## New Glucosides of a 4-Hydroxy-5-methylcoumarin and a Dihydro-α-pyrone from Gerbera jamesonii hybrida

Seiji Nagumo, Tatsunori Toyonaga, Takao Inoue, and Masahiro Nagai\*.b

Faculty of Pharmaceutical Sciences<sup>a</sup> and Institute of Medicinal Chemistry,<sup>b</sup> Hoshi University, Ebara 2–4–41, Shinagawa-ku, Tokyo 142, Japan. Received March 13, 1989

Two new glucosides, 2,  $C_{16}H_{18}O_9$ , mp 202—203 °C,  $[\alpha]_D^{20}$  – 96.4° (MeOH) and gerberin (5),  $C_{12}H_{18}O_8$ ,  $[\alpha]_D^{19}$  +11.2° (MeOH), were isolated from the aerial part of *Gerbera jamesonii hybrida* (Compositae). On methanolysis, 2 yielded 6-hydroxy-4-methoxy-5-methylcoumarin. The tetraacetate of 5 yielded tetraacetylparasorboside on catalytic hydrogenation. The chemical structures of 2 and 5 were established as 4- $\beta$ -D-glucopyranosyloxy-6-hydroxy-5-methylcoumarin and (6S)-5,6-dihydro-4- $\beta$ -D-glucopyranosyloxy-6-methyl-2*H*-pyran-2-one, respectively. The yield of gerberin (5) was quite high, amounting to about 3.7% based on the weight of the dried plant material.

**Keywords** Gerbera jamesonii hybrida; Compositae; 4-hydroxy-5-methylcoumarin; 5-methylcoumarin; dihydro- $\alpha$ -pyrone; gerberin

Gerbera (Gerbera jamesonii hybrida, Compositae) is widely cultivated as a garden plant. We have already reported the isolation of two 4-hydroxy-5-methylcoumarin glycosides in addition to two cyanogenic glycosides from the underground parts of this plant. In the previous paper we performed a biosynthetic study on  $4-\beta$ -D-glucopyranosyloxy-5-methylcoumarin (1), one of the constituents of this plant, and proposed that its coumarin skeleton is formed *via* a pentaketide derived from the acetatemalonate pathway. This paper deals with the structure determination of two new glycosides 2 and 5, the latter being named gerberin. These were isolated from the aerial

TABLE I. 13C-NMR Chemical Shifts

| Solvent            | 1<br>C <sub>5</sub> D <sub>5</sub> N | 2<br>DMSO-d <sub>6</sub> | 3<br>DMSO-d <sub>6</sub> | 5<br>C <sub>5</sub> D <sub>5</sub> N | 6 <sup>a)</sup><br>CDCl <sub>3</sub> | 7<br>CDCl <sub>3</sub> |
|--------------------|--------------------------------------|--------------------------|--------------------------|--------------------------------------|--------------------------------------|------------------------|
| C-2                | 162.0                                | 161.5                    | 161.7                    | 166.5                                | 166.0                                | 167.1                  |
| C-3                | 94.1                                 | 93.0                     | 91.4                     | 94.6                                 | 95.1                                 | 90.3                   |
| C-4                | 167.1                                | 166.8                    | 168.7                    | 170.4                                | 170.5                                | 172.6                  |
| C-5                | 137.7                                | 121.1                    | 121.1                    | 34.1                                 | 34.0                                 | 34.6                   |
| C-6                | 127.8                                | 151.7                    | 151.4                    | 72.0                                 | 72.9                                 | 72.2                   |
| C-7                | 131.7                                | 119.5                    | 119.1                    |                                      |                                      |                        |
| C-8                | 115.1                                | 114.4                    | 114.1                    |                                      |                                      |                        |
| C-9                | 155.2                                | 147.5                    | 148.1                    |                                      |                                      |                        |
| C-10               | 114.8                                | 114.4                    | 114.9                    |                                      |                                      |                        |
| CH <sub>3</sub> -C | 23.7                                 | 13.2                     | 13.2                     | 20.5                                 | 20.6                                 | 20.6                   |
| CH <sub>3</sub> -O |                                      |                          |                          |                                      |                                      | 55.9                   |
| Glucose-1          | 101.3                                | 99.8                     |                          | 100.5                                | 96.5                                 |                        |
| -2                 | 74.3                                 | 73.0                     |                          | 74.2                                 | 70.7                                 |                        |
| -3                 | $79.1^{b)}$                          | $77.3^{b)}$              |                          | $78.9^{b}$                           | $72.5^{b}$                           |                        |
| -4                 | 70.7                                 | 69.5                     |                          | 70.7                                 | 68.1                                 |                        |
| -5                 | $78.6^{b}$                           | $76.5^{b}$               |                          | $78.1^{b}$                           | $72.2^{b)}$                          |                        |
| -6                 | 61.9                                 | 60.6                     |                          | 62.0                                 | 61.7                                 |                        |

a) The chemical shifts of four acetyl groups were eliminated. b) Assignments may be interchanged in each column.

