

Marine Natural Products. XXXVII.¹⁾ Aragusteroketals A and C, Two Novel Cytotoxic Steroids from a Marine Sponge of *Xestospongia* sp.

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Aragusteroketals A (1) and C (2), two new marine steroids having dimethylketal structure, were isolated from an Okinawan marine sponge of *Xestospongia* sp. The chemical structures of 1 and 2 were determined on the basis of spectroscopic analysis and chemical evidence. Compounds 1 and 2 showed potent cytotoxic activity with the same IC₅₀ value of 4 ng/ml against KB cells.

Key words marine sponge; *Xestospongia*; steroid; cytotoxic; dimethylketal

In the past 20 years, a number of steroids with unusual structures have been isolated from marine invertebrates. In 1993, Yamada and his group³⁾ first reported aragusterol A (3) from an Okinawan marine sponge of *Xestospongia* sp.; 3 strongly inhibited the cell proliferation of tumor cells *in vitro* and also showed potent *in vivo* anti-tumor activities against P388 and L1210 in mice. Since then, several side-chain analogues, aragusterols B,⁴⁾ C (4),⁵⁾ and D⁴⁾ and xestokerols A, B, and C,⁶⁾ have been isolated. In our continuing search¹⁾ for new biologically active marine natural products, we have investigated the chemical constituents of an Okinawan marine sponge of *Xestospongia* sp. and have isolated two new aragusterol analogues with dimethylketal structures named aragusteroketals A (1) and C (2), together with aragusterols A (3) and C (4). Aragusteroketals A (1) and C (2) exhibited potent cytotoxic activity against KB cells. In this paper, we describe the structure elucidation of aragusteroketals A (1) and C (2).

The acetone extract of a marine sponge of *Xestospongia* sp. (collected on the reef (−10 m) of Iriomote Island, Okinawa Prefecture), which showed cytotoxic activity against KB cells, was subjected to bioassay-guided separation (based on cytotoxicity towards L1210 and KB cells). The acetone extract was partitioned into water–EtOAc mixture to give the cytotoxic EtOAc-soluble portion, which was then separated by repeated SiO₂ column chromatography (*n*-hexane–EtOAc and CHCl₃–MeOH–H₂O) to give two active fractions (fraction A and fraction B). Fraction A was separated by SiO₂ column

chromatography and reversed-phase HPLC to provide aragusteroketal A (1) (0.08% yield from the EtOAc-soluble portion) and aragusterol A (3) (0.28%). Fraction B was separated by SiO₂ column chromatography and reversed-phase HPLC to provide aragusteroketal C (2) (0.28%) and the major steroid, aragusterol C (4) (2%).

Aragusteroketal A (1) was obtained as an amorphous solid. The FAB-MS of 1 showed a quasi-molecular (M + Li)⁺ ion peak at *m/z* 511 and the molecular formula was determined as C₃₁H₅₂O₅ by HR FAB-MS and NMR analysis. The ¹H- and ¹³C-NMR spectra of 1 defined the presence of an ketal (δ_C 100.3), two secondary hydroxyls [δ 3.36 (dd, *J* = 4, 11 Hz), 3.49 (d-like, *J* = 10 Hz); δ_C 77.8 (d), 72.2 (d)], an epoxide [δ 2.91 (d, *J* = 4 Hz), 3.08 (d, *J* = 4 Hz); δ_C 50.5 (t), 65.7 (s)], a 1,2-disubstituted cyclopropane ring [δ 0.17, 0.25 (2H), 0.51], two methoxys [δ 3.14, 3.19; δ_C 47.8 (q) × 2], and four methyls [δ 0.70, 0.79, 0.95, 1.02 (3H each)]. The COSY and HMQC spectra of 1 revealed four partial structures (fragment A, C-1—C-2; fragment B, C-4—C-9 and C-11—C-12; fragment C, C-14—C-17; fragment D, C-22—C-29). The connectivity of these partial structures was figured out on the basis of the heteronuclear multiple bond connectivity (HMBC) correlations depicted in Fig. 1. For example, the ketal carbon at C-3 was correlated with OCH₃, H_{ab}-2, and H_b-4. The H₃-19 was correlated with C-1, C-5, and C-9. These correlations defined the A and B ring structure in 1. The correlations between the H₃-18 proton and C-12, C-13, C-14, and C-17 revealed the connectivity between rings C and D. Furthermore, the connectivity of fragment

