Reaction Products from Fischer's Methylation of 3-Deoxy-D-glycero-D-galacto-2-nonulosonic Acid (KDN)

Toshitsugu Kai,* Xue-Long Sun, Makoto Tanaka, Hiroaki Takayanagi, and Kimio Furuhata

School of Pharmaceutical Sciences, Kitasato University, 5-9-1 Shirokane, Minato-ku, Tokyo 108, Japan. Received April 6, 1995; accepted August 21, 1995

Several compounds were isolated from the Fischer's methylation reaction mixture of 3-deoxy-D-glycero-D-galacto-2-nonulosonic acid (KDN) with hydrochloric acid or methanesulfonic acid in methanol. The major product was methyl (methyl 3-deoxy- β -D-glycero-D-galacto-2-nonulopyranosid)onate (2), which we have already reported. New furan derivatives and a 4,8-anhydro product were also isolated. The structures of these compounds were elucidated by nuclear magnetic resonance spectroscopy and X-ray crystallographic analysis.

Key words sialic acid; Fischer's glycosylation; furan derivative; 4,8-anhydro product; crystalline KDN; β-elimination

Deaminoneuraminic acid, 3-deoxy-D-glycero-D-galacto-2-nonulosonic acid (KDN, 1), was isolated from polysialoglycoprotein (PSGP) of rainbow trout eggs. 1) It has been synthesized, 2,3) and we have also synthesized various analogues. 3-8) These KDN analogues are of particular relevance to the investigation of neuramidase inhibitors, 9) and they also provide interesting carbohydrate templates for further transformation. KDN methyl ester and its methyl β -glycoside are extremely useful materials for the synthesis of KDN analogues. In a previous paper, we described their synthesis under Fischer's conditions. 6)

OH OH OH OH HO 7 6 O 2 COOH

1
Fig. 1

In this report, we describe the isolation of minor products from the reaction mixture and determination of their structures by means of mass and proton nuclear magnetic resonance (¹H-NMR) spectroscopy and X-ray crystallographic analysis.

Results and Discussion

Synthesis of carbohydrates requires a large quantity of starting material, because the overall yield is usually low. We developed an improved chemical synthesis of KDN by base-catalyzed condensation of oxalacetic acid with D-mannose, obtaining $100 \, \mathrm{g}$ quantities, and we succeeded in crystallizing the β -pyranose form of 1.

Previously, we examined Fischer's methyl glycosylation of **1** using the strong cation exchange resin, Dowex-50 (H⁺), and we found an anomeric equilibrium between α - and β -furanose and pyranose.⁶⁾ Thus we suspected that the β -pyranoside of methyl (methyl 3-deoxy-D-glycero-D-galacto-2-nonulopyranosid)onate (**2**) would be the only product if **1** were treated with a strong acid, such as hydrochloric acid or methanesulfonic acid, in methanol



Chart 1

Chart 2. Possible Mechanism for the Formation of 6 and 7

and heated. We thus found a facile method for the preparation of 2 as new crystals. Furthermore, from the reaction mixture, new furan derivatives and a 4,8-anhydro compound were isolated as minor products.

Treatment of 1 with hydrochloric acid in methanol at 70 °C gave one major and five minor products. The major product, 2, was crystallized in 28% yield from the concentrated reaction mixture, and the filtrate was separated by column chromatography on silica gel to give three known compounds, the α -isomer of 2 (3), and methyl (methyl 3-deoxy- β - and - α -D-glycero-D-galacto-2nonulofuranosid)onates (4 and 5) in 26% total yield, together with two new compounds, methyl 5-(α - and β -Derythrofuranosyl)furan-2-carboxylates (6, 0.2% yield and 7, 0.9% yield). The structures of 6 and 7 were elucidated by ¹H-NMR spectral analysis. The configuration at C-6 of the furan derivatives (6 and 7) was deduced from nuclear Overhauser effect (NOE) experiments. That is, on irradiation of H-6, NOEs were observed at H-9 in both the α - and β -anomers (α -anomer 6, 1.1%; β -anomer 7, 1.7%). But, on irradiation of H-9, NOE was observed at H-6 only in the β -anomer (7, 2.3%), and not in the α-anomer. Furthermore, the structures of 6 and 7 were confirmed by comparison of the 1H-NMR spectral properties with those of methyl 5-(2',3'-di-O-acetyl-α- and $-\beta$ -D-erythrofuranosyl)pyrrole-2-carboxylates, which are thermal degradation products of sodium N-acetylneuraminate.10) The reaction mechanism is considered to involve the formation of a furan ring through acidcatalyzed β -elimination of hydroxy groups and nucleophilic attack of the hydroxy group at C-9 upon the C-6 position, as shown in Chart 2.

