, J = 11.0 Hz, 2-10 Hz, 3-H), 6.19 (1H, d, J = 11.0 Hz, 3-H), 6.19 (1H, d, J = 11.0 Hz, 2-10 Hz, 3-H), 6.19 (1H, d, J = 11.0 Hz, 6.10 Hz, 6 H). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (ϵ): 251 (17000). IR $\nu_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 3350, 1650, 1605, 1100, 1005, 920. MS m/z: 306 (M⁺), 288, 273, 270, 255, 245, 227, 137. High-resolution MS [Found (Calcd)] m/z: C₂₀H₃₄O₂ (M⁺), 306.25527 (306.25577). 12b, mp 77— 78 °C, $[\alpha]_D$ +140 ° (c=0.42, CHCl₃). ¹H-NMR δ : 0.84 (3H, d, J=7.0 Hz, 19-H), 1.13 and 1.17 (each 3H, d, J= 7.0 Hz, 16,17-H), 1.53 (3H, s, 20-H), 1.76 (3H, s, 18-H), 2.64 (1H, sept, J = 7.0 Hz, 15-H), 3.49 (1H, m, 7-H), 4.91 (1H, m, 7-H), $4.91 (1\text{H}, \text$ br d, J = 11.0 Hz, 14-H), 5.16 (1H, br t, J = 7.0 Hz, 11-H), 5.95 (1H, br d, J = 11.2 Hz, 3-H), 6.19 (1H, d, J = 11.2 Hz, 2-Hz, 2 H). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (ϵ): 252 (15600). IR $\nu_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 3350, 1005, 930, 840. MS m/z: 306 (M⁺), 291, 288, 273, 237, 224, 137, 109. 12'b, oil, $[\alpha]_D + 49^{\circ} (c = 0.70, CHCl_3)$. H-NMR $\delta : 0.77 (3H, d, J = 7.0 Hz, 19-H), 1.13 and 1.15 (each 3H, d, d) = 0.12'b, oil, <math>[\alpha]_D + 49^{\circ} (c = 0.70, CHCl_3)$. J = 7.0 Hz, 16,17-H, 1.56 (3H, s, 20-H), 1.71 (3H, s, 18-H), 2.46 (1H, sept, J = 7.0 Hz, 15-H), 3.58 (1H, brt, J = 6.0 Hz, 15-Hz)7-H), 4.71 (1H, br d, J = 7.8 Hz, 14-H), 5.14 (1H, br t, J = 7.0 Hz, 11-H), 6.15 (1H, d, J = 11.4 Hz, 2-H), 6.18 (1H, br d, J = 11.4 Hz, 2-H), 6.18 (1 J = 11.4 Hz, 3-H). UV $\lambda_{\text{max}}^{\text{EtOH}} \text{ nm}$ (ϵ): 252 (19500). IR $\nu_{\text{max}}^{\text{neat}} \text{ cm}^{-1}$: 3400, 1005, 975, 900. MS m/z: 306 (M⁺), 291, 288, 273, 237, 224, 137, 109.

Lithium Aluminum Deuteride Reduction of 2a—Compound 2a (1g) was treated with lithium aluminum deuteride (300 mg) as described above and the product was acetylated in the usual way, giving 250 mg of 6c.

Dehydration of 6a and 6c—(a) Phosphoryl chloride (200 µl) was added to a solution of 6a (190 mg) in pyridine

(3 ml) with cooling and the mixture was kept at room temperature. After 1.5 h, the excess reagent was decomposed with a small amount of H_2O and the reaction mixture was diluted with Et_2O . The Et_2O solution was washed with 5% HCl, H_2O , and saturated NaCl solution, then the solvent was evaporated off to give 170 mg of 1b with about 90% purity. A portion of this was purified over a column of 10% silver nitrate-impregnated silica gel with ethyl acetate-hexane (1:19) giving 1b, which was identical with natural 1b based on comparisons of IR and ¹H-NMR spectra, and also in several TLC systems. ¹H-NMR δ : 1.04 and 1.07 (each 3H, d, J=7.0 Hz, 16,17-H), 1.47 (3H, s, 19-H), 1.57 (3H, s, 20-H), 1.73 (3H, d, J=1.0 Hz, 18-H), 2.50 (1H, sept, J=7.0 Hz, 15-H), 4.98 (1H, brt, J=7.0 Hz, 7-H), 5.07 (1H, brt, J=7.0 Hz, 11-H), 6.03 (1H, dd, J=9.5, 4.4 Hz, 14-H), 6.10 (1H, br d, J=11.5 Hz, 3-H), 6.18 (1H, d, J=11.5 Hz, 2-H). The experiment was repeated on a reduced scale with 50 mg of 6c, giving 1b with 25% deuterium incorporation at C-7.

