Lipase-Catalyzed Asymmetric Synthesis of Desprenyl-carquinostatin A and Descycloavandulyl-lavanduquinocin

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An asymmetric synthesis of the core carbazole structure, 6-desprenyl-carquinostatin 3 and 6-descycloavandulyl-lavanduquinocin 3, toward a total synthesis of carquinostatin A (1) and lavanduquinocin (2), has been established. Lipase QLM (Meito) catalyzed enantioselective acetylation of the racemic alcohol 6 gave the (-)-acetate 7 and the (+)-alcohol 6 with high enantioselectivity. The absolute stereochemistry of the (-)- and (+)-alcohol 6 have been determined to be R- and S-configurations, respectively, by the advanced Mosher method. In the same manner, the (-)-acetate 13 and the (+)-alcohol 12 have been obtained from the racemic alcohol 12. The (R)-(-)-acetate 13, derived from the (R)-(-)-acetate 7, was the same as the (-)-acetate 13, which has been determined to be (R)-configuration. Oxidation of the (R)-(-)-acetate 13 followed by hydrolysis afforded (R)-(-)-6-desprenyl-carquinostatin [and (R)-(-)-6-descycloavandulyl-lavanduquinocin] 3. In addition, oxidation of the (S)-(+)-alcohol 12 provided (S)-(+)-3, which is the enantiomer of 6-desprenyl-carquinostatin A (R)-(-)-3.

Key words lipase QLM; enantioselective acetylation; advanced Mosher method; desprenyl carquinostatin A; descycloavandulyl lavanduquinocin

The carbazole-3,4-quinone alkaloids, carquinostatin A $(1)^{1)}$ and lavanduquinocin $(2)^{2)}$ were isolated from *Streptomyces exfoliates* 2419-SVT2 and *Streptomyces viridochromogenes* by Seto and co-workers in 1993 and 1995, respectively. The structures of the two alkaloids were elucidated to be the same carbazole-3,4-quinone moiety by NMR spectral analyses and other spectroscopic experiments. The absolute stereochemistry of the C-11 position of the two alkaloids was the same *R*-configuration. Carquinostatin A (1) and lavanduquinocin (2) were also shown to be a potent neuronal cell protecting substance which exhibits a free radical scavenging activity. Total syntheses of these alkaloids have recently been developed by the Knölker group.³⁻⁸⁾ The transition metalmediated and -catalyzed methodologies for the construction of the carbazole framework have been efficiently employed.

Throughout the course of this study, we have been interested in the synthetic development of biologically active condensed-heteroaromatic compounds, including natural products, by the thermal electrocyclic reactions of either conjugated hexatriene⁹⁻¹⁷⁾ or monoazahexatriene¹⁸⁾ systems incorporating one double bond from an aromatic or heteroaro-

matic portion. We recently reported the synthesis of the highly substituted carbazole alkaloids, carazostatin⁹⁾ and carbazoquinocins, ^{9,10)} by the construction of the appropriate carbazole framework based on the allene-mediated electrocyclic reaction of the 6π -electron system involving the indole 2,3-bond. In the present paper, we describe the asymmetric synthesis of 6-desprenyl-carquinostatin A (6-descycloavandulyl-lavanduquinocin) 3, which is a common carbazole framework of both alkaloids, based on a lipase-catalyzed esterification using a racemic alcohol 6 for the determination of the absolute stereochemistry of 3. We chose the 3-ethoxy-2-methyl-1-(trifluoromethylsulfonyloxy)carbazole (4), ^{9,10)} as a starting material, which was prepared in a six-step sequence from 3-iodoindole-2-carbaldehyde by the application of our methodology, as shown in the retrosynthetic Chart 1.

The required 1-allylcarbazole **5** was prepared from the triflate **4** and allyboronic acid pinacol ester in the presence of PdCl₂(dppf) in dimethylformamide (DMF) by the Suzuki– Miyaura reaction. ^{19,20)} Subsequent the Wacker reaction²¹⁾ of **5** in the presense of palladium chloride(II) and copper chloride under an oxygen atmosphere gave the acetonylcarbazole **8**

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followed by reduction of **8** with NaBH₄ provided the racemic alcohol **6** (Chart 2). Next, we investigated the enzymatic resolution of **6** with lipase PS (Amano) or lipase QLM (Meito). The results of the kinetic resolution by transesterification are shown in Table 1. Of the lipases, lipase QLM showed high enantioselectivities for this substrate. The influence of a solvent in the enantioselectivity was also examined, and disopropyl ether (i-Pr₂O) gave the best result. Enantiomeric excess of the (-)-acetate **7** and the (+)-alcohol **6** were measured by HPLC on a CHIRALPAC AD column.

