Fern Constituents: Cheilanthenetriol and Cheilanthenediol. Sesterterpenoids Isolated from the Leaves of Aleuritopteris khunii

Rumiko Kamaya and Hiroyuki AGETA*

Showa College of Pharmaceutical Sciences, 5-1-8 Tsurumaki, Setagaya-ku, Tokyo 154, Japan. Received July 5, 1989

Two sesterterpenoids, cheilanthenetriol (1) and cheilanthenediol (2), were isolated from the leaves of Aleuritopteris khunii together with hop-22(29)-ene, adiantone, hydroxyhopane, a mixture of sterols, 3,5-dihydroxy-7,4'-dimethoxyflavone, 5-hydroxy-3,7,4'-trimethoxyflavone and 4-methoxybenzoic acid. Cheilanthenetriol was found to be identical with authentic samples of cheilanthatriol and cheilarinosin isolated from Indian ferns. The new sesterterpenoid, cheilanthenediol, was established as (17Z)-13 α ,19-dihydroxycheilanth-17-ene.

Keywords fern; Aleuritopteris khunii; Pteridaceae; triterpenoid; sesterterpenoid; cheilanthenetriol; cheilanthenediol; flavonoid

The ferns belonging to the genus Aleuritopteris¹⁾ of Pteridaceae are widely distributed in the world as typical ferns on limestone, though only three species of them have been reported in Japan. Some ferns in this group are very difficult to classify morphologically, and the occurrence of diterpenoids, sesterterpenoids, triterpenoids, flavonoids and other chemical components is, therefore, of chemotaxonomic interest. This paper concerns the isolation and characterization of two sesterterpenoids from the leaves of Aleuritopteris khunii (MILDE) CHING (Cheilanthes khunii MILDE, miyama-urajiro in Japanese), which is a beautiful summer-green species growing in the limestone areas of central Japan.

Results and Discussion

The dried leaves of the fern were extracted with n-

hexane and the extract was separated into several fractions containing triterpenoids [hop-22(29)-ene (diploptene²⁾), adiantone,³⁾ and hydroxyhopane⁴⁾], sterols (a mixture of sitosterol, stigmasterol and campesterol, and a mixture of their fatty acid esters), flavonoids (3,5-di-hydroxy-7,4'-dimethoxyflavone and 5-hydroxy-3,7,4'-trimethoxyflavone),⁵⁾ 4-methoxybenzoic acid⁶⁾ and two sesterterpenoids, namely cheilanthenetriol (1) and a new compounds, cheilanthenediol (2).

Cheilanthenetriol (1), $^{7)}$ C₂₅H₄₄O₃, mp 179—180 °C, showed strong infrared (IR) absorption bands due to hydroxyl groups. The 1 H-nuclear magnetic resonance (1 H-NMR) spectrum (270 MHz) indicated the presence of five tertiary methyl groups, one olefinic methyl group (δ 1.780), one proton attached to the carbon bearing a hydroxyl group (δ 3.954), two protons of a hydroxymethyl group (δ 4.053 and

© 1990 Pharmaceutical Society of Japan

TABLE I. ¹H-Chemical Shifts of Compounds 1, 2 and Their Derivatives (100 or 270 MHz, CDCl₃, δ Value)

| Compound | H-20 | H-21 | H-22 | H-23 | H-24 | H-25 | H-19 | H-18 | Η-6β | ОСОСН |
|------------------------|-------|--------|--------|-------|----------------|-------------------|--|----------------------------|------------------------------|-------|
| 1 ^{a)} | 1.168 | 1.010 | 0.856 | 0.844 | 1.124 | 1.780 | 4.053 (dd, 11.7, 7.3) | 5.451 (dd, 7.3, 7.3) | 3.954 (ddd, 10.7, 10.7, 3.4) | |
| 3 | 1.174 | 1.017 | 0.857 | 0.857 | 1.129 | 1.794 | 4.130 (dd, 11.7, 7.3) 4.541 (dd, 11.7, 7.0) | 5.331 (dd, 7.0, 7.0) | 3.958 (ddd, 10.5, 10.5, 3.1) | 2.049 |
| 4 | 0.864 | 1.022 | 0.909 | 0.909 | 1.132 | 1.755 | 4.686 (dd, 11.7, 7.0) 4.447 (dd, 12.0, 7.1) | 5.306 (dd, 7.1, 7.1) | 5.246 (ddd, 11.0, 11.0, 3.5) | 2.044 |
| $5^{a)}$ | 1.159 | 1.015 | 0.851 | 0.878 | 1.132 | 0.921 | 4.683 (dd, 12.0, 7.1) 3.659 (dt, 10.5, 6.6) | | 3.965 (ddd, 10.8, 10.8, 3.5) | 2.044 |
| 64) | 1.159 | 1.015 | 0.853 | 0.882 | 1.131 | (d, 6.6) 0.860 | 3.716 (dt, 10.5, 6.6) 0.850 (t, 7.0) | | 3.965 (ddd, 10.7, 10.7, 3.7) | |
| 7 ^{a)} | 1.216 | 0.924 | 0.830 | 0.822 | 1.146 | (d, 6.5) 0.861 | 0.843 (t, 6.5) | | | |
| 8 | 1.171 | 1.028 | 0.879 | 0.862 | 1.145 | (d, 6.9) 2.000 | 9.943 (d, 8.0) | 5.847 (dd, 8.0, 1.5) | 3.973 (ddd, 10.5, 10.5, 3.5) | |
| 2 ^{a)} | 0.861 | 0.797 | 0.808 | 0.783 | 1.130 | (d, 1.5) 1.775 | 4.055 (dd, 12.0, 7.3) | 5.463 (ddd, 7.3, 7.3, 1.5) | | |
| 12 | 0.860 | 0.798 | 0.798 | 0.798 | 1.132 | (d, 1.5) 1.786 | 4.133 (dd, 12.0, 7.3) 4.505 (dd, 12.0, 7.5) | 5.315 (dd, 7.5, 7.5) | | 2.044 |
| 13 ^{a)} | 0.852 | 0.796 | 0.796 | 0.805 | 1.133 | 0.919 | 4.685 (dd, 12.0, 7.5) 3.664 (dt, 10.6, 6.6) | | | |
| 14 ^{a)} | 0.853 | 0.797 | 0.797 | 0.805 | 1.133 | (d, 6.3) 0.868 | 3.715 (dt, 10.6, 6.6) 0.855 (t, 6.7) | | | |
| 15 | 0.857 | 0.796 | 0.796 | 0.666 | 4.537 4.836 | (d, 6.8) 1.755 | 4.504 (d, 7.2) | 5.348 (t, 7.2) | | 2.044 |
| 16 | 0.860 | 0.801 | 0.801 | 0.826 | 1.451 | 1.787 | 4.566 (d, 7.2) | 5.336 (t, 7.2) | | 1.960 |
| 18 | 0.853 | 0.803 | 0.803 | 0.803 | 1.147 | 2.007 | 9.944 (d, 8.0) | 5.853 (dd, 8.0, 1.5) | | 2.049 |
| | H-23) | (H-24) | (H-25) | | | (d, 1.5) | | | | |
| | | 1.015 | | | | | | | | |
| | | 1.031 | | | | | | | | |
| | | 0.905 | | | | | | | | |
| | | | 0.818 | | | | | | | |