- (b) Compound 6a (50 mg) in 0.5 ml of pyridine was treated with 30 μ l of thionyl chloride with cooling. After 1 h, the mixture was worked up as in (a). TLC (ethyl acetate-hexane, 1:19) of the crude product showed virtually the same pattern as in (a) and 1b was the predominant product. The experiment was repeated on a reduced scale with 50 mg of 6c, giving 1b with 38% deuterium incorporation at C-7.
- (c) A solution of **6c** (50 mg) in 5 ml of benzene was evaporated to ca. 3 ml to remove moisture. Thionyl chloride (40 μ l) was added to the refluxing solution and the mixture was cooled immediately. Usual work-up followed by purification as in (a) gave 25 mg of 7. The ¹H-NMR spectrum was identical with that of **1b**, except for the lack of a signal at δ 4.98.

Chromic Acid Oxidation of 1a——In a typical run, Jones' reagent (40 ml) was added slowly to a solution of 1a (5 g) in Et₂O (75 ml) with cooling. The mixture was stirred for 5 min, then the Et₂O layer was separated. It was washed with H_2O , then with 1 N NaOH solution ($10 \text{ ml} \times 3$), H_2O , and saturated NaCl solution. Evaporation of the solvent gave 3.48 g of the neutral fraction. The 1 N NaOH extract was acidified with concentrated HCl and extracted with Et₂O. It was washed with H_2O , and saturated NaCl solution, then the solvent was evaporated off to give 0.67 g of the carboxylic acid mixture. A portion (1.67 g) of the carboxylic acid mixture was treated briefly with ethereal diazomethane solution and then the solvent was evaporated off. Column chromatography of the residue with ethyl acetate—hexane (1:19 to 1:4) gave 15a (53 mg), 14a (38 mg), and 13a (450 mg), in order of elution. Column chromatography of a portion (2.0 g) of the neutral fraction with ethyl acetate—hexane (1:19 to 1:4) gave 19 (50 mg), 18 (95 mg), 20a (98 mg), 16 (1.0 g) and 17 (34 mg) in order of elution.

Compound 13b—Oil, [α]_D + 3.6 ° (c = 1.00, CHCl₃). ¹H-NMR δ: 1.13 (6H, d, J = 6.8 Hz, 16,17-H), 1.34 (3H, s, 18-H), 1.59 (3H, br s), 1.68 (3H, br d, J = 0.7 Hz), 2.81 (1H, sept, J = 6.8 Hz, 15-H), 3.00 (2H, s, 13-H), 3.67 (3H, s), 5.14 and 5.26 (each 1H, br t, J = 7.0 Hz), 6.41 (1H, d, J = 15.6 Hz, 2-H), 6.88 (1H, d, J = 15.6 Hz, 3-H). UV $\lambda_{\text{max}}^{\text{EiOH}}$ nm (ε): 222 (12000). IR $\nu_{\text{max}}^{\text{neat}}$ cm $^{-1}$: 3450, 1735, 1690, 1668, 1625, 984. MS m/z: 350 (M $^{+}$), 335, 332, 317, 301, 289, 261. High-resolution MS [Found (Calcd)] m/z: $C_{21}H_{34}O_{4}$ (M $^{+}$), 350.2438 (350.2457).

