Purines. XL.¹⁾ Preparation of 9-(ω-Carboxyalkyl)-3-methyladenines

Tozo Fujii,* Tohru Saito, and Yukinari Kumazawa

Faculty of Pharmaceutical Sciences, Kanazawa University, Takara-machi, Kanazawa 920, Japan. Received November 17, 1989

With a view to supplying haptens to be connected to carrier proteins for raising antibodies to 3-methyl-2'-deoxyadenosine (1) and/or 3-methyladenine (3), (3-methyl-9-adenyl)acetic acid hydrochloride (15a) and 4-(3-methyl-9-adenyl)butyric acid hydrochloride (15b) have been prepared from 1-alkoxy-9-(\omega-carboxyalkyl)adenine salts (7a and 7b) through the intermediates 11a,b, 12a,b, 13a,b, and 14a,b.

Keywords 3,9-disubstituted adenine; 9-substituted adenine 1-oxide; *N*-oxide *O*-methylation; 9-substituted 1-alkoxyadenine; adenine ring fission; formamido group *N*-methylation; alkaline hydrolysis; hydrogenolytic dealkoxylation; amidine formamido cyclization

Recent advances in the characterization of deoxyribonucleic acid (DNA) components structurally modified by chemical carcinogens are remarkable.2) In particular, the modifications produced in DNA by the alkylating Nnitroso carcinogens have mostly been identified.³⁾ 3-Methyl-2'-deoxyadenosine (1)4) is believed to occur as an unstable part structure in methylated DNA molecules.5) Although loss of 3-methyladenine (3) from methylated DNA in vivo could be explained in terms of chemical depurinylation alone, active enzymatic excision has also been suggested. 5b) This suggestion led to the isolations of 3methyladenine-DNA glycosylase in partially purified form from both bacterial and mammalian sources. 5d,e,k,6) The detection of the presence of 1 or 3 could therefore be used to monitor a previous exposure of cells, tissues, and individuals to methylating carcinogens, and it could be quantitatively done by producing antibodies to 1 and/or 3

and setting up radioimmunoassays for them. In the preparation of the antigens, application of the most widely used sugar-cleavage oxidation method³⁾ to 3-methyladenosine $(2)^{4b,7)}$ would be inadequate because of the extraordinary instability of this nucleoside. The title compounds, 9- $(\omega$ -carboxyalkyl)-3-methyladenines (4), or 9- $(\omega$ -aminoalkyl)-3-methyladenines (5) would then provide a practical alternative source of haptens to be connected to carrier proteins³⁾ for raising antibodies to 1 and/or 3. In the present study, we synthesized (3-methyl-9-adenyl)acetic acid hydrochloride (15a) and 4-(3-methyl-9-adenyl)butyric acid hydrochloride (15b), both belonging to type 4, in response to a request⁸⁾ for such potential haptens.

The synthesis of the target compounds 15a and 15b was so designed that it becomes a 9-(ω -carboxyalkyl) version of our previous general synthesis⁹⁾ of 3,9-dialkyladenine salts, as shown in Chart 1. Treatment of 1-ethoxyadenine (6^{10}) with ethyl bromoacetate in AcNMe₂ at 30 °C for 24h produced 1-ethoxy-9-[(ethoxycarbonyl)methyl]adenine hydrobromide, which was isolated in the form of the perchlorate 7a in 56% overall yield (from 6). This result was in general agreement with those of our previous alkylations of 1-alkoxyadenines at the 9-position. ¹⁰⁾ The perchlorate 7a was converted into the free base by the use of Amberlite IRA-402 (HCO₃⁻), and the base was treated with H₂O at 40 °C for 7 h to furnish the formamidoimidazole 11a in 35% yield. Such a ready ring-opening under mild condi-

© 1990 Pharmaceutical Society of Japan

Chart 1

tions had already been observed by us for 1-alkoxy-9alkyladenines.¹¹⁾ On methylation with MeI and anhydrous K₂CO₃ in HCONMe₂ at room temperature for 2h, 11a afforded the N-methylformamido derivative 12a in 84% yield. Hydrolysis of 12a was effected in boiling 1 N aqueous NaOH for 15 min, and the product was isolated in 72% yield in the form of the dihydrochloride 13a. Hydrogenolysis of 13a using hydrogen and Raney Ni catalyst in H₂O at 1 atm and room temperature for 3h gave crude 14a, which was then cyclized with boiling formic acid for 5 h, affording the target compound 15a in 42% overall yield (from 13a). A similar cyclization of 1-methyl-5-(methylamino)-1 H-imidazole-4-carboxamidine hydrochloride (type 14a: Me in place of CH₂CO₂H) to give 3,9-dimethyladenine hydrochloride had previously been achieved by the use of diethoxymethyl acetate in HCONMe₂.9b)