Multiplicity and coupling constants (J in Hz) are shown in parentheses. Hydrogen numbers given in parentheses are those of the hopane group. a) Measured at 270 MHz.

4.130) and one proton of a trisubstituted double bond (δ 5.451) as shown in Table I.

Compound 1 was acetylated with Ac₂O-pyridine to give a monoacetate (3) and a diacetate (4). Catalytic hydrogenation of 1 gave 5 and 6, the latter being converted to a ketone (7). Oxidation of 1 with MnO₂ afforded an aldehyde (8) as a sole product, a part of which changed into its geometric isomer (8a) spontaneously in solution, until equilibrium was reached. In a comparison of the ¹H-NMR spectra of compounds 1, 4 and 7 with those of zeorin (9) and its derivatives (10 and 11) (Table I), the three corresponding methyl groups on ring A have very similar chemical shifts. The coupling pattern [δ 3.954 (ddd, J=10.7, 10.7, 3.4 Hz)] of a geminal proton of the secondary hydroxyl group in 1 suggests that the proton is an axial one, and it also resembles that in 98; therefore, one of the hydroxyl group of 1 can be located at C-6α. This compound was identical [mixture melting point, IR, mass spectrum (MS) and ¹H-NMR] with authentic samples of cheilarinosin⁹⁾ and cheilanthatriol¹⁰⁾ isolated from Indian socalled Cheilanthes farinosa.11) Thus, the structure of cheilarinosin, which has been reported to have the ophiobolane skeleton, should be revised to formula 1.

A new compound, cheilanthenediol (2),⁷⁾ mp 156—157 °C, C₂₅H₄₄O₂, which had a higher *Rf* value than 1 on thin-layer chromatography (TLC), showed IR absorption bands due to hydroxyl groups. The ¹H-NMR spectrum

(270 MHz) indicated the presence of five tertiary methyl groups, one olefinic methyl group (δ 1.775), two protons of a hydroxymethyl group (δ 4.055 and 4.133) and one proton of a trisubstituted double bond (δ 5.463): the latter two are coupled to each other (Table I). The MS of 2 and 12 showed fragment peaks a, b, c and d corresponding to those of 1 and 3 (Table II), suggesting that 2 has no hydroxyl group on ring B. This was confirmed by the absence of a geminal proton of a secondary hydroxyl group in the ¹H-NMR spectrum.

Compound 2 was acetylated under a mild condition (Ac₂O-pyridine) to yield only a monoacetate (12) which has a free hydroxyl group considered to be tertiary, and catalytic hydrogenation gave two products, 13 and 14. Dehydration of 2 with Ac₂O-NaOAc afforded a dehydrate (15), along with a diacetate (16), and furthermore ozonolysis of 15 gave a diketone (17). Oxidation of 2 with MnO₂ afforded an aldehyde (18) as a sole product, a part of which gradually changed into its geometric isomer (18a) until equilibrium was reached as observed in the case of 1.

The ¹H-chemical shifts of three methyl groups of 2 and its derivatives (12—16, 18) closely resembled those of hydroxyhopane (19) on ring A. By analogy of the chemical shifts and/or coupling patterns of the methyl group (C-24) and some protons of the side chain between each of the four pairs of 1 and 2, 3 and 12, 5 and 13, 8 and 18 (Table I), compound 2 is expected to have the same side chain as 1.

