Vol. 54, No. 3

New Neolignans and Lignans from Vietnamese Medicinal Plant *Machilus odoratissima* Nees

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Four new natural neolignans and lignans, which were given the trivial names odoratisols A—D (1—4), together with (—)-licarin A, kachirachirol B, obovatifol, and machilin-I were isolated from the air-dried bark of the Vietnamese medicinal plant *Machilius odoratissima* Nees (Lauraceae). Their absolute structures were determined on the basis of spectroscopic analyses including circular dichroism spectra.

Key words Machilus odoratissima; Lauraceae; neolignan; lignan; odoratisol; absolute structure

Machilus is a genus in the Lauraceae family which includes twelve species distributed throughout Vietnam. They are Machilus odoratissima Nees, M. thunbergii Sieb. et Zucc., M. velutina Champ. ex. Benth., M. oreophila Hance, M. robutus J. J. Son., M. bonii Lec., M. coriacea A. Chev., M. thunbergii var. condorensis Lec., M. parviflora Meissn., M. platycarpa Chun., M. macrophyla Hemsley, and M. cochinchinensis Lec. 1—3) Machilus odoratissima Nees (Vietnamese name Khao nham) is a timber tree growing up to a height of 8—10 m. The tree bark is used in the folk medicine as antiseptic and anti-inflammatory remedies. The leaves are used to treat snake bite and burn wounds. 4) We carried out the first systematic study on M. odoratissima dealing with the isolation and structural elucidation of four new and four known neolignans and lignans.

The bark of M. odoratissima was air-dried in the shadow, powdered, and extracted with MeOH at room temperature. The MeOH extract was partitioned between H_2O and solvents of increasing polarities to afford n-hexane-, CH_2Cl_2 -, EtOAc-, and 1-BuOH-soluble fractions. Column-chromatographic separation of the n-hexane- and CH_2Cl_2 -soluble fractions led to the isolation of four new neolignans and lignans, named odoratisol A—D (1—4) and four known ones, (—)-licarin A, kachirachirol B, obovatifol, and machilin-I, which were determined by comparing their physical ($[\alpha]_D$) and spectroscopic data with the literature values. $^{5-9}$

Odoratisol A (1) was isolated as an oil and its molecular formula was deduced to be C21H24O5 from negative-ion highresolution (HR)-FAB-MS. The IR spectrum indicated the presence of hydroxyl groups (3450 cm⁻¹) and aromatic rings (1609, 1517, 1458 cm⁻¹). The ¹H- (Table 1) and ¹³C-NMR (Table 2) spectroscopic data indicated that 1 had a planar structure of 5-methoxydehydrodiisoeugenol. The trans geometry of 1'-propenyl group was determined on the basis of the large coupling constant between H-7' and H-8' $(J=15.6 \,\mathrm{Hz})$. The trans relationship of H-7 and H-8 was established based on ¹H-NMR spectroscopic data characteristic of trans-7-aryl-8-methyl-7,8-dihydro-benzofuranoid-type neolignans [$\delta_{\rm H}$ 5.0 (1H, d, J=9.5 Hz, H-7), 3.37 (1H, dq, J=9.5, 6.6 Hz, H-8), 1.32 (3H, d, J=6.6 Hz, H-9)]. ^{10,11)} The positions of the methoxyl groups at $\delta_{\rm H}$ 3.81 (3H, s) and 3.82 (6H, s) were assigned using nuclear Overhauser effect spectroscopy (NOESY) (Fig. 2). 5-Methoxydehydrodiisoeugenol was reported previously from Myristica fragrans, 12) however, its stereochemistries at C-7 and C-8 have not been determined at that time. In this study the circular dichroism (CD) spectrum was used to determine the absolute configuration of 1 to be the 7*S*,8*S* stereoisomer of 5-methoxydehydrodisoeugenol from a positive Cotton effect at 242 nm (+4.35) and a negative one at 269 nm (-5.93), which were similar to those exhibited by (-)-licarin A.⁶⁾ This configuration agreed well with the same sign of the optical rotations of 1 ($[\alpha]_D^{25}$ -35.1°) and (-)-licarin A ($[\alpha]_D^{25}$ -44.0°).⁶⁾ Thus 1 was isolated for the first time from Nature and its absolute stereostructure was concluded to be (7*S*,8*S*)-7,8-dihydro-7-(4-hy-

