

Fluorine-18 Labeling of Methionine Derivatives in the Presence of Xenon Difluoride¹⁾

Kentaro HATANO

Department of Biofunctional Research, National Institute for Longevity Sciences, Gengo, Morioka, Obu, Aichi, 474–8522, Japan. Received June 1 1998; accepted June 30, 1998

Radiofluorination of protected methionine derivatives were examined in the presence of xenon difluoride (XeF₂). The simple reaction procedure with ¹⁸F-tetrabutylammonium fluoride and XeF₂ led *N*-Boc-methionine esters to corresponding ¹⁸F-fluoromethyl thioethers. The deprotection of the obtained labeled compounds was unsuccessful. This method expands the range of target molecules that can be labeled by fluorine-18 for noninvasive clinical measurements using positron emission tomography.

Key words fluorine-18; positron emission tomography; xenon difluoride

Noninvasive imaging of a specific biochemical or physiological function by positron emission tomography (PET) is a powerful tool in clinical diagnosis, neuroscience, and many other fields of medical and biological science. Among the short-lived positron emitting radionuclides, fluorine-18 (*t*_{1/2} = 110 min) is the most attractive from the viewpoint of radiosynthesis, radiopharmaceutical design, and radiology as well as clinical diagnosis.²⁾ Although some efforts have been made to examine the synthesis and the biological activity of ¹⁹F-fluorinated amino acids,³⁾ the number of reports on fluorine-18 labeled amino acids is limited.⁴⁾ Carbon-11 is another representative positron emitter with a half-life of 20.4 min, and methionine labeled with this nuclide is widely used in cancer diagnosis. Therefore ¹⁸F labeled methionine analogs should be promising.⁵⁾ The report of fluorination of methionine derivatives by xenon difluoride (XeF₂)⁶⁾ encouraged the author to examine its application to ¹⁸F chemistry.

Although the feasibility of stable XeF₂ is well established⁷⁾ there have been only limited examples of fluorine-18 labeled xenon difluoride (¹⁸F-XeF₂). Schrobilgen *et al.* reported the synthesis of ¹⁸F-XeF₂ by distillation of carrier added (CA) H¹⁸F (a radioisotope which was intentionally diluted by stable isotope) into XeF₂.⁸⁾ Chirakal and colleagues reported the reaction of CA ¹⁸F₂ and Xe to afford ¹⁸F-XeF₂.⁹⁾ These two methods ensured the synthesis of ¹⁸F-XeF₂ but required special synthetic apparatus. Patrick *et al.* reported on the fluorodecarboxylation reaction in the presence of XeF₂.¹⁰⁾ The simple work-up they employed was to mix the anhydrous fluorine-18 labeled tetrabutylammonium fluoride ((*n*-Bu)₄N¹⁸F), XeF₂, and a substrate in CH₂Cl₂. The present author applied this procedure to the fluorination of a methyl thioether moiety. *N*-Boc-protected methionine esters were prepared and allowed to react as shown in Fig. 1.

The incorporation of ¹⁸F proceeded with CA (*n*-Bu)₄N¹⁸F in CH₂Cl₂ (Table 1).¹¹⁾ The radiochemical yield of a methyl ester was superior to that of a *t*-butyl ester. Patrick *et al.* suggested that isotopic exchange between ¹⁸F-fluoride and XeF₂ would not occur under the present reaction conditions and the XeF₂ catalyzed generation of a cationic species which captures radioactivity.¹⁰⁾ This XeF₂-catalyzed radiofluorination mech-

anism could also be applied to the present result and CH₃CN might have an adverse effect on the generation of the active intermediate. The three previous studies employed CA fluorine-18, which reduces the specific radioactivity and therefore the sensitivity of measurement with the radiopharmaceutical thus obtained.^{8–10)} However, no carrier added (*n*-Bu)₄N¹⁸F was also found ineffective in this synthesis by direct comparison.

Janzen *et al.*⁶⁾ and others¹²⁾ reported on the instability of the CFH₂S– group with acid treatment. The deprotection of the labeled derivatives with CF₃COOH was also unsuccessful in the present experiment. Only a polar compound which was considered to be ¹⁸F-fluoride was observed with this treatment.