For the synthesis of the other target compound (15b), adenine (10) was first treated with ethyl 4-bromobutyrate in AcNMe₂ in the presence of anhydrous K₂CO₃ and 18crown-6 at 30 °C for 24 h, giving ethyl 4-(9-adenyl)butyrate (9) in 76% yield. 12) Oxidation of 9 with m-chloroperbenzoic acid in EtOH at room temperature furnished the 1-oxide 8 in 81% yield. Methylation of 8 with MeI was effected in AcNMe₂ at room temperature for 20 h, and the Omethylated product was isolated as the perchlorate 7b in 82% yield (from 8). The synthesis of 7b from 10 through 9 and 8 was 9-[ω-(ethoxycarbonyl)alkyl] version of our wellestablished general synthesis of 1-alkoxy-9-alkyladenine salts from 10.13,14) The succeeding steps beyond 7b were parallel to those described above for the a-series, producing 11b (63% yield), 12b (86%), 13b (crude), 14b (crude), and 15b [20% (from 12b)].

The correctness of the structures of 15a and 15b was supported by the way in which they had been generated, microanalytical data, and comparison of their ultraviolet (UV) and nuclear magnetic resonance (NMR) spectra with those of known 3,9-dialkyladenine salts. $^{9b-d}$ According to Professor A. E. Pegg (The Pennsylvania State University, U.S.A.), 15 his research group has not been able to obtain antibodies to 3-methyladenine (3) by using conjugates synthesized with the above precursors, 15a and 15b. Their lack of success in the antibody production may be attributable to the instability of the hapten moiety in the conjugates; facile ring-opening would have occurred under mild alkaline conditions because of the 3,9-disubstituted adenine structure. $^{9b-d}$

Experimental

General Notes All melting points were taken on a Yamato MP-1 capillary melting point apparatus and are corrected. See ref. 9b for details of instrumentation and measurements. Elemental analyses were performed by Mr. Y. Itatani and his associates at Kanazawa University. The following abbreviations are used: br=broad, d=doublet, m=multiplet, q=quartet, s=singlet, s=shoulder, t=triplet.

6-Amino-9H-purine-9-butanoic Acid Ethyl Ester (9) A mixture of adenine (10) (405 mg, 3 mmol), anhydrous K_2CO_3 (415 mg, 3 mmol), 18-crown-6 (79 mg, 0.3 mmol), and ethyl 4-bromobutyrate (1.17 g, 6 mmol) in AcNMe₂ (12 ml) was stirred at 30 °C for 24 h. The reaction mixture was concentrated to dryness in vacuo. The residue was washed with two 5-ml portions of hexane and the washings were removed by decantation. The insoluble solid was then extracted with hot benzene (2 × 10 ml, 4 × 5 ml). The benzene extracts were combined and concentrated to dryness in vacuo. The residue was chromatographed on a column packed with silica gel (35 g), and fractions eluted with CH_2Cl_2 -EtOH (10:1, v/v) gave 9 (565 mg, 76%)¹²⁾ as a colorless solid, mp 104-107 °C. Recrystallization from

benzene-hexane (2:1, v/v) yielded colorless needles, mp 110—111 °C (lit. mp 110—111 °C; ^{13b)} mp 108—109 °C¹⁶⁾). This sample was identical (by comparison of the IR spectrum and thin-layer chromatographic mobility) with authentic $9.^{13b)}$