Fig. 1. Absolute Structures of Compounds 1—4, (-)-Licarin A, Verrucosin, Austrobailignan-7, and Futokadsurin B

March 2006 381

Table 1. ¹H-NMR Spectroscopic Data of 1—4 (δ in ppm, J in Hz in Parentheses, 400 MHz, CDCl₃)

Н	1	2	3	4
2	6.59 s	6.84 d (2.0)	6.97 d (1.7)	6.95 d (1.7)
5		6.78 d (8.5)	6.84 d (8.3)	6.83 d (8.3)
6	6.59 s		6.90 dd (8.3, 1.7)	6.87 dd (8.3, 1.7)
7	5.0 d (9.5)	4.75 d (3.2)	4.31 d (9.3)	4.27 d (9.3)
8	3.37 dq (9.5, 6.6)	4.27 dq (3.2, 6.4)	1.70 br dq (9.3, 6.6)	1.68 br dq (9.3, 6.6)
9	1.32 d (6.6)	1.09 d (6.4)	0.98 d (6.6)	0.95 d (6.6)
2'	6.72 s	6.91 d (2.0)	6.78 d (1.7)	6.70 d (1.7)
5′		6.88 d (8.3)	6.80 d (8.0)	6.71 d (8.0)
6'	6.69 s	6.84 dd (8.3, 2.0)	6.74 dd (8.0, 1.7)	6.79 dd (8.0, 1.7)
7′	6.29 d (15.6)	6.28 d (15.6)	5.03 d (8.8)	5.01 d (8.8)
8'	6.03 dq (15.6, 6.3)	6.08 dq (15.6, 6.6)	2.14 br dq (8.8, 6.8)	2.13 br dq (8.8, 7.1)
9′	1.79 d (6.3)	1.80 d (6.6)	0.58 d (6.8)	0.59 d (7.1)
MeO-3	3.82 s	3.819 s	3.82 s	3.82 s
MeO-5	3.82 s			
MeO-3'	3.81 s	3.822 s	3.77 s	
OCH ₂ O				5.85 s

All assignments were made on the basis of heteronuclear single quantum correlation (HSQC) and NOESY experiments

Table 2. 13 C-NMR Spectroscopic Data of 1—4 (δ in ppm, 100 MHz, CDCl₃)

C	1	2	3	4
1	133.2	133.8	133.2	132.5
2	103.6	109.0	109.5	109.3
3	147.1	146.5	146.6	146.55
4	134.9	144.9	145.2	145.3
5	147.1	113.9	114.2	114.3
6	103.6	119.9	119.3	119.5
7	94.0	82.4	87.4	82.8
8	45.7	73.7	47.8	47.8
9	17.6	13.4	14.9	15.0
1'	132.3	131.9	132.8	135.2
2'	109.5	109.5	109.8	120.2
3′	144.2	145.6	146.2	147.4
4'	146.6	151.6	144.6	146.59
5′	136.4	119.1	113.9	107.7
6′	113.3	118.8	119.9	107.5
7′	131.0	130.5	83.1	87.5
8'	123.5	125.0	46.0	45.9
9′	18.3	18.5	15.0	14.9
MeO-3	56.4	55.8	55.86	55.9
MeO-5	56.4			
MeO-3'	56.0	55.9	55.9	
OCH ₂ O				100.9

droxy-3,5-dimethoxyphenyl)-3'-methoxy-8-methyl-1'-trans-propenylbenzofuran.