TABLE II. MS Data for Compounds 1, 2 and Their Derivatives

| | - | 1 | 3 | 2 | 12 | | | |
|----------------|---------------------------|-----------------|-----------|-----------|-----------|--|--|--|
| Fragment | - | m/z (rel. int.) | | | | | | |
| Molecular ion | M ⁺ | | | 376 (<1) | 418 (2) | | | |
| Molecular 1011 | $M^+ - H_2O$ | 374 (6) | 416 (1) | 358 (12) | 400 (5) | | | |
| | $M^+ - H_2O - Me$ | 359 (9) | | 348 (13) | 385 (3) | | | |
| | $M^+ - 2H_2O$ | 356 (26) | 398 (2) | 340 (13) | | | | |
| | $M^+ - 2H_2O - Me$ | 341 (14) | | 325 (7) | | | | |
| | $M^+ - 3H_2O$ | 338 (10) | | | | | | |
| | M ⁺ – AcOH | | 374 (4) | | 358 (27) | | | |
| | $M^+ - 3H_2O - Me$ | 327 (7) | | | | | | |
| | $M^+ - AcOH - Me$ | | | | 343 (11) | | | |
| | M^+ – AcOH – H_2O | | 356 (20) | | 340 (35) | | | |
| | $M^+ - AcOH - H_2O - Me$ | | 341 (11) | • | 325 (13) | | | |
| | $M^+ - AcOH - 2H_2O$ | | 338 (11) | | | | | |
| | $M^+ - AcOH - 2H_2O - Me$ | | 323 (9) | | | | | |
| Fragment a | $M^+ - 116$ | 276 (21) | 276 (19) | 260 (100) | 260 (89) | | | |
| Tagment a | a – Me | 261 (23) | 261 (36) | 245 (76) | 245 (100) | | | |
| | $a - H_2O$ | 258 (100) | 258 (100) | | | | | |
| | $a - H_2O - Me$ | 243 (12) | 243 (38) | | | | | |
| Fragment b | $M^+ - 117$ | 275 (16) | 275 (28) | 259 (54) | 259 (59) | | | |
| 1 ragment o | $b-H_2O$ | 257 (27) | 257 (22) | | | | | |
| Fragment c | $M^+ - 171$ | 221 (5) | 221 (6) | 205 (17) | 205 (17) | | | |
| i raginone o | $c-H_2O$ | 203 (18) | 203 (39) | | | | | |
| Fragment d | $M^+ - 185$ | 207 (13) | 207 (21) | 191 (68) | 191 (42 | | | |
| I lugilloni u | $d-H_2O$ | 189 (28) | 189 (55) | | | | | |

Derivatives (68 MHz, CDCl₃, δ Value)

| C | 1 | 4 | 7 | 2 | 12 | 13 | 14 | 15 | 16 |
|-------------------|-------|-------|-------|-------|-------|------|------|-------|------|
| 1 | 40.2 | 40.0 | 40.7 | 40.1 | 40.1 | 40.1 | 40.1 | 40.2 | 40.1 |
| 2 | 18.6 | 18.4 | 18.4 | 18.7 | 18.7 | 18.7 | 18.7 | 18.7 | 18.7 |
| 3 | 43.6 | 43.3 | 42.6 | 42.2 | 42.2 | 42.2 | 42.2 | 42.1 | 42.2 |
| 4 | 33.7 | 33.3 | 32.3 | 33.3 | 33.3 | 33.3 | 33.3 | 33.3 | 33.4 |
| 5 | 61.3 | 58.9 | 66.8 | 56.7 | 56.7 | 56.7 | 56.7 | 56.7 | 56.7 |
| 6 | 67.9 | 70.7 | 211.8 | 18.3 | 18.3 | 18.2 | 18.2 | 18.7 | 18.2 |
| 7 | 52.9 | 48.3 | 58.1 | 41.5 | 41.5 | 41.5 | 41.5 | 40.7 | 41.3 |
| 8 | 40.2 | 40.0 | 45.5 | 39.4 | 39.4 | 39.5 | 39.5 | 39.8 | 39.8 |
| 9 | 60.1 | 60.2 | 60.7 | 60.7 | 60.7 | 60.7 | 60.7 | 60.2 | 60.2 |
| 10 | 39.3 | 39.5 | 43.7 | 37.6 | 37.6 | 37.6 | 37.6 | 37.9 | 37.5 |
| 11 | 19.6 | 19.5 | 19.8 | 19.4 | 19.4 | 19.4 | 19.4 | 19.2 | 19.0 |
| 12 | 44.3 | 44.3 | 44.0 | 44.3 | 44.4 | 44.5 | 44.5 | 38.2 | 38.9 |
| 13 | 74.0 | 73.9 | 74.0 | 74.2 | 73.8 | 74.3 | 74.3 | 148.4 | 88.0 |
| 14 | 61.8 | 61.9 | 62.4 | 61.8 | 62.2 | 62.8 | 63.0 | 56.3 | 59. |
| 15 | 24.5 | 24.1 | 22.7 | 24.5 | 24.3 | 22.8 | 22.8 | 23.3 | 24.9 |
| 16 | 35.6 | 35.6 | 40.7 | 35.6 | 35.8 | 41.3 | 40.9 | 37.9 | 35. |
| 17 | 141.8 | 143.9 | 35.4 | 142.0 | 144.1 | 30.7 | 35.5 | 143.1 | 143. |
| 18 | 123.5 | 118.8 | 29.5 | 123.6 | 118.6 | 39.8 | 29.3 | 119.2 | 118. |
| 19 | 58.4 | 61.2 | 11.4 | 58.4 | 61.3 | 61.1 | 11.4 | 61.2 | 61. |
| 20 | 36.6 | 36.1 | 32.4 | 33.3 | 33.3 | 33.4 | 33.4 | 33.3 | 33. |
| 21 | 22.2 | 22.0 | 21.6 | 21.4 | 21.4 | 21.4 | 21.4 | 21.4 | 21. |
| 22 | 17.7 | 17.6 | 16.7 | 16.3 | 16.3 | 16.2 | 16.2 | 16.2 | 16. |
| 23 | 18.1 | 17.8 | 17.1 | 16.7 | 16.7 | 16.7 | 16.7 | 15.5 | 17. |
| 24 | 23.9 | 24.0 | 23.8 | 23.9 | 23.9 | 23.9 | 23.9 | 106.0 | 23. |
| 25 | 23.6 | 23.5 | 19.3 | 23.6 | 23.6 | 19.8 | 19.4 | 23.5 | 23. |
| COCH ₃ | 3 | 21.2 | | | 21.1 | | | 21.1 | 20. |
| | | 22.1 | | | | | | .= | 21. |
| COCH: | 3 | 170.3 | | | 170.8 | | | 170.8 | 170. |
| | | 170.9 | | | | | | | 171. |