6-Amino-9H-purine-9-butanoic Acid Ethyl Ester 1-Oxide (8) m-Chloroperbenzoic acid (80% purity) (32.36 g, 150 mmol) was added portionwise to a stirred solution of 9 (18.70 g, 75 mmol) in EtOH (600 ml), and the mixture was stirred at room temperature for 3 h. The precipitate that resulted was filtered off and then dissolved in 50% aqueous EtOH (1200 ml). The resulting solution was passed through a column of Amberlite IRA-402 (HCO₃⁻) (360 ml), and the column was eluted with H₂O (1000 ml). The eluate was concentrated to dryness in vacuo to leave 8 (16.20 g, 81%) as a colorless solid, mp 208—209 °C. Recrystallization from AcOEt furnished an analytical sample as colorless needles, mp 211—212 °C; MS m/z: 265 (M⁺); UV $\lambda_{max}^{95\%}$ aq. EIOH 235 nm (ϵ 41400), 263 (7200), 300 (1800); $\lambda_{\text{max}}^{\text{H}_{2}\text{O}}$ (pH 1) 214 (27800), 259 (11600); $\lambda_{\text{max}}^{\text{H}_{2}\text{O}}$ (pH 7) 232 (40000), 262 (7100), 292 (1600); $\lambda_{\text{max}}^{\text{H}_{2}\text{O}}$ (pH 13) 232 (29800), 267 (7900), 302 (3200); IR $v_{\text{max}}^{\text{Nujol}}$ 1730 cm⁻¹ (CO₂Et); NMR (Me₂SO- d_6) δ : 1.14 (3H, t, J=7 Hz, CO₂CH₂Me), 2.12 [2H, m, N(9)-CH₂CH₂], 2.30 (2H, m, CH₂CO₂Et), 4.00 $(2H, q, J=7 Hz, CO_2CH_2Me), 4.21 [2H, t, J=7 Hz, N(9)-CH_2], 8.24 (2H, t)$ br, NH₂), 8.29 [1H, s, C(8)-H], 8.61 [1H, s, C(2)-H]. Anal. Calcd for $C_{11}H_{15}N_5O_3$: C, 49.81; H, 5.70; N, 26.40. Found: C, 49.86; H, 5.49; N, 26.23

1-Ethoxy-1,6-dihydro-6-imino-9H-purine-9-acetic Acid Ethyl Ester Perchlorate (7a) A mixture of 1-ethoxyadenine (6)¹⁰ (21.50 g, 0.12 mol) and ethyl bromoacetate (100.2 g, 0.6 mol) in AcNMe₂ (370 ml) was stirred at 30 °C for 24 h. The reaction mixture was concentrated in vacuo, and the residual oil was triturated with ether (2 × 200 ml). The insoluble solid that resulted was filtered off, washed with ether, and dissolved in warm H₂O (80 ml), and then a solution of NaClO₄ (22.04 g, 0.18 mol) in H₂O (40 ml) was added. The precipitate that deposited was filtered off, washed with H₂O, and recrystallized from 80% aqueous EtOH to yield a first crop $(20.89 \,\mathrm{g}, \,48\%)$ of 7a, mp $215-217\,^{\circ}\mathrm{C}$ (dec.). A further crop from the mother liquor of the recrystallization raised the yield of 7a to 24.78 g (56%). Recrystallization of the first crop of crystals from H₂O gave an analytical sample as colorless prisms, mp 212—213 °C (dec.); UV $\lambda_{\max}^{95\% aq. EtOH}$ 258 nm (ϵ 12300); $\lambda_{\max}^{H_{2}O}$ (pH 1) 258 (12400); $\lambda_{\max}^{H_{2}O}$ (pH 7) 258 (12300); $\lambda_{\text{max}}^{\text{H}_{2}\text{O}}$ (pH 13) (unstable) 258 (12700), 265 (sh) (11400); IR $\nu_{\text{max}}^{\text{Nujol}}$ 1756 cm⁻¹ (CO₂Et); NMR (Me₂SO- d_6) δ : 1.24 (3H, t, J=7 Hz, CO_2CH_2Me , 1.43 [3H, t, J=7 Hz, N(1)-OCH₂Me], 4.21 (2H, q, J=7 Hz, CO_2CH_2Me), 4.44 [2H, q, J=7 Hz, N(1)-OC H_2Me], 5.25 [2H, s, N(9)-CH₂], 8.53 and 9.13 (1H each, s, purine protons), 9.65 and 10.36 (1H each, br, $= NH_2^+$ or $2 \times NH$). Anal. Calcd for $C_{11}H_{15}N_5O_3 \cdot HClO_4$: C, 36.13; H, 4.41; N, 19.15. Found: C, 36.15; H, 4.46; N, 19.05.