Odoratisol B (2) was isolated as an oil and had the molecular formula $C_{20}H_{24}O_5$ based on negative-ion HR-FAB-MS. The IR spectrum indicated the presence of hydroxyl groups (3448 cm⁻¹) and aromatic rings (1603, 1511, 1458 cm⁻¹). The ¹H- (Table 1) and ¹³C-NMR (Table 2) spectroscopic data of **2** were superimposable with those of machilin $C_1^{(13)}$ including the *erythro* stereochemistry between H-7 and H-8 as shown by a small coupling constant (J=3.2 Hz) between them. However, the optical rotation of **2** ($[\alpha]_D^{25} + 18.6^\circ$) was almost of the same in value but reverse in sign in comparison with that of machilin $C_1^{(13)} = 16.5^\circ$ showing the need to determine the absolute stereochemistry at two stereogenic elements at C-7 and C-8. The CD spectrum of **2** showed positive Cotton effect at 258 nm (+0.38) established the configu-

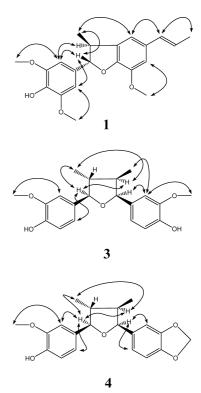


Fig. 2. NOESY Correlations of Compounds 1, 3, and 4

rations at C-7 an C-8 are 7*S* and 8*R* as in the cases of analogous neolignans of *erythro* series. ¹⁴⁾ On the basis of these data, the structure of **2** was concluded to be 7*S*,8*R*-*erythro*-4-hydroxy-3,2'-dimethoxy-4'-*trans*-propenyl-neolignan.

Odoratisol C (3) was isolated as an oil, $[\alpha]_D^{25} - 26.0^\circ$, and its molecular formula was characterized to be $C_{20}H_{24}O_5$ in negative-ion HR-FAB-MS. The IR spectrum indicated the presence of hydroxyl groups (3450 cm⁻¹) and aromatic rings (1607, 1514, 1457 cm⁻¹). The ¹H- (Table 1) and ¹³C-NMR (Table 2) spectroscopic data showed the structural resemblance of 3 and verrucosin, ¹⁵⁾ which, however, displayed a positive optical rotation ($[\alpha]_D + 14.8^\circ$). The *trans* H-7/H-8, *trans* H-8/H-8', and *cis* H-7'/H-8' relative stereochemistries of the tetrahydrofuran ring were determined by comparing

382 Vol. 54, No. 3

the ¹H-NMR data [$\delta_{\rm H}$ 0.98 (3H, d, J=6.6 Hz, H-9), 1.70 (1H, br dq, J=9.3, 6.6 Hz, H-8), 4.31 (1H, d, J=9.3 Hz, H-7); 0.58 (3H, d, J=6.8 Hz, H-9'), 2.14 (1H, br dq, J=8.8, 6.8Hz, H-8'), 5.03 (1H, d, J=8.8 Hz, H-7')] with those reported in literature for 7,8-trans-8,8'-trans-7',8'-cis-configurated tetrahydrofuran-type lignans. 15—17) The trans H-7/H-8 and cis H-7'/H-8' configurations were in agreement with the upfield shift ($\Delta \delta_{\rm H} - 0.4$) of methyl proton signals on going from C-9 to C-9' due to the anisotropic effect of the aromatic group in the case of the cis-configuration of the aryl group at C-7' and methyl substituent at C-8'. The stereochemical assignments were supported by the NOESY spectrum (Fig. 2) of 3, which showed NOEs between H₃-9 ($\delta_{\rm H}$ 0.98) and H-7 ($\delta_{\rm H}$ 4.31), between H_3 -9 and H-8' (δ_H 2.14), and between H-7 and H-8', between H_3 -9' (δ_H 0.58) and H-2' (δ_H 6.78). The locations of two 4-hydroxy-3-methoxyphenyl moieties were also confirmed by NOEs between H-6 [$\delta_{\rm H}$ 6.90 (dd, J=8.3, 1.7 Hz)] and H-7 and between H-2' [$\delta_{\rm H}$ 6.78 (d, J=1.7 Hz)] and H-7'. Since 3 had the same relative stereochemistry but opposite optical rotation in comparison with those of verrucosin, the absolute configurations at the C-7, C-8, C-7', and C-8' were deduced to be opposite to those of verrucosin. Thus the absolute structure of 3 was determined to be (7R,8R,7'S,8'R)-4-hydroxy-3-methoxy-4'-hydroxy-3'methoxy-7,7'-epoxylignan.

