Synthesis and Platelet Aggregation-Inhibitory Activities of Novel 3-(2-Oxopropylidene) azetidin-2-one Derivatives. ${\bf I}^{(1)}$

Yutaka Kawashima, Masakazu Sato, *. Yuuichi Hatada, Jun Goto, Yoshimoto Nakashima, Katsuo Hatayama and Shiroshi Shibuya b

Research Center, Taisho Pharmaceutical Co., Ltd., Yoshino-cho, Ohmiya, Saitama 330, Japan and Tokyo College of Pharmacy, 1432–1, Horinouchi, Hachioji, Tokyo 192–03, Japan. Received April 22, 1989

Treatment of (E)-3-(2-hydroxypropylidene)-4-methyl-1-phenylazetidin-2-one (11) with 10% Pd/C gave (E)-(12), (Z)-3-(2-oxopropylidene)-4-methyl-1-phenylazetidin-2-one (13), 3,4-cis-(14a) and 3,4-trans-3-(2-oxopropyl)-4-methyl-1-phenylazetidin-2-one (14b). Among them, 12 and 13 were found to show potent inhibitory activities against rabbit platelet-rich plasma aggregation induced by adenosine diphosphate or collagen. Ring-expanded homologous derivatives and an acyclic analogue of 12 were also synthesized and tested for the biological activities. The azetidin-2-one skeleton bearing a 2-oxoalkylidene moiety at the 3 position was found to be essential for the platelet aggregation inhibitory activities of these compounds.

Keywords 3-(2-oxopropylidene)-4-methyl-1-phenylazetidin-2-one; azetidin-2-one; platelet aggregation inhibition; adenosine diphosphate; collagen; structure-activity relationship

In the previous paper in this series, $^{2)}$ we have demonstrated that 1-aryl-3-(2-hydroxyalkylidene)azetidin-2-ones are easily converted to furo[3,2-c]quinolines or 2-anilinomethyl-2,3-butenolides under acidic conditions (Chart 1). In the course of the further investigation of 4-substituted-1-phenyl-2-azetidinones, we have synthesized (E)-3-(2-hydroxypropylidene)-4-methyl-1-phenylazetidin-2-one (11) to examine the acid catalyzed rearrangement. Although 11 was not converted to either the corresponding 4-methylfuro[3,2-c]quinoline or 2-(1-anilinoethyl)-4-methyl-2,3-butenolide under acidic conditions, the novel reactivity toward 10% Pd/C was found to give the enone (12) and isomerized Z form enone (13) accompanied with rearranged ketone derivatives (14a, b).

During the biological evaluation of these monocyclic 1-arylazetidin-2-ones, 12 and 13 were found to show potent inhibitory activities against rabbit platelet-rich plasma (PRP) aggregation induced by adenosine diphosphate (ADP) or collagen *in vitro*.

Since some β -lactam antibiotics such as nafcillin or penicillin G are reported to show platelet aggregation-inhibitory activities at higher concentrations, ³⁾ the ring-expanded homologous compounds such as 3-(2-oxopropylidene)-1-phenylpyrrolidin-2-one (21) or 3-(2-oxopropylidene)-1-phenylpyrrolidin-2-one (21)

R₁ OH H 1) H⁺ 2) Pd-C (-H₂)
$$R_1 = H$$
, Me, Et 1 $R_1 = H$, Ph R₁R₂ = + CH₂+4

pylidene)-1-phenylpiperidin-2-one (28), and an acyclic analogue (36) were also synthesized to examine the structure activity relationships.

Furthermore, oxidation of the enol acid (30) obtained by alkaline hydrolysis of 11 with activated MnO_2 was examined to prepare the acyclic analogue of 12, but the pyrrole derivative (32) was obtained instead of the corresponding enone. Synthesis of these compounds and the results of biological evaluations are described in this paper.

Chemistry The synthesis of 11 was effectively accomplished in a stereoselective manner according to the method reported previously, as shown in Chart 2. Lithiation of 4methyl-1-phenylazetidin-2-one (5)4) with lithium diisopropylamide (LDA) followed by condensation with the ester (6)⁵⁾ in tetrahydrofuran (THF) at -78 °C gave 3-acylazetidin-2-one derivatives (7a, b) as a mixture of inseparable diastereomers involving the SCH₃ group in 3:2 ratio. The stereochemistry of 7a and 7b was determined to be 3,4-trans based on the coupling constant (3 Hz) of 7a and 7b between C₃-H and C₄-H. Reduction of the ketone moiety of 7a, b by NaBH₄ in MeOH at $-78\,^{\circ}\text{C}$ proceeded in a stereoselective manner through the sodium cationchelated intermediate⁶⁾ to give the corresponding α alcohols (8a, b) in a 3:2 ratio in 89% yield, and the configuration of the hydroxy group was confirmed by the selective formation of the E form olefin 11 as described below. The major isomer (8a) was isolated by crystallization of the oily product from ether as a colorless amorphous solid in 33% yield, but the minor isomer (8b) obtained from the mother liquor as a viscous oil was contaminated with 8a, and could not be further purified. Then 8a was treated with methyl iodide in MeOH to give the sulfonium salt (9), which was treated successively with tert-BuOK at 0 °C to give 11 as a single isomer through ring cleavage of the oxirane intermediate (10). The geometry of the olefin moiety of 11 was determined to be E from the chemical shift of the olefinic proton signal (δ : 6.22) observed in the proton nuclear magnetic resonance (1H-NMR) spectrum.

Subsequently, the acid catalyzed rearrangement of 11 was examined by heating with CF₃CO₂H followed by treatment with Pd/C in toluene under reflux. Although neither the expected furo[3,2-c]quinoline nor the butenolide

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derivative was obtained by the acid treatment of 11, the oxidation of the allyl alcohol moiety to a mixture of the enone derivative (12) and its isomer (13), accompanied with the formation of the rearranged ketone derivatives (14a, b) proceeded upon the treatment of 11 with Pd/C in toluene under reflux. The geometry of the enone moiety of 12 was determined to be E and that of 13 to be Z based on the characteristic olefinic proton signals observed in their ¹H-NMR spectra. The olefinic proton of 12 resonated at lower field (δ 6.68 ppm) than that of 13 (δ 5.98 ppm) because of the deshielding effect of the carbonyl group of the azetidin-2-one ring. For the further confirmation of the structures of 12 and 13, alternative syntheses of 12 and 13 from 11 were investigated. Thus, the oxidation of 11 with 2,3-dichloro-5,6-dicyanoquinone (DDQ) was examined to yield 12 (48%) yield) and 13 (9.6% yield). The stereochemistry of 14a was determined as 3,4-cis and that of 14b was determined as 3,4trans based on the comparison of the coupling constant of 14a (5 Hz) and 14b (2 Hz) between C_3 -H and C_4 -H. The above conclusion was confirmed by the preparation of the cis isomer 14a from 12 or 13 by hydrogenation on Pd/C

(Chart 3).

