© Adis International Limited All rights reserved

## **Quinolone Activity Against Anaerobes**

Peter C. Appelbaum

Department of Pathology, Hershey Medical Center, Hershey, Pennsylvania, USA

#### **Abstract**

The first generation of fluoroquinolones such as ciprofloxacin and ofloxacin are inactive against most anaerobic bacteria. However, some broad-spectrum quinolones, which have recently become clinically available or are under active development, have significant antianaerobic activity. This review summarises the *in vitro* activity of currently available, as well as experimental, quinolones against clinically significant anaerobic bacteria. Quinolones with low activity against anaerobes include ciprofloxacin, ofloxacin, levofloxacin, fleroxacin, pefloxacin, enoxacin and lomefloxacin. Compounds with intermediate antianaerobic activity include sparfloxacin and grepafloxacin. Trovafloxacin, gatifloxacin and moxifloxacin yield low MICs against most groups of anaerobes. Quinolones with the greatest *in vitro* activity against anaerobes include clinafloxacin and sitafloxacin (DU-6859a).

### 1. Background

Anaerobes are established causes of serious human infections, especially in debilitated hosts. Classification of anaerobes has changed markedly in recent vears, and readers are referred to standard texts for full details.[1] A summary of anaerobes commonly encountered in human disease is presented in table I. Although infections caused by members of the Bacteroides fragilis group occur most commonly, infections caused by other Gram-negative anaerobic rods, as well as by Gram-positive cocci and rods, are increasingly encountered.<sup>[2]</sup> The susceptibility spectrum of clinically isolated anaerobes is changing: although βlactamase production, and concomitant resistance to β-lactams, is the rule in the B. fragilis group, both phenomena are increasingly encountered in non-B. fragilis group Bacteroides, Prevotella, Porphyromonas and Fusobacterium species. [3-5] B-Lactamase production has also been described in Clostridium butyricum, C. ramosum and C. clostridiiforme.[3] Metronidazole resistance is common amongst Gram-positive non-sporeforming rods, but has also been reported in peptostreptococci, non-C. perfringens clostridia and members of the B. fragilis group. Additionally, clindamycin resistance is not unusual amongst anaerobic Gram-negative rods.[6]

Commercially available quinolones such as ciprofloxacin, ofloxacin, fleroxacin, pefloxacin, enoxacin and lomefloxacin are inactive or marginally active against anaerobes, with MICs either higher

Table I. Shortened classification of clinically significant anaerobes

#### Gram-positive spore-forming rods (clostridia)

Clostridium perfringens

C. novyi

C. septicum C. difficile

#### Gram-positive non-spore-forming rods (microaerophils)

Propionibacteria

Actinomyces spp.

Fubacteria

Bifidobacteria

#### **Gram-positive cocci** Peptostreptococci

Gram-negative rods

Gram-negative rods
Bacteroides fragilis group

B. fragilis

B. thetaiotaomicron

B. distasonis

B. ovatus

Prevotella/Porphyromonas P. melaninogenica

P intermedia

P. bivia

P. disiens

Fusobacteria

Fusobacterium nucleatum

F. necrophorum

F. mortiferum F varium

r. varium Miscellaneous

Campylobacter gracilis (microaerophilic)

Bilophila wadsworthia

Sutterella wadsworthensis

#### Anaerobic Gram-negative cocci

Veillonella

**Table II.** *In vitro* activities of ciprofloxacin, ofloxacin and levofloxacin against anaerobes<sup>[7-10,16,18]</sup>

Group	MIC <sub>90</sub> (mg/L)				
	ciprofloxacin	ofloxacin	levofloxacin		
Bacteroides fragilis group	4.0-64.0	3.13-50.0	2.0-12.5		
Prevotella/Bacteroides/Porphyromonas spp.	1.0-32.0	2.0-32.0	6.25-8.0		
Fusobacterium spp.	2.0-32.0	1.56-64.0	0.39		
Peptostreptococcus spp.	0.5-4.0	0.78-16.0	0.39-12.5		
Non-spore-forming Gram-positive rods	2.0-16.0 <sup>a</sup>	4.0-8.0 <sup>a</sup>	No data		
Clostridium spp.	4.0-32.0 <sup>b</sup>	4.0-16.0 <sup>b</sup>	2.0		

- a Propionibacterium acnes: ciprofloxacin 0.5 to 1.0 and ofloxacin 0.5 to 2.0 mg/L.
- b Clostridium perfringens: ciprofloxacin 0.25 to 1.56 and ofloxacin 0.5 to 3.13 mg/L.

than, or clustering around breakpoints for aerobic bacteria. [7-9] However, trovafloxacin (recently released in the US) as well as some other quinolones currently under development have significant activity against anaerobes. There is a clear need for availability of nontoxic oral or injectable quinolones or naphthyridone compounds, which are active against clinically significant anaerobic bacteria. In view of the activity of all broad-spectrum quinolones against *Enterobacteriaceae*, such compounds could then potentially be used as single-drug therapy for mixed infections caused by aerobes and anaerobes, both in outpatients and inpatients.