parts of this plant together with 1,  $\alpha$ - and  $\beta$ -amyrin, prunasin, and amygdalin.

Compound 2,  $C_{16}H_{18}O_9$ , mp 202—203 °C,  $[\alpha]_D^{20} - 96.4^\circ$ (MeOH) showed absorption maxima at 225, 277 (sh), 284, 295 (sh) and 339 nm in its ultraviolet (UV) spectrum, and absorptions due to hydroxy groups (3310 cm<sup>-1</sup>), an aromatic ring (1610, 1560 cm<sup>-1</sup>), and an unsaturated lactone (1640 cm<sup>-1</sup>) in its infrared (IR) spectrum. The <sup>13</sup>C-nuclear magnetic resonance (13C-NMR) spectrum of 2 showed sixteen signals comprising a coumarin skeleton (six singlets and three doublets), a methyl group, and a  $\beta$ -glucopyranosyloxy group. These spectra are similar to those of 1, but 2 has an additional oxygen atom in the coumarin skeleton of 1. In the <sup>1</sup>H-NMR spectrum, 2 showed a one-proton doublet at  $\delta$  5.44 ppm (J=5.8 Hz, anomeric proton of the glucose residue), a three-proton singlet at  $\delta$  2.50 ppm, a one-proton singlet at  $\delta$  5.90 ppm, and a pair of orthocoupled aromatic protons at  $\delta$  7.06 and 7.12 ppm (J=8.9 Hz). These findings suggested that 2 is a 6- or 8hydroxylated 4-hydroxy-5-methylcoumarin glucoside.

On enzymatic hydrolysis using emulsin 2 afforded glucose and an aglycone (3),  $C_{10}H_8O_4$ , mp 258—260 °C. From the following observation, the glucosyl moiety of 2 could be deduced to be attached to C-4 of the aglycone (3). The <sup>1</sup>H-NMR spectrum of 3 showed a singlet at  $\delta$  5.52 ppm ascribable to the proton on C-3 (3-H) and this signal disappeared on addition of deuterium oxide, while the corresponding signal at  $\delta$  5.90 ppm in the spectrum of 2 was unaffected by the proton exchange. Moreover, the UV absorption maxima of 3 were observed at 221 and 288 nm, resembling those of 2 except for the appearance of bathochromic shifts with sodium acetate.

In order to clarify the position of the hydroxy group in 2, the chemical shifts of the aromatic carbons were calculated on the basis of the additivity rule of substituent effects for  $1.^{3)}$  The observed chemical shifts of 2 are in better agreement with the calculated ones for 6-hydroxylation of 1 rather than those for 8-hydroxylation. Namely, the hydroxylated aromatic carbon and its protonated *ortho* carbon were observed at  $\delta$  151.7 and 119.5 ppm, respectively: in 6-hydroxylation of 1, those carbon signals are expected to appear at  $\delta$  154.7 and 119.0 ppm; in 8-hydroxylation, at  $\delta$  142.0 and 129.2 ppm. In fact, treatment of 2 with *p*-TsOH in MeOH gave a methoxy derivative (4),  $C_{11}H_{10}O_4$ , mp

Table II. <sup>1</sup>H-NMR Data and Results of Spin Decoupling Experiments (400 MHz)<sup>a)</sup>

| Com-<br>pound          | 3-Н       | 5-H   | 6-H         | 6-CH <sub>3</sub> | CH <sub>3</sub> O- | Coupling constant  |
|------------------------|-----------|---|-------------|-------------------|--------------------|--|
| <b>6</b> <sup>a)</sup> | 5.29<br>d | H <sub>a</sub> 2.35<br>dd<br>H <sub>b</sub> 2.51<br>ddd |             | 1.45<br>d         |                    | $J_{3,5b} = 1.47, J_{5a,5b} = 17.33,$<br>$J_{5b,6} = 11.23, J_{5a,6} = 3.91,$<br>$J_{6,CH_3} = 6.34$ |
| 7                      | 5.14<br>d | H <sub>a</sub> 2.34<br>dd<br>H <sub>b</sub> 2.47<br>ddd | 4.53<br>ddq | 1.45<br>d         |                    | $J_{3,5b} = 1.47, J_{5a,5b} = 17.09, J_{5a,6} = 3.91, J_{5b,6} = 11.48, J_{6,CH_3} = 6.35$           |

a) Measured in CDCl<sub>3</sub>. b) The data for the tetraacetylglucosyl moiety are omitted.

