New Coumarins from Citrus Plant

Chihiro Ito, a Shiho Tanahashi, Mitsuo Omura and Hiroshi Furukawa*.a

Faculty of Pharmacy, Meijo University, a Tempaku, Nagoya 468, Japan and Okitsu Branch, Fruit Tree Research Station, Ministry of Agriculture Forestry and Fisheries, Okitsu, Shimizu, Shizuoka 424–02, Japan. Received February 9, 1989

New coumarins, kiyomal (1), casegravol isovalerate (3), and $trans\ O$ -methylgrandmarin (5) were isolated from several hybrid seedlings resulting from crosses of $Citrus\ unshiu \times C$. $sinensis \times C$. iyo and characterized by means of spectroscopic analysis and/or derivation from known coumarins.

Keywords Citrus; Rutaceae; coumarin; isovalerate; casegravol isovalerate; trans O-methylgrandmarin; kiyomal

During our phytochemical studies of Citrus plants, 1) new coumarins, kiyomal (1), casegravol isovalerate (3), and trans O-methylgrandmarin (5), were isolated from roots of several hybrid seedlings resulting from crosses of [Citrus unshiu MARC. (miyagawa-wase) \times C. sinensis OSBEC. (torobita orange)] (kiyomi) \times C. iyo HORT. ex TANAKA (iyo).

Structure of Kiyomal (1) Kiyomal (1) was obtained as a colorless amorphous solid with the molecular formula C₁₂- $H_{10}O_4$ as judged from the high-resolution mass spectrum (MS). In the proton nuclear magnetic resonance (1H-NMR) spectrum of kiyomal, a 3H singlet at δ 3.91 and two pairs of AB-type doublets at δ 7.66 and 6.27 (each 1H, J=9.4 Hz) and δ 7.43 and 6.89 (each 1H, J=8.7 Hz) together with a strong ultraviolet (UV) band at λ_{max} 321 nm and infrared (IR) absorptions at v_{max} 1610 and 1730 cm⁻¹ suggested the presence of a 7-methoxy-8-substituted coumarin nucleus in the molecule.2) Observations of a lower field 1H signal at δ 9.75 and a 2H signal at 3.98 coupled with each other ($J=1.0\,\mathrm{Hz}$), and a strong IR band at v_{max} 1730 cm⁻¹ revealed the structure of the side chain at C-8 as -CH₂-CHO. Next, 7-methoxy-8-allylcoumarin (2) from 7hydroxycoumarin was treated with water saturated ozonized oxygen³⁾ in ethyl acetate to give the aldehyde (1).⁴⁾ which was found to be identical with natural kiyomal by comparisons of the ¹H-NMR and IR spectra. On the basis of these results, kiyomal can be represented by formula 1, and has not previously been found as a natural product.

Structure of Casegravol Isovalerate (3) Casegravol isovalerate (3) was isolated as a colorless amorphous solid, $C_{20}H_{24}O_6$, $[\alpha]_D$ 0° (chloroform). The presence of the same 7methoxy-8-substituted coumarin nucleus as in 1 was shown by ¹H-NMR signals at δ 7.62 (1H, d, J=9.4 Hz), 7.32 (1H, d, J=8.7 Hz), 6.87 (1H, d, J=8.7 Hz), 6.26 (1H, d, J=8.7 Hz) 9.4 Hz), and 3.95 (3H, s), a characteristic UV band at λ_{max} 321 nm, and IR bands at v_{max} 1730 and 1600 cm⁻¹. An ABtype doublet at δ 7.07 and 6.90 with a characteristic coupling constant $(J=16.5 \,\mathrm{Hz})$ was assigned to a transdisubstituted double bond, a 3H singlet at δ 1.45 to a methyl group attached to a carbon bearing an oxygen atom, and two doublets at δ 4.23 and 4.10 (each 1H) coupled with each other (J=11.1 Hz) to methylene protons located between an oxygen and a quaternary carbon atom. These ¹H-NMR spectral features of this coumarin were similar to those of casegravol (4),5 except for a downfield shift of the methylene protons. The appearance of signals at δ 0.95 (6H, d, J=6.1 Hz), 2.11 (1H, m), and 2.26 (2H, m), an IR band at v_{max} 1730 cm⁻¹ and a mass fragment base peak at m/z 245 [M⁺-115 (C₆H₁₁O₂)] suggested the

presence of an isovaleryl moiety as an ester. From these spectral data, we proposed the structure (3) for casegravol isovalerate.