Odoratisol D (4) was isolated as an oil and its molecular formula was determined C₂₀H₂₂O₅ by means of negative-ion HR-FAB-MS. The IR spectrum indicated the presence of hydroxyl groups (3450 cm⁻¹) and aromatic rings (1608, 1517, 1442 cm⁻¹). The ¹H-NMR (Table 1) spectrum showed the presence of a 4-hydroxy-3-methoxyphenyl and a monosubstituted 3,4-methylenedioxyphenyl systems, which contained an aromatic methoxyl group [$\delta_{\rm H}$ 3.82 (3H, s)] and a methylenedioxy group [δ_H 5.85 (2H, s)], and a 2,5-disubstituted 3,4-dimethyltetrahydrofuran ring [$\delta_{\rm H}$ 0.95 (3H, d, J=6.6 Hz, H-9), 1.68 (1H, br dq, J=9.3, 6.6 Hz, H-8), 4.27 (1H, d, J=9.3 Hz, H-7); 0.59 (3H, d, J=7.1 Hz, H-9'), 2.13 (1H, br dq, J=8.8, 7.1 Hz, H-8'), 5.01 (1H, d, J=8.8 Hz, H-7')]. The ¹H- and ¹³C-NMR (Table 2) spectroscopic data of 4 resembled those of futokadsurin B¹⁸⁾ except for the lack of an additional methoxyl group. The trans H-7/H-8, trans H-8/H-8', and cis H-7'/H-8' relative stereochemistry of the tetrahydrofuran ring were conclusive on the basis of the comparison of the ¹H-NMR data with those reported in literature. ^{16—19)} Upfield shift ($\Delta\delta_{\rm H}$ -0.36) of methyl proton signals on going from C-9 to C-9' agreed with trans H-7/H-8 and cis H-7'/H-8' configurations. NOEs observed between H₃-9 ($\delta_{\rm H}$ 0.95) and H-7 $(\delta_{\rm H}$ 4.27), between H₃-9 and H-8' $(\delta_{\rm H}$ 2.13), and between H-7 and H-8', but not between H₃-9' ($\delta_{\rm H}$ 0.59) and H-7' ($\delta_{\rm H}$ 5.01) supported the stereochemical assignments.¹⁷⁾ Thus 4 was concluded to have the same relative stereochemistry as that of 3. NOESY spectrum (Fig. 2) of 4 showed the correlations between the methoxyl group ($\delta_{\rm H}$ 3.82) and H-2, between H-2 ($\delta_{\rm H}$ 6.95) and H-7 ($\delta_{\rm H}$ 4.27), between H-6 ($\delta_{\rm H}$ 6.87) and H-7, between H-2' ($\delta_{\rm H}$ 6.70) and H-7' ($\delta_{\rm H}$ 5.01), and between H-6' ($\delta_{\rm H}$ 6.79) and H-7' confirmed the assignments of the position of the 4-hydroxy-3-methoxyphenyl moiety at C-7 and the 3',4'-methylenedioxyphenyl moiety at C-7' as shown in Fig. 2. To establish the absolute structure of 4 the CD spectra of 3 and 4 were measured and compared. Similar CD curves of 3 and 4 were seen, namely, 4 displayed

the Cotton effects at 213 nm (-1.25), 240 nm (+0.95), and 285 nm (-0.40) assuring the absolute structure of 4 to be (7R,8R,7'S,8'R)-4-hydroxy-3-methoxy-3',4'-methylenedioxy-7,7'-epoxylignan.

Experimental

General Procedure Optical rotations were measured on a JASCO P-1030 polarimeter. FT-IR spectra were recorded on a Horiba FT-710 spectrophotometer. $^{1}\text{H-}$ (400 MHz) and $^{13}\text{C-NMR}$ (100 MHz) spectra were recorded using a JEOL JNM-α 400 NMR spectrometer with tetramethylsilane as an internal standard. Negative-ion HR-FAB-MS were measured on a JEOL SX-102 mass spectrometer with PEG-400 as a calibration matrix. HPLC was carried out with a JASCO PU-1580 pump and an UV-2075 Plus detector (set at 210 nm) on YMC ODS columns (150×4.6 mm i.d. in analytical and 150×20 mm i.d. in preparative scales) at the corresponding flow rates of 0.5 and 5 ml/min. Silica (Si) gel 60 (0.063—0.200 mm, Merck, Germany) and reversed-phase octadecyl Si (ODS) gel (YMC, Japan) were used for open-column chromatography. TLC was carried out on Merck TLC plates (Si gel 60 F254), and detected by spraying with 10% H2SO4 in 50% EtOH, followed by heating on a hot plate at 200 °C.