In the case of treatment with methanesulfonic acid, instead of hydrochloric acid, in methanol at 70°C, 1 was converted into a mixture of 2, 3, 4, and 5 in 67% total yield, and a new compound, methyl (methyl 4,8-anhydro-3-deoxy-α-D-glycero-D-galacto-2-nonulofuranosid)onate

Table 1. ¹H-NMR Data for 8 and 9

Proton	8	9
3-H	2.47 (dd, J=4.0, 14.5 Hz)	2.28 (dd, J=4.0, 14.5 Hz)
3-H'	2.85 (dd, J=2.0, 14.5 Hz)	2.60 (dd, J = 2.0, 14.5 Hz
4-H	4.27 (dd, J=2.0, 4.0 Hz)	4.22 (dd, J=2.0, 4.0 Hz)
5-H	4.69 (dd, J=2.0, 3.5 Hz)	4.45 (dd, J=2.0, 3.5 Hz)
6-H	4.23 (dd, J=3.5, 9.5 Hz)	5.14 (dd, J = 3.5, 9.5 Hz)
7-H	4.52 (t, J=9.5 Hz)	5.37 (t, J=9.5 Hz)
8-H	3.77 (dt, J = 3.0, 9.5 Hz)	3.56 (dt, J = 4.0, 9.5 Hz)
9-H	4.17 (dd, J = 3.0, 9.5 Hz)	4.08 (2H, d, J=4.0 Hz)
9-H'	4.53 (dd, J = 3.0, 9.5 Hz)	
2-OMe	3.36 (s)	3.27 (s)
2-COOMe	3.65 (s)	3.82 (s)
OAc	· · · · · · · · · · · · · · · · · · ·	2.03 (s), 2.06 (s), 2.10 (s)

Spectra were measured in C_5D_5N for **8** and in CDCl₃ for **9** at 300 MHz. Chemical shifts are given in δ (ppm). Coupling constants are given in parentheses

(8) was also isolated in 3% yield. Acetylation of 8 with acetic anhydride and pyridine afforded methyl (methyl 6,7,9-tri-O-acetyl-4,8-anhydro-3-deoxy- α -D-glycero-D-galacto-2-nonulopyranosid)onate (9). The structures of 8 and 9 were elucidated by ¹H-NMR spectral analysis, as shown in Table 1. The coupling constants between H-5 and H-6 (J=3.5 Hz), H-6 and H-7 (J=9.5 Hz), and H-7 and H-8 (J=9.5 Hz) indicate a manno-type proton network for 8. Further, the coupling between H-4 and H-5 (J=2.0 Hz) shows H-4 to be axial. Three singlet signals at δ 2.03, 2.06, and 2.10 of 9 indicate the presence of three newly introduced O-acetyl groups.

To confirm the stereochemistry of 9, X-ray crystallography was conducted. Figure 2 shows the crystal structure of 9. From this ORTEP view, the pyranoside takes chair form $({}^{7}C_{4})$, and the furanoside takes twist form $({}^{0}T_{4})$, and the anomeric configuration is α .

We assume that 8 is also formed from the open structure of KDN methyl ester in a multistep process, as shown in

210 Vol. 44, No. 1

Chart 3. The initial step could involve acid-catalyzed β -elimination of the hydroxy group at C-4 and the formation of an α,β -unsaturated keto moiety. Subsequent steps would include nucleophilic attack of the hydroxy group at C-8 on the C-4 position, resulting in a new anhydro ring through intramolecular Michael reaction, and the formation of the furanosyl ring. In this process, none of the original configuration of 8 is changed.