1,6-Dihydro-6-imino-1-methoxy-9H-purine-9-butanoic Acid Ethyl Ester Perchlorate (7b) A mixture of 8 (3.18g, 12 mmol) and MeI (4.26g, 30 mmol) in AcNMe₂ (24 ml) was stirred at room temperature for 20 h. The reaction mixture was concentrated in vacuo, and the residue was triturated with ether $(2 \times 20 \text{ ml})$. The insoluble solid that resulted was separated from the ethereal layer by decantation and dissolved in warm H₂O (5 ml), and then a solution of NaClO₄ (2.20 g, 18 mmol) in H₂O (3 ml) was added. The crystals that deposited were filtered off, washed with H_2O , and dried to give crude 7b (3.71 g, 82%), mp 138—141 °C. Recrystallization from H₂O afforded an analytical sample as colorless prisms, mp 153.5—154.5 °C; UV $\lambda_{\text{max}}^{95\% \text{ aq. EtOH}}$ 259 nm (ϵ 12500); $\lambda_{\text{max}}^{\text{H}_{2}\text{O}}$ (pH 1) 260 (12500); $\lambda_{\text{max}}^{\text{H}_{2}\text{O}}$ (pH 7) 260 (12500); $\lambda_{\text{max}}^{\text{H}_{2}\text{O}}$ (pH 13) (unstable) 258 (12900), 265 (sh) (11500); IR $v_{\text{max}}^{\text{Nujol}}$ 1735 cm⁻¹ (CO₂Et); NMR (Me₂SO-d₆) δ : 1.16 $(3H, t, J=7 Hz, CO_2CH_2Me)$, 2.11 [2H, m, N(9)-CH₂CH₂], 2.31 (2H, m, CH_2CO_2Et), 4.03 (2H, q, J=7 Hz, CO_2CH_2Me), 4.17 [3H, s, N(1)-OMe], 4.30 [2H, t, J=6.5 Hz, N(9)-CH₂], 8.56 and 9.15 (1H each, s, purine protons), 9.69 and 10.30 (1H each, br, $= NH_2^+$ or $2 \times NH$). Anal. Calcd for C₁₂H₁₇N₅O₃·HClO₄: C, 37.95; H, 4.78; N, 18.44. Found: C, 37.90; H, 4.93: N. 18.47.

4-(N'-Ethoxyamidino)-5-formamido-1H-imidazole-1-acetic Acid Ethyl Ester (11a) A solution of 7a (20.12 g, 55 mmol) in H₂O (1200 ml) was passed through a column of Amberlite IRA-402 (HCO₃⁻) (132 ml), and the column was eluted with H₂O. The eluate (2200 ml) was concentrated in vacuo to a volume of ca. 400 ml and kept at 40 °C for 7 h. The reaction mixture was concentrated to dryness in vacuo, and the residue was purified by column chromatography [silica gel (540 g), CH₂Cl₂-EtOH (10:1, v/v)] followed by two recrystallizations from AcOEt, furnishing 11a (5.43 g, 35%) as colorless minute prisms, mp 111—112 °C. Further recrystallization in the same manner yielded an analytical sample of 11a, mp 113—114 °C; MS m/z: 283 (M⁺); UV $\lambda_{\text{max}}^{\text{1850}}$ aq. EtOH 250 nm (sh) (ε 6700); $\lambda_{\text{max}}^{\text{H2O}}$ (pH 1) 253 (8100); $\lambda_{\text{max}}^{\text{H2O}}$ (pH 7) 219 (11900), 250 (sh) (6500); $\lambda_{\text{max}}^{\text{H2O}}$ (pH 13) 249

(11100); IR $v_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 1738 (CO₂Et), 1704 (CONHAr); NMR (Me₂SO- d_6) δ : 1.0—1.35 (6H, m, CO₂CH₂Me and NOCH₂Me), 3.86 (2H, q, J=6.8 Hz, NOCH₂Me), 4.14 (2H, q, J=7.1 Hz, CO₂CH₂Me), 4.74 and 4.83 [2H, s each, N(1)-CH₂], 5.5—5.7 (2H, br, NH₂), 7.65 (sh) and 7.66 [1H, C(2)-H], 8.01 (0.6H, d, J=10.5 Hz, trans-HCONH), 8.17 (0.4H, s, cis-HCONH), 9.34 (0.6H, d, J=10.5 Hz, trans-HCONH), 9.65 (0.4H, dull s, cis-HCONH). 17) Anal. Calcd for C₁₁H₁₇N₅O₄: C, 46.64; H, 6.05; N, 24.72. Found: C, 46.80; H, 6.04; N, 24.78.