The Z configuration of the Δ^{17} double bond of 1 has already been proposed from the carbon-13 nuclear magnetic resonance (¹³C-NMR) data. ^{10b)} This deduction was also supported by the following evidence: the ¹H-NMR signals of C-25 methyl groups in 8 and 18 appear at a higher field (-0.17 ppm in both) than those in their geometric

TABLE III. 13C-Chemical Shifts of Compounds 1, 2 and Their TABLE IV. Substituent Effects for the Cheilanthane and Hopane Groups

| | Chei | lanthane g | group | Н | Hopane group | | | |
|---------|-------|------------|--------|-------|--------------|--------|--|--|
| С - | 6α ОН | 6α OAc | 6 CO | 6α ОН | 6α OAc | 6 CO | | |
| 1 | +0.1 | -0.1 | +0.6 | 0 | -0.3 | +0.4 | | |
| 2 | -0.1 | -0.3 | -0.3 | -0.2 | -0.4 | -0.1 | | |
| 3 | +1.4 | +1.1 | +0.4 | +1.6 | +1.2 | +0.4 | | |
| 4 | +0.4 | 0 | -1.0 | +0.3 | -0.3 | -0.8 | | |
| 5 | +4.6 | +2.2 | +10.1 | +4.9 | +2.1 | +10.2 | | |
| 6 | +49.6 | +52.4 | +193.6 | +50.6 | +53.0 | +195.2 | | |
| 7 | +11.4 | +6.8 | +16.6 | +12.2 | +7.5 | +18.1 | | |
| 8 | +0.8 | +0.6 | +6.0 | +0.9 | +0.6 | +6.6 | | |
| 9 | -0.6 | -0.5 | 0 | -0.6 | -0.7 | +0.3 | | |
| 10 | +1.7 | +1.9 | +6.1 | +2.1 | +2.0 | +6.6 | | |
| 20 (23) | +3.3 | +2.8 | -1.0 | +3.3 | +2.7 | -1.0 | | |
| 21 (24) | +0.8 | +0.6 | +0.2 | +0.4 | +0.3 | +0.1 | | |
| 22 (25) | +1.4 | +1.3 | +0.4 | +1.2 | +1.1 | +0.7 | | |
| 23 (26) | +1.4 | +1.1 | +0.4 | +1.5 | +1.1 | +0.8 | | |

Carbon numbers given in parentheses are those of the hopane group.

isomers, 8a and 18a, which are affected more strongly by the anisotropy of the aldehyde group.

As shown in Table III, the ¹³C-chemical shifts of 2 are in good agreement with those of 1, except for C-3, C-4, C-5, C-6, C-7, C-8, C-9, C-10, C-20, C-21, C-22, and C-23. The ¹³C-chemical shift deviations of 1, 4 and 7 from 2, 12 and 14 show substituent group effects of 6α-hydroxy, 6α-acetoxy and 6-oxo in the cheilanthane group. These values are compatible with those for the corresponding positions in the hopane group (9, 10 and 11)12) (Table IV).

The stereochemistry of the cheilanthane skeleton was unequivocally determined on the basis of the following spectral data of 1, 2 and their derivatives: 1) The circular dichroism (CD) and optical rotatory dispersion (ORD) curves of compound 7 and zeorinone (11) showed the same negative Cotton effects. The differences of the respective

¹H-NMR solvent shift values $[\delta(C_6D_6) - \delta(CDCl_3)]$ of four corresponding methyl groups between 7 and 14 are +0.19(H-20), +0.12 (H-21), -0.04 (H-22) and -0.04 (H-23), which are characteristic of an isolated carbonyl group, excluding the solvent effect due to an additional hydroxy group (C-13 α OH); in the case of zeorinone (11) and hydroxyhopane (19), $^{13)}$ these differences are +0.21 (H-23), +0.10 (H-24), -0.04 (H-25) and -0.07 (H-26), showing a similar tendency to those mentioned above. 14) According to these indications, the cheilanthane group has the same absolute structure as the hopane group. 2) The ORD curve of 17 showed a positive Cotton effect similar to that of 20 derived from a diterpenoid, alepterolic acid. 15) This provided conclusive proof that the configuration of the side chain in 2 is β . This result is also reasonable from the biogenetic point of view.