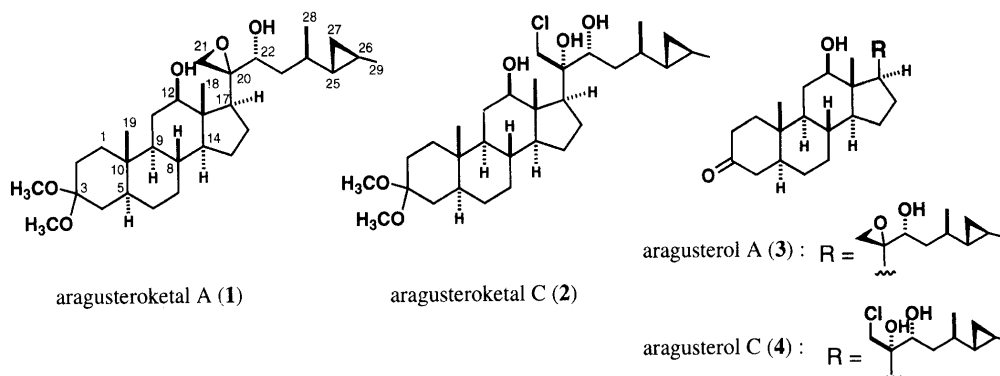
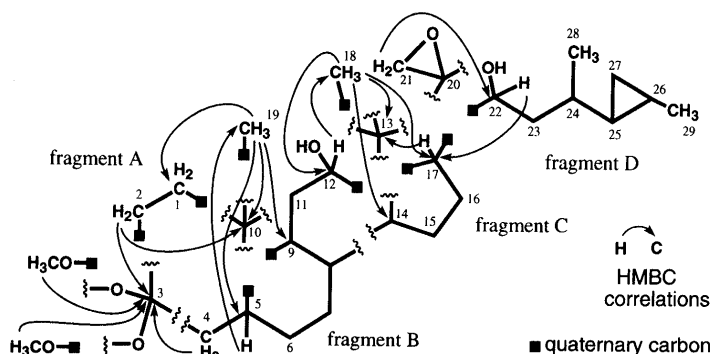


Chart 1

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Fig. 1. HMBC Correlations for Aragusteroketal A (**1**)Table 1. ^1H - and ^{13}C -NMR Data for Aragusteroketal A (**1**) and Aragusteroketal C (**2**)

| Atom No. | $\delta_{\text{C}}^{\text{a)}$ | 1 $\delta^{\text{b)}$ | $\delta_{\text{C}}^{\text{a)}$ | 2 $\delta^{\text{b)}$ |
|------------------|--------------------------------|-------------------------------------|--------------------------------|---|
| 1 | 34.9 (t) | a 1.07 (m), b 1.56 (m) | 35.0 (t) | a 1.09 (ddd, 3, 13, 14), b 1.57 (ddd, 3, 3, 13) |
| 2 | 28.3 (t) | a 1.42 (ddd, 4, 14, 18), b 1.90 (m) | 28.3 (t) | a 1.44 (ddd, 4, 14, 14), b 1.88 (m) |
| 3 | 100.3 (s) | | 100.3 (s) | |
| 4 | 35.4 (t) | a 1.28 (m), b 1.65 (m) | 35.3 (t) | a 1.31 (m), b 1.63 (m) |
| 5 | 42.4 (d) | 1.30 (m) | 42.3 (d) | 1.28 (m) |
| 6 | 28.3 (t) | a 1.20 (m), b 1.28 (m) | 28.3 (t) | a 1.20 (m), b 1.29 (m) |
| 7 | 31.3 (t) | a 0.88 (m), b 1.65 (m) | 31.3 (t) | a 0.88 (m), b 1.68 (m) |
| 8 | 33.8 (d) | 1.30 (m) | 33.8 (d) | 1.32 (m) |
| 9 | 52.4 (d) | 0.82 (m) | 52.6 (d) | 0.83 (m) |
| 10 | 35.7 (s) | | 35.6 (s) | |
| 11 | 29.3 (t) | a 1.28 (m), b 1.79 (m) | 29.7 (t) | a 1.27 (m), b 1.73 (m) |
| 12 | 77.8 (d) | 3.36 (dd, 4, 11) | 77.9 (d) | 3.43 (dd, 5, 11) |
| 13 | 48.9 (s) | | 49.0 (s) | |
| 14 | 54.5 (d) | 1.02 (m) | 54.2 (d) | 0.94 (m) |
| 15 | 23.7 (t) | a 1.28 (m), b 1.73 (m) | 23.5 (t) | a 1.25 (m), b 1.70 (m) |
| 16 | 26.8 (t) | a 1.58 (m), b 1.95 (m) | 23.5 (t) | a 1.68 (m), b 1.93 (m) |
| 17 | 48.2 (d) | 2.12 (dd, 10, 10) | 55.0 (d) | 1.92 (m) |
| 18 | 8.3 (q) | 0.70 (s) | 8.8 (q) | 0.95 (s) |
| 19 | 11.5 (q) | 0.79 (s) | 11.5 (q) | 0.81 (s) |
| 20 | 65.7 (s) | | 76.9 (s) | |
| 21 | 50.5 (t) | a 2.91 (d, 4), b 3.08 (d, 4) | 47.3 (t) | a 3.87 (s), b 3.88 (s) |
| 22 | 72.2 (d) | 3.49 (d-like, 10) | 71.3 (d) | 3.94 (dd, 11, 5) |
| 23 | 40.2 (t) | a 1.50 (m), b 1.62 (m) | 37.6 (t) | a 1.34 (m), b 1.68 (m) |
| 24 | 34.9 (d) | 0.94 (m) | 35.3 (d) | 0.98 (m) |
| 25 | 27.6 (d) | 0.25 (m) | 27.9 (d) | 0.26 (m) |
| 26 | 12.5 (d) | 0.51 (m) | 12.3 (d) | 0.53 (m) |
| 27 | 12.4 (t) | a 0.17 (m), b 0.25 (m) | 12.5 (t) | a 0.17 (m), b 0.26 (m) |
| 28 | 18.9 (q) | 0.95 ^{c)} | 18.8 (q) | 0.95 (s) |
| 29 | 19.1 (q) | 1.02 (d, 6) | 19.2 (q) | 1.02 (d, 6) |
| OCH ₃ | 47.8 (q) | 3.19 (s) | 47.5 (q) | 3.19 (s) |
| OCH ₃ | 47.8 (q) | 3.14 (s) | 47.5 (q) | 3.14 (s) |