5-Formamido-4-(N'-methoxyamidino)-1H-imidazole-1-butanoic Acid Ethyl Ester (11b) A solution of 7b (6.84 g, 18 mmol) in H_2O (300 ml) was passed through a column of Amberlite IRA-402 (HCO₃⁻) (45 ml), and the column was eluted with H₂O. The eluate (700 ml) was concentrated in vacuo to a volume of ca. 200 ml and kept first in a refrigerator for 3d and then at 30 °C for 48 h. The reaction mixture was worked up in a manner similar to that described above for 11a except that the eluent for silica gel (55 g) column chromatography was AcOEt, giving 11b (3.37 g, 63%), mp 125.5-126.5 °C. Recrystallization from AcOEt afforded an analytical sample as colorless prisms, mp 126—127 °C; UV $\lambda_{max}^{95\% \text{ aq. EiOH}}$ 250 nm (sh) (ϵ 6750); $\lambda_{\max}^{H_{2O}}$ (pH 1) 254 (8100); $\lambda_{\max}^{H_{2O}}$ (pH 7) 218.5 (11800), 250 (sh) (6700); $\lambda_{\max}^{H_{2O}}$ (pH 13) 254 (10900); IR ν_{\max}^{Nujol} cm⁻¹: 1732 (CO₂Et), 1696 (CONHAr); NMR (Me₂SO- d_6) δ : 1.17 (3H, t, J=7 Hz, CO₂CH₂Me), 1.90 [2H, m, $N(1)-CH_2CH_2$, 2.27 (2H, m, CH_2CO_2Et), 3.63 and 3.67 (3H, s each, NOMe), 3.84 [2H, m, N(1)-CH₂], 4.03 and 4.04 (2H, q each, J=7 Hz, CO₂CH₂Me), 5.59 and 5.64 (2H, dull s each, NH₂), 7.67 [1H, s, C(2)-H], 8.05 (0.5H, d, J = 10 Hz, trans-HCONH), 8.22 (0.5H, s, cis-HCONH), 9.39 (0.5H, d, J = 10 Hz, trans-HCONH), 9.63 (0.5H, s, cis-HCONH). Anal. Calcd for C₁₂H₁₉N₅O₄: C, 48.48; H, 6.44; N, 23.56. Found: C, 48.19; H, 6.52; N, 23.26.

4-(N'-Ethoxyamidino)-5-(N-methylformamido)-1H-imidazole-1-acetic Acid Ethyl Ester (12a) A mixture of 11a (5.13 g, 18.1 mmol) and anhydrous K₂CO₃ (3.76 g, 27.2 mmol) in HCONMe₂ (180 ml) was stirred at room temperature for 1 h. A solution of MeI (3.08 g, 21.7 mmol) in HCONMe₂ (20 ml) was then added, and the resulting mixture was stirred at room temperature for 2h. The reaction mixture was concentrated in vacuo to leave a brown jelly, which was extracted with boiling benzene (3 × 180 ml). The benzene extracts were combined and concentrated in vacuo, and the residue was recrystallized from hexane-AcOEt (2:1, v/v) to afford a first crop (4.07 g, 76%) of 12a, mp 101-102 °C. Concentration of the mother liquor from the recrystallization and purification of the residue by column chromatography [silica gel, benzene-EtOH (15:1, v/v)] gave a second crop (0.45 g) of 12a, mp 99-101 °C. The total yield of 12a was 4.52 g (84%). Further recrystallizations of crude 12a from hexane-AcOEt (2:1, v/v) yielded an analytical sample as colorless minute prisms, mp 103—104 °C; MS m/z: 297 (M⁺); UV $\lambda_{\text{max}}^{95\% \text{ aq. EtOH}}$ 250 nm (sh) (ε 5600); $\lambda_{\text{max}}^{\text{H}_2\text{C}}$ (pH 1) 249 (8490); $\lambda_{\max}^{H_2O}$ (pH7) 250 (sh) (5470); $\lambda_{\max}^{H_2O}$ (pH 13) 250 (sh) (5670); IR $\nu_{\max}^{N_{\text{ujol}}}$ cm⁻¹: 3520, 3410 (NH₂), 1749 (CO₂Et), 1678 (HCON); NMR (Me_2SO-d_6) δ : 1.15 (3H, t, J=7 Hz, $NOCH_2Me$ or CO_2CH_2Me), 1.20 (3H, t, J = 7 Hz, CO_2CH_2Me or $NOCH_2Me$), 2.93 (0.86 × 3H) and 3.19 $(0.14 \times 3H)$ (s each, NMe), 3.83 (2H, q, J = 7 Hz, NOC \underline{H}_2 Me), 4.16 (2H, q, J=7 Hz, CO_2CH_2Me), 4.7 (br) and 4.89 (dull s) [2H, N(1)-CH₂], 5.64 (2H, dull s, NH₂), 7.72 (0.14H) and 7.76 (0.86H) [s each, C(2)-H], 7.96 (0.86H) and 8.20 (0.14H) (s each, CHO). 17) Anal. Calcd for C₁₂H₁₉N₅O₄: C, 48.48; H, 6.44; N, 23.56. Found: C, 48.46; H, 6.48; N, 23.66.