All the above evidence leads to (17Z)- 13α ,19-dihydroxycheilanth-17-ene (2) for the structure of cheilanthenediol. Some new spectral data for 1, 2 and their derivatives also support the conclusion that the structure of cheilanthenetriol is (17Z)- 6α ,13 α ,19-trihydroxycheilanth-17-ene (1).

Cheilanthenediol is one of the most primitive tricyclic sesterterpenoids derived from geranyl farnesyl precursor. Further chemotaxonomical studies on some ferns belonging to *Aleuritopteris* and *Cheilanthes* are in progress.

Experimental

Melting points were measured with a Yanagimoto microapparatus and are uncorrected. The ¹H- and ¹³C-NMR spectra were recorded on JEOL JNM FX-100 and GX-270 spectrometers using tetramethylsilane as an internal stadard in CDCl₃ solution. Electron impact mass spectra (EI-MS) were measured at 70 eV (direct inlet) with a JEOL JMS D300 spectrometer and the relative intensities of peaks were reported with reference to the most intense peak higher than m/z 100. Gas chromatography (GC) was run on a Hitachi 163 using a glass column containing Chromosorb G HP coated with SE-30 (1.4%) at 260 °C in a flow of N₂. Cholestane was used as an internal reference; its retention time was set at 3.0 min. Gas chromatography mass spectra (GC-MS) were run on a JGC 20K-JMS D300 system using the same column as described above in a flow of He. TLC was carried on precoated Silica gel 60 plates (Merck) with n-hexane–EtOAc or CHCl₃–MeOH as the solvent system.

Plant Materials The leaves (6.7 kg) of Aleuritopteris khunii were collected in September at Shimonita, Gunma Prefecture, Japan. A voucher specimen has been deposited in the Herbarium of Showa College of Pharmaceutical Sciences, Tokyo.

Extraction and Separation The air-dried plant materials were extracted with *n*-hexane and the extract was treated with MeOH to remove aliphatic substances. The resulting extract was chromatographed on silica gel using *n*-hexane, C₆H₆ and Et₂O as eluents to be separated first into six fractions (frs. 1—6). By repeated chromatography and recrystallization, hop-22(29)-ene (0.094g) was obtained from fr. 1, sterol esters (0.134g) from fr. 2, adiantone (0.158g) from fr. 3, hydroxyhopane (0.268g), sterol mixture (1.474g), 3,5-dihydroxy-7,4'-dimethoxyflavone (1.210g) and 5-hydroxy-3,7,4'-trimethoxyflavone (0.020g) from fr. 4, cheilanthenediol (2) (2.747g) and 4-methoxybenzoic acid (0.007g) from fr. 5, and cheilanthenetriol (1) (6.030g) from fr. 6.

Hop-22(29)-ene Colorless needles (from EtOAc), mp 210—211 °C, $[\alpha]_D^{23}$ +60.5° (c=0.32, CHCl₃). IR v_{\max}^{KBr} cm⁻¹: 3035, 1645, 1630, 886. MS m/z (rel. int.): 410 (M+, 28), 395 (10), 376 (6), 299 (8), 218 (13), 205 (15), 204 (20), 203 (10), 191 (100), 189 (78). Identical (mixture melting point, IR and MS) with an authentic sample.²⁾

Adiantone Colorless needles (from CHCl₃–MeOH), mp 227–230 °C, $[\alpha]_D^{23}$ +78.0° (c =0.44, CHCl₃). IR $\nu_{\rm max}^{\rm KBr}$ cm $^{-1}$: 1700. MS m/z (rel. int.): 412 (M $^+$, 22), 397 (9), 369 (17), 191 (100). Identical (mixture melting point, IR and MS) with an authentic sample.³⁾

Hydroxyhopane Colorless needles (from EtOAc), mp 253—255 °C, $[\alpha]_{\rm D}^{23}$ +42.0° (c=0.38, CHCl₃). IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 3600, 3040, 1152. MS m/z (rel. int.): 428 (M⁺, 3), 410 (9), 395 (6), 369 (3), 207 (17), 205 (9), 191 (100),

189 (76), 177 (11), 163 (17). Identical (mixture melting point, IR and MS) with an authentic sample. 41

Sterol Esters Colorless flakes (from EtOAc), mp 76—78 °C. Hydrolysis of sterol esters with 5% KOH-EtOH under reflux for 2 h gave the sterol and fatty acid fractions. The GC and GC-MS of the sterol mixture showed R t_R 2.24 (6% of total peaks) (m/z 400, M⁺), 2.41 (16%) (412, M⁺) and 2.73 (78%) (414, M⁺), which correspond to campesterol, stigmasterol and sitosterol respectively. The fatty acid fraction consisted of palmitic acid and a small amount of stearic acid, which were identified as their methyl esters by GC and GC-MS.