In conclusion, we have developed a facile method for the preparation of crystalline KDN and its β -methyl glycoside, which are required for syntheses of sialic acid analogues. The formation of bicyclic derivatives (6, 7, and 8) on Fischer methylation is a consequence of the characteristic properties of KDN.

Experimental

Melting points were measured with a Yazawa BY-10 melting point apparatus without correction. Optical rotations were measured with a JASCO DIP-370 digital polarimeter. Thin layer chromatography (TLC) was performed on silica gel 60 F_{254} (Merck) plates, and spots were detected under ultraviolet (UV) light and by spraying with 5% sulfuric

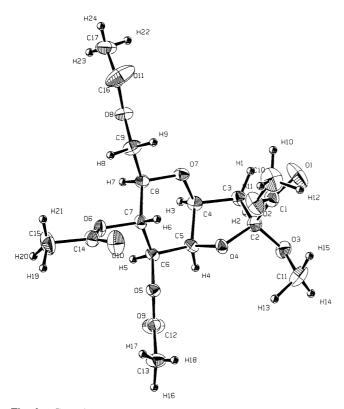


Fig. 2. Crystal Structure of 9

acid. Fast atom bombardment mass spectra (FAB-MS), and infrared (IR) spectra were measured with JEOL JMS-DX300 and JASCO A-102 instruments, respectively. The $^1\mathrm{H}\text{-NMR}$ spectra were measured with Varian VXR-300 spectrometer. Tetramethylsilane (TMS) in CDCl₃ and C_5D_5N or sodium 3-(trimethylsilyl)-1-propanesulfonate (DSS) in D_2O were used as internal references. Column chromatography was conducted on Silica gel 60 (70—230 mesh, Merck).

Purification of KDN A solution of 1 (10.0 g, 37.3 mmol) in water (100 ml) was passed over the anion exchange resin Dowex-1 (HCOO⁻, 350 ml), and 1 was eluted with 0—1 N formic acid. The eluate was concentrated under reduced pressure. The residue was left at room temperature for 7d, to afford colorless crystals (4.65 g, 47%). mp 144—147 °C. [α]_D³⁰ – 49.7° (c=1, H₂O). Anal. Calcd for C₉H₁₆O₉: C, 40.30; H, 6.01. Found: C, 40.39; H, 5.89. IR ν^{KB}_{max} cm⁻¹: 3440, 2960, 1730. ¹H-NMR (300 MHz, D₂O) δ: 1.75 (1H, dd, J=11.5, 12.5 Hz, 3-H_{ax}), 2.18 (1H, dd, J=5.0, 12.5 Hz, 3-H_{eq}), 3.51 (1H, t, J=9.5 Hz, 5-H), 3.58 (1H, dd, J=6.0, 11.5 Hz, 9-H), 3.67 (1H, ddd, J=3.0, 6.0, 9.0 Hz, 8-H), 3.79 (1H, dd, J=1.5, 9.0 Hz, 7-H), 3.79 (1H, dd, J=3.0, 11.5 Hz, 9-H'), 3.91 (1H, dd, J=1.5, 9.5 Hz, 6-H), 3.92 (1H, ddd, J=5.0, 9.5, 11.5 Hz, 4-H).

Esterification and Glycosylation of 1 by Fischer's Method Using Hydrochloric Acid and Methanol Acetyl chloride (22.08 g, 0.281 mol) was added slowly to methanol (500 ml), then KDN (1, 10.0 g, 37.3 mmol) was added to the solution. The mixture was heated at 70 °C for 6h, then evaporated, and the residual syrup was left at room temperature to afford methyl (methyl 3-deoxy-β-D-glycero-D-galacto-2-nonulopyranosid)onate (2, 3.04 g, 28%) as colorless crystals. These were filtered off, and the filtrate was evaporated to dryness. The residual syrup was purified on a column of silica gel with chloroform-methanol (10:1) to give a mixture of methyl 5-(α - and β -D-erythrofuranosyl)furan-2carboxylates (6 and 7, 216.9 mg, 3%), a mixture of methyl (methyl 3-deoxy- β - and - α -D-glycero-D-galacto-2-nonulopyranosid)onate (2 and 3) and a mixture of methyl (methyl 3-deoxy- β - and - α -D-glycero-Dgalacto-2-nonulofuranosid)onate (4 and 5) (2.9 g, 26%). The mixture of 6 and 7 was separated by silica gel TLC with 20:1 (v/v) chloroformmethanol (repeated 3 times) to give 6 (12.9 mg, 0.2%) and 7 (78.3 mg, 0.9%).