264—265 °C. The physicochemical data of **4** were identical with the reported values for 6-hydroxy-4-methoxy-5-methylcoumarin, which is one of the constituents of *Gerbera jamesonii*.<sup>4)</sup> Consequently **2** was concluded to be 4- $\beta$ -D-glucopyranosyloxy-6-hydroxy-5-methylcoumarin.

Gerberin (5), amorphous powder,  $[\alpha]_D^{19} - 11.2^{\circ}$  (MeOH) showed an absorption band at 232 nm in its UV spectrum and absorptions in the IR spectrum assignable to an  $\alpha$ ,  $\beta$ -unsaturated lactone (1680,  $1630\,\mathrm{cm}^{-1}$ ). Since 5 was acetylated to a tetraacetate (6),  $C_{20}H_{26}O_{12}$ , mp 175—176°C,  $[\alpha]_D^{21} + 11.6^{\circ}$  (MeOH), the molecular formula of 5 was determined as  $C_{12}H_{18}O_8$ . In the <sup>13</sup>C-NMR spectrum of 5, twelve carbon signals were observed, comprising a methyl, a methylene, a methine joined to oxygen, two olefinic carbons (a singlet and a doublet), a lactone carbonyl and six carbons of the  $\beta$ -glucopyranosyl moiety. These spectral data led us to conclude that 5 is a glucoside of a dihydro- $\alpha$ -pyrone such as osmundalin isolated from Osmunda japonica.<sup>5)</sup>

Spin-decoupling experiments on the tetraacetate (6) were performed and the results are summarized in Table II. A proton at  $\delta$  2.51 ppm (5-H<sub>b</sub>) coupled with three protons resonating at  $\delta$  2.35 (5-H<sub>a</sub>,  $^2J$ =17.33 Hz), 4.80 (6-H,  $^3J$ =11.23 Hz) and 5.29 ppm (3-H,  $^4J$ =1.47 Hz). This observation revealed that the chemical structure of the tetraacetate (6) of gerberin is as shown by 6 in Chart 1.

In order to derive it into a known compound, **6** was submitted to methanolysis in the presence of an acid catalyst, yielding a methoxy derivative 7,  $C_7H_{10}O_3$ , mp 59—60 °C,  $[\alpha]_D^{17} + 256^\circ$  (MeOH), whose <sup>1</sup>H- and <sup>13</sup>C-NMR spectra indicated that its chemical structure is 5,6-dihydro-4-methoxy-6-methyl-2*H*-pyran-2-one. The spectral data of 7 were coincident with the reported values of an authentic sample, which is known as a racemate.<sup>6)</sup> In the circular dichroism (CD) spectrum of **6** a positive Cotton effect was observed at 256 nm, which revealed that the  $\alpha,\beta$ -unsaturated lactone chromophore has left-handed chirality from Beecham's rule,<sup>7)</sup> and at the same time the absolute con-

figuration at C-6 should be S. Moreover, **6** was subjected to catalytic hydrogenation, and a dihydro derivative **8**,  $C_{20}H_{28}O_{12}$ , mp 150—151 °C,  $[\alpha]_D^{15}$  –16.1° (acetone), was isolated. The physicochemical properties of **8** were identical with those of tetraacetylparasorboside, an acetate of parasorboside isolated from *Sorbus aucuparia*.<sup>8)</sup> Accordingly, the chemical structure of gerberin (**5**) is (6S)-5,6-dihydro-4- $\beta$ -D-glucopyranosyloxy-6-methyl-2*H*-pyran-2-one.

Compounds related to 7 are biologically active in insects and fungi,<sup>9)</sup> so synthetic studies have been performed.<sup>6)</sup> Since this plant contains a large amount of gerberin (5) (in a 3.7% yield by dry weight), it might be of value as a natural resouce for synthetic studies.

