© Adis International Limited. All rights reserved.

# **Pancreatic Cancer**

# A Review of Emerging Therapies

# Lawrence Rosenberg

The Pancreatic Diseases Centre, Montreal General Hospital, McGill University Health Centre, and Department of Surgery, McGill University, Montreal, Quebec, Canada

# Contents

### **Abstract**

The incidence of adenocarcinoma of the pancreas has risen steadily over the past 4 decades. Since pancreatic cancer is diagnosed at an advanced stage, and because of the lack of effective therapies, the prognosis of such patients is extremely poor. Despite advances in our understanding of the molecular biology of pancreatic cancer, the systemic treatment of this disease remains unsatisfactory. Conventional chemotherapy has not produced dramatic improvements in response rates or patient survival. New treatment strategies are clearly needed. This paper reviews emerging therapies for pancreatic carcinoma. A more profound understanding of the molecular biology of cell growth and proliferation, as well as of neoplastic cell transformation, has led to advances in several areas, including the use of somatostatin analogues and antiandrogens as adjuvant therapy; inhibition of tumour growth and metastasis by inhibitors of matrix metalloproteinases and angiogenesis, and by small molecules such as retinoids, which interfere with

progression through the cell cycle; immunotherapy with monoclonal antibodies; disruption of intracellular signal transduction with farnesyltransferase inhibitors; and finally gene therapy with specifically designed vaccines.

### 1. Pancreatic Cancer

The incidence of adenocarcinoma of the pancreas has risen steadily over the past 4 decades. It currently stands at approximately 29 000 new cases per year in North America,[1] making it the second most common gastrointestinal malignancy and the fifth leading cause of adult deaths from cancer.[2] The disease is characterised by its aggressive nature. The diagnosis of pancreatic cancer is usually established at an advanced stage, and a lack of effective therapies leads to an extremely poor prognosis. A meta-analysis of 144 reported series including approximately 37 000 patients found the median survival time to be 3 months.[3] According to these findings, 65% of patients with pancreatic cancer die within 6 months from the time of diagnosis, and about 90% within 1 year. Surgical resection, if performed early enough, is currently the only effective form of curative therapy.<sup>[4]</sup> However, fewer than 15% of patients with pancreatic cancer are potential candidates for a curative resection<sup>[5]</sup> because of spread of the cancer to adjacent tissues or beyond.<sup>[6]</sup> Only 1 to 4% of patients with adenocarcinoma of the pancreas will survive 5 years after diagnosis.<sup>[7,8]</sup> Thus the incidence rates are virtually identical to mortality rates.

Approximately half of all patients with pancreatic cancer have metastatic disease at the time of diagnosis, [9,10] and most of the rest have locally advanced, unresectable disease. [11,12] Metastatic pancreatic cancer is one of the most chemotherapyresistant tumours, as evidenced by the fact that pancreatic cancer has the lowest 5-year survival rate (3%) of any cancer listed in the Surveillance, Epidemiology, and End Results (SEER) database of the National Cancer Institute. [13]

Computed tomography (CT) and magnetic resonance imaging (MRI) have made it easier to determine the diagnosis and to define the stage of the disease. CT guided needle biopsies and laparoscopy have resulted in fewer unnecessary laparoto-

mies, while biliary stenting can reduce the need for an invasive operative procedure in patients with advanced tumours. Unfortunately, these advances have not resulted in disease detection at an earlier stage. [14]

Reports of 5-year survival among patients managed with nonsurgical therapies remain anecdotal. Thus, of 150 patients who have survived for more than 10 years after their diagnosis of pancreatic cancer, only 12 have been cured by nonsurgical therapies.<sup>[3]</sup> Clearly, more effective therapies need to be developed.

#### 1.1 Risk Factors

A number of factors that may contribute to the pathogenesis of pancreatic cancer have recently been identified. These have been classified as environmental factors, pathological factors (e.g. chronic pancreatitis), genetic factors (e.g. familial pancreatic cancer) and occupational exposure. [15,16]

Currently, cigarette smoking is the most firmly established risk factor associated with pancreatic cancer. Pancreatic malignancies can be induced in animals through long term administration of tobaccospecific N-nitrosamines or by parenteral administration of other N-nitroso compounds. [17-19] Induction of pancreatic cancer in these experimental models can be influenced by additional factors, including changes in bile acid composition, cholecystokinin levels, diet, and pancreatic duct obstruction. [20-23]

Clinically, numerous case-control and cohort studies have reported an increased risk of pancreatic cancer for smokers in both the US and Europe, and current estimates suggest that approximately 30% of pancreatic cancer cases may be attributed to cigarette smoking.<sup>[24,25]</sup>

# 1.2 Pathology

Most malignant pancreatic tumours (95%) are believed to arise from the exocrine portion of the gland and have light microscopic features consistent with those of adenocarcinomas. Much more infrequent are tumours that arise from acinar cells or islet cells. Primary non-epithelial tumours of the pancreas (e.g. lymphomas or sarcomas) are exceedingly rare.

# 1.3 Natural History

Adenocarcinoma of the pancreas metastasises to regional lymph nodes at an early stage of the disease, and subclinical liver metastases are present in the majority of patients at the time of diagnosis, even though findings from imaging studies may be otherwise normal. Patients who undergo surgical resection for localised non-metastatic cancer of the head of the pancreas have a long term survival rate of approximately 20% and a median survival of 15 to 19 months. However, disease recurrence following a potentially curative Whipple resection is the norm.

