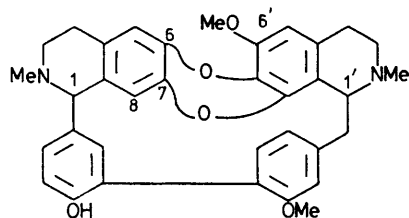


Absolute Configuration and Biosynthesis of Tiliacorine and Tiliacorinine

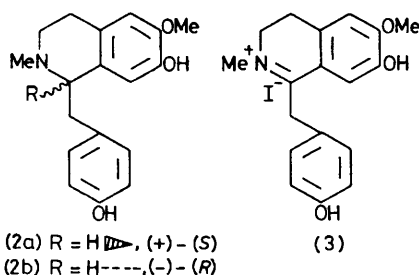
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Summary Biosynthetic experiments with (*R*)- and (*S*)-*N*-methylcoclaurines in *Tiliacora racemosa* Colebr. established that tiliacorine has the (*S*) and (*R*) configuration at the asymmetric centres C(1) and C(1'), respectively, and tiliacorinine has the (*SS*) configuration at both asymmetric centres.

THE diastereomeric bisbenzylisoquinoline alkaloids tiliacrine¹ and tiliacrinine¹ have been assigned the structure² (1). The absolute configuration at the asymmetric centres C(1) and C(1') in both the bases cannot be determined by the usual sodium-ammonia cleavage method³ because the two lower rings of these bases are linked through a direct carbon-to-carbon bond, rather than through the much more common diaryl ether bridge.⁴



(1)



According to established biogenetic theory^{5,6} both tiliacrine and tiliacrinine can be formed in nature by inter- and intra-molecular oxidative coupling of the two *N*-methylcoclaurine units. The 1,4-dioxan bridge present in (1) can be generated by loss of one MeO⁺ group from one of the *N*-methylcoclaurine (6) units.^{4,7}

TABLE. Tracer experiment on *T. racemosa* Colebr.

Precursor fed	% Incorporation into	
	Tiliacrine	Tiliacrinine
(±)-[U ¹⁴ C] Tyrosine ..	0.09	0.06
(±)-[1- ³ H] (4) ..	0.10	0.09
(±)-[3',5',8- ³ H ₃] (5) ..	0.14	0.12
(±)-[3',5',8- ³ H ₃] (6) ..	0.18	0.15
(±)-[3',5',8- ³ H ₃] (7) ..	0.0004	0.00045
[N- ¹⁴ CH ₃] (3) ..	0.12	0.10
(±)-[1- ³ H, N- ¹⁴ CH ₃] (6) ..	0.15	0.12
(±)-[1- ³ H, 6-O- ¹⁴ CH ₃] (6) ..	0.19	0.16
(-)-(R)-[3',5',8- ³ H ₃] (2b) ..	0.16	0.004
(+)-(S)-[3',5',8- ³ H ₃] (2a) ..	0.15	0.28

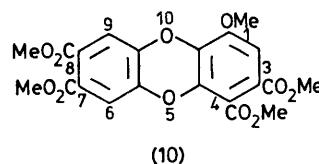
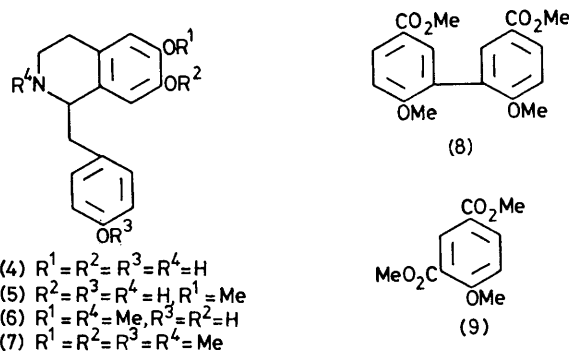
† The precursors (3)—(6) were prepared by standard methods [D. H. R. Barton, D. S. Bhakuni, G. M. Chapman, and G. W. Kirby, *J. Chem. Soc. (C)*, 1967, 2134]; (7) was obtained by treatment of (6) with diazomethane. Details of the counting method employed are given by D. S. Bhakuni, S. Tewari, and R. S. Kapil, *J.C.S. Perkin I*, 1977, 706. The precursors were fed by the wick feeding method.

‡ Tritium was introduced specifically into positions *ortho* to the phenolic hydroxy groups in the precursors by base catalysed exchange (G. W. Kirby and L. Ogunkoya, *J. Chem. Soc.*, 1965, 6914). Uniformity of the labelling in the *ortho* positions was established by degradation of the labelled precursor (6).

§ Taking into account the loss of tritium in oxidative coupling, the distribution of the radioactivity in (8) and (10) formed by oxidative degradation of the biosynthetic bases derived from specifically and uniformly labelled (6) demonstrated that (±)-(6) is incorporated into both units of the dimers.

Feeding of (±)-tyrosine (Table), (±)-norcoclaurine (4) (±)-coclaurine (5), (±)-¹N-methylcoclaurine (6), (±)-*N*OOC trimethylcoclaurine (7), and didehydro-*N*-methylcoclaurinium iodide (3) established that (4), (5), and (6) are efficient precursors of tiliacrine and tiliacrinine in *Tiliacora racemosa* Colebr. (Menispermaceae). The efficient incorporation of (3) is probably due to prior reduction *in vivo* to (6). As expected (7) was not incorporated into tiliacrine and tiliacrinine.