In addition, the isomerization of 12 to 13, and the dehydrogenation of 14a or 14b to 12 or 13 were not observed under the same reaction conditions as used to synthesize them from 11.

The synthesis of ring expanded homologous derivatives of 12 was examined. Lithiation of 1-phenylpyrrolidin-2one (15) with LDA followed by condensation with the ester $(16)^{7}$ in THF at -78 °C afforded the ketone (17). The ketone moiety of 17 was reduced by NaBH₄ in MeOH at 0 °C to give a separable mixture of isomers (18a; 33% yield) and (18b; 64% yield). The relative configuration of these isomers was determined to be as shown in Chart 5 based on the comparison of the signals of methine protons bearing a hydroxy group in their ¹H-NMR spectra and on the observation of intramolecular hydrogen bonding in the infrared (IR) spectrum of 18b. The methine proton of the polar isomer (18a) resonated at lower field (δ 4.33 ppm) than that of the less polar isomer (18b) (δ 3.80 ppm) because of the deshielding effect of the carbonyl group of the pyrrolidin-2-one February 1990 395

ring. The presence of intramolecular hydrogen bonding between the hydroxy group and carbonyl group of 18b is indicated by the low wave number shift of the carbonyl and hydroxy absorption bands in the IR spectrum. These spectral differences between the two isomers seemed to be caused by the inhibition of free rotation of the substituent at the 3-position based on the steric hindrance between the bulky acetal group and the lactam ring. Each of 18a and 18b was treated with CH₃SO₂Cl to give the corresponding mesylate (19a) and (19b). Each of 19a or 19b was converted to 3-[(2,2ethylenedioxy)propylidene]-1-phenylpyrrolidin-2-one (20) as a single isomer by treatment with excess 1.8-diazabicyclo[5.4.0]undec-7-ene (DBU) in benzene under reflux. The geometry of the enone moiety of 20 was found to be E based on the low field olefinic proton signals $(\delta 6.59)$ in the ¹H-NMR spectrum. Deprotection of the acetal of 20 to the exo olefin (21) was done by a short treatment with 80% CF₃CO₂H at 0 °C. Upon treatment with CF₃CO₂H at room temperature, 21 was isomerized to the more stable endo isomer (22) in 77% yield (Chart

4).

Chart 5

The piperidin-2-one homologue (28) was also prepared from 1-phenylpiperidin-2-one (23) in the same way as described for the synthesis of 21. Reduction of the ketone (24) with NaBH₄ in MeOH gave a mixture of separable isomers (25a; 15% yield) and (25b; 37% yield). The spectral data of the polar isomer (25a) and the less polar isomer (25b) were quite similar to those of 18a and 18b, respectively. Mesylation of 25a or 25b followed by elimination with DBU gave 27 as a single product in good yield. Deprotection of 27 with 80% CF₃CO₂H at 0 °C for 0.5 min afforded the enone (28) in 31% yield. When this reaction was continued for 1.5 h, the *endo* isomer (29) was formed in 36% yield (Chart 6).

Finally, the synthesis of acyclic analogues possessing the same chromophor as 12 or 13 was examined. An attempt to cleave the azetidin-2-one ring of 12 under basic conditions failed, but the hydrolysis of 11 by aqueous KOH proceeded smoothly to give the desired amino acid derivative (30) in good yield. Oxidation of the allyl alcohol moiety of 30 was examined, but the enone derivative (31) was not isolated

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and ring closure to the pyrrole derivative (32) proceeded even under mild conditions such as oxidation with Attemburrow's activated MnO_2^{8} in ether at room temperature. The structure of 32 was confirmed by direct comparison with authentic 32 prepared by condensation of the diketone (33)⁹ with aniline. The 3-benzoylacrylamide derivative (36) was synthesized from 35¹⁰ and aniline by N,N'-dicyclohexylcarbodiimide (DCC) condensation, but in low yield (Chart 7).

Pharmacological Results The platelet aggregation inhibitory activities of the compounds synthesized were tested on rabbit PRP *in vitro* by the method of Born, and the results are summarized in Table I. 3-(2-Oxopropylidene) azetidin-2-one derivatives (12 and 13) both dosedependently inhibited the first wave of ADP aggregation and collagen aggregation of rabbit PRP (Fig. 1). The IC₅₀ values of 12 for ADP or collagen aggregation were calculated to be 29 or $27 \mu M$, and those of 13 were 52 or $19 \mu M$.

Fig. 1. Platelet Aggregation-Inhibitory Activities of 12 and 13 on Rabbit PRP

Table I. Platelet Aggregation Inhibitory Activities on Rabbit PRP Aggregation Induced by ADP (5 μ m) or Collagen (5 μ g/ml) in Vitro

Compound	IC ₅₀ (μ _M)	
	ADP	Collager
11	> 100	> 100
12	29.0	27.0
13	52.0	19.0
14a	> 100	>100
14b	>100	> 100
21	>100	> 100
22	>100	> 100
28	> 100	> 100
29	> 100	> 100
32	>100	> 100
36	> 100	> 100
Aspirin	> 100	35.0

However, the other compounds synthesized showed no apparent inhibitory activity at the concentration of $100 \,\mu\text{M}$. From these observations, it was clear that the azetidin-2-one skeleton bearing a 2-oxoalkylidene moiety at the 3 position was essential for the platelet aggregation inhibitory activities of 12 and 13.