Local recurrence occurs in up to 85% of patients who undergo surgery alone, and local-regional tumour control may be improved by combined modality therapy involving both chemoradiation and surgery. Liver metastases then become the dominant form of tumour recurrence and occur in 50 to 70% of patients after potentially curative combined modality treatment.

Patients with locally advanced, non-metastatic disease have a median survival of 6 to 10 months, whereas those with metastatic disease have a short survival (3 to 6 months), the length of which depends on the extent of disease and performance status.

Because of the prognosis and the patterns of treatment failure associated with adenocarcinoma of the pancreas, any proposed treatment must not be worse than the disease. The low cure rate and modest median survival following a Whipple's resection mandate that treatment-related morbidity must be low and treatment-related death be rare. A recent report of the experience from Johns Hopkins University, Baltimore, Maryland, USA, demonstrates that this can be achieved by careful selection of patients who undergo therapy.<sup>[26]</sup> In addition, however, the development of innovative treatment

strategies directed at the known sites of tumour recurrence should be focused on improvements in patient survival and quality of life.

# 2. Historical Overview of Pancreatic Cancer Therapies

# 2.1 Phase II Trials

Single agent phase II trials in patients with advanced pancreatic cancer reported large variations in response rates. [27,28] In evaluating these studies, it is important to realise that clinical trial methodology and the criteria for judging objective response have changed with time. [27] Phase II trials in the 1970s often included patients with a variety of different tumours in a single trial, and therefore the published response rates were frequently based on a small number of patients with a particular cancer. As a result, these studies were difficult to interpret from a statistical point of view.

Prior to 1985, trials relied primarily on an estimation of tumour size by physical examination and responses were defined as shrinkage of a palpable abdominal mass by 50% or more, or a reduction in the palpable liver span by 30% or more. [28] The inherent inaccuracy of these techniques, intra-observer and inter-observer variability, and the influence of confounding factors on the size of the measured lesions all contributed to the initial reports of high response rates for drugs such as fluorouracil (5-FU), chlorambucil and mitomycin, as well as the failure to confirm these promising response rates in subsequent trials, especially when CT scans were used to determine tumour response. [27]

### 2.2 Phase III Trials

Two kinds of comparative studies have been carried out in patients with advanced pancreatic cancer: those that compared active treatment with best supportive care (to determine whether chemotherapy made any difference in the outcome of these patients); and those that compared multi-agent regimens with single agent chemotherapy (to determine whether combinations of drugs with distinct mechanisms of action could improve outcome com-

pared with that achieved by single agents).<sup>[27]</sup> Of the three trials that compared active treatment with best supportive care, two demonstrated no significant difference, <sup>[29,30]</sup> and the third suggested a substantial survival advantage in favour of a 5-drug regimen. <sup>[31]</sup> However, subsequent trials of the regimen failed to duplicate the promising results of the original study. <sup>[32]</sup>

Despite a number of potentially interesting preclinical findings and promising phase II studies, virtually no progress has been made in the chemotherapy for advanced pancreatic cancer during the past 30 years.

# 2.3 Current Approach to Single Agent Chemotherapy

The thymidylate synthase inhibitor fluorouracil remains the most extensively evaluated chemotherapeutic agent for pancreatic cancer. [33-35] Despite numerous trials, however, its efficacy remains questionable.

Between 1991 and 1994, 25 investigational new drugs were evaluated in phase II trials for the treatment of pancreatic cancer. The median response rate in these trials was 0% (range 0 to 14%) and the median survival was 3 months.<sup>[28]</sup> Inactive drugs that have undergone evaluation over the past 5 years include iproplatin,<sup>[36]</sup> trimetrexate,<sup>[37]</sup> edatrexate,<sup>[38]</sup> fazarabine,<sup>[39]</sup> diaziquone,<sup>[40]</sup> mitoguazone,<sup>[40]</sup> and amonafide.<sup>[41]</sup> One trial conducted during this period focused on gemcitabine (2',2'-difluoro-2'-deoxycytidine).<sup>[42]</sup>

Gemcitabine is a deoxycytidine analogue with structural similarities to cytarabine. As a prodrug, gemcitabine must be phosphorylated to its active metabolites, gemcitabine diphosphate and gemcitabine triphosphate. In both preclinical and clinical testing, gemcitabine demonstrated greater activity against solid tumours than did cytarabine. These observations have been explained by the following properties of gemcitabine: (i) it is 3 to 4 times more lipophilic than cytarabine, resulting in greater membrane permeability and cellular uptake; (ii) it has higher affinity for deoxycytidine kinase; and

(iii) the intracellular retention of gemcitabine triphosphate, an active metabolite, is prolonged.<sup>[43]</sup>

Following a phase I study, [44] gemcitabine was evaluated in a multicentre trial of 44 patients with advanced pancreatic cancer. [45] Although the objective response rate to this drug was only 11% and median survival was 5.6 months, a number of potentially important observations were made in this trial. The 1-year survival rate was a remarkably high 23%, and the responses observed appeared to be somewhat prolonged, i.e. from 4 to more than 20 months. However, perhaps the most unexpected outcome of this study was the impact of gemcitabine on tumour-related symptoms.

