

evidence in favor of this assumption, although the presence of two partial structures shown by heavy lines in **1a** (Fig. 1) could be ascertained. The molecular ion (HR-MS) of **2** [mp 265–266°C, $[\alpha]_D^{23} +50.3^\circ$ ($c=0.5$, CHCl_3); $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3450, 1120 (OH), m/z (rel. int.): 427 (M^+-CH_3 , 2), 424 ($\text{M}^+-\text{H}_2\text{O}$, 33), 409 ($\text{M}^+-\text{H}_2\text{O}-\text{CH}_3$, 7), 406 (13), 232 (9), 219 (11), 191 (100), 174 (34), 137 (14), 123 (11)] at m/z 442.3800 ($\text{C}_{30}\text{H}_{50}\text{O}_2$, 5), which is 18 mass units higher than that of **1**, indicated that **2** might be a hydration product of **1**. Its ^1H and ^{13}C NMR spectra (Tables 1 and 2) showed the presence of six tertiary methyl groups (in place of five in **1**), one hydroxymethine, and one hydroxy quaternary carbon in the molecule. No signals for the exo-methylene group could, however, be observed. Its ^{13}C NMR data (Table 2) clearly revealed a resemblance between the structures of **1** and **2**. The structure of **2** could finally be established based on its HMBC spectrum.

Table 1. ^1H Chemical Shifts^{a)} (δ ppm, CDCl_3 , 500 MHz) of **1–4**

	H ₃ -23	H ₃ -24	H ₃ -25	H ₃ -26	H ₃ -27	H ₃ -28/H-28	H ₂ -29	H ₃ -30/H ₂ -30	H-13 _B	H-17 _B	H-21
1	0.849	0.790	0.814	0.994	1.111	4.311 (dd, 10.4, 6.7)	1.933	4.539 (m)	1.53	1.32	2.471 (dd, 5.8, 2.1)
2	0.852	0.791	0.822	0.995	1.131	4.237 (dd, 10.4, 5.5)	1.28 (β) 1.84 (α)	1.338	1.55	1.37	1.84
3	0.835	0.783	0.806	0.994	0.800	10.016	4.495 4.908	1.790	1.69	1.72	2.672 (ddd, 8.7, 8.7, 4.0)

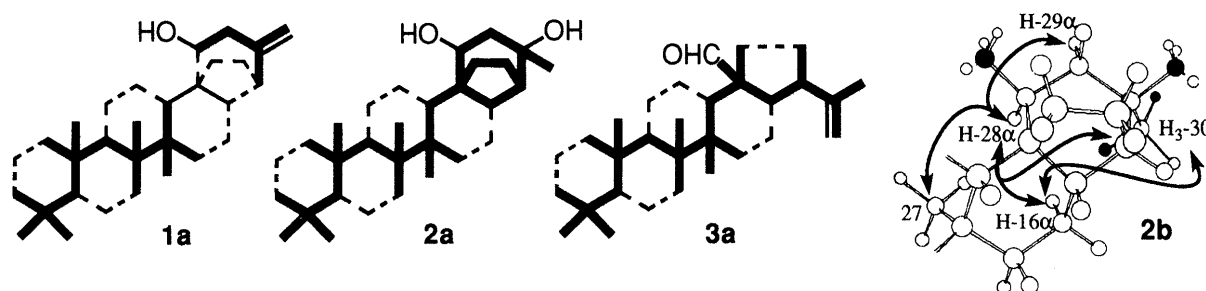
a) Figures in parentheses are coupling constants in Hz. Chemical shifts were assigned on the basis of ^1H - ^1H COSY, HSQC, and HMBC spectral analyses.

Table 2. ^{13}C Chemical Shifts (δ , CDCl_3 , 125 MHz) of **1–4**

	C-1	C-2	C-3	C-4	C-5	C-6	C-7	C-8	C-9	C-10
1	40.29	18.66	42.07	33.25	55.96	18.62	33.57	42.51	50.81	37.32
2	40.28	18.62	42.03	33.22	56.02	18.62	33.66	42.35	51.01	37.32
3	40.24	18.65	42.00	33.23	56.07	18.61	33.90	41.63	50.35	37.38
4	40.31	18.70	42.10	33.25	56.10	18.70	33.25	41.90	50.37	37.39

	C-11	C-12	C-13	C-14	C-15	C-16	C-17	C-18	C-19	C-20
1	21.86	27.20	50.76	42.63	32.52	20.31	53.22	47.91	31.17	29.87
2	22.00	27.20	51.25	42.39	33.40	22.46	51.58	47.11	31.37	26.20
3	20.93	23.94	51.92	42.08	32.76	21.43	53.15	59.63	35.79	27.52
4	20.91	23.99	49.42	42.07	33.61	21.67	54.88	44.80	41.90	27.39