Experimental

Melting points were determined on a Mettler FP-60 melting point apparatus. IR spectra were taken on a Jasco X-1A spectrometer. $^1\text{H-NMR}$ spectra were recorded with a Varian XL-200 spectrometer (Me_4Si as an internal standard, δ value), and the following abbreviations are used: singlet (s), broad singlet (br s), doublet (d), double doublet (dd), double doublet (ddd), triplet (t), quartet (q), double quartet (dq), double triplet (dt), multiplet (m). Mass spectra (MS) were taken on a Hitachi M-80A spectrometer. Microanalytical data were obtained by using a Carlo Elba 1106R or a Perkin-Elmer 240C elemental analyzer. For column chromatography, Wakogel 200 (Wako Pure Chemical) was used, and thin layer chromatography was performed on silica gel pre-coated plates (Merck, Kieselgel 60F-254).

4-Methyl-3-(2-methylthio)propionyl-1-phenylazetidin-2-one (7) A solution of 4-methyl-1-phenylazetidin-2-one (5) (96.7 g, 0.60 m) in THF (505 ml) was added to a solution of LDA prepared from diisopropylamine $(60.6\,\mathrm{g},\ 0.60\,\mathrm{M})$ and $n\text{-BuLi}\ (385\,\mathrm{ml}\ \mathrm{of}\ 1.6\,\mathrm{N}\ \mathrm{hexane}\ \mathrm{solution},\ 0.67\,\mathrm{M})$ in THF (720 ml) at -78 °C. The reaction mixture was stirred for 30 min, then a solution of ethyl 2-(methylthio)propionate (6) (88.8 g, 0.60 m) in THF (250 ml) was added dropwise over 1 h. The whole was stirred for 30 min at the same temperature, then water was added, and the reaction mixture was extracted with CHCl₃. The extract was dried (Na₂SO₄), and evaporated in vacuo, and the residue was purified by passing it through a short silica gel column (benzene) to give (7a, b) (78 g, yield: 50%) as a mixture of two diastereomers in a 3:2 ratio. A colorless viscous oil. IR (film): 1745 (NC = O), 1695 (C = O), 1600 (C = C) cm⁻¹. 1 H-NMR (CDCl₃) δ : 1.40 and 1.48 (3H, d, J = 7 Hz, CH₃CHSCH₃), 1.56 and 1.58 (3H, d, J = 7 Hz, C₄- CH_3), 1.94 and 1.96 $\overline{(3H)}$, s, SCH_3), 3.50 and 3.70 (1H, q, $J=7\,Hz$, $\underline{\text{CH}}$ SCH₃), 4.20 and 4.52 (1H, d, J = 3 Hz, C₃-H), 4.40 and 4.55 (1H, dq, $J_a = 7 \text{ Hz}$, $J_d = 3 \text{ Hz}$, C_4 -H), 7.10 (1H, m, aromatic proton), 7.28 (4H, m, aromatic protons).

3-[1-Hydroxy-(2-methylthio)propyl]-4-methyl-1-phenylazetidin-2-one (8a, b) A solution of the mixture of 7a and 7b (31 g, 0.118 M) in MeOH (150 ml) was added dropwise to a stirred solution of NaBH₄ (8.3 g, 0.219 M) in MeOH (700 ml) at -78 °C over 2 h. The reaction mixture was stirred for 1h at -78 °C, then AcOH (32 ml, 0.56 M) was added, and the whole was allowed to warm to room temperature. Water was added and the reaction mixture was extracted with CHCl₃. The extract was washed with aqueous NaHCO₃ and brine, dried (MgSO₄), and evaporated *in vacuo* to give a mixture of two diastereomers (27.8 g, 89%) in a 3:2 ratio as a viscous oil. Crystallization of the oily product from ether gave a major isomer 8a (10.5 g, yield: 33%) as a colorless amorphous solid, mp 109.5—110.5 °C. IR (KBr): 3400 (OH), 1725 (NC=O), 1600 (C=C) cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.41 (3H, d, J=7 Hz, CH₃CHSCH₃), 1.56 (3H, d, J=7 Hz, CH₂CH₃), 2.16 (3H, s, SCH₃), 2.84 ($\overline{1}$ H, d, J=5 Hz, OH), 2.90 (1H, q, J=7 Hz, CHSCH₃), 3.15 (1H, dd, J=5, 3 Hz, C₃-H), 3.98 (1H, dt, J₁=

5 Hz, $J_{\rm d}$ =7 Hz, CHOH), 4.29 (1H, dt, $J_{\rm t}$ =7 Hz, $J_{\rm d}$ =3 Hz, C₄-H), 7.10 (1H, m, aromatic proton), 7.3—7.5 (4H, m, aromatic protons). EIMS m/z: 265 (M⁺). Anal. Calcd for C₁₄H₁₉NSO₂: C, 63.36; H, 7.22; N, 5.28. Found: C, 63.28; H, 7.26; N, 5.27. The mother liquor yielded a minor isomer **8b** contaminated with the major isomer as a viscous oil. ¹H-NMR (CDCl₃) δ : 1.43 (3H, d, J=7 Hz, CH₃CHSCH₃), 1.55 (3H, d, J=7 Hz, C₄-H), 2.11 (3H, s, SCH₃), 3.15 (2H, m, C₃-H), 3.24 (1H, br s, OH), 3.85 (1H, dd, J=8, 5 Hz, CHOH), 4.25 (1H, dt, $J_{\rm t}$ =7 Hz, $J_{\rm d}$ =3 Hz, C₄-H), 7.10 (1H, m, aromatic proton), 7.3—7.5 (4H, m, aromatic protons).