## **Experimental**

All melting points were taken on a Yanagimoto melting point determination apparatus and are uncorrected. Optical rotations were measured with a JASCO DIP-181 automatic polarimeter in a 1 dm tube. IR spectra were obtained with a Shimadzu IR-400 spectrometer. UV spectra were recorded on a Shimadzu UV-250. Nuclear magnetic resonance (NMR) spectra were recorded with a JEOL JNM-FX-100 or JNM-GX-400 spectrometer with tetramethylsilane as an internal standard; chemical shifts are given on the  $\delta$  scale (ppm), coupling constants (J values) are expressed in herz (Hz), and the following abbreviations are used. sesinglet, d=doublet, t=triplet, q=quartet and m=multiplet. Mass spectra (MS) were recorded with a JEOL JMS-D 300 machine. The CD spectrum was measured with a JASCO J-40A in a 1 cm tube. Thin-layer chromatography (TLC) was performed on Kieselgel 60  $F_{254}$  pre-coated plates (Merck) and detection was carried out by UV absorption.

Isolation and Separation Commercial Gerbera jamesonii hybrida in bloom was used as the plant material. The air-dried aerial parts of the plant (272 g) were extracted with MeOH (21×7) under reflux. The total MeOH solution was concentrated under reduced pressure. Hot water (1.41) was added to the residue (81.7g), and the mixture was separated into a water-soluble part and an insoluble part. The insoluble part (10 g) afforded a mixture of  $\alpha$ - and  $\beta$ -amyrin on silica gel column chromatography and the two compounds were identical with authentic samples on gas chromatography. The hot water-soluble part was applied to a column of polyamide (polyamide C-200, Wako Pure Chemical Industry). The column was eluted with water to give fractions (frs.) 1-20 (each 11). Fractions 6-20 were combined and concentrated under reduced pressure to about 1 l, giving 4-β-D-glucopyranosyloxy-5-methylcoumarin (1, 1.5 g) as a precipitate. The total aqueous solution of frs. 1-5 was applied to a column of Amberlite XAD-2 and the column was eluted with water (5 l) and then with MeOH (51). The water eluate was further applied to a column of Amberlite XAD-4. After being washed with water, the column was eluted with MeOH. The residue obtained on concentration of the MeOH eluate was chromatographed over Sephadex LH-20 with water to give 5 (9.8 g). The MeOH eluate of the Amberlite XAD-2 column was repeatedly chromatographed over silica gel, affording 2 (103.6 mg), amygdalin (66.5 mg) and prunasin (663 mg). The known compounds (1, amygdalin, and prunasin) were identified by direct comparison with authentic samples

**Compound 2** Colorless needles (MeOH), mp 202—203 °C,  $[\alpha]_{20}^{20}-96.4^{\circ}$  (c=1.0, MeOH). Anal. Calcd for  $C_{16}H_{18}O_{9}$ : C, 54.24; H, 5.12. Found: C, 54.10; H, 5.10. IR  $v_{max}^{KBr}$  cm<sup>-1</sup>: 3310 (OH), 1640 (C=O), 1610, 1560 (arom.). UV  $\lambda_{max}^{MeOH}$  nm (log  $\varepsilon$ ): 225 (4.29), 277 (sh, 4.05), 284 (4.09), 295 (sh, 3.96), 339 (3.55). (unchanged on addition of NaOAc). ¹H-NMR (DMSO- $d_{6}$ ) δ: 2.50 (3H, s, CH<sub>3</sub>-) 5.44 (1H, d, J=5.8, 1-H of glucose moiety), 5.90 (1H, s, 3-H), 7.06, 7.12 (each 1H, d, J=8.9, 7 or 8-H), 9.64 (1H, s, 6-OḤ, disappeared on addition of D<sub>2</sub>O).

Enzymatic Hydrolysis of 2 An EtOH solution (5 ml) of 2 (36 mg) was added to a solution of emulsion in Na<sub>2</sub>HPO<sub>4</sub>-citric acid buffer (pH 5.0, 15 ml), and the mixture was stirred for 24 h at 37 °C. The EtOH in the mixture was removed *in vacuo*. The resulting aqueous solution was extracted with EtOAc (10 ml × 4). The total EtOAc solution was washed with H<sub>2</sub>O, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated to afford 3 (11 mg). The water-soluble fraction of the hydrolysate was passed through a column of Amberlite MB-3, and concentrated to a small volume. Glucose was detected on TLC (Cellulose F<sub>254</sub> (Merck), BuOH–AcOH–H<sub>2</sub>O (6:1:2), visualized with aniline–H<sub>3</sub>PO<sub>4</sub>). 3, pale yellow needles