The absolute configuration of these compounds 6 was ex-

Table 1. Kinetic Resolution of (±)-6 by Lipase-Catalyzed Acetylation

Chart 2

amined by the advanced Mosher method^{22,23)} using 2methoxy-2-(1-naphthyl)propionic acid (M α NP acid). The (-)-acetate 7 was hydrolyzed with aqueous 1 M K₂CO₃/ MeOH to give the (-)-alcohol 6. Both alcohols ((-)-6 and (+)-6) were treated with (R)- and (S)-M α NP acid in the presence of dicyclohexylcarbodiimide (DCC) and 4-dimethylaminopyridine (DMAP) in CH₂Cl₂ to produce the four $M\alpha NP$ esters (9a, b and 10a, b). The absolute stereochemistry can be elucidated by the difference of calculated chemical shift on their NMR spectra. In the M α NP ester 9a, b $((-)-6+M\alpha NP \text{ acid})$, the 10-H resonated at upper field in the (R)-ester **9a** than in the (S)-ester **9b** ($\Delta \delta$, -0.32 ppm and -0.04 ppm. $\Delta \delta = \delta_R - \delta_S$). Moreover, 12-H appeared upfield for the (S)-ester **9b** than for the (R)-ester **9a** ($\Delta \delta$, +0.13 ppm). The other M α NP ester **10a**, **b** ((+)-**6**+M α NP acid) was completely a result of the contrariety of ester 9a, b. Based on the analyses of their NMR spectra, the absolute stereochemistry of (+)- and (-)-alcohol 6 were determined to be S- and R-configurations, respectively (Chart 3).

Cleavage of the ethyl ether of **8** with boron tribromide (BBr₃) afforded the 3-hydroxycarbazole **11**, which was reduced with NaBH₄ to provide the racemic alcohol **12**. Subsequent lipase-catalyzed enantioselective esterification of **12** with lipase QLM and vinyl acetate in *i*-Pr₂O gave the (-)-acetate **13** (98% ee) and (+)-alcohol **12** (97% ee) (Chart 4). The spectral data and the retention time by HPLC of (-)-acetate **13** were identical with those of the (R)-(-)-acetate **13** prepared by treatment of (R)-(-)-7 with BBr₃ in all respects. Consequently, the absolute stereochemistry of (-)-13 and (+)-12 was determined to be R- and S-configuration, respec-

Run	Lipase	Solvent	Time (h)	Acetate 7 % (% e		cohol (% ee)
1	Lipase PS	CH ₂ Cl ₂	24		98	_
2	Lipase PS	THF	24		96	_
3	Lipase PS	t-BuOMe	48	19 (98) 75	(8)
4	Lipase PS	Et ₂ O	48	12 (10	71	(9)
5	Lipase PS	CH ₃ CN	48	6 (13) 56	_
6	Lipase PS	i-Pr ₂ O	48	4 (97	90	_
7	Lipase QLM	t-BuOMe	24	40 (99	54	(70)
8	Lipase QLM	i-Pr ₂ O	24	49 (99) 40	(78)

The enantiomeric excess (%ee) was determined by HPLC on CHIRALPAC AD.

(-)-7
$$\frac{1 \text{M K}_2 \text{CO}_3}{\text{MeOH}}$$
 $\frac{(F)-\text{M}\alpha \text{NP acid}}{\text{and/or}}$ $\frac{(S)-\text{M}\alpha \text{NP acid}}{\text{DCC}, \text{DMAP}}$ $\frac{(S)-\text{M}\alpha \text{NP acid}}{\text{DCC}, \text{DMAP}}$ $\frac{(S)-\text{M}\alpha \text{NP acid}}{\text{DCC}, \text{DMAP}}$ $\frac{\text{Me}}{\text{Me}}$ $\frac{(S)-\text{M}\alpha \text{NP acid}}{\text{DCC}, \text{DMAP}}$ $\frac{\text{Me}}{\text{Me}}$ $\frac{\text{Me}}{\text{O}}$ $\frac{\text{Me}}{$

Chart 3

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tively. Finally, oxidative treatment of the (-)-acetate 13 with benzeneseleninic anhydride¹⁰ [(PhSeO)₂O] gave the carbazole-3,4-quinone (R)-(-)-14, which was hydrolyzed with aqueous 1 M K₂CO₃/MeOH to yield the desprenyl-carquinostatin A (descycloavandulyl-lavanduquinocin) (R)-(-)-3. In the same way, treatment of the (S)-(+)-alcohol 12 with (PhSeO)₂O also gave its enantiomer (S)-(+)-3 (Chart 5).

In conclusion, a synthesis of the functionalized core carbazole, (R)-(-)-6-desprenyl-carquinostatin A [(R)-(-)-6-descycloavandulyl-lavanduquinocin] **3** together with its enantiomer (S)-(+)-**3**, have been completed by the lipase-catalyzed enantioselective esterification, followed by the determination of the absolute stereochemistry based on the advanced Mosher method, by using the key compound, 3-ethoxy-2-methyl-1-(trifluoromethylsulfonyloxy)carbazole (**4**). In addition, it has been demonstrated that a new synthetic route toward a total synthesis of carquinostatin A (**1**) and lavanduquinocin (**2**) has been provided.