This review summarises current information on the *in vitro* susceptibility of anaerobes to available and experimental quinolones, as well as current clinical information on activity of certain quinolones against anaerobic infections in humans.

# 2. *In Vitro* Antianaerobic Activity of Quinolones

Results of published papers on *in vitro* activity of a broad range of quinolone/naphthyridone compounds against the major anaerobe groups [7-24] are summarised in tables II-VI. Previous broad-spectrum quinolones with excellent *in vitro* activity against anaerobes (e.g. Win 57273, Bay y3118) [14,15] proved to be too toxic to permit further development. It should be noted that, apart from trovafloxacin (susceptible  $\leq$ 2.0 mg/L, intermediate 4.0 mg/L, resistant  $\geq$ 8.0 mg/L) [25] there are currently no recommended breakpoints for quinolones against anaerobes, so results are presented throughout as MIC90s (mg/L) against the major anaerobe groups.

Ciprofloxacin and ofloxacin (table II) are only active against *Propionibacterium acnes* and some *C. perfringens* strains and usually yield MIC<sub>90</sub>s ≥4.0 mg/L for all other organisms. Levofloxacin generally has MICs 1 or 2 doubling dilutions lower than those for ofloxacin (table II). By comparison, fleroxacin,

lomefloxacin, pefloxacin and enoxacin are inactive against anaerobes (table III).

Among other quinolones, grepafloxacin yields MICs that are one or 2 dilutions lower than those for ciprofloxacin (table IV), but values are still too high to suggest clinical efficacy. Sparfloxacin yields MICs slightly lower than those of grepafloxacin (table IV). Trovafloxacin has been found to be very active against anaerobes, with most workers reporting an overall MIC90 of 1.0 mg/L and all groups of strains susceptible at an MIC90 of  $\leq 1.0$  mg/L. Some authors have reported trovafloxacin MICs of 4.0 mg/L, but this would still be in the therapeutic range with the intravenous formulation (table V). Gatifloxacin and moxifloxacin have been shown to yield MICs that are a little higher than those of trovafloxacin, but no

**Table III.** *In vitro* activities of fleroxacin, lomefloxacin, pefloxacin and enoxacin against anaerobes<sup>[11-13]</sup>

Group	MIC <sub>90</sub> (mg/L)
Bacteroides fragilis group	8.0->64.0
Prevotella/Bacteroides/	8.0-32.0
Porphyromonas sp.	00.0 04.0
Fusobacterium spp. Peptostreptococcus spp.	32.0->64.0 8.0-16.0
Non-spore-forming Gram-positive rods	8.0-32.0
Clostridium spp.	16.0->64.0 <sup>a</sup>

a Clostridium perfringens: fleroxacin and lomefloxacin 1.0 to 4.0 mg/L.

MIC<sub>90</sub> = concentration required to inhibit 90% of strains.

**Table IV.** *In vitro* activities of grepafloxacin and sparfloxacin against anaerobes<sup>[13-15,17,19]</sup>

Group	MIC <sub>90</sub> (mg/L)		
	grepafloxacin	sparfloxacin	
Bacteroides fragilis group	3.13-25.0	1.0-4.0	
Prevotella/Bacteroides/ Porphyromonas spp.	4.0-25.0	4.0-16.0	
Fusobacterium spp.	8.0	4.0-16.0	
Peptostreptococcus spp.	1.0-3.13	0.5-4.0	
Non-spore-forming Gram-positive rods	25.0	0.25-8.0	
Clostridium spp.	12.5	0.5-8.0	

MIC<sub>90</sub> = concentration required to inhibit 90% of strains.