(E)-3-(2-Hydroxypropylidene)-4-methyl-1-phenylazetidin-2-one (11) MeI (4.82 g, $0.034\,\mathrm{M}$) was added to a solution of 8a (9 g, $0.034\,\mathrm{M}$) in MeOH (330 ml), and the mixture was heated under reflux for 6 h. After removal of the solvent in vacuo, the resulting sulfonium salt (9) was dissolved in absolute EtOH (120 ml), and tert-BuOK (7.6 g, 67.7 mm) was added in small portions at 0 °C. The reaction mixture was stirred at room temperature for 1 h, then evaporated in vacuo, and the resulting residue was extracted with CHCl3. The extract was washed with water, aqueous NaHCO3 and brine successively, dried (MgSO4) and evaporated in vacuo to give 11 (4.6 g, yield: 62%), which was recrystallized from ether to give a colorless amorphous solid, mp 89.5—91.5 °C. IR (KBr): 3440 (OH), 1725 (N=CO), 1590 (C=C) cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.37 (3H, d, J=7Hz, CH_3 CHOH), 1.64 (3H, d, J = 7 Hz, C_4 - CH_3), 2.28 (1H, br s, OH), 4.60 (1H, $\overline{\text{m, CHOH}}$), 4.75 (1H, q, J = 7 Hz, C_4 -H), 6.22 (1H, d, J = 5 Hz, HC = C), 7.10 (1H, m, aromatic proton), 7.38 (4H, m, aromatic protons). EIMS m/z: 217 (M⁺). Anal. Calcd for C₁₃H₁₅NO₂: C, 71.87; H, 6.96; N, 6.44. Found: C, 71.88; H, 6.90; N, 6.38.

The Reaction of 11 with 10% Pd/C in Toluene under Reflux A mixture of 11 (6.1 g, 0.028 M), 10% Pd/C (3 g), and toluene (120 ml) was heated under reflux for 3 h. After removal of the Pd/C by filtration, the filtrate was evaporated in vacuo and the oily residue was submitted to silica gel column chromatography (CH $_2$ Cl $_2$) to give 12 (1.0 g, yield: 16.7%), 13 (0.2 g, yield: 3.3%), **14a** (1.45 g, yield: 23.8%) and **14b** (0.95 g, yield: 11%). **12**: Yellow prisms from EtOH, mp 120—122 °C. IR (KBr): 1745 (NC=O), 1650 (C= CC=O), 1590 (C=C) cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.63 (3H, d, J=7 Hz, C_4 - CH_3), 2.37 (3H, s, $CH_3C = O$), 4.98 (1H, dq, $J_d = 2$, $J_q = 7$ Hz, C_4 -H), 6.67 (1H, d, J=2 Hz, HC=C), 7.15 (1H, m, aromatic proton), 7.42 (4H, m, aromatic protons). EIMS m/z: 215 (M $^+$). Anal. Calcd for $C_{13}H_{13}NO_2$: C, 72.54; H, 6.09; N, 6.50. Found: C, 72.54, H, 6.10; N, 6.49. 13: Pale yellow needles from ether, mp 114-115.5 °C. IR (KBr): 1740 (C=O), 1660 (C=CC=O), 1590 (C=C) cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.61 (3H, d, J=7 Hz, C₄-CH₃), 2.74 (3H, s, CH₃C=O), 4.68 (1H, q, J=7 Hz, C₄-H), 5.98 (1H, s, HC=C), 7.20 (1H, m, aromatic proton), 7.45 (4H, m, aromatic protons). EIMS m/z: 215 (M⁺). Anal. Calcd for $C_{13}H_{13}NO_2$: C_1 72.54; H, 6.09; N, 6.50. Found: C, 72.61; H, 6.14; N, 6.44. 14a: Colorless needles from EtOH, mp 114.5—116.5 °C. IR (KBr): 1745 (NC=O), 1705 (C=O), 1590 (C=C) cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.32 (3H, d, J=7 Hz, C_4 -CH₃), 2.24 (3H, s, CH₃C=O), 2.84 (1H, dd, J=20, 12 Hz, HCC=O), 3.04 (1H, dd, J=20, 5 Hz, HCC=O), 3.82 (1H, dt, $J_t=5$ Hz, $J_d=12$ Hz, C_3 -H), 4.40 (1H, dq, $J_q = 7$ Hz, $J_d = 5$ Hz, C_4 -H), 7.10 (1H, m, aromatic protons), 7.38 (4H, m, aromatic protons). EIMS m/z: 217 (M+). Anal. Calcd for C₁₃H₁₅NO₂: C, 71.87; H, 6.96; N, 6.44. Found: C, 71.88; H, 6.98; N, 6.55. 14b: A colorless viscous oil, IR (film): 1745 (NC=O), 1705 (C=O), 1590 (C=C) cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.62 (3H, d, J=6 Hz, C_4 -CH₃), 2.21 (3H, s, CH₃C=O), 2.70 (1H, dd, J=18, 12 Hz, HCC=O), 2.95 (1H, ddd, J=12, 3.5, 2Hz, C_3 -H), 3.13 (1H, dd, J=18, 3.5Hz, HCC=O), 3.79 (1H, dq, $J_q = 6 \text{ Hz}$, $J_d = 2 \text{ Hz}$, C_4 -H), 7.10 (1H, m, aromatic proton), 7.35 (4H, m, aromatic protons). EIMS m/z: 217 (M⁺).

Oxidation of 11 with DDQ A mixture of 11 (20.3 mg, 0.093 mM), DDQ (28 mg, 0.12 mM), and dioxane (0.35 ml) was stirred at room temperature for 20 h. The reaction mixture was evaporated *in vacuo* and separated by silica gel column chromatography (CH₂Cl₂) to give 12 (16.2 mg, yield: 48%) and 13 (3.2 mg, yield: 9.6%).

Hydrogenation of 12 A mixture of **12** (50 mg, 0.23 mM), 10% Pd/C (30 mg), and EtOH (50 ml) was stirred at room temperature in a hydrogen atmosphere under normal pressure to give **14a** (42 mg, yield: 83%).

3-[2,2-(Ethylenedioxy)propionyl]-1-phenylpyrrolidin-2-one (17) A solution of 1-phenylpyrrolidin-2-one (15) (10 g, 62 mm) in THF (10 ml) was added dropwise to a stirred solution of LDA, prepared from diisopropylamine (8.94 ml, 62 mm) and n-BuLi (38.3 ml of 1.6 n hexane solution, 62 mm) in THF (90 ml) at -78 °C. The reaction mixture was stirred for 10 min, then a solution of ethyl 2,2-ethylenedioxypropionate (16) (9.9 g, 62 mm) in THF (10 ml) was added. The whole was stirred for 30 min and then poured into water, and extracted with CHCl₃. The extract was washed with brine, dried (MgSO₄) and evaporated *in vacuo* to give 17 (13.8 g, yield: 81%), which was recrystallized from CHCl₃-ether to give

colorless prisms, mp 84—86 °C. IR (KBr): 1730 (C=O), 1680 (NC=O) cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.58 (3H, s, CH₃), 2.38 (2H, m, C₄-H), 3.92 (2H, m, C₅-H), 4.10 (4H, m, OCH₂CH₂O), 4.37 (1H, dd, J=8, 9 Hz, C₃-H), 7.18 (1H, m, aromatic proton), 7.39 (2H, m, aromatic protons), 7.60 (2H, m, aromatic protons). EIMS m/z: 275 (M⁺). *Anal*. Calcd for C₁₅H₁₇NO: C, 65.44; H, 6.22; N, 5.09. Found: C, 65.42; H, 6.22; N, 5.06.