Compound 14b—Oil, [α]_D 0° (c = 0.70, CHCl₃). ¹H-NMR δ: 1.32 (3H, s, 18-H), 1.58 (3H, br s), 1.68 (6H, br s), 1.70 (3H, s), 2.98 (2H, br s, 13-H), 3.67 (3H, s), 5.15 (1H, br t, J = 7.0 Hz), 5.28 (1H, br t, J = 7.0 Hz), 5.91 (1H, d, J = 5.9 Hz, 2-H), 6.26 (1H, d, J = 5.9 Hz, 3-H). ¹³C-NMR δ: C-1 (153.6), C-2,3 (121.7, 136.4), C-4 (90.8), C-5,9 (39.3, 40.6), C-6 (22.8), C-7 (124.7), C-8 (134.6), C-10 (26.8), C-11 (129.2), C-12 (128.1), C-13 (44.9), C-14 (172.6), C-15 (98.5), C-18 (26.1), C-16,17,19,20 (15.9, 16.3, 17.0, 18.9). UV $\lambda_{\text{max}}^{\text{EIOH}}$ nm (ε): 276 (10000). IR $\nu_{\text{max}}^{\text{neat}}$ cm $^{-1}$: 1736, 1680, 1580, 1157, 847. MS m/z: 332 (M⁺), 317, 314, 301, 123. High-resolution MS [Found (Calcd)] m/z: C₂₁H₃₂O₃ (M⁺), 332.2333 (332.2351).

Compound 15b—Oil. ¹H-NMR δ : 1.64 and 1.71 (each 3H, br s), 2.14 (3H, s, 18-H), 2.99 (2H, s, 13-H), 3.67 (3H, s), 5.09 and 5.24 (each 1H, br t, $J=7.0\,\text{Hz}$). IR $v_{\text{max}}^{\text{neat}}$ cm⁻¹: 1736, 1715, 1158. MS m/z: 252 (M⁺), 194, 178, 151, 135, 125, 107. High-resolution MS [Found (Calcd)] m/z: $C_{15}H_{24}O_3$ (M⁺), 252.1717 (252.1725).

Compound 16—Oil, $[\alpha]_D$ – 3.0° (c = 1.00, CHCl₃). ¹H-NMR δ: 1.13 (6H, d, J = 6.8 Hz, 16,17-H), 1.34 (3H, s, 18-H), 1.60 (3H, br s), 1.67 (3H, br d, J = 0.7 Hz), 2.82 (1H, sept, J = 6.8 Hz, 15-H), 3.03 (2H, br d, J = 2.5 Hz, 13-H), 5.14 (1H, br t, J = 6.8 Hz), 5.28 (1H, br t, J = 7.1 Hz), 6.40 (1H, d, J = 15.6 Hz, 2-H), 6.88 (1H, d, J = 15.6 Hz, 3-H), 9.96 (1H, t, J = 2.5 Hz, 14-H). ¹³C-NMR δ: C-1 (204.0), C-2 (130.4), C-3 (152.0), C-4 (73.3), C-5 (41.9), C-6 (22.7), C-7,11 (124.7, 124.3), C-8 (135.3), C-9 (39.2), C-10 (26.6), C-12 (126.4), C-13 (54.2), C-14 (200.6), C-15 (39.2), C-16,17 (18.3), C-18 (27.9), C-19,20 (16.0, 16.9). UV λ_{\max}^{EiOH} nm (ε): 227 (10000). IR ν_{\max}^{neat} cm⁻¹: 3450, 1725, 1694, 1667, 1630, 985. MS m/z: 320 (M⁺), 305, 302, 259, 231, 205, 165. High-resolution MS [Found (Calcd)] m/z: C₂₀H₃₂O₃ (M⁺), 320.2347 (320.2351).