Experimental

All melting points were measured on a Yanagimoto micro-melting point apparatus MP-500D and were uncorrected. IR spectra were recorded on a Shimadzu FT-IR-8500 spectrophotometer using attenuated total reflection (ATR) method. ¹H- and ¹³C-NMR spectra were taken with a JEOL AL-300 instrument using tetramethylsilane as an internal standard. Mass spectra (MS) were determined by a Shimadzu QP5050 and/or Shimadzu 9020DF spectrometers. All air sensitive reactions were run under an argon atmosphere. Solvents were distilled by normal methods (THF was dried over sodium benzophenone ketyl, CH₂Cl₂ was dried over P₂O₅, DMF was dried over CaH₂). Silica gel 60PF₂₅₄ (60—100 mesh, Merck Art 7744) was used

for column chromatography.

1-Allyl-3-ethoxy-2-methylcarbazole (5) 2-Allyl-4,4,5,5-tetramethyl-1,3,2-dioxaboralane (187 μ l, 0.99 mmol) was added to a mixture of the triflate 4 (246 mg, 0.66 mmol), 3 M NaOH (0.64 ml, 1.98 mmol) and PdCl₂(dppf) (49 mg, 0.06 mmol) in THF (10 ml) under Ar atmosphere. The stirred mixture was heated at 85 °C for 2 h. After being cooled to r.t., the mixture was quenched with water, and was then extracted with EtOAc. The EtOAc layer was washed with brine, dried over Na2SO4 and concentrated. The residue was purified by column chromatography (silica gel, 10 g) using EtOAc-hexane (3:97, v/v) as an eluent to give the allylcarbazole 5 (134 mg, 77%): mp 83—85 °C (from hexane). 1 H-NMR (CDCl₃) δ : 1.50 (3H, t, $J=6.9\,\mathrm{Hz}$), 2.36 (3H, s), 3.69 (2H, dt, J=1.8, 1.8, 5.4 Hz), 4.14 (2H, q, J=6.9 Hz), 5.05 (1H, dq, J=1.8, 1.8, 17.2 Hz), 5.09 (1H, dq, J=1.8, 1.8, 10.2 Hz), 6.03 (1H, ddd, J=5.4, 10.2, 17.2 Hz), 7.18 (1H, dt, J=1.1, 8.1 Hz), 7.34 (1H, dt, J=1.1, 8.1 Hz), 7.41 (1H, d, J=8.1 Hz), 7.42 (1H, s), 7.84 (1H, br s), 7.98 (1H, d, J=8.1 Hz). MS m/z: 265 (M⁺). HR-MS m/z: 265.1471 (Calcd for C₁₈H₁₉NO: 265.1467).

1-Acetonyl-3-ethoxy-2-methylcarbazole (8) A mixture of PdCl₂ (17 mg, 0.094 mmol) and CuCl (93 mg, 0.94 mmol) in DMF–H₂O (7:1, 4 ml) was stirred at r.t. for 30 min under an O₂ atmosphere. A solution of allylcarbazole **5** (249 mg, 0.94 mmol) in DMF–H₂O (7:1, 6 ml) was added to a reaction mixture at the same temperature, and was then stirred at the same temperature for 4 h under an O₂ atmosphere. The reactant was quenched with 10% HCl, and was then extracted with EtOAc. The EtOAc layer was washed with water and brine, dried over Na₂SO₄ and concentrated. The residue was purified by column chromatography (silica gel, 10 g) using EtOAc–hexane (3:7, v/v) as an eluent to give the acetonylcarbazole **8** (215 mg, 81%): mp 169—170 °C (from pet. Et₂O). IR (ATR) v: 1708 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.51 (3H, t, J=7.0Hz), 2.17 (3H, s), 2.48 (3H, s), 4.01 (2H, s), 4.16 (2H, q, J=7.0Hz), 7.18 (1H, t, J=6.9Hz), 7.37 (1H, t, J=6.9Hz), 7.45 (1H, d, J=6.9Hz), 7.46 (1H, s), 7.97 (1H, d, J=6.9Hz), 8.34 (1H, brs). MS m/z: 281 (M⁺). HR-MS m/z: 281.1425 (Calcd for $C_{18}H_{19}NO_2$: 281.1416).

3-Ethoxy-1-(2-hydroxypropyl)-2-methylcarbazole (6) NaBH₄ (64 mg, 1.70 mmol) was added to a solution of acetonylcarbazole **8** (400 mg, 1.42 mmol) in EtOH (50 ml) under ice-cooled water, and was then stirred at the same temperature for 1 h. The reaction mixture was quenched with water, and was then extracted with EtOAc. The EtOAc layer was washed with brine, dried over Na₂SO₄ and concentrated. The residue was purified by column chromatography (silica gel, 10 g) using EtOAc-hexane (3: 7, v/v) as an eluent to give the alcohol **6** (375 mg, 93%): mp 141—142 °C (from Et₂O-pet. Et₂O); IR (ATR) v: 3374 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.38 (3H, d, J=6.2 Hz), 1.50 (3H, t, J=6.9 Hz), 2.36 (3H, s), 3.03 (1H, dd, J=8.1, 14.3 Hz), 3.15 (1H, dd, J=3.3, 14.3 Hz), 4.15 (2H, q, J=6.9 Hz), 4.20—4.25 (1H, m), 7.15 (1H, dt, J=1.1, 8.1 Hz), 7.35 (1H, dt, J=1.1, 8.1 Hz), 7.42 (1H, d, J=8.0 Hz), 7.43 (1H, s), 7.97 (1H, d, J=8.0 Hz), 8.42 (1H, br s). MS m/z: 283 (M⁺). HR-MS m/z: 283.1579 (Calcd for C₁₈H₂₁NO₂: 283.1572).