Sterol Mixture Colorless plates (from CHCl₃–MeOH), mp 137.5—139 °C. GC and GC-MS: Rt_R 2.24 (9% of total peaks) (m/z 400, M^+), 2.41 (4%) (412, M^+), 2.73 (87%) (414, M^+), which correspond to the three sterols mentioned above.

3,5-Dihydroxy-7,4'-dimethoxyflavone Yellow needles (from C_6H_6), mp 182—184 °C. IR $v_{\rm max}^{\rm KBr}$ cm $^{-1}$: 3295, 1650, 1600, 1510, 1325, 1260, 830. MS m/z (rel. int.): 314 (M $^+$, 100), 313 (9), 299 (10), 285 (5), 271 (10), 243 (5), 135 (5). ¹H-NMR (100 MHz) δ : 3.875 (3H, s), 3.885 (3H, s), 6.358 (1H, d, J = 2.2 Hz), 7.020 (1H, d, J = 9.2 Hz), 8.154 (1H, d, J = 9.2 Hz). Anal. Calcd for $C_{17}H_{14}O_6$: C, 64.96; H, 4.49. Found: C, 64.94; H, 4.49. (lit. mp 178—180 °C). Methylation of this compound (0.089 g) with diazomethane gave 5-hydroxy-3,7,4'-trimethoxyflavone, yellow needles (0.032 g) (from C_6H_6), mp 149—149.5 °C. IR $v_{\rm max}^{\rm KBr}$ cm $^{-1}$: 3910, 1665, 1510, 1350, 1260, 1180, 830. MS m/z (rel. int.): 328 (M $^+$, 100), 327 (81), 313 (11), 299 (16), 285 (37), 135 (16). 1 H-NMR (100 MHz) δ : 3.858 (3H, s), 3.872 (3H, s), 3.897 (3H, s), 6.354 (1H, d, J = 2.2 Hz), 6.441 (1H, d, J = 2.2 Hz), 7.020 (1H, d, J = 9.2 Hz), 8.074 (1H, d, J = 9.2 Hz).

5-Hydroxy-3,7,4'-trimethoxyflavone Yellow needles (from C_6H_6), mp 149—149.5 °C. Identical (mixture melting point, IR, MS and ¹H-NMR) with the compound given above.

4-Methoxybenzoic Acid Colorless needles (from C_6H_6), mp 180—185 °C. IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 3000—2500, 2800, 1675, 1600, 1425, 1295, 1255, 1165, 1025, 840. MS m/z (rel. int.): 152 (M⁺, 99), 135 (100), 107 (13), 92 (16), 77 (21). ¹H-NMR (100 MHz) δ : 3.870 (3H, s), 6.941 (2H, d, J= 9.1 Hz). 8.062 (2H, d, J=9.1 Hz). Identical (mixture melting point, IR, MS and ¹H-NMR) with an authentic sample.

Cheilanthenetriol (1) Colorless needles (from Me₂CO), mp 179—180 °C, [α]₂²³ + 30.2° (c = 0.37, CHCl₃). IR ν ^{KBB}_{max} cm⁻¹: 3350, 2950, 1660, 1450, 1395, 1000, 980, 860. *Anal*. Calcd for C₂₅H₄₄O₃: C, 76.48; H, 11.30. Found: C, 76.11; H, 11.08. HRMS: m/z 374.3225 (M⁺ – H₂O, Calcd for C₂₅H₄₂O₂: 374.3185.)

Acetylation of 1 1(0.15 g) in pyridine (2.5 ml) and Ac₂O (1.3 ml) was allowed to stand at 5 °C for 0.5 h. Half of the solution was worked up, and the reaction mixture was chromatographed on silica gel to give 3 (0.03 g), colorless plates (from *n*-hexane–Me₂CO), mp 126—128.5 °C. IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 3540, 3450, 1730, 1240, 1010. The rest of the solution was left at room temperature overnight. After the same work-up and separation as described above, the main product was 4 (0.04 g), colorless gum. MS m/z (rel. int.): 416 (M⁺ – AcOH, 6), 401 (14), 398 (14), 389 (17), 356 (13), 341 (18), 338 (19), 329 (70), 316 (42), 301 (22), 261 (32), 259 (25), 258 (100), 257 (42), 243 (32), 203 (43), 191 (83), 189 (59), 187 (40).

Catalytic Hydrogenation of 1 1 (0.2 g) was hydrogenated in the presence of PtO₂ (0.12 g) in AcOH (8 ml) and EtOAc (4 ml) at room temperature for 1.5 h. The reaction mixture was chromatographed on silica gel to yield 5 (0.054 g) and 6 (0.06 g). 5: colorless needles (from EtOAc), mp 176—177.5 °C. IR $v_{\rm max}^{\rm RB}$ cm⁻¹: 3450, 3330, 1130, 1010. MS m/z (rel. int.): 394 (M⁺, 1), 376 (26), 361 (7), 358 (9), 343 (10), 306 (28), 305 (44), 261 (6), 243 (15), 223 (22), 205 (20), 203 (18), 191 (100), 189 (23), 163 (30). 6: colorless needles (from n-hexane—Et₂O), mp 194—195 °C. IR $v_{\rm max}^{\rm RB}$ cm⁻¹: 3520, 1110, 1040. MS m/z (rel. int.): 378 (M⁺, 1), 360 (3), 345 (10), 342 (8), 327 (12), 307 (37), 289 (100), 275 (11), 261 (47), 258 (15), 243 (17), 207 (34), 203 (25), 191 (74), 190 (39), 189 (29), 163 (89).