Compound 17—Oil, [α]_D 0 ° (c = 1.0, CHCl₃). ¹H-NMR δ: 1.13 (6H, d, J = 6.8 Hz, 16,17-H), 1.35 (3H, s, 18-H), 1.59 (3H, br s, 19-H), 1.97 and 2.17 (total 1H, br d, J = 1.2 Hz, 20-H), 2.81 (1H, sept, J = 6.8 Hz, 15-H), 5.15 (1H, br t, J = 7.3 Hz, 11-H), 5.88 (1H, br d, J = 8.1 Hz, 13-H), 6.40 (1H, d, J = 15.6 Hz, 2-H), 6.88 (1H, d, J = 15.6 Hz, 3-H), 9.99 and 9.92 (total 1H, d, J = 8.1 Hz, 14-H). ¹³C-NMR δ: C-1 (203.9), C-2 (130.8), C-3 (151.8), C-4 (73.2), C-5 (41.9), C-6 (22.6), C-7 (124.7), C-8 (134.9, 132.4), C-9 (39.0, 39.3), C-10 (26.8), C-11 (40.0, 25.2), C-12 (164.2, 164.7), C-13 (124.7, 128.8), C-14 (190.7, 191.2), C-15 (39.3), C-16,17 (18.3), C-18 (28.0), C-19 (15.9), C-20 (17.5, 24.9). UV $\lambda_{\text{max}}^{\text{EIOH}}$ nm (ε): 234 (12000). IR $\nu_{\text{max}}^{\text{neat}}$ cm $^{-1}$: 3350, 1670, 1630, 982. MS m/z: 320 (M $^+$), 302, 259, 231, 207. High-resolution MS [Found (Calcd)] m/z: $C_{20}H_{32}O_3$ (M $^+$), 320.2346 (320.2351).

Compound 18—Oil. ¹H-NMR δ : 1.13 (6H, d, J=6.8 Hz, 16,17-H), 1.57 and 1.61 (each 3H, s, 19,20-H), 2.13

(3H, s, 18-H), 2.80 (1H, sept, J=6.8 Hz, 15-H), 3.25 (2H, br s, 13-H), 4.9—5.2 (2H, m, 7,11-H), 6.25 (1H, d, J=2.0 Hz, 2-H), 7.23 (1H, d, J=2.0 Hz, 3-H). 13 C-NMR δ : C-1 (126.6), C-2 (108.6), C-3 (140.1), C-4 (208.5), C-5 (43.7), C-6,10 (22.5, 26.6), C-7,11 (122.7, 125.6), C-8 (136.0), C-9 (39.4), C-12 (132.2), C-13 (36.2), C-14 (147.4), C-15 (24.4), C-16,17 (23.8), C-18 (29.9), C-19,20 (16.0). UV $\lambda_{\text{max}}^{\text{EiOH}}$ nm (ϵ): 214 (7000). IR $\nu_{\text{max}}^{\text{neat}}$ cm $^{-1}$: 1715, 1672, 1605. MS m/z: 302 (M $^+$), 223, 205, 189, 177, 176, 149. High-resolution MS [Found (Calcd)] m/z: $C_{20}H_{30}O_2$ (M $^+$), 302.2228 (302.2225).

Compound 19—Oil. ¹H-NMR and IR spectra were identical with those described in our previous report. ¹³⁾ Compound 20a—Oil, $[\alpha]_D$ – 133 ° (c = 0.63, CHCl₃). ¹H-NMR δ : 1.02 (3H, s, 18-H), 1.03 and 1.17 (each 3H, d, J = 6.8 Hz, 16,17-H), 1.56 and 1.58 (each 3H, t, J = 1.5 Hz, 19,20-H), 2.46 (1H, dd, J = 13.6, 5.5 Hz, 13-H), 4.70 (1H, dq, J = 5.5, 1.0 Hz, 3-H), 4.86 (1H, br t, J = 5.5 Hz, 14-H), 5.12 (1H, br d, J = 9.2 Hz), 5.28 (1H, br d, J = 10.3 Hz), 5.41 (1H, q, J = 1.0 Hz, 2-H). ¹³C-NMR δ : C-1 (151.0), C-2 (131.2), C-3 (87.2), C-4 (74.7), C-5,9 (38.6, 39.8), C-6,10 (22.1, 25.4), C-7,11 (128.5, 119.4), C-8 (132.4), C-12 (129.8), C-13 (43.1), C-14 (84.5), C-15 (26.6), C-16,17 (21.0, 22.1), C-18 (23.8), C-19,20 (15.7, 17.7). IR $\nu_{\text{max}}^{\text{neat}}$ cm ⁻¹: 3460, 1085, 975, 955. MS m/z: 304 (M⁺), 286. High-resolution MS [Found (Calcd)] m/z: $C_{20}H_{32}O_2$ (M⁺), 304.2402 (304.2416).