(-)-1-(2-Acetoxypropyl)-3-ethoxy-2-methylcarbazole (-)-(7) and (+)-3-Ethoxy-1-(2-hydroxypropyl)-2-methylcarbazole (+)-(6) iPr₂O (10 ml), vinyl acetate (98 μ l, 1.06 mmol), and the alcohol 6 (30 mg, 0.11 mmol) were added to the lipase QLM (60 mg). The mixture was stirred at 32 °C for 24 h. The reaction mixture was filtered and the volatiles were removed under reduced pressure to give the products. The crude product was purified by preparative TLC using EtOAc–hexane (3:7, v/v) as an eluent to give the acetate (-)-7 (17 mg, 49%) and an alcohol (+)-6 (12 mg, 40%).

(-)-7: The optical purity was 99% ee by HPLC (CHIRALPAC AD; 10% iPrOH-hexane). $[\alpha]_D^{23}$ -99.6° (c=0.11, CHCl₃). IR (ATR) v: 1716 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.29 (3H, d, J=6.2 Hz), 1.51 (3H, t, J=6.9 Hz), 2.17

$$(R) - (-) - 13 \qquad \underbrace{(PhSeO)_2O}_{THF} \qquad \underbrace{(PhSeO)_2O}_{(R) - (-) - 14} \qquad \underbrace{(PhSeO)_2O}_{N} \qquad \underbrace{(R) - (-) - 14}_{N} \qquad \underbrace{(PhSeO)_2O}_{N} \qquad \underbrace{(PhSeO$$

Chart 5

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(3H, s), 2.38 (3H, s), 3.11 (1H, dd, J=9.9, 13.2 Hz), 3.32 (1H, dd, J=2.9, 13.2 Hz), 4.14 (2H, q, J=6.9 Hz), 5.05—5.13 (1H, m), 7.16 (1H, dt, J=1.1, 8.0 Hz), 7.36 (1H, dt, J=1.1, 8.0 Hz), 7.42 (1H, s), 7.50 (1H, d, J=8.1 Hz), 7.97 (1H, d, J=8.1 Hz), 8.42 (1H, br s). MS m/z: 325 (M $^+$). HR-MS m/z: 325.1691 (Calcd for $C_{20}H_{23}NO_3$: 325.1678).

(+)-6: The optical purity was 78% ee by HPLC (CHIRALPAC AD; 10% iPrOH-hexane). $[\alpha]_0^{23} + 10.0^{\circ}$ (c=0.12, CHCl₃). mp 146—147 °C (CHCl₃-hexane). IR (ATR) v: 3370 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.37 (3H, d, J=6.2 Hz), 1.50 (3H, t, J=6.9 Hz), 2.36 (3H, s), 3.03 (1H, dd, J=8.1, 14.3 Hz), 3.14 (1H, dd, J=3.3, 14.3 Hz), 4.15 (2H, q, J=6.9 Hz), 4.20—4.25 (1H, m), 7.15 (1H, dt, J=1.1, 8.1 Hz), 7.35 (1H, dt, J=1.1, 8.1 Hz), 7.42 (1H, d, J=8.0 Hz), 7.43 (1H, s), 7.97 (1H, d, J=8.0 Hz), 8.42 (1H, br s). MS m/z: 283 (M⁺). HR-MS m/z: 283.1580 (Calcd for C₁₈H₂₁NO₂: 283.1572).

(-)-3-Ethoxy-1-(2-hydroxypropyl)-2-methylcarbazole (-)-(6) An acetate (-)-7 (30 mg, 0.092 mmol) was hydrolyzed with aqueous 1 m $\rm K_2CO_3$ (1 ml) and MeOH (3 ml) at r.t. for 1.5 h to give an (-)-alcohol. The reaction mixture was quenched with water, and was then extracted with EtOAc. The EtOAc layer was washed with brine, dried over $\rm Na_2SO_4$ and concentrated. The residue was purified by column chromatography (silica gel, 10 g) using EtOAc–hexane (3:7, v/v) as an eluent to give the alcohol (-)-6 (25 mg, 96%): mp 143—144 °C (from CHCl₃—hexane). $[\alpha]_D^{23}$ -22.9° (c=0.120, CHCl₃). IR (ATR) v: 3370 cm⁻¹. 1 H-NMR (CDCl₃) δ : 1.38 (3H, d, J=6.2 Hz), 1.50 (3H, t, J=6.9 Hz), 2.36 (3H, s), 3.03 (1H, dd, J=8.1, 14.3 Hz), 3.14 (1H, dd, J=3.3, 14.3 Hz), 4.15 (2H, q, J=6.9 Hz), 4.20—4.25 (1H, m), 7.15 (1H, dt, J=1.1, 8.1 Hz), 7.35 (1H, dt, J=1.1, 8.1 Hz), 7.42 (1H, d, J=8.0 Hz), 7.43 (1H, s), 7.97 (1H, d, J=8.0 Hz), 8.42 (1H, br s). MS m/z: 283 (M⁺). HR-MS m/z: 283.1576 (Calcd for $C_{18}H_{21}\rm{NO}_2$: 283.1572).