Table V. In vitro activities of trovafloxacin, gatifloxacin and moxifloxacin against anaerobes<sup>[19-23]</sup>

Group	MIC <sub>90</sub> (mg/L)			
	trovafloxacin	gatifloxacin	moxifloxacin	
Bacteroides fragilis group	0.5-1.0	2.0	4.0	
Prevotella/Bacteroides/Porphyromonas spp.	1.0	4.0	0.5-2.0	
Fusobacterium spp.	1.0-2.0	4.0	0.5-8.0	
Peptostreptococcus spp.	0.25-1.0	1.0	0.25	
Non-spore-forming Gram-positive rods	1.0-4.0	1.0	0.25 (few strains)	
Clostridium spp.	0.25-1.0	2.0	0.5-1.0	

breakpoints have yet been established for these compounds (table V).

The most active quinolones currently under development against anaerobes are clinafloxacin and sitafloxacin (DU-6859a), with MICs ≤0.5 mg/L for most groups (table VI). MICs of clinafloxacin and sitafloxacin are significantly lower than those of any other quinolones currently under development. No breakpoints are available for either compound, and their clinical potential will depend on their pharmacokinetic/pharmacodynamic properties (see below) as well as their toxicology.

Factors other than MIC play a role in the potential therapeutic usefulness of quinolones against anaerobes. Activity of quinolones has been proved to be dose related. Craig<sup>[26]</sup> has reported that, in order to be active, 24-hour AUC (area under the plasma concentration-time curve)/MIC ratios for quinolones must be at least 25 after therapeutic doses, and at least 125 for seriously ill patients. Current data suggest that trovafloxacin, gatifloxacin, moxifloxacin, clinafloxacin and sitafloxacin all yield AUC/MIC values of at least 25 against most anaerobes, with clinafloxacin and sitafloxacin, followed by trovafloxacin, having values of ≥125, indicating potential use in seriously ill, hospitalised patients.

Time-kill studies performed in our laboratory have demonstrated that bacteriostatic/bactericidal concentrations of trovafloxacin after 48 hours for 6 Gram-positive and -negative anaerobes were 0.03 to 1.0/0.03/1.0 mg/L. Table VII shows bacteriostatic/bactericidal concentrations of trovafloxacin compared

**Table VI.** In vitro activities of clinafloxacin and sitafloxacin (DU-6859a) against anaerobes

roup	MIC <sub>90</sub> (mg/L)			
	clinafloxacin	sitafloxacin		
acteroides fragilis group	0.125-2.0	0.06-0.5		
revotella/Bacteroides/ orphyromonas spp.	0.125-0.5	0.25-0.39		
usobacterium spp.	0.06-1.0	0.06-0.5		
eptostreptococcus spp.	0.125-0.5	0.06-0.1		
on-spore-forming ram-positive rods	0.06	0.25		
lostridium spp.	0.5-2.0	0.06-0.5		

with other quinolone and nonquinolone antibacterial agents. Against 11 different anaerobes, sitafloxacin was bactericidal at the MIC (0.06 to 0.5 mg/L) against all strains after 48 hours (table VIII). Time-kill studies have not been reported for other quinolones with improved antianaerobic activity. [27,28] However, the clinical significance of *in vitro* bactericidal activity of drugs against anaerobes is unknown at the present time

Other factors may play a role in the in vivo susceptibility of anaerobes to the quinolone group. Appelbaum and co-workers<sup>[29]</sup> have found ciprofloxacin to reduce counts of anaerobic organisms in saliva of human volunteers, 91.3 to 100% of strains inhibited for all anaerobe groups. Goldstein and Citron<sup>[30]</sup> have found that ofloxacin + metronidazole usually showed additive or indifferent, but no antagonistic activity, when tested against anaerobes isolated from intra-abdominal infections. Metronidazole + fleroxacin have been shown to be at least as active as clindamycin + gentamicin against all anaerobe species in an experimental intra-abdominal abscess model, and significantly more effective against clostridia.[31] In in vitro and animal studies, fleroxacin and sparfloxacin have been found to be bactericidal against Escherichia coli under anaerobic conditions, and also bactericidal against B. fragilis.[32,33]

#### 3. Clinical Studies

Clinical results for quinolone treatment of mixed aerobic-anaerobic infections are only available for clinafloxacin and trovafloxacin. Wilson<sup>[34]</sup> compared clinafloxacin versus imipenem/cilastatin in the treatment of intra-abdominal infection in a multicentre trial. At the major testing facility (University of California at Irvine), clinafloxacin inhibited 126 anaerobic isolates from 44 patients (100%) at a concentration of <1 mg/L and imipenem inhibited 124 strains (98.4%) at <4.0 mg/L. Of clinically evaluable patients, clinafloxacin at a dosage of 200mg every 12 hours resulted in cure in 76.5% and failure in 22.5%, with 1.0% not assessed. Corresponding results for imipenem/cilastatin (500mg every 6 hours) were 71.5, 27.6 and 0.9%. [34]