Compound 20b—A mixture of 20a (61.5 mg), dimethylaminopyridine (3 mg), and triethylamine (0.1 ml) was treated with Ac₂O (0.1 ml) at 110 °C for 1.5 h, then extracted with Et₂O and worked up as usual. Silica gel column chromatography of the crude product with ethyl acetate-hexane (1:40) gave 29.2 mg of 20b, mp 62—63 °C, $[\alpha]_D$ –97 ° (c=0.76, CHCl₃). ¹H-NMR δ : 1.03 and 1.19 (each 3H, d, J=7.0 Hz, 16,17-H), 1.27 (3H, s, 18-H), 1.55 and 1.56 (each 3H, s, 19,20-H), 2.30 (1H, br sept, J=7.0 Hz, 15-H), 4.75 (1H, br d, J=5.3 Hz, 3-H), 4.85 (1H, br t, J=5.5 Hz, 14-H), 5.15 (1H, br d, J=8.4 Hz), 5.31 (1H, m), 5.32 (1H, br s, 2-H). IR ν_{max}^{neat} cm⁻¹: 1735, 1245, 1087, 847, 822. MS m/z: 346 (M⁺), 303, 286, 243, 218, 150.

Compound 21——A solution of 16 (150 mg) in 0.5% KOH in MeOH (3 ml) was kept at room temperature for 1 h. Extraction with Et₂O followed by usual work-up gave a mixture, which, on silica gel column chromatography (Et₂O–CHCl₃, 1:4), gave 50 mg of 17. It was further treated at room temperature with 0.5% KOH in MeOH for 2 h. Extraction with Et₂O, usual work-up, and chromatography in the same way as above gave 21 as an oil, [α]_D + 3 ° (c = 1.00, CHCl₃). ¹H-NMR δ: 1.33 (3H, s, 18-H), 1.57, 1.68, 1.70 (each 3H, s, 16,17,19-H), 2.16 and 1.97 (total 3H, d, J = 1.2 Hz, 20-H), 5.13 (1H, br t, J = 7.0 Hz, 7-H), 5.88 (1H, br d, J = 8.2 Hz, 13-H), 5.91 and 6.27 (each 1H, d, J = 6.0 Hz, 2,3-H), 9.93 and 9.99 (total 1H, J = 8.2 Hz, 14-H). ¹³C-NMR δ: C-1 (153.4), C-2,3 (121.7, 136.2), C-4 (90.6), C-5 (40.5), C-6 (22.7), C-7 (125.3), C-8 (133.6, 133.9), C-9 (39.0, 39.2), C-10 (26.8), C-11 (25.3, 40.0), C-12 (164.0, 164.5), C-13 (127.3, 128.5), C-14 (190.6, 191.1), C-15 (98.5), C-16,17,19 (15.7, 17.0, 18.8), C-18 (26.1), C-20 (17.5, 25.5). UV $\lambda_{\text{max}}^{\text{EtoH}}$ nm (ε): 240 (12000), 276 (8000). IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 1715, 1672, 1627, 1580, 950, 845. MS m/z: 302 (M⁺), 287, 273, 205, 137, 123. High-resolution MS [Found (Calcd)] m/z: C₂₀H₃₀O₂ (M⁺), 302.2247 (302.2225).

Chromic Acid Oxidation of 1b—Jones' reagent (8 ml) was added slowly to a solution of 1b (2 g) in 30 ml of Et_2O and the mixture was stirred at room temperature for 22 h. The Et_2O layer was separated, washed with H_2O , 2 N NaOH solution (5 ml × 3), H_2O , and saturated NaCl solution. Evaporation of the solvent gave 1.6 g of neutral fraction. Acidification of the NaOH extract followed by extraction with Et_2O gave 64 mg of acid mixture. Silica gel column chromatography of the neutral fraction with Et_2O -CHCl₃ (1:9) gave first unreacted 1b (0.78 g), then 22a (50 mg), an unidentified material (70 mg), and 23a (290 mg) in order of elution.