Synthesis of the Ester 9a from (–)-6 and (R)-MαNP Acid A mixture of the alcohol (–)-6 (5 mg, 17.6 μmol), (R)-MαNP acid (4.9 mg, 21.2 μmol), DCC (6.2 mg, 29.9 μmol), and DMAP (1.1 mg, 8.8 μmol) in CH₂Cl₂ (1 ml) was stirred at r.t. for 24 h under a N₂ atmosphere. The reaction mixture was quenched with water, and was then extracted with CH₂Cl₂. The CH₂Cl₂ layer was washed with brine, dried over Na₂SO₄ and concentrated. The residue was purified by column chromatography (silica gel, 5 g) using EtOAc-hexane (1:9, v/v) as an eluent to give the ester 9a (7.5 mg, 86%). ¹H-NMR (CDCl₃) δ: 1.15 (3H, d, J=6.2 Hz), 1.46 (3H, t, J=7.0 Hz), 191 (3H, s), 2.22 (3H, s), 2.72 (1H, dd, J=8.8, 14.0 Hz), 3.06 (3H, s), 3.13 (1H, dd, J=4.8, 14.0 Hz), 4.10 (2H, q, J=7.0 Hz), 5.11—5.17 (1H, m), 7.17 (1H, t, J=7.0 Hz), 7.34—7.65 (7H, m), 7.85—7.90 (2H, m), 7.96 (1H, d, J=7.7 Hz), 8.44—8.47 (1H, m), 9.28 (1H, br s). MS m/z: 495 (M⁺).

Synthesis of the Ester 9b from (–)-6 and (*S*)-MαNP Acid The same procedure as above was carried out using the alcohol (–)-6 (5 mg, 17.6 μ mol), (*S*)-MαNP acid to give the ester 9b (8 mg, 91%). ¹H-NMR (CDCl₃) δ: 1.03 (3H, d, J=6.2 Hz), 1.48 (3H, t, J=7.0 Hz), 1.78 (3H, s), 2.31 (3H, s), 3.00—3.09 (1H, m), 3.04 (3H, s), 3.18 (1H, dd, J=5.8, 14.0 Hz), 4.12 (2H, q, J=7.0 Hz), 5.07—5.15 (1H, m), 7.18 (1H, t, 7.0 Hz), 7.32—7.54 (7H, m), 7.81—7.89 (2H, m), 7.97 (1H, d, J=8.0 Hz), 8.26 (1H, d, J=7.5 Hz), 8.88 (1H, br s). MS m/z: 495 (M⁺).

Synthesis of the Ester 10a from (+)-6 and (*R*)-MαNP Acid The same procedure as above was carried out using the alcohol (+)-6 (5 mg, 17.6 μmol), (*R*)-MαNP acid to give the ester 10a (8 mg, 91%). ¹H-NMR (CDCl₃) δ: 1.03 (3H, d, J=6.2 Hz), 1.48 (3H, t, J=7.0 Hz), 1.79 (3H, s), 2.32 (3H, s), 3.00—3.09 (1H, m), 3.04 (3H, s), 3.18 (1H, dd, J=5.8, 14.0 Hz), 4.13 (2H, q, J=7.0 Hz), 5.07—5.16 (1H, m), 7.18 (1H, t, J=7.0 Hz), 7.32—7.54 (7H, m), 7.81—7.89 (2H, m), 7.98 (1H, d, J=8.0 Hz), 8.26 (1H, d, J=7.5 Hz), 8.88 (1H, br s). MS m/z: 495 (M⁺).

Synthesis of the Ester 10b from (+)-6 and (S)-MαNP Acid The same procedure as above was carried out using the alcohol (+)-6 (5 mg, 17.6 μmol), (S)-MαNP acid to give the ester 10b (8.5 mg, 97%). 1 H-NMR (CDCl₃) δ: 1.16 (3H, d, J=6.2 Hz), 1.46 (3H, t, J=7.0 Hz), 1.93 (3H, s), 2.22 (3H, s), 2.73 (1H, dd, J=8.8, 14.0 Hz), 3.06 (3H, s), 3.13 (1H, dd, J=4.8, 14.0 Hz), 4.10 (2H, q, J=7.0 Hz), 5.11—5.18 (1H, m), 7.17 (1H, t, J=7.0 Hz), 7.34—7.65 (7H, m), 7.85—7.90 (2H, m), 7.96 (1H, d, J=7.7 Hz), 8.44—8.47 (1H, m), 9.28 (1H, br s). MS m/z: 495 (M $^{+}$).