(MeOH–H<sub>2</sub>O), mp 258–260 °C. *Anal.* Calcd for  $C_{10}H_8O_4$ : C, 62.50; H, 4.20. Found: C, 62.36; H, 4.14. MS m/z: 192 (M<sup>+</sup>), 150 (M<sup>+</sup> – CH<sub>2</sub>CO), 122 (150–CO). IR  $v_{max}^{KBr}$  cm<sup>-1</sup>: 3150, 1630, 1610, 1460. UV  $\lambda_{max}^{MeOH}$  nm (log  $\varepsilon$ ): 221 (4.50), 288 (4.07);  $\lambda_{max}^{MeOH+NaOAc}$  nm: 297 (bathochromic shift). <sup>1</sup>H-NMR (DMSO- $d_6$ )  $\delta$ : 5.52 (1H, s, 3-H, disappeared on addition of D<sub>2</sub>O) 6.98, 7.11 (each 1H, d, J=9.0, 7- or 8-H), 9.48 (1H, s, 6-OH, disappeared on addition of D<sub>2</sub>O).

Methanolysis of 2 A solution of 2 (20 mg) and p-TsOH (120 mg) in anhydrous MeOH (8 ml) was refluxed for 4 h. After concentration to dryness, the residue was chromatographed over silica gel. Fractions eluted with CHCl<sub>3</sub> gave 4 (6 mg). 4, pale yellow needles (MeOH-H<sub>2</sub>O), mp 264—265 °C. High-resolution MS m/z: Calcd for  $C_{11}H_{10}O_4$  (M \*): 206.058. Found: 206.057. UV  $\lambda_{\max}^{\text{MeOH}}$  nm: 222, 273 (sh), 280, 290, 332. <sup>1</sup>H-NMR (DMSO- $d_6$ )  $\delta$ : 2.46 (3H, s, CH<sub>3</sub>-arom.), 3.95 (CH<sub>3</sub>-O, 5.75 (1H, s, 3-H), 7.05, 7.11 (each 1H, d, J = 8.9, 7- or 8-H), 9.50 (6-OH). The MS, IR, and <sup>1</sup>H-NMR data of 4 were identical with those reported for 6-hydroxy-4-methoxy-5-methylcoumarin. <sup>4</sup>)

**Gerberin (5)** Colorless amorphous powder,  $[\alpha]_D^{19}-11.2^{\circ}$  (c=1.2, MeOH). IR  $\nu_{\rm max}^{\rm KBr}$  cm $^{-1}$ : 3390 (OH), 1680, 1630 ( $\alpha,\beta$ -unsaturated lactone). UV  $\lambda_{\rm max}^{\rm MeOH}$  nm ( $\log \varepsilon$ ): 232 (4.05).

Acetylation of 5 5 (500 mg) was acetylated with Ac<sub>2</sub>O-pyridine. After work-up in the usual manner, the tetraacetate (6, 490 mg) was obtained. 6, colorless needles (MeOH), mp 175—176 °C, [α] $_{21}^{21}$  +11.6° (c=1.0, MeOH). Anal. Calcd for C<sub>20</sub>H<sub>26</sub>O<sub>12</sub>: C, 52.40; H, 5.72. Found: C, 52.40; H, 5.72. CI-MS m/z: 459 (M<sup>+</sup>+H). UV  $\lambda_{\rm meOH}^{\rm meOH}$  nm (log  $\varepsilon$ ): 229 (4.04). CD (c=5.8 × 10<sup>-3</sup>, MeOH) [ $\theta$ ] (nm): +2.57 × 10<sup>4</sup> (256).

Acid Hydrolysis of 5 A solution of 5 (500 mg) in 10% HCl (10 ml) was refluxed for 30 min. The solution was passed through a column of Amberlite MB-3, and processed in the same manner as in the case of 2. Glucose was identified as a sugar product.