 (R^*) -3- $[(R^*)$ -2,2-(Ethylenedioxy)-1-hydroxypropyl]-1-phenylpyrrolidin-2-one (18a) and (R^*) -3- $[(S^*)$ -2,2-(Ethylenedioxy)-1-hydroxypropyl)]-1phenylpyrrolidin-2-one (18b) A solution of 17 (11 g, 40 mm) in MeOH (50 ml) was added dropwise to a solution of NaBH₄ (1.8 g, 48 mm) in MeOH (100 ml) at 0 °C. The mixture was stirred at 0 °C for 1 h, then AcOH (8 ml) was added and the whole was poured into water and extracted with CH2Cl2. The extract was washed with water and brine, dried (MgSO₄), and evaporated in vacuo and the residue was separated by silica gel column chromatography (CH₂Cl₂) to give 18a (3.65 g, yield: 33%) and 18b (7.1 g, yield: 64%). 18a: Colorless prisms from CHCl₃ether, mp 129—130°C. IR (KBr): 3380 (OH), 1670 (C=O) cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.38 (3H, s, CH₃), 2.18 and 2.50 (each 1H, m, C₄-H), 2.40 (1H, d, J=4 Hz, OH), 2.85 (1H, brt, J=9.5 Hz, C_3 -H), 3.81 (2H, dd, J=9.5, 5 Hz, C₅-H), 4.04 (4H, m, OCH₂CH₂O), 4.33 (1H, dd, J=4, 2 Hz, HOCH), 7.15 (1H, m, aromatic proton), 7.38 (2H, m, aromatic protons), 7.69 (2H, m, aromatic protons). EIMS m/z: 277 (M⁺). Anal. Calcd for C₁₅H₁₉NO₄: C, 64.97; H, 6.91; N, 5.05. Found: C, 64.96; H, 6.91; N, 5.05. **18b**: Colorless prisms from CHCl₃-ether, mp 94—95 °C. IR (KBr): 3350 (OH), 1650 (NC=O) cm⁻¹. 1 H-NMR (CDCl₃) δ : 1.42 $(3H, s, CH_3)$, 2.12 and 2.30 (each 1H, m, C₄-H), 2.78 (1H, dt, $J_d = 13$ Hz, $J_t = 8 \text{ Hz}, C_3 - H), 3.80 \text{ (1H, dt, } J_d = 2.5 \text{ Hz}, J_t = 10 \text{ Hz}, \text{ HO}\underline{\text{CH}}), 3.85 \text{ (2H, } J_t = 10 \text{ Hz})$ m, C₅-H), 4.03 (4H, m, OCH₂CH₂O), 5.78 (1H, d, J=2 Hz, OH), 7.20 (1H, m, aromatic proton), 7.40 (2H, m, aromatic protons), 7.61 (2H, m, aromatic protons). EIMS m/z: 277 (M⁺). Anal. Calcd for $C_{15}H_{19}NO_4$: C, 64.97; H, 6.91; N, 5.05. Found: C, 64.95; H, 6.94; N, 5.06.

 (R^*) -3-[(R^*) -2,2-(Ethylenedioxy)-1-methylsulfonylpropyl]-1-phenylpyrrolidin-2-one (19a) A solution of CH₃SO₂Cl (1.1 ml, 14 mm) in CH₂Cl₂ (5 ml) was added dropwise to a solution of 18a (3.3 g, 12 mm) and Et₃N (3.3 ml, 24 mm) in CH₂Cl₂ (50 ml) at 0 °C. The reaction mixture was stirred at room temperature for 1 h, then poured into water, washed with aqueous NaHCO₃ and brine, dried (MgSO₄), and evaporated *in vacuo*. The residue was purified by silica gel column chromatography (CH₂Cl₂: ether = 10: 1) to give 19a (3.1 g, yield: 74%) as a colorless oil. Rfilm): 1685 (NC=O) cm⁻¹. H-NMR (CDCl₃) δ: 1.50 (3H, s, C-CH₃), 2.40 (2H, m, C₄-H), 3.10 (1H, m, C₃-H), 3.18 (3H, s, SO₂CH₃), 3.80 (2H, m, C₅-H), 4.05 (4H, m, OCH₂CH₂O), 4.93 (1H, d, J=5 Hz, HCOSO₂CH₃), 7.18 (1H, m, aromatic proton), 7.38 (2H, m, aromatic protons), 7.60 (2H, m, aromatic protons). EIMS m/z: 355 (M⁺).

(*R**)-3-[(*S**)-2,2-(Ethylenedioxy)-1-methylsulfonylpropyl]-1-phenylpyrrolidin-2-one (19b) Prepared from 18b in the same manner as used for 19a. Colorless prisms from ether, mp 105—107 °C. IR (KBr): 1690 (NC=O) cm⁻¹. ¹H-NMR (CDCl₃) δ: 1.43 (3H, s, CCH₃), 2.40 (2H, m, C₄-H), 3.03 (3H, s, SO₂CH₃), 3.12 (1H, m, C₃-H), 3.85 (2H, dd, J=10, 7 Hz, C₅-H), 4.10 (4H, m, OCH₂CH₂O), 5.25 (1H, d, J=2 Hz, HCOSO₂CH₃), 7.18 (1H, m, aromatic proton), 7.38 (2H, m, aromatic protons), 7.65 (2H, m, aromatic protons). EIMS m/z: 355 (M⁺).