160 patients with newly diagnosed, unresectable pancreatic cancer were recruited in a phase III trial. [46] A total of 126 patients completed a period during which pain was stabilised and then they were randomised to treatment with gemcitabine or fluorouracil. 15 patients (23.8%) treated with gemcitabine achieved a clinically beneficial response compared with only 3 patients (4.8%) treated with fluorouracil (p = 0.002). The median duration of the clinically beneficial response for gemcitabinetreated patients was 18 weeks compared with 13 weeks for the fluorouracil-treated patients. Gemcitabine also proved superior to fluorouracil in terms of the trial's secondary end-points. Median survival for gemcitabine-treated patients was 5.6 months compared with 4.4 months for fluorouracil-treated patients. In addition, the probability of survival at 1 year was 18% in the gemcitabine group, significantly greater than the 2% in the fluorouracil group. Few objective responses, however, were observed in either treatment arm.

A subsequent phase II study enrolled 63 patients with pancreatic cancer that had progressed despite treatment with fluorouracil. [47] To be eligible for the trial, patients had to have a significant degree of tumour-related symptoms. In this study, 17 patients (27%) experienced a clinically beneficial response to gemcitabine, the median duration of which was 14 weeks. Median survival of all patients treated in this trial was 3.8 months. Objective responses were

seen in 6 (10.5%) of the 57 patients with measurable disease.

While these results could suggest that gemcitabine should become the accepted first-line therapy for patients with advanced pancreatic adenocarcinoma, the median survival for patients with metastatic disease was still less than 6 months, with few patients achieving long term disease stabilisation. Furthermore, some of the effects attributed to chemotherapy may not be substantially different from what can be achieved with aggressive supportive care alone. In fact the use of clinical benefit response as a valid means to determine the efficacy of gemcitabine has itself been questioned. Thus 85% did not survive 7 months, and 8 patients had extension to regional organs or lymph nodes without distant metastases.

The quality of life one seeks to evaluate by defining net patient benefit must take into account the duration of remaining survival available to the patient. Gelber<sup>[48]</sup> has suggested that performing treatment comparisons based on the amount of time patients spend in clinical health states characterised by relatively good quality of life might be a better indicator of net patient benefit than defining a percentage of patients who achieve some criterion of response. The fact that treatments which produce higher response rates do not always yield better survival also argues against putting too much emphasis in estimating response rates as a guide to net benefit.

17 patients had a clinically beneficial response in the phase II study, and the investigators claimed that the treatment was generally well tolerated. [47] However, although the toxicities were reported as moderate, more patients had some noticeable adverse experiences than achieved a clinical benefit response. The evidence for substantial benefit for gemcitabine is not overwhelming, and additional studies are required to more fully define its role in the treatment of pancreatic cancer.

Preclinical studies have also shown enhanced radiosensitivity of gemcitabine-treated pancreatic cell lines, [49] and several combination therapies with gemcitabine are in phase I and II trials. The

initial reports of gemcitabine as a radiosensitiser have been notable for marked toxicity at relatively low dose levels. Hoffman et al. [50] treated patients with localised, resectable pancreatic cancer with combined gemcitabine and radiation therapy before resection. Eight patients required hospitalisation (5 for cholangitis, 1 for pulmonary embolus, 1 for liver abscess, and 1 for gastrointestinal haemorrhage from a perforated gastric ulcer). Wolff et al.[51] are conducting an ongoing phase I study of gemcitabine and radiotherapy in patients with locally advanced pancreatic cancer. Gemcitabine was administered weekly at a starting dose of 400 mg/ m<sup>2</sup> with 300 cGy/day radiation. At 400 mg/m<sup>2</sup> gemcitabine, 4 of 9 patients required hospitalisation for treatment-related toxicity. At 500 mg/m<sup>2</sup> gemcitabine, 3 of 3 patients required hospitalisation for nausea, vomiting and dehydration. The appropriate phase II dose has not been established.

In summary, despite a rather meagre effect on survival, gemcitabine remains the only chemotherapeutic agent at this time that can be considered as first-line therapy for this disease.

# 3. Emerging Approaches to Pancreatic Cancer

Despite advances in our understanding of the molecular biology of pancreatic cancer, the systemic treatment of metastatic disease remains unsatisfactory. Systemic chemotherapy and the administration of biologically active molecules such as tumour necrosis factor or interferons have not resulted in significant improvements in response rates or patient survival. [52,53]

A number of more general areas of investigation may yield more promising results. One of these involves interruption or modulation of growth factors and signal transduction pathways. One example is the successful treatment of carcinoma of the breast that has been achieved by endocrine manipulation. The presence of estrogen receptors on neoplastic breast tissue is correlated with response to ovarian ablation and/or antiestrogen treatment. A similar approach to the treatment of pancreatic cancer seems justified because of the presence of

estrogen receptors in pancreatic carcinoma<sup>[54-57]</sup> as well as in normal pancreatic tissue.<sup>[58,59]</sup> In fact, the use of tamoxifen in 80 patients with ductal adenocarcinoma of pancreas has been reported in a casecontrol study to increase the median survival time from 3 months to 7 months.<sup>[60,61]</sup> However, steroid hormones may not be the most important regulator of pancreatic cell proliferation. Other potential influences include insulin-like growth factor (IGF)-1 and the growth inhibitor somatostatin.

# 3.1 Biological Basis for Somatostatin-Based Therapies

Somatostatin is a tetradecapeptide that elicits a variety of biological processes including inhibition of hormonal secretion and cell proliferation. [62] In some patients, analogue therapy leads to an inhibition of tumour growth. [63-65] However, the use of native somatostatin is limited because of its very short plasma half-life and the need for continuous infusion. The recent development of long-acting somatostatin analogues, such as vapreotide (RC-160) and octreotide (SMS-201995), however, has made clinical trials possible.