Compound 22a—Oil, [α]_D +6° (c=1.00, CHCl₃). ¹H-NMR δ: 1.09 and 1.16 (each 3H, d, J=7.0 Hz, 16,17-H), 1.61 and 1.66 (each 3H, br s), 2.05 (3H, s), 2.14 (3H, s, 18-H), 4.92—5.28 (2H, m), 5.90 (1H, d, J=7.8 Hz, 2-H), 5.99 (1H, dd, J=7.8, 5.6 Hz, 14-H), 10.19 (1H, d, J=7.8 Hz, 3-H). UV $\lambda_{\text{max}}^{\text{EIOH}}$ nm (ε): 234 (16000). IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 1740, 1714, 1668. MS m/z: 302 (M⁺ – AcOH), 284, 259. High-resolution MS [Found (Calcd)] m/z: C₂₀H₃₀O₂ (M⁺ – AcOH), 302.2261 (302.2245).

Compound 23a—Oil, [α]_D -33° (c = 1.00, CHCl₃). ¹H-NMR δ: 0.81 and 0.91 (each 3H, d, J = 6.8 Hz, 16,17-H), 1.44 (3H, s, 18-H), 1.62 and 1.77 (each 3H, br s), 2.06 (3H, s), 5.14 (1H, dd, J = 9.0, 6.2 Hz, 14-H), 5.26 (1H, br t, J = 7.0 Hz), 5.34 (1H, br t, J = 8.0 Hz), 5.74 and 6.11 (each 1H, d, J = 16.1 Hz, 2,3-H). ¹³C-NMR δ: C-1 (79.2), C-2,7,11 (128.4, 128.6, 129.0), C-3 (137.1), C-4 (72.2), C-5,13 (43.7, 42.4), C-6,10 (22.4, 23.7), C-8,12 (132.6, 132.7), C-9 (39.0), C-14 (71.5), C-15 (32.9), C-16,17 (16.2, 16.7), C-18 (27.9), C-19,20 (14.8, 15.4), OAc (21.9, 169.8). IR $v_{\text{max}}^{\text{neat}}$ cm $^{-1}$: 3500, 1730, 1720, 980. MS m/z: 364 (M +), 346, 286, 261, 243. High-resolution MS [Found (Calcd)] m/z: $C_{22}H_{34}O_3$ (M + $-H_2O$), 346.2518 (346.2508).

Conversion of 22a to 18—A solution of 22a (20 mg) in 0.5% KOH in MeOH solution (2 ml) was kept at 40 °C for 30 min and then poured into water. The mixture was extracted with Et_2O , the extract was worked up as usual, and the solvent was evaporated off to give 15 mg of residue. The residue (10 mg) was dissolved in 0.1 N HCl in MeOH and kept at room temperature for 30 min. It was extracted with Et_2O , and worked up as usual. Silica gel column chromatography of the evaporation residue with ethyl acetate-hexane mixture gave 8 mg of oil which was shown to be identical with 18 by comparisons of IR and 1H -NMR spectra, and by TLC.

Conversion of 23a to 16—A solution of 23a (30 mg) in 0.5% KOH in MeOH solution (2 ml) was kept at 40 °C for 30 min and then poured into H_2O . The mixture was extracted with Et_2O , the extract was worked up as usual, and the solvent was evaporated off to give 20 mg of 23b. ¹H-NMR δ : 0.77 and 0.96 (each 3H, d, J=6.6 Hz, 16,17-H), 1.35 (3H, s, 18-H), 1.54 and 1.64 (each 3H, br s), 3.98 (1H, dd, J=9.0, 5.9 Hz, 14-H), 5.0—5.4 (2H, m), 5.69 and 6.01 (each 1H, d, J=16.1 Hz, 2,3-H). Jones' reagent (1 ml) was added slowly to a solution of 23b (10 mg) in Et_2O (2 ml) and the

mixture was stirred at room temperature for 10 min. The mixture was diluted with Et₂O, and the Et₂O layer was washed with H₂O, and saturated NaCl solution, then the solvent was evaporated off. Silica gel column chromatography of the residue with ethyl acetate—hexane mixture gave 6 mg of oil, shown to be identical with 16 by comparisons of IR and ¹H-NMR spectra, and by TLC.

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