Oxidation of 6 with CrO₃ A solution of 6 (0.045 g) in pyridine (2 ml) was combined with CrO₃ (0.1 g) in pyridine (3 ml) and the mixture was stirred at room temperature for 2.2 h. The product was subjected to silica gel chromatography to yield 7 (0.032 g), colorless plates (from *n*-hexane), mp 119—120 °C. IR v_{\max}^{KBr} cm⁻¹: 3450, 1710, 1070. MS m/z (rel. int.): 376 (M⁺, 7), 361 (6), 358 (14), 343 (14), 305 (11), 291 (8), 277 (25), 206 (39). ORD and CD: [M] (nm): -550° (325) (trough), $+700^{\circ}$ (292) (peak); [θ] (nm): -1470 (310) (negative maximum) (c=0.084, MeOH) {zeorinone (11), [M] (nm): -1070° (324) (trough), $+1790^{\circ}$ (288) (peak); [θ] (nm): -2450 (360) (negative maximum) (c=0.037, MeOH))}. 1 H-NMR solvent shift values [δ (C_oD_o) $-\delta$ (CDCl₃)]: +0.24 (H-20), +0.17 (H-21), -0.08 (H-22), -0.10 (H-23).

Oxidation of 1 with MnO₂ 1 (0.15 g) in dry Me₂CO (15 ml) was treated

with activated MnO₂ (1.2 g) at room temperature for 4 h. The product (8), which showed one spot on TLC was recrystallized from *n*-hexane–EtOAc to give colorless plates, mp 159—160 °C, indicating an additional weak, lower Rf spot. 8: IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 3480, 1670, 1175. MS m/z (rel. int.): 390 (M⁺, 1), 375 (37), 372 (22), 357 (59), 339 (19), 329 (48), 289 (41), 275 (19), 271 (42), 261 (68), 258 (26), 257 (37), 243 (30), 203 (51), 191 (100), 189 (58). The density of the lower Rf spot gradually increased after standing in the CHCl₃ solution. The ¹H-NMR spectrum of this mixture exhibited signals at δ 2.174 (3H, d, J=1.0Hz) and 10.012 (1H, d, J=8.0Hz) due to 8a in addition to the signals of 8 shown in Table I.

Cheilanthenediol (2) Colorless plates (from Me₂CO), mp 156—157 °C, $[\alpha]_D^{23} + 5.1$ ° $(c = 0.60, \text{CHCl}_3)$. IR v_{max}^{KBr} cm⁻¹: 3300, 1095, 1010, 800. HRMS m/z: 376.3331 (M⁺, Calcd for C₂₅H₄₄O₂: 376.3339).

Acetylation of 2 2 (0.062 g) in pyridine (1 ml) and $Ac_2O(0.5 \text{ ml})$ was allowed to stand at room temperature for 1 h. The product was chromatographed on silica gel to give 12(0.044 g) as a colorless gum.

Catalytic Hydrogenation of 2 2 (0.102 g) was hydrogenated in the presence of PtO₂ (0.14 g) in AcOH (8 ml) and EtOAc (4 ml) at room temperature for 2 h. Chromatography on silica gel of the reaction mixture followed by recrystallization afforded 13 (0.016 g), 14 (0.025 g) and the starting material (0.010 g). 13: colorless needles (from *n*-hexane–Me₂CO), mp 124—125 °C. IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 3350, 3330, 1135, 1090. HRMS m/z: 378.3491 (M⁺, Calcd for C₂₅H₄₆O₂: 378.3496), MS m/z (rel. int.): 378 (M⁺, 30), 363 (11), 360 (23), 345 (28), 307 (68), 289 (9), 260 (8), 259 (16), 245 (51), 205 (25), 204 (44), 193 (63), 192 (100), 191 (84), 177 (28). 14: colorless needles (from *n*-hexane), mp 125—127.5 °C, IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 3500, 1140, 1105, 1010. MS m/z (rel. int.): 362 (M⁺, 5), 344 (30), 329 (20), 291 (5), 260 (15), 259 (51), 245 (12), 205 (24), 204 (16), 192 (70), 191 (100), 177 (45). ¹H-NMR solvent shift values $[\delta(C_6D_6) - \delta(CDCl_3)]$: +0.05 (H-20), +0.05 (H-21), -0.04 (H-22), -0.06 (H-23).

Dehydration of 2 2 (0.13 g) was refluxed with Ac_2O (5 ml) and AcONa (0.2 g) for 5 h. The product was chromatographed on silica gel to yield the following compounds; **15** (0.015 g), an amorphous powder; $IR \ \nu_{\text{max}}^{\text{KBr}} \text{ cm}^{-1}$: 1735, 1645, 1230, 1020, 885, and **16** (0.04 g), colorless needless (from *n*-hexane), mp 103.5—104 °C, $IR \ \nu_{\text{max}}^{\text{KBr}} \text{ cm}^{-1}$: 1730, 1250, 1020, MS m/z (rel. int.): 400 (M⁺ – AcOH, 2), 385 (7), 340 (19), 325 (27), 272 (37), 260 (66), 259 (52), 245 (30), 205 (23), 204 (24), 191 (100), 177 (39), 163 (31).