Table VII. Bacteriostatic/bactericidal concentrations (mg/L) of trovafloxacin compared to other agents after 48 hours

Drug	Bacteroides fragilis	Bacteroides thetaiotaomicron	Prevotella melaninogenica	Fusobacterium mortiferum	Peptostreptococcus magnus	Clostridium perfringens
Trovafloxacin	0.25/0.5	0.25/0.5	1.0/1.0	0.5/1.0	0.015/0.06	0.03/0.03
Ciprofloxacin	16.0/32.0	8.0/16.0	4.0/4.0	2.0/2.0	1.0/2.0	0.25/0.25
Sparfloxacin	2.0/8.0	0.5/1.0	2.0/4.0	0.5/1.0	1.0/1.0	0.06/0.06
Metronidazole	1.0/1.0	2.0/2.0	1.0/1.0	1.0/2.0	64.0/64.0	1.0/1.0
Cefoxitin	16.0/32.0	1.0/1.0	0.5/1.0	2.0/2.0	4.0/8.0	0.125/0.125
Piperacillin	64.0/64.0	16.0/32.0	8.0/8.0	1.0/1.0	0.25/0.5	0.125/0.125
Piperacillin/tazobactam	2.0/8.0	2.0/4.0	0.125/0.25	1.0/1.0	0.25/0.5	0.06/0.125

Donahue and co-workers, [35] in a double-blind multicenter study, compared 2 sequential antibacterial regimens following surgical intervention of a documented intra-abdominal infection with a mixed aerobic/anaerobic flora. These were intravenous alatrofloxacin 300mg daily followed by oral trovafloxacin 200mg daily or imipenem/cilastatin 1g 3 times daily intravenously followed by oral amoxicillin/clavulanic acid 500mg 3 times daily for up to 14 days. At study end, cure or improvement occurred in 83% (129/156)

and 84% (127/152) of clinically evaluable patients in the trovafloxacin and comparative groups, respectively. [35] Pathogen eradication rates and adverse event profiles were comparable between groups.

#### 4. Conclusions

In summary, quinolones with low *in vitro* activity against anaerobes include ciprofloxacin, ofloxacin, levofloxacin, fleroxacin, pefloxacin, enoxacin and

Table VIII. Comparative sitafloxacin time-kill results after 24 and 48 hours. Number of anaerobic strains showing reduction of 1, 2 and 3 log<sub>10</sub> cfu/ml in bacterial numbers after exposure to sitafloxacin and other antibacterial agents

Drug	24 hours			48 hours			
	-1 <sup>a</sup>	-2 <sup>a</sup>	-3ª	-1	-2	-3	
Sitafloxacin							
4 x MIC	11	10	9	11	11	11	
2 x MIC	11	10	7	11	11	11	
MIC	11	9	5	11	11	11	
Ciprofloxacin							
4 x MIC	11	10	7	11	11	11	
2 x MIC	11	9	4	11	11	11	
MIC	11	8	0	11	11	11	
Levofloxacin	• •	•	-	• •	•	• •	
4 x MIC	11	11	8	11	11	11	
2 x MIC	11	10	6	11	11	11	
MIC	11	10	4	11	11	11	
Sparfloxacin		10	7			••	
4 x MIC	11	10	7	11	11	11	
2 x MIC	10	10	5	11	11	11	
MIC	10	8	3	11	11	11	
Piperacillin	10	O	3	11	11	11	
4 x MIC	11	8	6	11	11	11	
2 x MIC	10	8	6	11	11	11	
MIC	10	7	5	11	11	11	
Piperacillin/tazobactam	10	,	3	11	11	11	
4 x MIC	11	7	6	11	11	11	
2 x MIC	10	7		11	11	11	
			6				
MIC	10	7	5	11	11	11	
Imipenem		_	_				
4 x MIC	11	9	7	11	11	11	
2 x MIC	11	8	7	11	11	11	
MIC	11	8	6	11	11	11	
Clindamycin							
4 x MIC	11	11	8	11	11	11	
2 x MIC	11	9	5	11	11	11	
MIC	11	3	4	11	11	11	
Metronidazole							
4 x MIC	11	9	4	11	11	11	
2 x MIC	10	7	4	11	11	11	
MIC	8	6	3	11	11	11	