4-(N'-Methoxyamidino)-5-(N-methylformamido)-1H-imidazole-1butanoic Acid Ethyl Ester (12b) A mixture of 11b (2.97 g, 10 mmol) and anhydrous K₂CO₃ (2.07 g, 15 mmol) in HCONMe₂ (50 ml) was stirred at room temperature for 1 h. A solution of MeI (1.70 g, 12 mmol) in HCONMe₂ (5 ml) was then added, and the resulting mixture was stirred at room temperature for 2 h. The reaction mixture was concentrated in vacuo, and the residue was triturated with H₂O (5 ml). The insoluble solid that resulted was filtered off, washed with H₂O, and dried to give 12b (2.68 g, 86%), mp 90.5-91.5 °C. Recrystallization from H₂O provided an analytical sample as colorless prisms, mp 90.5—91.5 °C; UV λ_{max} 250 nm (sh) (ϵ 5560); $\lambda_{\text{max}}^{\text{H}_{2}\text{O}}$ (pH 1) 251.5 (8120); $\lambda_{\text{max}}^{\text{H}_{2}\text{O}}$ (pH 7) 250 (sh) (5720); $\lambda_{\text{max}}^{\text{H}_{2}\text{O}}$ (pH 13) 250 (sh) (5760); IR $\nu_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 3435, 3315 (NH₂), 1730 (CO₂Et), 1692 (HCON); NMR (Me₂SO- d_6) δ : 1.17 (3H, t, J=7 Hz, CO₂CH₂Me), 1.92 [2H, m, N(1)-CH₂CH₂], 2.33 (2H, m, CH₂CO₂Et), 3.01 (0.83 \times 3H) and 3.26 (0.17 \times 3H) (s each, NMe), 3.60 (0.83 \times 3H) and 3.66 (0.17 \times 3H) (s each, NOMe), 3.86 [2H, m, N(1)-CH₂], 4.03 and 4.05 (2H, q each, J=7 Hz, CO₂CH₂Me), 5.16 (sh) and 5.67 (2H, br, NH₂), 7.74 (0.17H) and 7.78 (0.83H) [s each, C(2)-H], 8.01 (0.83H) and 8.25 (0.17H) (s each, CHO).¹⁷⁾ Anal. Calcd for C₁₃H₂₁N₅O₄: C, 50.15; H, 6.80; N, 22.49. Found: C, 50.03; H. 7.01: N. 22.56

4-(N'-Ethoxyamidino)-5-(methylamino)-1H-imidazole-1-acetic Acid Dihydrochloride (13a) A solution of 12a (892 mg, 3 mmol) in 1 N aqueous

NaOH (15 ml) was heated under reflux for 15 min. After cooling, the reaction mixture was passed through a column of Dowex 50W-X8 (H+) (40 ml), and the column was washed with H₂O until the eluate became neutral. The column was then eluted with 5% aqueous NH₃ (400 ml), and the ammoniacal eluate was applied to a column of Amberlite IRA-402 (OH⁻) (20 ml), which was then eluted successively with H₂O (300 ml) and 2% aqueous HCl (300 ml). The acid eluate was concentrated in vacuo, and the residual solid was washed with acetone (10 ml) to give 13a (680 mg, 72%), mp 172-173 °C (dec.). Recrystallization of crude 13a by dissolving it in H₂O and adding acetone to the resulting aqueous solution produced an analytical sample as colorless needles, mp 178—180 °C (dec.); IR $v_{\text{max}}^{\text{Nujol}}$ 1772 cm⁻¹ (CO₂H); NMR (Me₂SO-d₆) δ : 1.26 (3H, t, J= 7 Hz, $NOCH_2Me$), 2.70 (3H, s, NMe), 4.02 (2H, q, J = 7 Hz, $NOCH_2Me$), 4.91 (2H, s, CH₂CO₂H), 8.17 [1H, s, C(2)-H]. Anal. Calcd for C₉H₁₅N₅O₃·2HCl: C, 34.41; H, 5.45; N, 22.29. Found: C, 34.07; H, 5.50; N. 22.06.