(E)-3-[(2,2-Ethylenedioxy)propylidene]-1-phenylpyrrolidin-2-one (20) A mixture of 19a (2.5 g, 7 mM), DBU (3.2 g, 21 mM), and benzene (50 ml) was heated under reflux for 2 h, washed twice with water, dried (MgSO₄), and evaporated *in vacuo* to give 20 (1.5 g, 82%), which was recrystallized from hexane-CHCl₃ to give colorless prisms, mp 110—112 °C. IR (KBr): 1685 (C=O) cm⁻¹. 14 -NMR (CDCl₃) δ : 1.56 (3H, s, CH₃), 3.05 (2H, dt, J_d =2 Hz, J_t =7 Hz, C_4 -H), 3.90 (2H, t, J=7 Hz, C_5 -H), 4.00 (4H, m, OCH₂CH₂O), 6.59 (1H, t, J=2 Hz, C=CH), 7.19 (1H, m, aromatic proton), 7.41 (2H, m, aromatic protons), 7.75 (2H, m, aromatic protons). EIMS m/z: 324 (M⁺). Anal. Calcd for $C_{15}H_{17}NO_3$: C, 69.48; H, 6.60; N, 5.39. Found: C, 69.44; H, 6.60; N, 5.38.

(E)-3-(2-Oxopropylidene)-1-phenylpyrrolidin-2-one (21) Compound 20 (1.5 g, 4.67 mm) was added as a single portion to 80% CF₃CO₂H at 0°C. After being stirred for 1 min, the reaction mixture was poured into 5% Na₂CO₃ solution and extracted with CH₂Cl₂. The extract was washed with water and brine, dried (MgSO₄), and evaporated *in vacuo* to give 21 (1.2 g, yield: 100%), which was recrystallized from CH₂Cl₂-ether to give pale yellow prisms, mp 152—153 °C. IR (KBr): 1660 (NC=O), 1630 (C=O) cm⁻¹. H-NMR (CDCl₃) δ : 2.37 (3H, s, CH₃), 3.32 (2H, dt, J_1 =7 Hz, J_4 =3 Hz, C₄-H), 3.96 (2H, t, J=7 Hz, C₅-H), 7.08 (1H, t, J=3 Hz, C=CH), 7.23 (1H, m, aromatic protons). EIMS m/z: 215 (M⁺). Anal. Calcd for C₁₃H₁₃NO₂: C, 72.54; H, 6.09; N, 6.50. Found: C, 72.68; H, 6.32; N, 6.55.

3-(2-Oxopropyl)-2-oxo-1-phenyl-3-pyrroline (22) A mixture of 21 (560 mg, 2.6 mM) and 80% CF₃CO₂H (5 ml) was stirred at room temperature for 2h, diluted with CH₂Cl₂, washed with aqueous NaHCO₃ and brine, dried (MgSO₄), and evaporated *in vacuo*. The residue was purified by silica gel column chromatography (CH₂Cl₂) to give 22 (430 mg, yield: 77%), which was recrystallized from CH₂Cl₂-iso-Pr₂O to give yellow needles, mp 95—96 °C. IR (KBr): 1710 (C=O), 1660 (NC=O) cm⁻¹. ¹H-NMR (CDCl₃) δ : 2.30 (3H, s, CH₃), 3.52 (2H, d, J = 2 Hz, CH₂C=O), 4.23 (2H, m, C₅-H), 7.10 (1H, m, C=CH), 7.16 (1H, m, aromatic proton), 7.40 (2H, m, aromatic protons), 7.86 (2H, m, aromatic protons). EIMS m/z: 215 (M⁺). *Anal.* Calcd for C₁₃H₁₃NO₃: C, 72.54; H, 6.09; N, 6.50. Found: C, 72.26; H, 6.08; N, 6.35.

Compounds 24 to 29 were prepared in the same manner as described above

3-(2,2-Ethylenedioxypropionyl)-1-phenylpiperidin-2-one (**24**): A colorless viscous oil. IR (KBr): 1730 (C=O), 1635 (NC=O) cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.53 (3H, s, CH₃), 1.95 (2H, m, C₄-H), 2.15 (2H, m, C₅-H), 3.65 (2H, m, C₆-H), 4.00 (4H, m, OCH₂CH₂O), 4.20 (1H, m, C₃-H), 7.20—7.50 (5H, m, aromatic protons). EIMS m/z: 289 (M⁺).

 (R^*) -3-[(R^*) -2,2-(Ethylenedioxy)-1-hydroxypropyl)]-1-phenylpiperidin-2-one (**25a**): Colorless prisms from CH₂Cl₂, mp 69—70 °C. IR (KBr): 3400 (OH), 1625 (C=O) cm⁻¹. ¹H-NMR (CDCl₃) δ: 1.38 (3H, s, CH₃), 2.10 (4H, m, C₄-H and C₅-H), 2.80 (1H, dt, J_d =3 Hz, J_t =9 Hz, C₃-H), 2.96 (1H, d, J=7 Hz, OH), 3.65 (2H, m, C₄-H), 4.04 (4H, m, OCH₂CH₂O), 4.50 (1H, dd, J=7, 3 Hz, <u>HC</u>OH), 7.20—7.50 (5H, m, aromatic protons). EI MS m/z: 291 (M⁺). Anal. Calcd for C₁₆H₂₁NO₄: C, 65.96; H, 7.26; N, 4.80. Found: C, 65.76; H, 7.13; N, 4.93.

(*R**)-3-[(*S**)-2,2-(Ethylenedioxy)-1-hydroxypropyl)]-1-phenylpiperidin-2-one (**25b**): A colorless viscous oil. IR (film): 3320 (OH), 1620 (C=O) cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.41 (3H, s, CH₃), 1.80—2.10 (4H, m, C₄-H and C₅-H), 2.70 (1H, m, C₃-H), 3.65 (2H, m, C₆-H), 3.82 (1H, dd, *J*=8, 5 Hz, <u>HC</u>OH), 4.00 (4H, m, OCH₂CH₂O), 5.43 (1H, d, *J*=5 Hz, OH), 7.20—7.50 (5H, m, aromatic protons). EIMS *m/z*: 291 (M⁺).