6: $[\alpha]_{\rm b}^{30}$ -7.6° (c=1, MeOH). HRMS m/z Calcd for C₁₀H₁₂NaO₆: 251.0532 (M⁺ + Na). Found: 251.0533 (M⁺ + Na). IR $\nu_{\rm max}^{\rm film}$ cm⁻¹: 3420, 1740, 1610. ¹H-NMR (300 Mz, C₅D₅N) δ : 3.74 (3H, s, -COOCH₃), 4.22 (1H, dd, J=6.5, 8.5 Hz, 9-H), 4.30 (1H, dd, J=6.5, 8.5 Hz, 9-H'), 4.67 (1H, t, J=4.5 Hz, 7-H), 4.72 (1H, dt, J=4.5, 6.5 Hz, 8-H), 5.27 (1H, d, J=4.5 Hz, 6-H), 6.90 (1H, d, J=3.5 Hz, 4-H), 7.30 (1H, d, J=3.5 Hz, 3-H).

7: $[\alpha]_D^{30} - 59.4^{\circ}$ (c = 1, MeOH). HRMS m/z Calcd for $C_{10}H_{12}NaO_6$: 251.0532 (M⁺ + Na). Found: 251.0544 (M⁺ + Na). IR $\nu_{\rm max}^{\rm film}$ cm⁻¹: 3450, 2980, 1740, 1660, 1540. ¹H-NMR (300 Mz, C_5D_5N) δ : 3.76 (3H, s, -COOCH₃), 4.26 (1H, dd, J = 2.5, 9.0 Hz, 9-H), 4.42 (1H, dd, J = 5.0, 9.0 Hz, 9-H'), 4.65 (1H, dt, J = 2.5, 9.0 Hz, 8-H), 4.80 (1H, t, J = 5.0, 7.0 Hz, 7-H), 5.34 (1H, d, J = 7.0 Hz, 6-H), 6.66 (1H, d, J = 3.5 Hz, 4-H), 7.24 (1H, d, J = 3.5 Hz, 3-H).

Esterification and Glycosylation of 1 by Fischer's Method Using Methanesulfonic Acid and Methanol A solution of 1 (5.1 g, 19.0 mmol) and methanesulfonic acid (1 ml) in methanol was stirred for 8 h at 70 °C, then neutralized with dry Dowex-1 (OH $^-$) and filtered. The filtrate was evaporated and the residual syrup was chromatographed on a column of silica gel with chloroform—methanol (4:1) to give methyl (methyl

Chart 3. Possible Mechanism for the Formation of 8

4,8-anhydro-3-deoxy- α -D-glycero-D-galacto-2-nonulofuranosid)onate (8, 140 mg, 3%) and a mixture of 2, 3, 4, and 5 (3.8 g, 67%).

8: $[\alpha]_{0}^{23}$ -53.6° (c=1, MeOH). HRMS m/z Calcd for $C_{11}H_{19}O_{8}$: 279.1080 (M⁺+H). Found: 279.1104 (M⁺+H). IR $\nu_{max}^{CHCl_{3}}$ cm⁻¹: 3450, 1745. ¹H-NMR data are given in Table 1.