64 Appelbaum

lomefloxacin. Compounds with intermediate anaerobe activity include sparfloxacin and grepafloxacin. Quinolones very active against anaerobes include clinafloxacin and sitafloxacin, followed by trovafloxacin, gatifloxacin and moxifloxacin. More pharmacokinetic, toxicological and clinical studies are necessary before firmer conclusions can be drawn as to the therapeutic value of quinolones against anaerobic infections in humans. For clinafloxacin and trovafloxacin, clinical studies have been performed which document the usefulness of these compounds as single-drug therapy for intra-abdominal infections caused by a mixed aerobic/anaerobic flora.

#### References

- Manual of clinical microbiology. Murray PR, Baron EJ, Pfaller MA, et al., editors. Washington, DC: ASM Press, 1995
- Finegold SM, George WL, editors. Anaerobic infections in humans. New York: Academic Press, 1989
- Nord CE. Mechanisms of β-lactam resistance in anaerobic bacteria. Rev Infect Dis 1986; 8 Suppl. 5: S543-8
- Appelbaum PC, Spangler SK, Jacobs MR. B-lactamase production and susceptibilities to amoxicillin, amoxicillin-clavulanate, ticarcillin, ticarcillin/clavulanate, cefoxitin, imipenem, and metronidazole of 320 non-Bacteroides fragilis Bacteroides isolates and 129 fusobacteria from 28 US centers. Antimicrob Agents Chemother 1990; 34: 1546-50
- Jacobs MR, Spangler SK, Appelbaum PC. β-lactamase production, β-lactam sensitivity and resistance to synergy with clavulanate of 737 Bacteroides fragilis group organisms from thirty-three US centres. J Antimicrob Chemother 1990; 26: 361-70
- Appelbaum PC, Spangler SK, Jacobs MR. Susceptibility of 539 Grampositive and Gram-negative anaerobes to new agents, including RP 59500, biapenem, trospectomycin and piperacillin/tazobactam. J Antimicrob Chemother 1993; 32: 223-31
- Prabhala RH, Rao B, Marshall R, et al. In vitro susceptibility of anaerobic bacteria to ciprofloxacin (Bay o 9867). Antimicrob Agents Chemother 1984: 26: 785-6
- Wise R, Andrews JM, Danks G. In-vitro activity of enoxacin (CI-919), a new quinolone derivative, compared with that of other antimicrobial agents. Antimicrob Agents Chemother 1984; 13: 237-44
- Goldstein EJC, Citron DM. Comparative activity of the quinolones against anaerobic bacteria isolated at community hospitals. Antimicrob Agents Chemother 1985; 657-9
- Jones BM, Geary I, Lee ME, et al. Activity of perfloxacin and thirteen other antimicrobial agents in vitro against isolates from hospital and genitourinary infections. J Antimicrob Chemother 1986; 17: 739-46
- Wüst J, Hardegger U. Comparative in vitro activity of cefetamet and fleroxacin against anaerobic bacteria. Eur J Clin Microbiol 1987; 6: 688-90
- Chin N-X, Novelli A, Neu HC. In vitro activity of lomefloxacin (SC-47111; NY-198), a difluoroquinolone 3-carboxylic acid, compared with those of other quinolones. Antimicrob Agents Chemother 1988; 32: 656-62
- Barry AL, Fuchs PC. In vitro activities of sparfloxacin, tosufloxacin, ciprofloxacin and fleroxacin. Antimicrob Agents Chemother 1991; 35: 955-60
- Goldstein EJC, Citron DM. Comparative activity of ciprofloxacin, ofloxacin, sparfloxacin, temafloxacin, CI-960, CI-990, and Win 57273 against anaerobic bacteria. Antimicrob Agents Chemother 1992; 36: 1158-62
- Bauernfeind A. Comparative in vitro activities of the new quinolone, Bay y 3118, and ciprofloxacin, sparfloxacin, tosufloxacin, CI-960 and CI-990. J Antimicrob Chemother 1993; 31: 505-22
- Fernandes PB, Shipkowitz N, Bower RR, et al. *In-vitro* and *in-vivo* potency of five new fluoroquinolones against anaerobic bacteria. J Antimicrob Chemother 1986; 18: 693-701

 Wexler HM, Molitoris E, Finegold SM. In vitro activities of three of the newer quinolones against anaerobic bacteria. Antimicrob Agents Chemother 1902; 36: 239-43