 (R^*) -3-[(R^*) -2,2-(Ethylenedioxy)-1-methylsulfonyloxy)]-1-phenylpiperidin-2-one (**26a**): A colorless viscous oil. IR (film): 1680 (C=O). ¹H-NMR (CDCl₃) δ: 1.40 (3H, s, C-CH₃), 2.10 (4H, m, C₄-H and C₅-H), 2.95 (1H, m, C₃-H), 3.22 (3H, s, SO₂CH₃), 3.65 (2H, m, C₆-H), 4.05 (4H, m, OCH₂CH₂O), 4.95 (1H, d, J=5 Hz, HCOSO₂CH₃), 7.2—7.5 (5H, m, aromatic protons). EIMS m/z: 369 (M⁺).

 (R^*) -3-[(S^*)-2,2-(Ethylenedioxy)-1-methylsulfonyloxy)]-1-phenylpiperidinone (**26b**): A colorless viscous oil. IR (film): 1680 (C=O). ¹H-NMR (CDCl₃) δ: 1.41 (3H, s, C-CH₃), 2.10 (4H, m, C₄-H and C₅-H), 3.01 (1H, m, C₃-H), 3.05 (3H, s, SO₂CH₃), 3.65 (2H, m, C₆-H), 4.07 (4H, m, OCH₂CH₂O), 5.33 (1H, d, J=2 Hz, $HOSO_2CH_3$), 7.2—7.5 (5H, m, aromatic protons). EIMS m/z: 369 (M^+).

(E)-3-(2,2-Ethylenedioxypropylidene)-1-phenylpiperidin-2-one (27): Colorless prisms from iso- $Pr_2O-CH_2Cl_2$, mp 118—119 °C. IR (KBr): 1655 (C=O) cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.57 (3H, s, CH₃), 2.02 (2H, m, C₅-H), 2.92 (2H, m, C₄-H), 3.72 (2H, m, C₆-H), 3.90 and 3.98 (each 2H, m, OCH₂CH₂O), 6.90 (1H, t, J=1 Hz, C=CH), 7.28 (3H, m, aromatic protons), 7.40 (2H, m, aromatic protons). EIMS m/z: 273 (M⁺). Anal. Calcd for C₁₆H₁₉NO₃: C, 70.31; H, 7.01; N, 5.12. Found: C, 70.33; H, 7.01; N, 5.16.

(E)-3-(2-Oxopropylidene)-1-phenylpiperidin-2-one (28): Pale yellow prisms from CH₂Cl₂-ether, mp 96—98 °C. IR (KBr): 1675 (C=O), 1635 (NC=O) cm⁻¹. ¹H-NMR (CDCl₃) δ: 2.04 (2H, m, C₅-H), 2.34 (3H, s, CH₃), 3.17 (2H, dt, J_d =2 Hz, J_t =6 Hz, C₄-H), 3.78 (2H, t, J=6 Hz, C₆-H), 7.30 (3H, m, aromatic protons), 7.39 (1H, d, J=3 Hz, C=CH), 7.43 (2H, m, aromatic protons). EIMS m/z: 229 (M⁺). *Anal.* Calcd for C₁₄H₁₅NO₂: C, 73.34; H, 6.59; N, 6.11. Found: C, 72.91; H, 6.67; N, 5.82.

5,6-Dihydro-2-oxo-3-(2-oxopropyl)-1-phenyl-3-piperine (**29**): Colorless prisms from CH₂Cl₂-ether, mp 87—88 °C. IR (KBr): 1685 (C=O), 1655 (NC=O) cm⁻¹. ¹H-NMR (CDCl₃) δ : 2.10 (2H, m, C₅-H), 2.36 (3H, s, CH₃), 2.72 (2H, dt, J_d =2 Hz, J_t =6 Hz, C₄-H), 3.76 (2H, t, J=6 Hz, C₆-H), 6.15 (1H, t, J=2 Hz, C=CH), 7.25—7.35 (3H, m, aromatic protons), 7.40 (2H, m, aromatic protons). CIMS m/z: 230 (M+H). *Anal.* Calcd for C₁₄H₁₅NO₂: C, 73.34; H, 6.59; N, 6.11. Found: C, 72.99; H, 6.58; N, 5.84.

(E)-4-Hydroxy-2-(1-phenylaminoethyl)pent-2-enoic Acid (30) A solution of KOH (1.1 g, 16 mm) in water (9 ml) was added to a solution of 11 (0.5 g, 2.3 mm) in dioxane (1 ml) at room temperature. After being stirred for 16 h, the reaction mixture was neutralized by addition of 1 n HCl and extracted twice with EtOAc. The extract was washed with brine, dried (MgSO₄), and evaporated in vacuo, and the residue was purified by silica gel column chromatography (CH₂Cl₂: MeOH: AcOH = 200: 20: 1) to give

30 (470 mg, yield: 87%) as a viscous oil. IR (film): 3600 (COOH), 1700 (C=O), 1600 (C=C) cm⁻¹. 1 H-NMR (CDCl₃) δ : 1.26 (3H, d, J=6.5 Hz, CH₃CHOH), 1.45 (3H, d, J=7 Hz, CH₃CHNHPh), 4.75 (1H, q, J=7 Hz, CHOH), 6.04 (3H, br s, COOH, OH, NH), 6.55—6.75 (3H, m, aromatic protons), 6.80 (1H, d, J=8.5 Hz, HC=C), 7.05—7.2 (2H, m, aromatic protons). CIMS m/z: 236 (M+H).

2,6-Dimethyl-1-phenylpyrrole-3-carboxylic Acid (32) Method 1: Attemburrow's MnO₂ (1.74 g, 20 mm) was added to a stirred solution of 30 (220 mg, 1 mm) in ether (60 ml). The reaction mixture was stirred for 20 min at room temperature, filtered through Celite, decolorized with charcoal, and evaporated *in vacuo* to give 32 (100 mg, yield: 46%), which was recrystallized from iso-Pr₂O to give colorless prisms, mp 207—208 °C. IR (KBr): 3000 (OH), 1640 (C=O) cm⁻¹. ¹H-NMR (DMSO- d_6) δ : 1.90 and 2.20 (each 3H, s, CH₃ × 2), 3.26 (1H, br s, COOH), 6.02 (1H, s, C₄-H), 7.0—7.65 (5H, m, aromatic protons). EIMS m/z: 215 (M⁺). *Anal.* Calcd for C₁₃H₁₃NO₂: C, 72.54; H, 6.09; N, 6.50. Found: C, 72.47; H, 6.08; N, 6.39.