Acetylation of 8 Acetic anhydride (165 mg, 1.62 mmol) was added to a solution of 8 (50.0 mg, 0.180 mmol) in pyridine (128 mg, 1.62 mmol) at room temperature. The mixture was stirred for 16 h at room temperature, then concentrated, and the residue was purified on a column of silica gel with *n*-hexane–ethyl acetate (1:1) to give methyl (methyl 6,7,9-tri-O-acetyl-4,8-anhydro-3-deoxy- α -D-glycero-D-galacto-2-nonulo-furanosid)onate (9, 30 mg, 41%) as colorless prisms. mp 133—135 °C, $[\alpha]_{\rm D}^{23}$ –73.2° (c=1, MeOH). Anal. Calcd for $C_{17}H_{24}O_{11}$: C, 50.50; H, 5.94. Found: C, 50.49; H, 6.02. IR $v_{\rm max}^{\rm CCl_4}$ cm $^{-1}$: 1750. 1 H-NMR data are given in Table 1.

Crystal Data for 9 A crystal with the dimensions of $0.3 \times 0.3 \times 0.3 \times 0.3$ mm³ was used for the structure determination. The cell dimensions and diffraction intensities were measured on a Rigaku four-circle diffractometer (AFC-5R), using graphite-monochromated CuK_{α} radiation.

Crystal Data: $C_{17}H_{24}O_{11}$, orthorhombic, space group $P2_12_12_1$, a=9.625(4), b=25.930(9), c=7.802(5) Å, V=1947(1) Å³, Z=4, $D_{\rm calcd}=1.379$ g/cm³. In total, 2053 independent reflections in the range of $2\theta=139.6^{\circ}$ were collected by use of the $2\theta-\omega$ scan mode with a scanning rate of 16° min⁻¹ (ω). In total, 1629 independent reflections with $I>3.00\sigma(I)$ were obtained and corrected for Lorentz and polarization factors, but not for absorption. The structure was solved by direct methods using the program MITHRIL. ¹¹ The positions of all hydrogen atoms were located in the difference Fourier map. Atomic scattering factors were taken from the International Table for X-Ray Crystallography. ¹² All calculations were performed using the TEXSAN ¹³ crystallographic software package of Molecular Structure Corporation. The final R value was 5.3%.

References

- Nadano D., Iwasaki M., Endo S., Kitajima K., Inoue S., Inoue Y., J. Biol. Chem., 261, 11550—11557 (1986).
- 2) Shirai R., Ogura H., Tetrahedron Lett., 30, 2263—2264 (1989).
- Nakamura M., Furuhata K., Yamasaki T., Ogura H., Chem. Pharm. Bull., 39, 3140—3144 (1991).
- Nakamura M., Furuhata K., Ogura H., Chem. Pharm. Bull., 36, 4807—4813 (1988).
- Nakamura M., Furuhata K., Ogura H., Chem. Pharm. Bull., 37, 821—823 (1989).
- Nakamura M., Takayanagi H., Furuhata K., Ogura H., Chem. Pharm. Bull., 40, 879—885 (1992).
- Nakamura M., Fujita S., Ogura H., Chem. Pharm. Bull., 41, 21—25 (1993).
- 8) Nakamura M., Takeda K., Takayanagi H., Asai N., Ibata N., Ogura H., Chem. Pharm. Bull., 41, 26-30 (1993).
- von Itzstein M., Wu W.-Y., Kok G. B., Pegg M. S., Dyason J. C., Jin B., Phan T. V., Smythe M. L., White H. F., Oliver S. W., Colman P. M., Varghese J. N., Ryan D. M., Woods J. M., Bethell R. C., Hotham V. J., Cameron J. M., Penn C. R., Nature (London), 363, 418—423 (1993).
- Sugiyama N., Saito K., Fujikura K., Sugai K., Yamada N., Goto M., Ban C., Hayasaka E., Tomita K., Carbohydr. Res., 212, 25—36 (1991).
- Gilmore C. J., MITHRIL, an integrated direct methods computer program, J. Appl. Cryst., 17, 42—46, Univ. of Glasgow, Scotland, 1984.
- 12) "International Tables for X-Ray Crystallography," Vol. IV, Kynoch Press, Birmingham, 1974, pp. 72—149.
- TEXSAN-TEXRAY Structure Analysis Package, Molecular Structure Corporation, 1985.