1-Acetonyl-3-hydroxy-2-methylcarbazole (11) A solution of BBr₃ (60 μl, 0.63 mmol) in CH₂Cl₂ (2 ml) was added to a stirred solution of 3-ethoxycarbazole **8** (89 mg, 0.32 mmol) in CH₂Cl₂ (4 ml) at -78 °C under a N₂ atmosphere. After being gradually warmed to r.t., the mixture was extracted with EtOAc. The EtOAc layer was washed with brine, dried over Na₂SO₄ and concentrated. The residue was purified by column chromatography (silica gel, 10 g) using EtOAc–hexane (3:17, v/v) as an eluent to give the 3-hydroxycarbazole **11** (64 mg, 80%): mp 169—170 °C (from EtOAc–hexane). IR (ATR) v: 3397, 1697 cm⁻¹. ¹H-NMR (CDCl₃) δ: 2.17 (3H, s), 2.49 (3H, s), 4.00 (2H, s), 4.66 (1H, br s), 7.15 (1H, dt, J=2.0,

6.9 Hz), 7.36 (1H, dt, J=2.0, 6.9 Hz), 7.41 (1H, s), 7.43 (1H, dd, J=2.0, 6.9 Hz), 7.93 (1H, d, J=6.9 Hz), 8.29 (1H, br s). MS m/z: 253 (M $^+$). HR-MS m/z: 253.1097 (Calcd for $C_{16}H_{15}NO_2$: 253.1103).

3-Hydroxy-1-(2-hydroxypropyl)-2-methylcarbazole (12) NaBH₄ (16 mg, 0.43 mmol) was added to a solution of acetonylcarbazole **11** (54 mg, 0.21 mmol) in EtOH (10 ml) under ice-cooled water, and was then stirred at the same temperature for 1 h. The reaction mixture was quenched with water, and was then extracted with EtOAc. The EtOAc layer was washed with brine, dried over Na₂SO₄ and concentrated. The residue was purified by column chromatography (silica gel, 10 g) using EtOAc-hexane (1:1, v/v) as an eluent to give the alcohol **12** (54 mg, 99%): mp 133—135 °C (from EtOAc-hexane). IR (ATR) v: 3384 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.38 (3H, d, J=6.4 Hz), 2.37 (3H, s), 3.04 (1H, dd, J=8.0, 14.3 Hz), 3.15 (1H, dd, J=3.6, 14.3 Hz), 4.18—4.26 (1H, m), 4.62 (1H, br s), 7.16 (1H, dt, J=1.1, 8.1 Hz), 7.35 (1H, dt, J=1.1, 8.1 Hz), 7.37 (1H, s), 7.43 (1H, d, J=8.1 Hz), 7.93 (1H, d, J=8.1 Hz), 8.41 (1H, br s). MS m/z: 255 (M⁺). HR-MS m/z: 255.1263 (Calcd for C₁₆H₁₇NO₂: 255.1259).

1-(2-Acetoxypropyl)-3-hydroxy-2-methylcarbazole (-)-(13) and 3-Hydroxy-1-(2-hydroxypropyl)-2-methylcarbazole (+)-(12) i-Pr $_2$ O (10 ml), vinyl acetate (198 μ l, mmol), and the alcohol 12 (23 mg, 0.09 mmol) were added to the lipase QLM (60 mg). The mixture was stirred at 32 °C for 48 h. The reaction mixture was filtered and the volatiles were removed under reduced pressure to give the products. The crude product was purified by preparative TLC using EtOAc—hexane (3:7, v/v) as an eluent to give the acetate (-)-13 (12 mg, 45%) and the alcohol (+)-12 (10 mg, 43%).

(-)-13: The optical purity was 98% ee by HPLC (CHIRALCEL OD; 15% iPrOH-hexane). $[\alpha]_{2}^{D^{3}}$ -122.8° (c=0.12, MeOH). mp 152—154°C (from EtOAc-hexane). IR (ATR) v: 3351, 1708 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.30 (3H, d, J=6.3 Hz), 2.17 (3H, s), 2.40 (3H, s), 3.10 (1H, dd, J=10.0, 13.2 Hz), 3.33 (1H, dd, J=3.3, 13.2 Hz), 5.02—5.12 (1H, m), 7.15 (1H, t, J=7.7 Hz), 7.37 (1H, s), 7.37 (1H, t, J=7.7 Hz), 7.50 (1H, d, J=7.7 Hz), 7.93 (1H, d, J=7.7 Hz), 9.49 (1H, s). MS m/z: 297 (M⁺). HR-MS m/z: 297.1368 (Calcd for C₁₈H₁₀NO₃: 297.1365).