Labelled tiliacrine and tiliacrinine derived from (±)-[3',5',8-³H₃]-*N*-methylcoclaurine (6)† feedings were separately converted into *O*-methyltiliacrine dimethiodide¹ and *O*-methyltiliacrinine dimethiodide¹ by treatment with methyl iodide-sodium methoxide. Alkaline permanganate oxidation of the dimethiodides,¹ followed by methylation with diazomethane of the acids so formed yielded, in each case, (8) and (10). Oxidation of labelled *O*-methyltiliacrine dimethiodide¹ (molar activity 2.63 × 10⁶ dis. min⁻¹ mmol⁻¹) gave (8) (molar activity 1.78 × 10⁵ dis. min⁻¹ mmol⁻¹) and (10) (molar activity 8.36 × 10⁴ dis. min⁻¹ mmol⁻¹) having essentially 2/3 and 1/3 radioactivity, respectively, of the parent base.‡ Similar results were obtained for the degradation of *O*-methyltiliacrinine.



Feeding of (±)-[1-³H, N-¹⁴CH₃] (6) gave tiliacrine and tiliacrinine labelled both with ¹⁴C and ³H. The ratios of these radioatoms in the precursor and biosynthetic bases

were practically unchanged.¶ Feeding of (\pm)-[1- 3 H, 6-O- 14 CH $_3$]-**(6)** also yielded tiliacrine and tiliacrinine labelled with 14 C and 3 H, but the 14 C and 3 H ratios in the precursor was 1:30 and in the biosynthetic tiliacrine and tiliacrinine was 1:61 and 1:59.5, respectively.**

Parallel feeding experiments with (-)-(*R*)-*N*-methylcoclaurine (**2b**) and (+)-(*S*)-*N*-methylcoclaurine (**2a**) gave, in each case, radioactive tiliacrine. Labelled tiliacrine derived from the (+)-(*S*)-form (**2a**) was converted into tiliacrine dimethiodide¹ by treatment with methyl iodide with practically no loss of radioactivity. Alkaline permanganate oxidation which destroys the phenolic ring of the tiliacrine dimethiodide¹ (molar activity 10.65×10^5 dis. min⁻¹ mmol⁻¹) gave (**9**)¹ (inactive) and (**10**) (molar activity 5.19×10^5 dis. min⁻¹ mmol⁻¹). Tiliacrine derived from the (-)-(*R*)-form (**2b**) was converted into tiliacrine dimethiodide¹ (molar activity 1.75×10^5 dis. min⁻¹ mmol⁻¹) and was similarly degraded to (**9**) (molar activity 1.5×10^5 dis. min⁻¹ mmol⁻¹) and (**10**) (radioinactive). These results thus established the (*S*)- and (*R*)-configuration at the asymmetric centres C(1) and C(1'), respectively, in tiliacrine.

Parallel feeding of the (+)-(*S*) and (-)-(*R*) isomers (**2a**) and (**2b**) demonstrated that the former was incorporated into tiliacrinine 70 times more efficiently than the latter. Labelled tiliacrinine derived from the (+)-(*S*)-form (**2a**) was converted into *O*-methyltiliacrinine dimethiodide¹ by treatment with methyl iodide-sodium methoxide. Alkaline permanganate oxidation of the *O*-methyltiliacrinine dimethiodide (molar activity 1.8×10^5 dis. min⁻¹ mmol⁻¹) followed by methylation with diazomethane of the acids so formed gave (**8**) (molar activity 1.22×10^5 dis. min⁻¹ mmol⁻¹) and (**10**) (molar activity 5.72×10^4 dis. min⁻¹ mmol⁻¹) having essentially 2/3 and 1/3 radioactivity, respectively. The results thus established the (*SS*) configuration at the two asymmetric centres C(1) and C(1'), respectively, in tiliacrinine.

Trapping experiments by feeding (\pm)-tyrosine to *T. racemosa* Colebr plants showed fairly high incorporation (0.50%) into (**6**). Thus (**6**) is a true precursor of (**1**).

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¶ Since there is no loss of hydrogen from C(1) in the precursor in the biotransformation and stereospecificity is maintained in the biosynthesis of the bisbenzylisoquinoline alkaloid from the 1-benzyltetrahydroisoquinoline precursor [D. H. R. Barton, G. W. Kirby, and A. Wiechers, *J. Chem. Soc. (C)*, 1966, 266], this demonstrates that the stereochemistry of these asymmetric centres remain unchanged during biosynthesis.

** The loss of the MeO⁺ group from one of the *N*-methylcoclaurine units in the formation of the 1,4-dioxan bridge in cocsulin has been confirmed (D. S. Bhakuni, V. M. Labroo, A. N. Singh, and R. S. Kapil, *J.C.S. Perkin I*, 1977, in the press).

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³ Y. Inubushi, K. Momura, and M. Miyawaki, *J. Pharm. Soc., Japan*, 1963, 83, 282.

⁴ M. Shamma, 'The Isoquinoline Alkaloids,' Academic Press, New York, 1972, p. 138.

⁵ F. Faltis and H. Fraendorfer, *Ber.*, 1930, 63, 806.

⁶ D. H. R. Barton and T. Cohen, 'Festschrift A. Stoll,' Birkhauser, Basel, 1957, p. 117.

⁷ A. R. Battersby in 'Oxidative coupling of Phenols,' eds. A. R. Battersby and W. I. Taylor, Marcel Dekker, New York, 1967.