These properties of somatostatin form the basis for the treatment of hormone-producing pituitary or gastroenteropancreatic tumours by long-acting analogues of the native hormone. [63] Thus, hormonal suppression is produced in patients with acromegaly or with neuroendocrine tumours such as insulinoma, glucagonoma, gastrinoma, vipoma (diarrhoeogenic tumour) or carcinoid syndrome by somatostatin analogues, resulting in symptomatic relief. [63]

Somatostatin can exert an antiproliferative effect either by indirectly inhibiting angiogenesis or hormone and growth factor release, or by acting directly on neoplastic cells. [62-65] For example, a number of gastrointestinal hormones, including gastrin and cholecystokinin, have trophic effects on pancreatic tissue [66] and can stimulate the growth of pancreatic tumours. Somatostatin suppresses the secretion and action of these peptides, and this may also contribute to its antiproliferative activity. [67,68] In addition, somatostatin and its analogues may act

by reducing levels of growth factors such as epidermal growth factor (EGF) and IGF-1, which are thought to be important in neoplastic processes.<sup>[69-73]</sup> This latter possibility is of considerable interest because both tamoxifen<sup>[73]</sup> and octreotide<sup>[74]</sup> have been shown to lower circulating levels of IGF-1, and the combination has recently been reported to lower IGF-1 levels more substantially than either agent alone.<sup>[75]</sup> This observation raises the possibility of therapeutic synergy if the two agents were used together, a suggestion which is supported by a report that tamoxifen and octreotide were effective treatment for human pancreatic cancers growing in nude mice.<sup>[76]</sup> However, appealing as this suggestion is, Klijn et al.<sup>[77]</sup> measured insulin, IGF-1 and EGF levels in a clinical study of the use of octreotide for pancreatic cancer. Long term treatment with octreotide had no effect on EGF levels, and although these investigators observed early significant decreases in insulin and IGF-1, the levels of both growth factors had returned to pretreatment values by 5 days and 4 weeks, respectively. This may have been due to the down-regulation of the receptors responsible for inhibiting the release of these trophic factors. This study does not support suppression of trophic peptides as an important mechanism by which somatostatin inhibits pancreatic cancer, but it does not eliminate the possibility that these hormones may influence pancreatic tumour growth.

The direct actions of somatostatin are mediated by specific receptor, <sup>[62,63,78-88]</sup> and the antiproliferative action of somatostatin and its analogues on pancreatic cancer, in particular, has been demonstrated both *in vitro* and *in vivo*. <sup>[65,89]</sup> This being said, though, the inhibition of tumour growth by somatostatin and its analogues is rather complex and many gaps remain in our knowledge. <sup>[78,82-86,88,90-93]</sup>

Only a few studies have so far addressed the potential benefit of combined treatment with octreotide and various chemotherapeutic agents. Lamberts et al. [94] combined octreotide with vincristine, methotrexate, fluorouracil or suramin sodium and found an additive interaction.

More recently, Weckbecker et al.[95] demonstrated the inhibitory effect of octreotide in combination with the chemotherapeutic agents paclitaxel, fluorouracil, doxorubicin and mitomycin on the growth of AR42J pancreatic cancer cells in vitro. The dose-dependent antiproliferative effects of mitomycin, doxorubicin and paclitaxel were synergistically enhanced by octreotide. Combinations of octreotide and fluorouracil resulted either in additive or, at high concentrations of the chemotherapeutic agent, in synergistic interactions. Similar effects were observed for a cytotoxic analogue of somatostatin containing methotrexate. [96] These experiments suggest a modulatory role for octreotide in combination with widely used anticancer drugs. The additive to synergistic interaction of octreotide with these chemotherapeutic agents in in vitro and in vivo models warrants clinical studies to explore the potential of such combinations in the treatment of pancreatic cancer.

# 3.1.1 Clinical Studies of Somatostatin and its Analogues

Preliminary results of somatostatin analogue therapy in patients with cancer other than pancreatic have been encouraging.[97-100] The rationale for the use of somatostatin and its analogues can be briefly summarised as follows. Somatostatin decreases growth of the normal pancreas, [101] and at least one somatostatin analogue, octreotide, inhibits the growth of human pancreatic adenocarcinoma in nude mice.[89] The demonstration of somatostatin receptors in exocrine pancreatic adenocarcinomas<sup>[102]</sup> and the large body of evidence[76,102-108] demonstrating antiproliferative activity of somatostatin and somatostatin analogues on experimental pancreatic neoplasms in vitro and in vivo justify clinical studies on the potential therapeutic role of these drugs in pancreatic cancer.

The first clinical study on somatostatin analogue treatment was published by Klijn et al.<sup>[109]</sup> This group went on to treat 14 patients who had metastatic pancreatic cancer with 3 daily subcutaneous injections of octreotide (100 to 200µg per injection) for an average of 7 weeks and observed no antitumour effect.<sup>[77]</sup>

Friess et al.[110,111] and Ebert et al.,[112] using octreotide at a low dose level  $(3 \times 100 \text{ to } 200 \,\mu\text{g/day})$ and a high dose level  $(3 \times 2000 \,\mu\text{g/day})$  suggested that the effects of this somatostatin analogue were dose dependent. These observations are in accord with the dependent relationship of somatostatin analogues on the proliferation in breast cancer cell lines.[113] In the Friess high dose study, the median survival increased from 4 to 6 months with symptomatic and clinical improvement. A positive effect on the course of disease was confirmed by a randomised trial of octreotide versus best supportive care<sup>[114]</sup> based on a low dose (300 to 600 µg/day) therapy given 5 days per week. In this trial a significant advantage in duration of survival and in percentage of stable disease was observed for the octreotide-treated patients although no objective response was reached.