3,6-Dihydro-6-imino-3-methyl-9H-purine-9-acetic Acid Hydrochloride (15a) A solution of 13a (220 mg, 0.7 mmol) in H_2O (15 ml) was hydrogenated over Raney Ni W-2 catalyst¹⁸⁾ (240 mg) at atmospheric pressure and room temperature for 3 h. The catalyst was removed by filtration and washed with H₂O (10 ml). The filtrate and washings were combined and concentrated to dryness in vacuo to leave a pale greenish glass (203 mg), presumed to be 4-amidino-5-(methylamino)-1H-imidazole-1acetic acid dihydrochloride (14a). The glass was suspended in formic acid (of over 98% purity) (7 ml), and the suspension was heated under reflux for 5 h. The reaction mixture was concentrated in vacuo, and the residue was triturated with ether $(2 \times 10 \text{ ml})$. The insoluble solid that resulted was separated from the ethereal layer by decantation and recrystallized by dissolving it in warm 5% aqueous HCl (5 ml) and adding acetone (70 ml) to the resulting acid solution, affording 15a [72 mg, 42% (from 13a)]. Further recrystallization in a similar manner gave an analytical sample as colorless prisms, mp 264—265 °C (dec.); UV $\lambda_{\max}^{95\%}$ aq. Ei0H 271.5 nm (ϵ 15900); $\lambda_{\max}^{H_{2}O}$ (pH 1) 270 (16200); $\lambda_{\max}^{H_{2}O}$ (pH 7) 270 (16200); $\lambda_{\max}^{H_{2}O}$ (pH 13) unstable; IR $v_{\text{max}}^{\text{Nujol}}$ 1735 cm⁻¹ (CO₂H); NMR (Me₂SO-d₆) δ : 4.07 [3H, s, N(3)-Me], 5.51 (2H, s, CH₂CO₂H), 8.41 [1H, s, C(8)-H], 8.65 [1H, s, C(2)-H], 9.25 and 9.30 (1H each, br s, =N H_2^+). Anal. Calcd for $C_8H_9N_5O_2$ HCl: C, 39.44; H, 4.14; N, 28.74. Found: C, 39.25; H, 4.12; N, 28.74.

3,6-Dihydro-6-imino-3-methyl-9H-purine-9-butanoic Acid Hydrochloride (15b) A suspension of 12b (2.18 g, 7 mmol) in 1 N aqueous NaOH (35 ml) was heated under reflux for 20 min. After cooling, the reaction mixture was worked up in a manner similar to that described above for 13a, producing a yellow glass presumed to be 4-(N'-methoxyamidino)-5-(methylamino)-1 H-imidazole-1-butanoic acid dihydrochloride (13b). The glass was then hydrogenated in H₂O (70 ml) over Raney Ni W-2 catalyst¹⁸⁾ (1.2 g) for 6 h as described above for 15a, affording a greenish glass (866 mg), presumed to be 4-amidino-5-(methylamino)-1 H-imidazole-1-butanoic acid dihydrochloride (14b). Cyclization of the total amount of crude 14b with boiling formic acid (40 ml) for 4 h, work-up of the reaction mixture, and recrystallization of the product 15b H₂O [411 mg, 20% (from 12b)] also followed those described above for 15a. After drying over P_2O_5 at 3 mmHg and room temperature for 24 h, an analytical sample of 15b H₂O was obtained as colorless minute prisms, mp 245—246 °C (dec.); UV $\lambda_{\max}^{95\%,aq.EiOH}$ 271.5 nm (ϵ 15900); $\lambda_{\max}^{H_{2}O}$ (pH 1) 271 (16300); $\lambda_{\max}^{H_{3}O}$ (pH 7) 271 (16300); $\lambda_{\max}^{H_{3}O}$ (pH 13) unstable; IR ν_{\max}^{Nujol} 1736 cm⁻¹ (CO₂H); NMR (Me_2SO-d_6) δ : 2.06 [2H, m, N(9)-CH₂CH₂], 2.40 (2H, t, J=6Hz, CH_2CO_2H), 4.19 [3H, s, N(3)-Me], 4.50 [2H, t, J=7 Hz, N(9)-CH₂], 8.44 [1H, s, C(8)-H], 8.64 [1H, s, C(2)-H], 9.12 and 9.19 (1H each, s, $=NH_2^+$), 12.31 (1H, br, CO₂H). Anal. Calcd for C₁₀H₁₃N₅O₂·HCl·H₂O: C, 41.46; H, 5.57; N, 24.17. Found: C, 41.39; H, 5.61; N, 24.46.