(+)-12: The optical purity was 97% ee by HPLC (CHIRALCEL OJ; 15% iPrOH-hexane). $[\alpha]_2^{12} + 42.0^{\circ}$ (c=0.14, CHCl₃). mp 134—136 °C (EtOAchexane). IR (ATR) v: 3390 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.38 (3H, d, J=6.2 Hz), 2.37 (3H, s), 3.04 (1H, dd, J=8.0, 14.3 Hz), 3.15 (1H, dd, J=3.7, 14.3 Hz), 4.17—4.28 (1H, m), 4.73 (1H, br s), 7.16 (1H, t, J=7.0 Hz), 7.36 (1H, t, J=7.0 Hz), 7.38 (1H, s), 7.39 (1H, d, J=7.0 Hz), 7.93 (1H, d, J=7.0 Hz), 8.44 (1H, s). MS m/z: 255 (M⁺). HR-MS m/z: 255.1250 (Calcd for C₁₆H₁₇NO₂: 255.1259).

(R)-1-(2-Acetoxypropyl)-3-hydroxy-2-methylcarbazole (R)-(-)-(13) A solution of BBr₃ (21 µl, 0.22 mmol) in CH₂Cl₂ (2 ml) was added to a stirred solution of carbazole (R)-(-)-7 (36 mg, 0.11 mmol) in CH₂Cl₂ (2 ml) at -78 °C under a N₂ atmosphere. After being gradually warmed to r.t., the mixture was stirred at r.t. for 3h and extracted with EtOAc. The organic layer was washed with brine, dried over Na2SO4 and concentrated. The residue was purified by column chromatography (silica gel, 10 g) using EtOAc-hexane (3:17, v/v) as an eluent to give the acetate (R)-(-)-13 (10 mg, 30%). The absolute stereochemistry of the acetate (-)-13, derived from the alcohol 12 as described above procedure, was determined to be Rconfiguration by HPLC (CHIRALCEL OD; 15% iPrOH-hexane), because the retention time of the acetate (-)-13 has shown the same retention time in the comparison with the acetate (R)-(-)-acetate 13. 1 H-NMR (CDCl₃) δ : 1.30 (3H, d, J=6.3 Hz), 2.17 (3H, s), 2.40 (3H, s), 3.10 (1H, dd, J=10.0, 13.2 Hz), 3.33 (1H, dd, J=3.3, 13.2 Hz), 5.03—5.13 (1H, m), 7.15 (1H, t, J=7.7 Hz), 7.37 (1H, s), 7.37 (1H, t, J=7.7 Hz), 7.50 (1H, d, J=7.7 Hz), 7.94 (1H, d, J=7.7 Hz), 9.50 (1H, s). MS m/z: 297 (M⁺).

(*R*)-1-(2-Acetoxypropyl)-2-methylcarbazole-3,4-dione (*R*)-(-)-(14) A solution of 3-hydroxycarbazole (-)-13 (22 mg, 0.074 mmol) in THF (2 ml) was added to a stirred suspension of 70% (PhSeO)₂O (76 mg, 0.15 mmol) in THF (2 ml). The mixture was stirred at 50 °C for 30 min. After being cooled to r.t., the reaction mixture was diluted with EtOAc. The mixture was washed with aqueous 10% Na₂CO₃ solution, water and brine. The organic layer was dried over Na₂SO₄ and concentrated. The residue was purified by column chromatography (silica gel, 10 g) using EtOAc-hexane (1:1, v/v) as an eluent to give the carbazole-3,4-quinone (*R*)-(-)-14 (17 mg, 74%). mp 221-223 °C (from EtOAc-hexane). IR (ATR) v: 1735, 1662 cm⁻¹. 1 H-NMR (CDCl₃) δ : 1.41 (3H, d, J=6.2 Hz), 2.06 (3H, s), 2.25 (3H, s), 2.88-3.02 (2H, m), 4.86-4.95 (1H, m), 7.25-7.31 (2H, m), 7.46-7.49 (1H, m), 8.10-8.13 (1H, m), 11.10 (1H, s). MS m/z: 311 (M⁺). HR-MS m/z: 311.1175 (Calcd for $C_{18}H_{17}NO_4$: 311.1158).

(R)-1-(2-Hydroxypropyl)-2-methylcarbazole-3,4-dione (R)-(-)-(3) A carbazole-3,4-quinone (-)-14 (17 mg, 0.055 mmol) was hydrolyzed with

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aqueous 1 M K₂CO₃ (2 ml) and MeOH (3 ml) at r.t. for 3 h to give an (-)-alcohol. The reaction mixture was quenched with water, and was then extracted with EtOAc. The EtOAc layer was washed with brine, dried over Na₂SO₄ and concentrated. The residue was purified by column chromatography (silica gel, 10 g) using EtOAc–hexane (7:3, v/v) as an eluent to give the alcohol (R)-(-)-3 (14 mg, 95%): mp 201—203 °C (EtOAc). IR (ATR) v: 3208, 1716, 1619 cm $^{-1}$. 1 H-NMR (CDCl₃) &: 1.25 (3H, d, J=6.3 Hz), 1.94 (3H, s), 2.77—2.79 (2H, m), 3.93—4.00 (1H, m), 4.90 (1H, br s), 7.22—7.25 (2H, m), 7.51—7.53 (1H, m), 7.84—7.87 (1H, m), 11.10 (1H, s). MS m/z: 269 (M^{\pm}). HR-MS m/z: 269.1059 (Calcd for C₁₆H₁₈NO₃: 269.1052).