Structure of trans O-Methylgrandmarin (5) trans O-Methylgrandmarin (5) was obtained as a colorless amorphous solid, $C_{16}H_{18}O_6$, $[\alpha]_D - 13.4^{\circ}$ (chloroform). A 5,7oxygenated 8-substituted coumarin nucleus was indicated by the ¹H-NMR (acetone-d₆), IR, and UV spectral data as follows: δ 8.04 and 6.15 (coupled with each other, 1H, d, $J=9.8\,\mathrm{Hz})$ and 6.31 (1H, s), v_{max} 1730, 1635, and 1610 cm⁻¹, and λ_{max} 259 and 328 nm.²⁾ In the ¹H-NMR (acetone- d_6) spectrum, the appearance of methoxy signals at δ 3.96 and 3.64, geminal dimethyl signals at δ 1.43 and 1.48, and a 1H signal at δ 3.98 partially overlapped with a methoxy signal and coupled with protons at 4.43 (1H, d, J=3.0 Hz) and 4.47 (1H, d, J=5.4 Hz, disappeared with D₂O) suggested the presence of a 2',2'-dimethyl-3'hydroxy-4'-methoxydihydropyran ring system, which was also supported by the occurrence of a mass fragment base peak at m/z 235 (M⁺ – 71) due to the loss of (CH₃)₂C= CH(OH) radical from the molecular ion with a hydrogen transfer. 6) The differential chemical shift value between two geminal methyls at C-2' ($\Delta\delta$ 0.05 ppm) and the coupling constant $(J_{3',4'}=3.0 \text{ Hz})$ suggested the trans orientation of substituents at C-3' and 4' in the dimethylpyran ring system.^{6,7)} These findings led us to convert 5-methoxyseselin (7) into trans O-methylgrandmarin. Treatment of 5methoxyseselin $(7)^{8}$ isolated from C. grandis with mchloroperbenzoic acid (m-CPBA) in methanol at room temperature for 1 h gave two diastereomeric isomers 5 and

$$CH_3O$$
 CH_3O
 CH_3

© 1989 Pharmaceutical Society of Japan

205 (85).

6 after preparative silica gel thin-layer chromatography (TLC) in 50 and 15% yields, respectively. Easy cleavage of an oxiran ring of the intermediate followed by reaction with methanol at the benzylic C-4′ was considered to be a consequence of the effect of the methoxy group located *para* to the benzylic position. From analyses of the 1 H-NMR spectra, taking account of the $J_{3',4'}$ values and the differential chemical shift values of 2′-geminal dimethyls, structures **5** and **6** can be deduced for these products (see Experimental). The synthetic *trans* isomer **5** was found to be identical with natural *trans O*-methylgrandmarin by comparisons of the 1 H-NMR and IR spectra. Consequently, the structure of *trans O*-methylgrandmarin was established as formula **5** except for the absolute stereochemistry.

Experimental

Åll melting points were measured on a micromelting point hot-stage apparatus (Yanagimoto). $^1\text{H-}$ and $^{13}\text{C-NMR}$ spectra were recorded on GX-270 (JEOL) and GX-400 (JEOL) spectrometers, respectively, in CDCl₃, unless otherwise stated. Chemical shifts are shown in δ -values (ppm) with tetramethylsilane (TMS) as an internal reference. Electron impact mass spectra (EI-MS) were taken with an M-52 (Hitachi) spectrometer having a direct inlet system, and high-resolution MS with an M-80 (Hitachi) spectrometer. UV spectra were determined in methanol and IR spectra were recorded in CHCl₃.

Extraction and Separation The roots of several hybrid seedlings resulting from crosses of [Citrus unshiu MARC. (miyagawa-wase) \times C. sinensis OSBEC. (torobita orange)] (kiyomi) \times C. iyo HORT. ex TANAKA (iyo) were used as materials. These Citrus plants were grown in the orchard of Okitsu Branch, Fruit Tree Research Station, Ministry of Agriculture, Forestry, and Fisheries, Shimizu, Shizuoka.