- Chemother 1992; 36: 239-43

  18. Fu KP, Lafredo SC, Foleno B, et al. *In vitro* and *in vivo* antibacterial activities of levofloxacin (<u>I</u>-ofloxacin), an optically active ofloxacin. Antimicrob Agents Chemother 1992; 36: 860-6
- Spangler SK, Jacobs, MR, Appelbaum, PC. Activity of CP 99,219 compared with those of ciprofloxacin, grepafloxacin, metronidazole, cefoxitin, piperacillin, and piperacillin-tazobactam against 489 anaerobes. Antimicrob Agents Chemother 1994; 38: 2471-6
- Ednie LM, Jacobs MR, Appelbaum PC. Activities of gatifloxacin compared to those of seven other agents against anaerobic organisms. Antimicrob Agents Chemother 1998; 42: 2459-62
- Kato N, Kato H, Tanaka-Bandoh K, et al. Comparative in-vitro and invivo activity of AM-1155 against anaerobic bacteria. J Antimicrob Chemother 1997: 40: 631-7.
- Aldridge, KE, Ashcraft, DS. Comparison of the *in vitro* activities of Bay 12-8039, a new quinolone, and other antimicrobials against clinically important anaerobes. Antimicrob Agents Chemother 1997; 41: 709-11
- MacGowan AP, Bowker KE, Holt HA, et al. Bay 12-8039, a new 8-methoxy-quinolone: comparative in-vitro activity with nine other antimicrobials against anaerobic bacteria. J Antimicrob Chemother 1997; 40: 503.0
- Wexler HM, Molitoris E, Reeves D, et al. In vitro activity of DU-6859a against anaerobic bacteria. Antimicrob Agents Chemother 1994; 38: 2504-0
- National Committee for Clinical Laboratory Standards. Methods for antimicrobial susceptibility testing of anaerobic bacteria; approved standard. 4th ed. M11-A4. Wavne. PA: NCCLS. 1997
- Craig WA. Pharmacokinetic/pharmacodynamic parameters: rationale for antibacterial dosing of mice and men. Clin Infect Dis 1998; 26: 1-12
- Spangler SK, Jacobs MR, Appelbaum PC. Time-kill study of the activity
  of trovafloxacin compared with ciprofloxacin, sparfloxacin, metronidazole, cefoxitin, piperacillin and piperacillin/tazobactam against six
  anaerobes. J Antimicrob Chemother 1997; 39 Suppl. B: 23-7
- Spangler SK, Jacobs MR, Appelbaum PC. Bactericidal activity of DU-6859a compared to activities of three quinolones, three β-lactams, clindamycin, and metronidazole against anaerobes as determined by time-kill methodology. Antimicrob Agents Chemother 1997; 41: 847-9
- Appelbaum PC, Spangler SK, Strauss M. Reduction of oral flora with ciprofloxacin in human volunteers. J Antimicrob Chemother 1988; 21: 243-9
- Goldstein EJC, Citron DM. Susceptibility of anaerobic bacteria isolated from intra-abdominal infections to ofloxacin and interaction of ofloxacin with metronidazole. Antimicrob Agents Chemother 1991; 35: 2447-9
- Pefanis A, Thauvin-Eliopoulos C, Holden J, et al. Activity of fleroxacin alone and in combination with clindamycin or metronidazole in experimental intra-abdominal abscesses. Antimicrob Agents Chemother 1994; 38: 252-5
- Griggs DJ, Piddock LJV, Wise R. The killing activity of fleroxacin upon Bacteroides fragilis. J Antimicrob Chemother 1989; 23: 53-8
- Cooper MA, Andrews JM, Wise R. Bactericidal activity of sparfloxacin and ciprofloxacin under anaerobic conditions. J Antimicrob Chemother 1991: 28: 399-405
- Wilson SE. Current issues in surgical and antimicrobial management of intraabdominal infection [abstract 2.006]. Proceedings of the 8th International Congress on Infectious Diseases;1998 May; Boston, MA: 5
- Donahue PE, Smith DL, Yellin AE, et al. Trovafloxacin in the treatment of intra-abdominal infections: results of a double-blind, multicenter comparison with imipenem/cilastatin. Trovafloxacin Surgical Group. Am J Surg 1998: 176 Suppl. 6A: 53S-61S

Correspondence and reprints: Dr *Peter C. Appelbaum*, Department of Pathology, Hershey Medical Center, P.O. Box 850, Hershey, PA 17033, USA.