Method 2: A stirred solution of **38** (0.9 g, 4 mm) in MeOH (10 ml) was treated with 1 N NaOH (1 ml). The mixture was stirred for 3 h at room temperature, then 1 N HCl (3 ml) was added, and the reaction mixture was extracted twice with EtOAc. The extract was washed with brine, dried (MgSO₄), and evaporated *in vacuo* to give **32** (0.5 g, yield: 60%).

Methyl 2,6-Dimethyl-1-phenylpyrrole-3-carboxylate (34) A mixture of 33 (1.72 g, 10 mm), aniline (0.93 g, 10 mm) and pyridinium p-toluene-sulfonate (125 mg, 0.5 mm) in benzene (50 ml) was refluxed for 1 h with removal of the resulting water under azeotropic conditions. The reaction mixture was evaporated in vacuo and the residue was purified by silica gel column chromatography (CH₂Cl₂) to give 34 (0.96 g, yield: 42%), which was recrystallized from hexane to give colorless needles, mp 51–54 °C. IR (KBr): 1710 (C=O). ¹H-NMR (CDCl₃) δ : 1.97 (3H, s, C₅-H), 2.27 (3H, s, C₂-H), 3.73 (3H, s, CO₂CH₃), 6.23 (1H, s, C₄-H), 7.0—7.5 (5H, m, aromatic protons). EIMS m/z: 229 (M⁺). Anal. Calcd for C₁₄H₁₅NO₂: C, 73.34; H, 6.59; N, 6.11. Found: C, 73.26; H,6.61; N, 6.09.

(E)-N-Phenyl 4-Oxopent-2-enamide (36) DCC (1.5 g, 8.4 mm) in CH₂Cl₂ (50 ml) was added to a mixture of (E)-4-oxo-2-pentenoic acid (35) (1.14 g, 8.4 mm) and aniline (0.93 g, 10 mm) in CH₂Cl₂ (50 ml) at 0 °C. The mixture was stirred for 1 h at the same temperature, and the resulting precipitate was filtered off. The filtrate was washed with 1 n HCl, aqueous NaHCO₃ and brine successively, and evaporated in vacuo. The residue was purified by silica gel column chromatography (CH₂Cl₂: ether = 10: 1) to give 36 (120 mg, 6.6%), which was recrystallized from ether to give a yellow amorphous solid, mp 138—140 °C. IR (KBr): 3300 (NH), 1655 (C=O), 1620 (NC=O) cm⁻¹. ¹H-NMR (CDCl₃) δ : 2.38 (3H, s, CH₃), 6.97 (1H, d, J=15 Hz, C₃-H), 7.18 (1H, m, aromatic proton), 7.17 (2H, m, aromatic protons), 7.22 (1H, d, J=15 Hz, C₂-H), 7.64 (2H, m, aromatic

protons), 8.20 (1H, br s, NH). EIMS m/z: 319 (M⁺). Anal. Calcd for $C_{11}H_{11}NO_2$: C, 69.83; H, 5.86; N, 7.40. Found: C, 69.91; H, 5.88; N, 7.52.

Preparation of PRP Blood was taken from the carotid artery of a male New Zealand White rabbit into a plastic syringe containing 10% volume of 3.2% sodium citrate dihydrate solution under ether anesthesia. The citrated blood was centrifuged at $150\,g$ for $15\,\text{min}$ at room temperature to obtain PRP. The sediment was further centrifuged at $1500\,g$ for $10\,\text{min}$ to obtain platelet-poor plasma (PPP). The platelet count was adjusted to approximately $5\times10^5-6\times10^5/\mu\text{l}$ by adding PPP.

Platelet Aggregation Test in Vitro Platelet aggregation was measured by a Aggrecoder PA-3210 (Kyoto Daiichi Kagaku) at 37 °C under stirring at 1000 rpm. The agents used were ADP (Shigma Chemical Co., final concentration, 5 μ M), and collagen (Kyoto Daiichi Kagaku Co., Ltd., final concentration, 5 μ g/ml). The compound to be tested was dissolved in DMSO, diluted with saline, and added in a volume of 25 μ l to 275 μ l of PRP 3 min before addition of the aggregating agents. Platelet aggregation was measured for 5 min and the IC₅₀ value was calculated from the maximum decrease in absorbancy of PRP in comparison with that of vehicle-treated PRP.

References and Notes

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- S. Kano, S. Shibuya and T. Ebata, Heterocycles, 15, 1011 (1981); S. Kano, T. Ebata and S. Shibuya, J. Chem. Soc., Perkin Trans. 1, 1980, 2105; S. Kano, T. Ebata, Y. Yuasa and S. Shibata, Heterocycles, 14, 589 (1980).
- G. D. Qureshi, C. A. Fletcher, S. C. Choi, and R. J. Duma, Clin. Res., 31, 849A (1983); K. H. Jakobs, G. Schultz, B. Gauglar, and T. Pfeuffer, Eur. J. Biochem., 134, 351 (1983).
- T. Kano, S. Shibuya and T. Ebata, J. Chem. Soc., Perkin Trans. 1, 1982, 257.
- K. Tanaka, N. Yamagishi, R. Tanikaga and A. Kaji, *Bull. Chem. Soc. Jpn.*, 52, 3619 (1979).
- F. A. Bouffard and B. G. Christensen, J. Org. Chem., 46, 2208 (1981);
 R. S. Glass, D. R. Deardorff and K. Hengar, Tetrahedron Lett., 21, 2467 (1980).
- A. Holland, J. M. Inglis and R. Slack, Brit. Patent 951115 (1964)
 [Chem. Abstr., 60, 15874h (1964)].
- J. Attemburrow, A. F. B. Cameron, J. H. Chapman, R. M. Evans, B. A. Hems, A. B. A. Jansen and T. Walker, J. Chem. Soc., 1952, 1094.
- 9) C. D. Hurd and K. Wilkinson, J. Am. Chem. Soc., 70, 739 (1948).
- 10) R. Scheffold and P. Dubs, Helv. Chim. Acta, 50, 798 (1967).
- G. V. R. Born, *Nature* (London), **194**, 927 (1962); G. V. R. Born and M. Cross, *J. Physiol.* (London), **168**, 178 (1963).