The dried roots of *Citrus* plant (kiyomi × iyo) were extracted with acetone. The acetone extract was subjected to silica gel column chromatography. Successive elutions with benzene, benzene—isopropyl ether [(3:1)—(1:3)], and benzene—acetone (3:1) each gave 3—6 fractions. Each fraction was subjected repeatedly to preparative silica gel TLC developed with appropriate combinations of acetone, benzene, hexane, isopropyl ether, and chloroform to obtain kiyomal (1), casegravol isovalerate (3), and *trans O*-methylgrandmarin (5) in 0.0008, 0.0021, 0.001% yields, respectively, as well as known coumarins and other components. Details of the procedures of separation and characterization of known components will be reported

Kiyomal (1) Colorless amorphous solid. High-resolution MS: Calcd for C₁₂H₁₀O₄: 218.0578. Found: 218.0602. UV λ_{max} nm: 205, 219, 248, 257, 321. IR ν_{max} cm⁻¹: 1730, 1610. ¹H-NMR δ: 9.75 (1H, t, J= 1.0 Hz, -CHO), 7.66 (1H, d, J= 9.4 Hz, H-4), 7.43 (1H, d, J= 8.7 Hz, H-5), 6.89 (1H, d, J= 8.7 Hz, H-6), 6.27 (1H, d, J= 9.4 Hz, H-3), 3.98 (2H, d, J=1.0 Hz, benzylic-CH₂), 3.91 (3H, s, OCH₃). MS m/z (%): 218 (M⁺, 40), 190 (22), 189 (100), 131 (64).

Ozonolysis³⁾ of 7-Methoxy-8-allylcoumarin (2) Ozonized oxygen was bubbled through H_2O and then through a solution of 2 (15 mg) in EtOAc (10 ml) at room temperature for 3 min. The solvent was evaporated off and the residue was subjected to preparative TLC (benzene: iso- $Pr_2O = 1:2$) to give 1 as a colorless oil (3.5 mg). This was found to be identical with kiyomal by IR and ¹H-NMR comparisons.

Casegravol Isovalerate (3) Colorless amorphous solid, $[\alpha]_D 0^\circ$ (CHCl₃). High-resolution MS: Calcd for $C_{20}H_{24}O_6$: 360.1572. Found: 360.1591. UV $\lambda_{\rm max}$ nm: 212, 245, 254, 273, 283, 321. IR $\nu_{\rm max}$ cm⁻¹: 3500 (br), 1730, 1600.

¹H-NMR δ: 7.62 (1H, d, J=9.4 Hz, H-4), 7.32 (1H, d, J=8.7 Hz, H-5), 7.07 (1H, d, J=16.5 Hz, H-1′), 6.90 (1H, d, J=16.5 Hz, H-2′), 6.87 (1H, d, J=8.7 Hz, H-6), 6.26 (1H, d, J=9.4 Hz, H-3), 4.23 (1H, d, J=11.1 Hz, H-4′), 4.10 (1H, d, J=11.1 Hz, H-4′), 3.95 (3H, s, OCH₃), 2.38 (1H, br, OH), 1.45 (3H, s, 3-CH₃), [2.26 (2H, m), 2.11 (1H, m), 0.95 (6H, d, J=6.1 Hz); isovaleryl]. MS m/z (%): 360 (M⁺), 258 (7), 246 (17), 245 (100), 203 (21). *trans O-Methylgrandmarin* (5) Colorless amorphous solid, [α]_D – 13.4° (c=0.067, CHCl₃). High-resolution MS: Calcd for C₁₆H₁₈O₆: 306.1102. Found: 306.1186. UV λ_{max} nm: 209, 225, 250, 259, 328. IR ν_{max} cm⁻¹: 3400 (br), 1730, 1635, 1610. ¹H-NMR δ: 7.97 (1H, d, J=9.8 Hz, H-4), 6.23 (1H, s, H-6), 6.17 (1H, d, J=9.8 Hz, H-3), 4.48 (1H, d, J=3.4 Hz, H-4′), 3.88

(1H, br, H-3'), 3.87 (3H, s, OCH₃), 3.71 (3H, s, OCH₃), 1.49 (3H, s, CH₃), 1.45 (3H, s, CH₃). MS m/z (%): 306 (M⁺, 40), 235 (100), 234 (55), 219 (30),