(S)-1-(2-Hydroxypropyl)-2-methylcarbazole-3,4-dione (S)-(+)-(3) A solution of alcohol (+)-12 (9 mg, 0.035 mmol) in THF (1 ml) was added to a stirred suspension of 70% (PhSeO)₂O (36 mg, 0.071 mmol) in THF (1 ml). The mixture was stirred at 50 °C for 30 min. After being cooled to r.t., the reaction mixture was diluted with EtOAc. The mixture was washed with aqueous 10% Na₂CO₃ solution, water and brine. The organic layer was dried over Na₂SO₄ and concentrated. The residue was purified by column chromatography (silica gel, 10 g) using EtOAc–hexane (7:3, v/v) as an eluent to give the carbazole-3,4-quinone (S)-(+)-3 (9 mg, 95%) mp 198—201 °C (from EtOAc). IR (ATR) v: 3208, 1716, 1619 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.25 (3H, d, J=6.3 Hz), 1.93 (3H, s), 2.76—2.78 (2H, m), 3.95—3.97 (1H, m), 4.89 (1H, br s), 7.21—7.25 (2H, m), 7.51—7.54 (1H, m), 7.84—7.87 (1H, m), 12.25 (1H, br s). MS m/z: 269 (M⁺). HR-MS m/z: 269.1066 (Calcd for $C_{16}H_{15}NO_3$: 269.1052).

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References and Notes

- Shin-ya K., Tanaka M., Furihata K., Hayakawa Y., Seto H., Tetrahedron Lett., 34, 4943—4944 (1993).
- Shin-ya K., Shimizu S., Kunigami T., Furihata K., Furihata K., Seto H., J. Antibiot., 48, 574—578 (1995).
- 3) Knölker H.-J., Fröhner W., Synlett, 1997, 1108—1110 (1997).
- 4) Knölker H.-J., Reddy K. R., Synlett, 1999, 596—598 (1999).

- Knölker H.-J., Baum E., Reddy K. R., Tetrahedron Lett., 41, 1171— 1174 (2000).
- Czerwonka R., Reddy K. R., Baum E., Knölker H.-J., Chem. Commun., 2006, 711—713 (2006).
- 7) Knölker H.-J., Fröhner W., Tetrahedron Lett., 39, 2537—2540 (1998).
- 8) Knölker H.-J., Baum E., Reddy K. R., Chirality, 12, 526—528 (2000).
- Choshi T., Sada T., Fujimoto H., Nagayama C., Sugino E., Hibino S., Tetrahedron Lett., 15, 2593—2596 (1996).
- Choshi T., Sada T., Fujimoto H., Nagayama C., Sugino E., Hibino S., J. Org. Chem., 62, 2535—2543 (1997).
- Hagiwara H., Choshi T., Fujimoto H., Sugino E., Hibino S., *Tetrahedron*, 56, 5807—5811 (2000).
- Hagiwara H., Choshi T., Fujimoto H., Sugino E., Hibino S., Chem. Pharm. Bull., 46, 1948—1949 (1998).
- Hagiwara H., Choshi T., Nobuhiro J., Fujimoto H., Hibino S., *Chem. Pharm. Bull.*, 49, 881—886 (2001).
- 14) Hirayama M., Choshi T., Kumemura T., Tohyama S., Nobuhiro J., Hibino S., *Heterocycles*, **63**, 1765—1770 (2004).
- Nobuhiro J., Hirayama M., Choshi T., Kamoshita K., Maruyama S., Sukenaga Y., Ishizu T., Fujioka H., Hibino S., *Heterocycles*, 70, 491— 499 (2006).
- Tohyama S., Choshi T., Matsumoto K., Yamabuki A., Ikegata K., Nobuhiro J., Hibino S., Tetrahedron Lett., 46, 5263—5264 (2005).
- Yamabuki A., Fujinawa H., Choshi T., Tohyama S., Matsumoto K., Ohmura K., Nobuhiro J., Hibino S., *Tetrahedron Lett.*, 47, 5859—5861 (2006)
- Kumemura T., Choshi T., Yukawa J., Hirose A., Nobuhiro J., Hibino S., Heterocycles, 66, 87—90 (2005) and related references cited therein.
- 19) Oh-e T., Miyaura N., Suzuki A., Synlett, 1990, 221—223 (1990).
- Miyaura N., Ishiyama T., Sasaki H., Ishikawa M., Sato M., Suzuki A., J. Am. Chem. Soc., 111, 314—321 (1989).
- 21) Tsuji J., Nagashima H., Nemoto H., Org. Synth., 62, 9—13 (1984).
- Harada N., Watanabe M., Kuwahara S., Sugio A., Kasai Y., Ichikawa A., Tetrahedron Asymmetry, 11, 1249—1253 (2000).
- Tsuji H., Kasai Y., Sugio A., Kuwahara S., Watanabe M., Harada N., Ichikawa A., Chirality, 14, 81—84 (2002).