References and Notes

- Paper XXXIX in this series, T. Fujii, T. Saito, T. Date, and Y. Nishibata, Chem. Pharm. Bull., 38, 912 (1990).
- a) N. C. Yang and C.-W. Chang, Proc. Natl. Acad. Sci. U.S.A., 82, 5250 (1985), and references cited therein; b) G. M. Blackburn and B. Kellard, Chem. Ind. (London), 1986, 607.
- For a review, see R. Müller and M. F. Rajewsky, J. Cancer Res. Clin. Oncol., 102, 99 (1981).
- a) T. Fujii, T. Saito, and T. Nakasaka, J. Chem. Soc., Chem. Commun., 1980, 758; b) Idem, Chem. Pharm. Bull., 37, 2601 (1989).
- a) P. D. Lawley and P. Brookes, Biochem. J., 89, 127 (1963); b) G. P. Margison and P. J. O'Connor, Biochim. Biophys. Acta, 331, 349 (1973); c) A. M. Maxam and W. Gilbert, Proc. Natl. Acad. Sci. U.S.A., 74, 560 (1977); d) S. Riazuddin and T. Lindahl, Biochemistry, 17, 2110 (1978); e) B. Singer and T. P. Brent, Proc. Natl. Acad. Sci.

- U.S.A., 78, 856 (1981); f) M. D. Mamet-Bratley and B. Karska-Wysocki, Biochim. Biophys. Acta, 698, 29 (1982); g) P. Karran, T. Hjelmgren, and T. Lindahl, Nature (London), 296, 770 (1982); h) P. E. Gallagher and T. P. Brent, Biochemistry, 21, 6404 (1982); i) M. Szyf, Y. Gruenbaum, S. Urieli-Shoval, and A. Razin, Nucleic Acids Res., 10, 7247 (1982); j) J. Hindley, "DNA Sequencing," Elsevier Biochemical Press, Amsterdam, 1983; k) P. E. Gallagher and T. P. Brent, Biochim. Biophys. Acta, 782, 394 (1984).
- a) J. Laval, Nature (London), 269, 829 (1977); b) P. E. Gallagher and T. P. Brent, Biochem. Biophys. Res. Commun., 101, 956 (1981); c) L. Thomas, C.-H. Yang, and D. A. Goldthwait, Biochemistry, 21, 1162 (1982)
- 7) T. Saito and T. Fujii, J. Chem. Soc., Chem. Commun., 1979, 135.
- A. E. Pegg, The Pennsylvania State University, personal communication, 1982.
- a) T. Fujii, T. Saito, and M. Kawanishi, Tetrahedron Lett., 1978, 5007;
 b) T. Fujii, T. Itaya, T. Saito, K. Mohri, M. Kawanishi, and T. Nakasaka, Chem. Pharm. Bull., 37, 1504 (1989);
 c) T. Fujii, T. Saito, and T. Nakasaka, ibid., 37, 3243 (1989);
 d) T. Fujii, T. Saito, T. Nakasaka, and K. Kizu, ibid., 38, 99 (1990).

- 10) T. Fujii and T. Itaya, Tetrahedron, 27, 351 (1971).
- a) T. Fujii, T. Itaya, C. C. Wu, and F. Tanaka, Tetrahedron, 27, 2415 (1971); b) T. Itaya, F. Tanaka, and T. Fujii, ibid., 28, 535 (1972); c) Earlier papers cited in ref. 9b.
- Our previous procedure, in which 18-crown-6 was not used, 136) gave
 in only 45% yield.
- a) T. Fujii, S. Sakurai, and T. Uematsu, Chem. Pharm. Bull., 20, 1334 (1972);
 b) T. Fujii and T. Saito, ibid., 33, 3635 (1985).
- a) T. Fujii, C. C. Wu, and T. Itaya, Chem. Pharm. Bull., 19, 1368 (1971);
 b) T. Fujii, T. Saito, K. Kizu, H. Hayashibara, Y. Kumazawa, and S. Nakajima, Heterocycles, 24, 2449 (1986), and earlier papers cited therein.
- 15) Personal communication, 1987.
- 16) N. J. Leonard and K. Ito, J. Am. Chem. Soc., 95, 4010 (1973).
- 17) The observed complexity of the proton signals is interpretable in terms of cis-trans isomerism of the formamido moiety, as we have experienced in similar structures. 4b,9b-d,11a)
- 18) R. Mozingo, "Organic Syntheses," Coll. Vol. III. ed. by E. C. Horning, John Wiley & Sons, New York, 1955, p. 181.