a) 125 MHz in CDCl_3 ; b) 500 MHz in CDCl_3 , J value in Hz. c) Observed at δ 1.08 (d, 7) in $\text{C}_5\text{D}_5\text{N}$.

D and the epoxide was defined by the HMBC correlations between H-21 and C-22; H-22 and C-17. Thus, the plane structure of aragusteroketal A has been constructed as **1**, which is presumed to be a 3-dimethylketal analogue of aragusterol A (**3**). This presumption was supported by the following evidence. When aragusteroketal A (**1**) was kept in CDCl_3 at room temperature, **1** was gradually converted to aragusterol A (**3**), the physicochemical properties of which were identical with those of the compound isolated from the sponge. From these findings, the absolute stereostructure of aragusteroketal A was confirmed to be **1**.

Aragusteroketal C (**2**), a colorless amorphous solid, gave a quasi-molecular $(\text{M} + \text{Na})^+$ ion peak at m/z 563 in the FAB-MS, the composition being defined as $\text{C}_{31}\text{H}_{53}$ -

ClNaO_5 by HR FAB-MS analysis. The ^1H - and ^{13}C -NMR spectra of **2** also showed signals ascribable to a ketal (δ_{C} 100.3), two secondary hydroxyls [δ 3.43 (dd, $J=5$, 11 Hz), 3.94 (dd, $J=5$, 11 Hz); δ_{C} 77.9 (d), 71.3 (d)], a 1,2-disubstituted cyclopropane ring [δ 0.17, 0.26 (2H), 0.53], two methoxyls [δ 3.14, 3.19; δ_{C} 47.5 (q) \times 2], and four methyls [δ 0.81, 0.95, 0.98, 1.02 (3H, each)]. The NMR spectra of **2** lacked the oxymethylene signal assignable to the epoxide in **1** and showed characteristic signals at δ 3.88 and 3.87 (both 1H, s) and δ_{C} 47.3 (t), which were similar to those of the chlorohydrin moiety in aragusterol C (**4**). Furthermore, aragusteroketal C (**2**) was gradually converted in CDCl_3 to aragusterol C (**4**). Consequently, aragusteroketal C (**2**) has been elucidated to be the 3-dimethylketal analogue of aragusterol C (**4**).

Aragusteroketals A (**1**) and C (**2**) have also been obtained from the acetone extract of the sponge by use of a separation procedure not involving methanol. Aragusteroketals A (**1**) and C (**2**) are rare examples of natural products having a dimethylketal structure. Aragusteroketals A (**1**) and C (**2**) and aragusterols A (**3**) and C (**4**) exhibited potent cytotoxic activities with IC_{50} values of 0.004, 0.004, 0.03, and 0.02 $\mu\text{g/ml}$ against KB cell, respectively.

Experimental

The IR spectra were obtained with a JASCO FT-IR 5300 spectrometer. Optical rotations were measured with a JASCO DIP-370 digital polarimeter. The ^1H - and ^{13}C -NMR spectra were measured with a JEOL GX-500 spectrometer and with TMS as the internal standard. The FAB-MS were recorded on a JEOL JMS SX-102 mass spectrometer.