	C-21	C-22	C-23	C-24	C-25	C-26	C-27	C-28	C-29	C-30
1	47.35	148.32	33.39	21.59	15.93	17.00	17.50	71.10	39.61	105.85
2	50.37	72.01	33.32	21.54	16.07	16.81	16.93	69.98	45.73	29.98
3	46.66	146.03	33.35	21.57	15.91	16.61	17.90	208.15	112.60	25.39
4	46.47	148.78	33.41	21.60	15.84	16.70	16.75	16.07	110.06	25.02



Thus, the two- and three-bond correlations of the methyl protons, the carbinyl proton (H-28), and some methylene protons (H₂-19 and H₂-20) with those of the neighboring carbons clearly showed the presence of the partial structure as in **2a** (Fig. 1). The carbon-carbon connectivities shown by broken lines could be deduced from ¹H-¹H COSY and HSQC spectral analyses. The stereochemistry at the chiral centers C-22 and C-28 could be established from the NOE interactions, viz. H₃-27 ↔ H-28α, H-28α ↔ H-29α, H-28α ↔ H₃-30, H-28α ↔ H-16α (δ 1.68), and H-16α ↔ H₃-30, observed in the NOESY spectrum of the compound as depicted in **2b** (Fig. 1). The coupling constants (10.4, 5.5 Hz, Table 1) of the ¹H NMR signal for the carbinyl proton (H-28α) of **2** indicated that the new carbocyclic ring F assumes a chair conformation and the hydroxyl group is equatorially oriented. The coupling constants of the H-28 signal of **1** were also very close to those of **2**. Based on the above observations, **1** and **2** may be represented as (28*S*)-28,29-cyclohop-22(30)-en-28-ol and (22*R*,28*S*)-28,29-cyclohopane-22,28-diol, respectively.

The high-resolution mass spectrum of **3** [mp 217–220°C, [α]_D²³ +145.6° (*c*=0.5, CHCl₃); ν_{max}^{KBr} cm⁻¹: 1700 (CHO), 1640, 890 (>C=CH₂), *m/z* (rel. int.): 409 (M⁺-CH₃, 13), 395 (M⁺-CHO, 7), 381 (M⁺-C₃H₇, 3), 353 (14), 312 (4), 271 (4), 233(8), 219 (18), 191 (100), 147 (42), 137 (20)] showed a molecular ion peak at *m/z* 424.3713 (13) corresponding to the molecular formula C₃₀H₄₈O. Its ¹H-NMR spectrum (Table 1) revealed the presence of six tertiary methyl groups, one exo-methylene group, and one aldehyde proton. The ¹³C chemical shifts (Table 2) of the compound were found to be very close to those of **4** except for the carbons C-13, C-18, C-19, and C-28. Since C-18 suffered a strong down-field shift by ~15 ppm compared to that of **4**, the aldehyde group of **3** must be located at this carbon. The assigned structure of the compound was finally corroborated by its 2D NMR spectra.

Biogenetically, the cyclohopane triterpenoids **1** and **2** may be formed from **3**, as shown in Chart 1. The double bond at Δ²²⁽²⁹⁾ may attack the aldehyde carbon forming the cyclohopane cation. Either a proton may be lost from C-30 of the cyclohopane cation to give **1** or a hydroxyl ion may attack the cation center at C-22 to yield **2**.

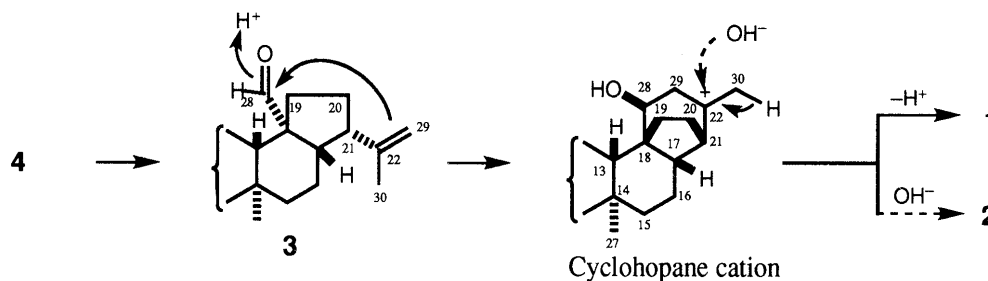


Chart 1

Many hopane-type triterpenoids, for example, oxygenated hopane triterpenoids from this species¹⁾ and bacteriohopane tetrol from *Acetobacter xylinum*,⁵⁾ and hopane-29-(2')-pentanone from sediments of various origins⁶⁾ with extra carbons at C-29 or C-30, have been reported from natural sources. Cyclohopane triterpenoids **1** and **2** are the first examples with an extra carbon ring in a hopane skeleton.

References and Notes

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(Received January 27, 1998; accepted February 27, 1998)