Methanolysis of 6 A solution of 6 (200 mg) and *p*-TsOH (50 mg) in absolute MeOH was refluxed for 7 h and treated in the same way as in the case of 2. The product (7, 85 mg) was purified by sublimation. 7, mp 59—60 °C, [α]<sub>1</sub><sup>17</sup> +256° (c=0.5, MeOH). UV  $\lambda_{\rm max}^{\rm MeOH}$  nm (log  $\varepsilon$ ): 233 (4.04). Highresolution MS m/z: Calcd for  $C_7H_{10}O_3$  ( $M^+$ ): 142.063. Found: 142.063. IR  $\nu_{\rm max}^{\rm KBr}$  cm<sup>-1</sup>: 1690, 1625. CD (c=3.9 × 10<sup>-3</sup>, MeOH) [ $\theta$ ]<sup>20</sup> (nm): +3.65 × 10<sup>4</sup> (253). The UV, IR, and <sup>1</sup>H-NMR spectral data of 7 were coincident with those of dl-5,6-dihydro-4-methoxy-6-methyl-2H-pyran-2-one.<sup>6)</sup>

Catalytic Hydrogenation of 6 A solution of 6 (1 g) in EtOAc (20 ml) was hydrogenated at atmospheric pressure in the presence of 5% Pd-C

(100 mg) until absorption of hydrogen ceased. After removal of the catalyst by filtration, the solvent was evaporated off *in vacuo*. The residue was recrystallized from 50% EtOH to give **8** (180 mg). **8**, mp 150—151 °C,  $[\alpha]_D^{21}-20.7^\circ$  (c=0.9, MeOH),  $[\alpha]_D^{20}-16.1^\circ$  (c=0.6, acetone). *Anal.* Calcd for  $C_{20}H_{28}O_{12}$ : C, 52.19; H, 6.13. Found: C, 52.03; H, 6.22. <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.42 (3H, d, J=6.10, 6-CH<sub>3</sub>), 1.57 (1H, ddd, J=10.25, 11.71, 13.67, 5-Ha or Hb), 2.23 (1H, dddd, J=1.46, 2.70, 5.05, 13.67, 5-Hb or Ha), 2.59 (1H, dd, J=8.30, 17.58, 3-Ha or Hb), 2.98 (1H, ddd, J=1.46, 6.35, 17.58, 3-Hb or Ha), 4.15 (1H, m, 4-H), 4.33 (1H, ddq, J=2.70, 6.10, 11.71, 6-H). The MS, IR, and <sup>1</sup>H-NMR data of **8** coincided with those of tetraacetylparasorboside. (lit.<sup>8</sup>) mp 157—158 °C,  $[\alpha]_D^{20}-17^\circ$  (acetone)).

## References and Notes

- S. Nagumo, K. Imamura, T. Inoue, and M. Nagai, Chem. Pharm. Bull., 33, 4803 (1985).
- T. Inoue, Toyonaga, S. Nagumo, and M. Nagai, *Phytochemistry*, in press (1989).
- 3) P. Diehl, Helv. Chim. Acta, 44, 829 (1961).
- A. F. Halim, E. M. Marwan, and F. Bohlmann, *Phytochemistry*, 19, 2496 (1980).
- 5) K. H. Hollenbeak and M. E. Kuehne, Tetrahedron, 30, 2307 (1974).
- a) R. Bacardit and M. Moreno-Mañas, Tetrahedron Lett., 1980, 551;
  b) R. Hänsel, H. Rimpler, and L. Langhammer, Fresenius Zeitschrift fur Analytische Chemie, 218, 346 (1966);
  c) L. Cattel, O. Caputo, G. Nobili, and G. Biglino, Ann. Chim. (Rome), 62, 655, (1972);
  d) R. Hänsel, L. Langhammer, and H. Rimpler, Arch. Pharm. Ber. Dtsch. Pharm. Ges., 300, 157 (1967);
  e) A. Ceruti and P. Amistadi, Ann. Chim. (Rome), 62, 607 (1972).
- 7) A. F. Beecham, Tetrahedron, 28, 5543 (1972).
- 8) a) R. Tschesche, H.-J. Hoppe, G. Snatzke, G. Wulff, and H.-W. Fehlhaber, Chem. Ber., 104, 1420 (1971); b) The melting point of 7 is lower by six degrees than that reported for tetraacetyl-parasorboside, <sup>8a</sup> though their spectral data and optical rotations coincided with each other. As the original specimen of tetraacetylparasorboside was unavailable to the authors, direct comparison could not be carried out.
- 9) a) R. Hänsel, D. Weiss, and B. Schmidt, *Planta Med.*, Heft 1, 1 (1966); b) A. Numata, K. Hokimoto, T. Takemura, T. Katsuno, and K. Yamamoto, *Chem. Pharm. Bull.*, 32, 2815 (1984).