The only objective response reported for somatostatin analogues in pancreatic cancer was observed by Canobbio et al., [115] who administered lanreotide (BIM-23014) in dosages between 250 and 1000 µg/day to 18 evaluable patients. However, only one partial response was observed at the highest dose level. Huguier et al., [116] in a randomised prospective study of 86 patients who were given a similar treatment regimen, demonstrated no significant increase in median survival rates using life-table analysis.

Weckbecker et al.[117] has demonstrated potentiation of the anticancer effect of tamoxifen by concomitant infusion of high dose octreotide in the rat DMBA mammary cancer model. To further define a possible beneficial role for combination hormonal therapy with octreotide and tamoxifen, Rosenberg et al.[118] studied the effect of long term administration of these two inhibitory agents on survival of a prospective series of 12 consecutive patients with biopsy-proven ductal adenocarcinoma of the pancreas followed up from 1990 to 1993. Five of these patients had resectable disease, and in the remaining 7 disease was deemed unresectable. Treatment consisted of octreotide 100µg subcutaneously three times daily and tamoxifen 10mg orally twice daily. The major outcome measured was the

median duration of patient survival in months from the time of diagnosis. The outcome of this group of patients was compared with that of a cohort of 68 patients with a biopsy-documented diagnosis of ductal adenocarcinoma of the pancreas treated between 1985 and 1990.

The median survival times of the octreotide/ tamoxifen-treated group and the historical cohort were 12 months and 3 months, respectively, and the 1-year actuarial survival was 59% and 16%, respectively. The median survival times of the 5 resected and the 7 unresected patients were, respectively, 20 and 12 months and the 1-year actuarial survival was 80% and 31%. The median survival times of the resected octreotide/tamoxifen-treated group and the resected historical cohort were 20 and 12 months, respectively, and the 1-year actuarial survival was 80% and 44%. The median survival times of the 7 patients in the unresected octreotide/tamoxifen-treated group and the 59 in the unresected historical cohort were 12 and 2.5 months, respectively, and the 1-year actuarial survival was 31% and 11%. Cox's proportional hazards analysis confirmed that treatment and resection both independently predicted a longer survival. The significance of a possible interaction between treatment and resection could not be fully determined because of the small sample size. CT scanning examinations of the 12 patients were performed at intervals of 6 to 8 weeks, and although no objective responses were seen in terms of tumour regression, the data were consistent with a slowing of tumour progression.

The most recent report is the phase II study by Fazeny et al., [119] which was designed to investigate the efficacy and toxicity of octreotide combined with goserelin in patients with advanced pancreatic cancer. Octreotide was injected subcutaneously in dosages increasing weekly, starting with 50µg twice daily, until the level of maintenance therapy of 500µg three times daily was reached. In addition, 3.8mg goserelin was administered subcutaneously at monthly intervals. A median of 7 cycles (range 1 to 27) were applied. In comparison to the 40% of patients who had no change in their disease while

on high dose octreotide, octreotide 500µg three times daily in combination with goserelin resulted in one partial response and no disease progression in 70% of participants. The observations suggest that combining octreotide with a gonadorelin (luteinising hormone-releasing factor) analogue might be of therapeutic benefit in patients with pancreatic cancer and could compensate for the potential advantage of a higher dose of octreotide. Overall, however, the regimen under investigation did not meet the criteria for sufficient antitumour effectiveness. Nevertheless, this study reinforces the concept that pancreatic cancer is in principle responsive to endocrine therapy and, therefore, further investigation of hormonal manipulation seems worth while in the future.

In summary, while it is clear from the numerous studies conducted on experimental neoplasms *in vitro* and *in vivo* that somatostatin analogues inhibit growth of exocrine pancreatic cancers, clinical studies have demonstrated that somatostatin analogue therapy probably does not produce an adequate clinical response in patients with advanced pancreatic cancer.

# 3.2 Other Hormonal Strategies

Testosterone may also have a positive effect on the growth of pancreatic carcinoma. The concept is supported by the presence of androgen receptors within human pancreatic cancer tissue<sup>[120]</sup> together with the enzymes aromatase and 5α-reductase, which, respectively, convert testosterone into estradiol or a more active androgen, dihydrotestosterone.[121] Confirmatory evidence for a central role of testosterone came with the demonstration of its growth potentiating action on human pancreatic adenocarcinoma xenografts grown in nude mice, together with the inhibiting action of antiandrogen.[122] To assess whether flutamide, a pure androgen receptor blocking agent, improved survival in patients with pancreatic cancer, a prospective, randomised, double-blind, placebo-controlled trial was conducted.[123] 24 patients received flutamide and 25 received placebo. Analysis of patients at 6 months and 1 year demonstrated 88% and 50% survival, respectively, in the flutamide group compared with 50% and 5% in the placebo-control group. Median survival for all patients was 8 months in the flutamide group and 4 months in the placebo group. Therefore, this early study supports the concept that testosterone is a growth factor for pancreatic carcinoma and that blockade of androgen receptors offers a potentially new approach to treatment.