Extraction and Isolation The frozen sponge of *Xestospongia* sp. (5 kg), which was collected in July, 1993 off Iriomote Island, Okinawa, was initially steeped in acetone. The residue obtained by evaporation of the solvent under reduced pressure was partitioned into EtOAc– H_2O mixture (1:1), and the EtOAc layer was taken and evaporated to give the EtOAc-soluble portion (15 g). The EtOAc-soluble portion (6 g) was separated by SiO_2 column (*n*-hexane:EtOAc=3:1) chromatography to give the cytotoxic fraction (750 mg). This fraction was further separated on an SiO_2 column (CHCl_3 :MeOH: H_2O =100:3:1, lower phase) to give fractions A (109 mg) and B (310 mg). Fraction A (109 mg) was then separated by SiO_2 column (*n*-hexane:acetone=4:1) chromatography and reversed-phase HPLC [Cosmosil 5C₁₈ AR 10 \times 250 mm, flow rate 3 ml/min; eluent MeOH– H_2O – CH_2Cl_2 (500:100:6)] to afford aragusteroketal A (**1**, 5 mg) and aragusterol A (**3**, 17 mg). Fraction B (310 mg) was also separated by SiO_2 column (*n*-hexane:acetone=4:1) chromatography and reversed-phase HPLC [Cosmosil 5C₁₈ AR 10 \times 250 mm, flow rate 4 ml/min; eluent MeOH– H_2O – CH_2Cl_2 (900:100:10)] to give aragusteroketal C (**2**, 17 mg) and aragusterol C (**4**, 119 mg).

Aragusteroketal A (**1**): Amorphous solid, $[\alpha]_D^{25} +25.3^\circ$ ($c=0.12$, CHCl_3 , 25 $^\circ\text{C}$). IR ν_{max} (KBr): 3300, 2945 cm^{-1} . FAB-MS (LiCl) m/z :

511 ($\text{M}+\text{Li}$) $^+$. HR FAB-MS m/z : Calcd for $\text{C}_{31}\text{H}_{52}\text{LiO}_5$: 511.3975. Found: 511.3957. ^1H -NMR (500 MHz, CDCl_3 , δ), ^{13}C -NMR (125 MHz, CDCl_3 , δ_C): as shown in Table 1. HMBC correlations of **1**: C-1/ H_a -2, H_3 -19; C-3/ H_{ab} -2, $\text{OCH}_3 \times 2$, H_b -4; C-4/ H_b -2; C-5/ H_b -6, H_3 -19; C-8/ H_b -11; C-9/ H -8, H_{ab} -11, H_3 -19; C-12/ H_b -11, H_3 -18; C-13/ H -17, H_3 -18; C-14/ H_a -15, H_3 -18; C-17/ H_3 -18, H -22; C-18/ H -12, H -14; C-20/ H_{ab} -21; C-22/ H_{ab} -21; C-23/ H_3 -28; C-24/ H_{ab} -23, H -25, H -26, H_{ab} -27, H_3 -28; C-25/ H_{ab} -27, H_3 -28, H_3 -29; C-26/ H_a -27, H_3 -29; C-28/ H_{ab} -23; C-29/ H -25, H_{ab} -27.

Aragusteroketal C (**2**): Amorphous solid, $[\alpha]_D^{25} +8^\circ$ ($c=1.5$, CHCl_3 , 25 $^\circ\text{C}$). IR ν_{max} (KBr): 3289, 2947 cm^{-1} . FAB-MS m/z : 565, 563 ($\text{M}+\text{Na}$) $^+$. HR FAB-MS m/z : Calcd for $\text{C}_{31}\text{H}_{52}^{37}\text{ClNaO}_5$: 565.3450 and $\text{C}_{31}\text{H}_{52}^{35}\text{ClNaO}_5$: 563.3479. Found: 565.3446, 563.3473. ^1H -NMR (500 MHz, CDCl_3 , δ), ^{13}C -NMR (125 MHz, CDCl_3 , δ_C): as shown in Table 1. HMBC correlations of **2**: C-1/ H_a -2, H_3 -19; C-3/ H_b -1, H_{ab} -2, $\text{OCH}_3 \times 2$; C-5/ H_{ab} -4, H_3 -19; C-8/ H_b -7; C-9/ H -8, H_{ab} -11, H_3 -19; C-10/ H_{ab} -1, H_b -11, H_3 -19; C-12/ H_{ab} -11; C-13/ H_b -11, H_b -15, H_3 -18; C-14/ H -8, H_3 -18; C-17/ H_3 -18, H_{ab} -21; C-18/ H -12; C-20/ H -17, H_{ab} -21; C-22/ H_{ab} -21; C-23/ H -22, H_3 -28; C-24/ H -22, H_{ab} -27, H_3 -28; C-25/ H_{ab} -27, H_3 -29; C-26/ H -25, H_{ab} -27, H_3 -29; C-28/ H_a -23; C-29/ H_{ab} -27.

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References and Notes

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