Plant Material The air-dried bark (2.0 kg) of *M. odoratissima* was collected in Province Thai Nguyen, Vietnam, and identified by Dr Nguyen Hoanh Coi of the Military Institute of Drug Controls (Hanoi, Vietnam), in June 2000. A voucher specimen (no. HCTN 2000-6) is deposited in the Laboratory of Chemistry of Natural Products, Faculty of Chemistry, Vietnam National University, Hanoi, Vietnam.

Extraction and Isolation of 1—10 The powdered air-dried bark of *M. odoratissima* (2.0 kg) was extracted with MeOH by percolation at room temperature (3 times, for 3 d each). After concentration by evaporation under reduced pressure, the resultant MeOH extract was suspended in H₂O and sequentially extracted with *n*-hexane, CH₂Cl₂, EtOAc, and 1-BuOH. The *n*-hexane-soluble fraction (5.9 g) was separated on a Si gel open column using mixtures of *n*-hexane in EtOAc (10:1, 4:1, 2:1, 1:1). Five pooled fractions were collected on the basis of TLC pattenrs. Fraction 2 (1.8 g), fraction 3 (0.5 g), and fraction 4 (0.4 g) underwent the same treatment, first separation on an ODS gel open column (MeOH—H₂O, 3:1, 4:1), then purification on ODS preparative HPLC (MeOH—H₂O, 3:1) to give odoratisol A (1, 12.2 mg), odoratisol B (2, 7.0 mg), odoratisol C (3, 27.9 mg), odoratisol D (4, 40.2 mg), (-)-licarin A (0.48 g), machilin-I (13.4 mg), kachirachirol B (16.1 mg), and obovatifol (17.8 mg). Similar procedure was used to separate the CH₂Cl₂-soluble fraction yielding 1 (7.9 mg), (-)-licarin A (55.8 mg), and kachirachirol B (56.4 mg).

and kachirachirol B (56.4 mg). Odoratisol A (1): Oil, $[\alpha]_{2}^{25}$ – 35.1° (c=1.22, CHCl₃). UV λ_{max} (MeOH) nm ($\log \varepsilon$): 270 (4.18), 219 (4.43). IR ν_{max} (film) cm⁻¹: 3450, 2957, 2926, 2854, 1609, 1517, 1493, 1458, 1375, 1078. CD (MeOH): $\Delta \varepsilon$ (nm): –2.14 (218), +4.35 (242), –5.93 (269) (c=2.0×10⁻⁵ M). ¹H- and ¹³C-NMR: see Tables 1 and 2. Negative-ion HR-FAB-MS: m/z 355.1544 [M–H]⁻ (Calcd for $C_{21}H_{23}O_5$: 355.1545).

Odoratisol B (2): Oil, $[\alpha]_D^{25}$ +18.6° (c=0.70, CHCl₃). UV $\lambda_{\rm max}$ (MeOH) nm (log ε): 258 (3.68), 218 (3.87). IR $\nu_{\rm max}$ (film) cm⁻¹: 3448, 2957, 2925, 2854, 1603, 1511, 1458, 1377, 1061. CD (MeOH): $\Delta\varepsilon$ (nm): -0.74 (219), +0.38 (258), +0.18 (350) (c=4.2×10⁻⁵ M). ¹H- and ¹³C-NMR: see Tables 1 and 2. Negative-ion HR-FAB-MS: m/z 343.1523 [M-H]⁻ (Calcd for $C_{20}H_{23}O_5$: 343.1545).