# 3.3 Farnesyltransferase Inhibitors

Beyond conventional chemotherapy, efforts are focusing on systemic therapies based on a rational approach to the biological properties of the pancreatic cancer cell itself. The *RAS* oncogene and the proteins encoded by it have been the subject of considerable research during the past 10 years. RAS proteins are membrane-bound guanosine triphosphate-binding proteins that act as molecular switches in mitogenic signal transduction. Mutations of the *RAS* gene lead to proteins that are permanently in the active state. These mutations occur in approximately 90% of pancreatic cancers. [124]

RAS proteins require a post-translational addition of a 15-carbon farnesyl group to become attached to the cellular membrane. If this farnesyl addition is blocked, the protein cannot attach to the membrane and remains inactive.[125] Farnesyltransferase inhibitors can block the addition of the farnesyl group and prevent membrane attachment of the RAS protein, leaving it inactive. Various farnesyl transferase inhibitors have been synthesised and found to have in vitro and in vivo activity. Farnesyltransferase inhibitors entered the clinic for phase I testing in 1998, and phase II and III studies are planned in the near future. These agents are generally given continuously by mouth and have minimal toxicity. Pancreatic cancers will be a prime target for this new class of agent because of the high incidence of RAS mutation.

### 3.4 Cell Cycle Inhibitory Factors

Another target for anticancer therapy is the cell cycling machinery itself. [126] The antiproliferative activity of a new retinoid, mofarotene (RO-408757), on 9 pancreatic cancer cell lines has been examined

for its effects on various cell cycle-regulating factors, including cyclins D1, E and A, cyclin-dependent kinases (2 and 4), cyclin-dependent kinase inhibitors (p21 and p27) and retinoblastoma protein. Mofarotene showed half-maximal inhibition of cell proliferation at concentrations that produced little cytotoxicity. A marked increase in the fraction of cells in G1 phase of the cell cycle was observed in association with up-regulation of p21/p27 and a shift of retinoblastoma protein into the hypophosphorylated form. Therefore, mofarotene and other similar molecules could represent a new approach to pancreatic cancer therapy.

# 3.5 Gene Therapy

Novel gene therapy strategies that also target cell cycle progression are under development. One of the more promising approaches is that reported by Joshi et al. [127] and is based on the p21WAF-1 gene and an adenovirus vector. Dergham et al.[128] have shown a significant survival advantage in patients treated with conventional therapies whose tumours expressed p21WAF-1. The p21WAF-1 protein is an important inhibitor of cell-cycle progression. Preliminary studies showed a significant dose-dependent increase in p21WAF-1 protein expression in pancreatic cancer cell lines infected with a recombinant adenovirus-p21WAF-1 construct (rAD-p21), with concomitant cell-growth arrest at G0/G1.[129] Use of the rAD-p21 construct also resulted in significant growth inhibition of pancreatic cell lines in tissue culture.[129] The same group has gone on to conduct preclinical trials of rAD-p21 using a model of human pancreatic adenocarcinoma implanted in the pancreas of SCID (severe combined immunodeficiency) mice.[130] Daily intratumoral injection significantly reduced tumour mass when compared with injections of phosphate-buffered saline or adenovirus alone.

However, there are drawbacks to the use of such viral vector constructs, including the low transduction rate and the induction of a severe host immune response caused by high titres of adenoviral vectors. [131] Replication-restricted viruses, such as G207, may offer an alternative to adenovirus-

retrovirus-based transduction vectors. Replication restriction denotes a viral gene vector that is capable of replication only within a desired cell type, such as cancer cells. G207 is an attenuated and multimutated form of herpes simplex virus (HSV) type 1.[132,133] The specific mutations ensure viral replication specifically in targeted cells, thereby making it ideal for cancer therapy. Lee at al.[131] have recently shown that G207 exhibited a dose-dependent cytotoxicity against three human pancreatic cell lines, AsPC-1, MIAPaCa-2 and BxPC-3. It is also important to note that following cell infection, expression of early viral genes is supported. This opens up the possibility of introducing different genes into cells by inserting such genes into the nonessential region of the HSV early genome, which will present opportunities for more specific gene therapies. Furthermore, in the study by Lee et al., viral growth was supported by each of the cell lines tested.<sup>[131]</sup> As for *in vivo* application, support for viral production eliminates the need for repeated cell transduction by retrovirus vectors carrying the HS-tk gene, for example, a well known 'suicide' gene.[134]

At this point, the host range of replication restriction for G207 has not been defined in humans. Further therapeutic evaluation of this promising viral vector seems warranted.

# 3.6 Vaccines

Immunotherapy as a potential alternative systemic treatment for adenocarcinoma of the pancreas has recently been reviewed by Yeo. [135] The advantage of this approach over radiation and chemotherapy is that it can act specifically against the tumour, without causing damage to normal tissues. Vaccines are one form of immunotherapy, which can also provide active immunisation that allows amplification of the immune response. In addition, vaccines can generate a memory immune response. Recent advances in the understanding of the mechanisms of immune system activation have revealed that any cellular protein (expressed in virally infected cells or cancer cells, including pancreatic cancer cells) can be recognised by the immune sys-

tem if those proteins are presented to the immune system in a form that results in activation rather than ignorance or tolerance to that antigen. In addition, T cells rather than B cells are responsible for this recognition.