Reaction of 5-Methoxyseselin (10)7) with m-CPBA A methanolic solution (2 ml) of 7 (15 mg) and m-CPBA (30 mg) was stirred at room temperature for 1 h under dry conditions, then aqueous NaHCO₃ solution was added, and the mixture was extracted with CH2Cl2. The CH2Cl2 extract was washed with H2O, dried with anhydrous MgSO4, and evaporated to dryness. The residue was subjected to preparative silica gel TLC (solvent, benzene: acetone = 10:1) to afford 5 (8.9 mg) and 6 (2.6 mg). 5: Colorless needles from acetone, mp 174—177 °C. ¹H-NMR (acetone- d_6) δ : 8.04 (1H, d, J = 9.7 Hz, H-4), 6.31 (1H, s, H-6), 6.15 (1H, d, J = 9.7 Hz, H-3), 4.47 (1H, d, J = 5.4 Hz, OH), 4.43 (1H, d, J = 3.0 Hz, H-4'), 3.98 (1H, dd, J = 5.4, 3.0 Hz, H-3'), 3.96 (3H, s, 5-OCH₃), 3.64 (3H, s, 4'-OCH₃), 1.48 and 1.43 (each 3H, s, 2'-CH₃). $\Delta\delta$ of 2'-(CH₃)₂ = 0.05 ppm. Synthetic 5 was found to be identical with natural trans O-methylgrandmarin by IR and ¹H-NMR comparisons. 6: Colorless needles from acetone, mp 182-185 °C. UV λ_{max} nm: 210, 227, 251, 259, 330. IR v_{max} cm⁻¹: 3450 (br), 1730, 1630, 1605. ¹H-NMR δ : 7.98 (1H, d, J=9.4 Hz, H-4), 6.21 (1H, s, H-6), 6.17 (1H, d, J=9.4 Hz, H-3), 4.65 (1H, d, J=5.1 Hz, H-4'), 3.87 (3H, s, OCH_3), 3.82 (1H, dd, J = 5.1, 7.4 Hz, H-3'), 3.81 (3H, s, OCH_3), 2.94 (1H, d, J = 7.4 Hz, OH), 1.42 (6H, s, $2 \times \text{CH}_3$). ¹H-NMR (acetone- d_6) δ : 8.02 (1H, d, J=9.8 Hz, H-4), 6.28 (1H, s, H-6), 6.14 (1H, d, J=9.8 Hz, H-3),4.58 (1H, d, J=4.7 Hz, H-4'), 4.16 (1H, d, J=7.7 Hz, OH), 3.94 (3H, s, H-4') OCH_3), 3.88 (1H, dd, J=4.7, 7.7 Hz, H-3'), 3.72 (3H, s, OCH_3), 1.43 (3H, s, CH₃), 1.41 (3H, s, CH₃). $\Delta \delta$ of 2'-(CH₃)₂ = 0.02 ppm.

Acknowledgement We thank Mr. K. Masuda of Meijo University for measurements of high-resolution MS. This work was supported in part by a Grant-in-Aid (No. 63571011) for Scientific Research from the Ministry of Education, Science and Culture of Japan.

References

- M. Ju-ichi, H. Kaga, M. Muraguchi, M. Inoue, I. Kajiura, M. Omura and H. Furukawa, *Heterocycles*, 27, 2197 (1988), and cited therein.
- 2) R. D. H. Murray, J. Mendez and S. A. Brown, "The Natural Coumarins," John Wiley & Sons Ltd., New York, 1982, p. 27.
- 3) F. M. Dean, D. R. Randell and G. Winfield, J. Chem. Soc., 1961,
- 4) M. F. Grundon and I. S. McColl, Phytochemistry, 14, 143 (1975).
- 5) S. K. Talapatra, S. Goswami, N. C. Granguly and B. Talapatra, *Chem. Ind.* (London), 1980, 154.
- F. Bohlmann, V. S. B. Rao and M. Grenz, Tetrahedron Lett., 1968, 3947.
- H. D. Schroeder, W. Bencze, O. Halpern and H. Schmid, *Chem. Ber.*,
 92, 2338 (1959); J. Lemmich, E. Lemmich and B. E. Nielsen, *Acta Chem. Scand.*, 20, 2497 (1966).
- T.-S. Wu, C.-S. Kuoh and H. Furukawa, *Phytochemistry*, 22, 1493 (1983).