In summary, incorporation of ¹⁸F into the methylthioether moiety of the protected methionine derivatives was observed in the presence of XeF₂. Deprotection with CF₃COOH to obtain radiofluorinated methionine was unsuccessful. But the method employed here is very simple and easily accessible as it requires no special treatment (*i.e.*, introduction of a leaving group) to the substrates. ¹⁸F labeling of methionine containing small peptides such as substance P¹³⁾ appears promising. This new reaction should enlarge the range of target molecules that can be labeled for the imaging of biological processes with

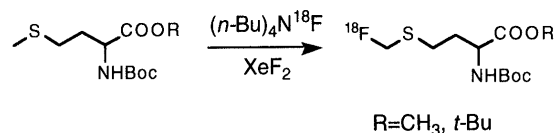


Fig. 1. Reaction Scheme

Table 1. Radiochemical Yield of ¹⁸F-Fluorinations

(<i>n</i> -Bu) ₄ N ¹⁸ F	Solvent	Substrate	Radiochemical yield
NCA	CH ₃ CN	<i>N</i> -Boc-methionine methyl ester	No reaction
CA	CH ₃ CN	<i>N</i> -Boc-methionine methyl ester	No reaction
NCA	CH ₂ Cl ₂	<i>N</i> -Boc-methionine methyl ester	No reaction
CA	CH ₂ Cl ₂	<i>N</i> -Boc-methionine methyl ester	12.3%
CA	CH ₂ Cl ₂	<i>N</i> -Boc-methionine <i>t</i> -butyl ester	1.9%

The Reaction mixture was stirred at room temperature for 60 min. NCA, no carrier added; CA, carrier added.

PET.

Acknowledgment The author is grateful to the members of Nishina Memorial Cyclotron Center, Japan Radioisotope Association where the present work was carried out.

References and Notes

- 1) This work was presented at the 35th annual meeting of the Japanese Society of Nuclear Medicine (Yokohama, 1995).
- 2) a) Kilbourn M. R., "Fluorine-18 labeling of radiopharmaceuticals," National Academy Press, Washington, D. C., 1990; b) Filler R., Kobayashi Y., Yagupolskii L. M. (eds.), "Organofluorine compounds in medicinal chemistry and biomedical applications," Elsevier Science Publishers, Amsterdam, 1993.
- 3) Kukhar V. P., Soloshonok V. A. (eds.), "Fluorine-containing amino acids. Synthesis and properties," John Wiley and Sons, Chichester, 1995.
- 4) Lemaire C., "PET studies on amino acid metabolism and protein synthesis," Mazoyer B. M. (ed.), Kluwer Academic Publishers, Amsterdam, 1993, pp. 89—108.
- 5) Ishiwata K., Kasahara C., Hatano K., Ishii S. I., Senda M., *Ann. Nucl. Med.*, **11**, 115—122 (1997).
- 6) Janzen A. F., Wang P. M. C., Lemaire A. E., *J. Fluorine Chem.*, **22**, 557—559 (1983).
- 7) a) Filler R., *Israel J. Chem.*, **17**, 71—79 (1978); b) Bardin V. V., Yagupolskii Y. L., "New fluorinating agents in organic synthesis," German L., Zemkov S. (eds.), Springer-Verlag, Berlin, 1989, pp. 1—34.
- 8) Schrobilgen G., Firnau G., Chirakal R., Garnett S., *J. Chem. Soc., Chem. Commun.*, **1981**, 198—199 (1981).
- 9) Chirakal R., Firnau G., Schrobilgen G. J., McKay J., Garnett E. S., *Int. J. Appl. Radiat. Isot.*, **35**, 401—404 (1984).
- 10) Patrick T. B., Johri K. K., White D. H., Bertrand W. S., Mokhtar R., Kilbourn M. R., Welch M. J., *Can. J. Chem.*, **64**, 138—141 (1986).
- 11) Aqueous solution of ^{18}F -fluoride was obtained by proton irradiation of 20% ^{18}O -enriched water (Enritech, Israel) using an MCY-1750 cyclotron (Shimadzu, Japan) and circulating water target system (NKK, Japan). To the ^{18}F -fluoride solution tetrabutylammonium hydroxide (5 μmol , Wako Pure Chemical, Japan) or tetrabutylammonium fluoride (5 μmol , Sigma-Aldrich, USA) was added to obtain NCA or CA ($n\text{-Bu}$) $_4\text{N}^{18}\text{F}$, respectively. The mixture was dried at 80 °C under a nitrogen gas stream and *in vacuo*. The residue was cooled to -20 °C and XeF_2 (12 μmol , Sigma-Aldrich, U.S.A.) and a substrate (20 μmol) were added. The reaction vessel was sealed and warmed to ambient temperature. The reaction mixture was analyzed by radio TLC. The TLC plate (Kieselgel 60 F_{254} , Merck, Germany) was cut into pieces and radioactivity was counted with a gamma counter (ARC-2000, Aloka, Japan) after development in diethylether:*n*-hexane (3:7).
- 12) a) Houston M. E., Honek J. F., *J. Chem. Soc., Chem. Commun.*, **1989**, 761—762 (1989); b) Sufrin J. R., Spiess A. J., Alks V., *J. Fluorine Chem.*, **49**, 177—182 (1990).
- 13) Franzen H. M., Ragnarsson U., Nagren K., Langstrom B., *J. Chem. Soc., Perkin Trans. 1*, **1987**, 2241—2247 (1987).