There are generally two ways in which pancreatic and other tumour cells are currently being genetically modified to more efficiently present their tumour antigens to the immune system, resulting in potent activation of a systemic antitumour immune response. In the first model, tumour cells are genetically modified to express cytokines (proteins normally expressed as paracrine factors by immune cells to orchestrate immune responses) that attract professional antigen presenting cells (APCs) such as macrophages or dendritic cells to the site of the tumour cell. These APCs have the ability to activate both helper and killer T cells. Evidence suggests that the helper T cells can significantly potentiate killer T-cell growth factors. In the second model, tumour cells are genetically modified to express costimulatory surface molecules or cytokines that can directly attract and activate killer T cells, often bypassing the helper T cell arm. Both of the above approaches have been tested in preclinical models.[82,136]

Genetically modified tumour cell vaccines that express various cytokines are already being tested in phase I studies, but for tumours other than pancreatic adenocarcinoma.[137,138] One study evaluated autologous renal tumour cells that had been genetically modified to express granulocyte-macrophage colony-stimulating factor (GM-CSF) in patients with stage 4 renal cell carcinoma.[137] Immune responses in the form of delayed type hypersensitivity reactions against autologous tumour cells after vaccination were observed in 3 of 3 patients. Interestingly, 1 of the 3 patients demonstrated an associated partial response of pulmonary metastases. These early studies have also revealed several problems that limit the feasibility of autologous tumour vaccines. First, it is technically difficult to isolate and expand in vitro autologous tumour cells for vaccine production for most histological tumour types including pancreatic cancers. Second, an autologous vaccine implies that it is individual therapy and therefore not generalisable to all patients with the same cancer. Third, it is very expensive to produce an individual vaccine for each patient.

To overcome these limitations, Jaffee et al.[139] have developed an allogeneic vaccine approach. Allogeneic vaccine cells that have been genetically modified to express GM-CSF should be feasible because GM-CSF attracts the intermediate professional APCs for the purpose of activating helper and killer T cells. Therefore, the tumour cell itself does not have to be major histocompatibility complex compatible with the host's T cells to activate an immune response. Additionally, in malignant melanoma, which is the only human tumour for which many tumour antigens have been identified, it has been shown that the majority of antigens recognised by T cells are shared antigens expressed by over 50% of other patients' tumours. Therefore, it should be possible to vaccinate patients with a histologically similar set of established tumour lines and still activate systemic antitumour immunity. This approach is currently being tested in patients with stage 1, 2 and 3 pancreatic adenocarcinoma at Johns Hopkins Hospital, Baltimore, Maryland, USA.[140]

Other more potent vaccines continue to be developed in preclinical models. These approaches use tumour-associated antigens either in the form of proteins or peptides mixed with defined adjuvants administered systemically. Antigen-based vaccines eliminate the need for the genetic manipulation of tumour cells. This simplifies the vaccine production process and should result in more generalised vaccines. In addition, antigen-based vaccines allow greater control over the amount of antigen formulated in the vaccine, which should translate into improved efficacy. In fact, some of these strategies have been demonstrated in preclinical models to be over 10-fold more potent than the whole cell vaccine approach. Several pancreatic cancerassociated antigens have been identified, including mutated K-ras and p53, reactivated carcinoembryonic antigen (CEA), the altered mucin MUC-1, and overexpressed HER-2/neu. Several of these antigens are currently undergoing phase I testing either mixed with defined antigens or pulsed directly onto autologous dendritic cells (professional APCs) that are isolated from the peripheral blood of each patient, expanded and activated *in vitro*, and then given back by adoptive transfer. Future antigenbased vaccine approaches will undoubtedly use newly identified pancreas-associated antigens that are delivered in recombinant viral vectors that also contain other immune stimulatory genes to further enhance antitumour immunity.

# 3.7 Monoclonal Antibodies

Another immunotherapy approach to the treatment of pancreatic cancer involves the use of monoclonal antibodies (MAbs). This subject has recently been reviewed extensively by Friess et al.<sup>[140]</sup>

The MAb edrecolomab (17-1A) is a murine immunoglobulin (Ig)G2a isotype, which was developed by immunising mice with the supernatant of colorectal carcinoma cell line SW1038. This MAb is immunoreactive with a variety of gastrointestinal malignancies and was originally used in clinical trials enrolling patients with colorectal, pancreatic and gastric cancers.[141] Following binding to a 37kDa glycoprotein, edrecolomab initiates antibody-dependent cellular cytotoxicity (ADCC) in the presence of monocytes or macrophages, [142] meaning that binding of the antibody to cancer cells leads to the activation of immune cells, which will destroy antibody-labelled cells. In nude mice, administration of edrecolomab inhibited the growth of human tumour and caused an increase in activated macrophages in the tumour mass.[142] In 4 clinical trials[143-147] involving 100 patients with pancreatic cancer, complete response, partial response and stable disease were reported in 1, 5 and 23 patients, respectively.

BW-494 is a murine IgG1 isotype that recognises a membrane and cytoplasmic 200kDa carbohydrate antigen which is expressed in pancreatic cancer cells.<sup>[148]</sup> The first clinical trials with BW-494 against pancreatic carcinoma were based on the following characteristics:

 the antibody shows strong binding capacity to well and moderately differentiated human pan-

creatic cancer cells *in vitro* and *in vivo*; immunohistochemical analysis demonstrated a binding sensitivity to more than 90% of pancreatic adenocarcinomas<sup>[148-150]</sup>

- the antibody mediates ADCC with human mononuclear cells against <sup>51</sup>Cr-labelled pancreatic cancer target cells<sup>[148]</sup>
- in vitro, the antibody inhibits specific functions of pancreatic cancer cells, such as endocytosis, superoxide anion generation, and the release of lysosomal enzymes<sup>[151,152]</sup>
- single injection of <sup>131</sup>I-labelled antibody leads to growth suppression of human pancreatic tumours transplanted into nude mice. <sup>[153]</sup>

Passive immunotherapy using BW-494 was carried out in 145 patients with pancreatic cancer in 2 phase I and 2 phase II trials. [154-158] In 1 of 75 patients a partial response and in 25 of 74 patients stable disease were reported. However, in a controlled, randomised trial of 61 patients after Whipple resection, comparable survival times in patients with and without BW-494 treatment led to the termination of further clinical trials with this antibody.