Ozonolysis of 15 A gas mixture of 3% ozone/O₂ was passed into a solution of **15** (0.015 g) in CHCl₃ (6 ml) for 0.5 h. After evaporation of the solution, the resluting ozonide was treated with AcOH (6 ml) in the presence of Zn powder (0.15 g). The product was chromatographed on silica gel to give **17** (0.008 g), colorless gum, IR v_{\max}^{film} cm⁻¹: 1710, MS m/z (rel. int.): 332 (M⁺, 20), 317 (100), 299 (18), 191 (44). ¹H-NMR (100 MHz) δ : 0.732, 0.816, 0.848, 0.877, 2.095 (3H,each), ORD: [M] (nm): +7330° (308) (peak), -8630° (270) (trough) (c=0.058, MeOH) {**20**; [M] (nm): +6750° (318) (peak), -7050° (280) (trough) (c=0.073, MeOH)}.

Oxidation of 2 with MnO₂ **2** (0.108 g) in dry Me₂CO (12 ml) was treated with activated MnO₂ (0.9 g) for 2.5 h at room temperature. The reaction product, **18**, IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 3400, 1660, 1100. MS m/z (rel. int.): 374 (M⁺, 10), 359 (20), 356 (13), 341 (16), 291 (61), 273 (36), 259 (31), 245 (63), 191 (100), showed one spot on TLC, but the CHCl₃ solution of the above compound after standing for a while showed two spots on TLC, the

lower Rf compound of which is the geometric isomer (18a). The ¹H-NMR spectrum of this mixture exhibited signals at δ 2.181 (3H, d, J=1.0 Hz) and 9.988 (1H, d, J=8.0 Hz) due to 18a in addition to the signals of 18 shown in Table I.

Acknowledgements We are grateful to Dr. Sukh DeV (India) for sending a sample (cheilanthatriol) and a plant specimen of the original material extracted, Professor S. Rangaswami (India) for supplying a sample (cheilarinosin), and Mr. T. Satomi (Shimonita) for his kind advice on the plant materials. Thanks are due to Mr. Y. Takase of this college for recording ¹H- and ¹³C-NMR, and mass spectra.

References and Notes

- 1) Y. Saiki, J. Phytogeog. & Taxon., 32, 1, 91 (1984).
- H. Ageta, K. Iwata, and K. Yonezawa, Chem. Pharm. Bull., 11, 408 (1963).
- G. Berti, F. Botarii, A. Marsili, J. M. Lehn, P. Witz, and G. Ourisson, *Tetrahedron Lett.*, 1963, 1283.
- H. Ageta, K. Iwata, and Y. Otake, Chem. Pharm. Bull., 11, 407 (1963).
- H. Erdtman, L. Novotny, and M. Romanuk, *Tetrahedron*, Suppl., 8, Part I, 71 (1966).
- J. D. Cocker, T. G. Halsall, and A. Bowers, J. Chem. Soc., 1956, 4259.
- 7) We used a different numbering system, according to the Triterpenoid Numbering System [M. Benson, W. D. Nes, and W. R. Landery, J. Natural Products, 46, 274 (1983)] shown in Chart 1, from the system proposed by Sukh DeV in order to compare the NMR data with those of the hopane group.
- 8) The ¹H-NMR (270 MHz) signal of H-6 β in zeorin (9) appears at δ 3.961 (1H, ddd, J=10.4, 10.4, 4.3 Hz).
- R. T. Iyer, K. N. N. Ayengar, and S. Rangaswami, *Indian. J. Chem.*, 10, 482 (1972).
- a) H. Khan, A. Zaman, G. L. Chatty, A. S. Gupta, and S. DeV, Tetrahedron Lett., 1971, 4443; b) A. S. Gupta, S. DeV, M. Sangare, B. Septe, and G. Lucas, Bull. Soc. Chim. Fr., 1976, 1879.
- 11) The fern, from which cheilanthatriol (1) was isolated, was identified as *Aleuritopteris pulveracea* FEE by Professor Y. Saiki.
- J. A. Elix, A. A. Whitton, and A. J. Jones, Aust. J. Chem., 35, 641 (1982); A. L. Wilkin, K. J. Ronaldson, P. M. Jager, and P. W. Bird, ibid., 40, 1713 (1987).
- 13) The ¹H-NMR (270 MHz) solvent shift values $[\delta(C_6D_6)-\delta(CDCl_3)]$ of the methyl groups in zeorinone (11) are +0.28 (H-23), +0.17 (H-24), +0.01 (H-25), and -0.04 (H-26), while those in hydroxyhopane (19) are +0.07 (H-23), +0.07 (H-24), +0.05 (H-25) and +0.03 (H-26).
- 14) D. H. Williams and D. A. Wilson, J. Chem. Soc. (B), 1966, 144.
- H. Ageta, K. Shiojima, and K. Masuda, The 95th Annual Meeting of Pharmaceutical Society of Japan, Nishinomiya, April 1975; H. Ageta, K. Masuda, and Y. Tanaka, Shoyakugaku Zasshi, 35, 259 (1981).