Odoratisol C (3): Oil, $[\alpha]_D^{25} - 26.0^{\circ}$ (c=2.79, CHCl₃). UV λ_{max} (MeOH) nm ($\log \varepsilon$): 280 (3.71), 232 (4.05). IR v_{max} (film) cm⁻¹: 3450, 2958, 2926, 2854, 1607, 1514, 1457, 1377, 1033. CD (MeOH): $\Delta \varepsilon$ (nm): -4.37 (211), +0.31 (258), -0.38 (281) (c=4.3×10⁻⁵ M). ¹H- and ¹³C-NMR: see Tables 1 and 2. Negative-ion HR-FAB-MS: m/z 343.1542 [M-H]⁻ (Calcd for $C_{20}H_{23}O_5$: 343.1545).

Odoratisol D (4): Oil, $[\alpha]_{\rm D}^{25}$ –12.8° (c=4.0, CHCl₃). UV $\lambda_{\rm max}$ (MeOH) nm (log ε): 282 (3.80), 234 (3.97). IR $\nu_{\rm max}$ (film) cm⁻¹: 3450, 2959, 2927, 2873, 1608, 1517, 1488, 1442, 1377, 1036. CD (MeOH): $\Delta\varepsilon$ (nm): –1.25 (213), +0.95 (240), -0.40 (285) (c=8.8×10⁻⁵ m). ¹H- and ¹³C-NMR: see Tables 1 and 2. Negative-ion HR-FAB-MS: m/z 341.1401 [M-H]⁻ (Calcd for C₂₀H₂₁O₅: 341.1389).

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March 2006

urements on its 400 MHz NMR instrument.

Reference

- 1) Pham H. H., "Illustrated Flora of Vietnam," Tom 1, Fascile 1, Published by the Author, Montreal, 1991, pp. 488—492.
- Le K. K., "Common Flora of Vietnam," Vol. 2, Science and Technique, Hanoi, 1971, pp. 265—266.
- Do T. L., "Medicinal Plants and Remedies of Vietnam," Science and Technique, Hanoi, 1995, pp. 666—668.
- 4) Nguyen D. M., "Anti-infective Drugs from Plants of Vietnam," Medicine, Ho Chi Minh city, 1995, pp. 105—109.
- Aiba C. J., Campos Corrêa R. G., Gottlieb O. R., *Phytochemistry*, 12, 1163—1164 (1973).
- Nascimento I. R., Lopes L. M. X., Davin L. B., Lewis N. G., *Tetrahedron*, 56, 9181—9193 (2000).
- Ito K., Ichino K., Iida J., Lai J., Phytochemistry, 23, 2643—2645 (1984).
- 8) Tsai I. L., Hsieh C. F., Duh C. Y., *Phytochemistry*, **48**, 1371—1375 (1998)
- 9) Shimomura H., Sashida Y., Oohara M., Phytochemistry, 27, 634—636

(1988).

Benevides P. J. C., Sartorelli P., Kato M. J., *Phytochemistry*, **52**, 339—343 (1999).

383

- Nascimento I. R., Lopes L. M. X., Phytochemistry, 52, 345—350 (1999).
- Forrest J. E., Heacock R. A., Forrest T. P., J. Chem. Soc. Perkin Trans. I, 1974, 205—209 (1974).
- Shimomura H., Sashida Y., Oohara M., *Phytochemistry*, 26, 1513— 1515 (1987).
- 14) Matsuda N., Kikuchi M., Chem. Pharm. Bull., 44, 1676—1679 (1996).
- Hattori M., Hada S., Kawata Y., Tezuka Y., Kikuchi T., Namba T., *Chem. Pharm. Bull.*, 35, 3315—3322 (1987).
- Kraft C., Jenett-Siems K., Köhler I., Tofern-Reblin B., Siems K., Bienzle U., Eich E., *Phytochemistry*, 60, 167—173 (2002).
- Urzúa A., Freyer A. J., Shamma M., Phytochemistry, 26, 1509—1511 (1987).
- Konishi T., Konoshima T., Daikonya A., Kitanaka S., Chem. Pharm. Bull., 53, 121—124 (2005).
- Hada S., Hattori M., Tezuka Y., Kikuchi T., Namba T., *Phytochemistry*, 27, 563—568 (1988).