Clearly, treatment of pancreatic cancer using MAbs is a challenging task that must await the development of new and more potent MAbs, such as chimeric and humanised MAbs which may make the application of higher dosages tolerable and clinically feasible.[159,160] To further improve immunotherapy with MAbs, immune response modifiers such as interleukin-2, colony-stimulating factors, or cytotoxic substances coupled to antibodies will need to be considered.[161-165] Therefore, further clinical studies are necessary to investigate the efficiency of new MAbs and MAb treatment in combination with immunomodulators in patients with pancreatic cancer. One potential new development has been the demonstration of the efficacy of an antibody (trastuzumab) to the HER2/neu oncogene in prolonging survival in metastatic breast cancer in patients whose tumours overexpress HER2/ neu. This strategy has been postulated for pancreatic cancer, but it remains to be investigated.[166]

# 3.8 Matrix Metalloproteinase Inhibitors

Pancreatic cancer is characterised by local invasion of adjacent structures, perineural invasion, early metastases to lymph nodes and liver, and an intense desmoplastic stromal reaction. [167-169] The molecular and cellular processes underlying the epithelial-stromal interactions are of great importance not only for understanding these biological characteristics of the disease, but also because they may represent novel therapeutic targets. This subject has been recently reviewed by Bramhall. [170]

The ability of malignant epithelial cells and induced desmoplastic fibroblasts to degrade adjacent extracellular matrix (ECM) is considered an essential step in the processes of invasion and metastasis.[171-173] The principal component of the basement membrane is type IV collagen, providing the scaffold on which the other major components consisting of laminin and heparan sulfate proteoglycan and the minor components of the ECM are assembled.[174] Loss of basement membrane integrity in breast and colorectal cancers has been shown to be associated with an increased metastatic potential and poor prognosis. [175,176] In pancreatic cancer, there is an absence of basement membrane proteoglycans and a discontinuity or absence of basement membrane type IV collagen.[177,178]

These findings suggest that matrix metalloproteinase (MMP) activity is likely to play an important role in the malignant phenotype of pancreatic cancer.

The MMPs are a family of 17 proteolytic enzymes that share common characteristics. Each degrades at least one component of the ECM, contains a zinc ion and is secreted as a proenzyme, which is activated by cleavage of defined peptide sequences. [179] All MMPs share sequence homologies and are inhibited by specific tissue inhibitors of the metalloproteinases (TIMPs). All of the MMPs are involved in normal remodelling processes such as embryonic development and wound healing, but also play a major role in pathological processes, including tumour invasion and metastasis. [173,180]

Data from a variety of studies (reviewed by Nagakawa et al.<sup>[167]</sup>) strongly support the hypothesis that the expression of MMPs 2, 7, 9, 11 and MT-MMP1 (membrane-type MMP1) is important in the phenotype of pancreatic cancer. These studies also suggest that MMP2 expressed in pancreatic cancer by both epithelial and stromal elements may be activated by the MT-MMP1 expressed predominantly by malignant pancreatic epithelial cells. These findings are added to by the possibility that reduced expression of the favoured inhibitor of activated MMP2 (TIMP2) could contribute to the aggressive phenotype, desmoplastic response, and discontinuous basement membrane type TV collagen and proteoglycans seen in pancreatic cancer.

Because of the importance of MMPs in tumour invasion and metastasis, low molecular weight inhibitors of the MMPs have been developed for clinical application. The first synthetic compound to enter clinical trials in cancer patients was a broad spectrum MMP inhibitor (MMPI) batimastat (BB-94), developed by British Biotech Pharmaceuticals (Oxford, UK), which has inhibitory activity against all of the MMPs in the low nanomolar concentrations. The major drawback of batimastat was its low solubility, but further development by British Biotech has led to a related low molecular weight analogue (marimastat; BB-2516) with greater solubility, which was until recently undergoing clinical trials.

Marimastat has now been administered to more than 150 patients with inoperable pancreatic cancer in the UK and US in a phase II clinical trial. Approximately 60% of these patients had stage IV disease, with the remainder having stage II or III disease. The median survival in patients with stage IV disease treated with marimastat was 94 days, and in those with stage II/III disease median survival had not been reached at 3 months. The adverse effects from marimastat tended to be primarily musculoskeletal, and dependent on dose and duration of treatment. Final results of phase III clinical trials are not yet published.

### 3.9 New Chemotherapy

Taxanes are a new class of anticancer agents believed to exhibit toxicity by disrupting microtubular assembly. Paclitaxel was the first taxane to be widely evaluated in clinical trials. A phase II trial of paclitaxel with granulocyte colony-stimulating factor was undertaken in 35 eligible patients with advanced pancreatic cancer. [181] Stable disease was observed in some patients. There was one response documented. Docetaxel, a semisynthetic taxoid, has also been studied in trials against a wide range of solid tumours. [182] In a phase II trial, 26 patients with advanced untreated pancreatic adenocarcinoma were treated with a 1-hour infusion of docetaxel 100 mg/m². [183] Five of 26 evaluable patients (17%) had a partial response with a median response duration of 3 months.

Another new drug under investigation is rubitecan (9-nitrocamptothecin; RFS-2000).<sup>[184]</sup> This drug appears to be well tolerated and efficacious as first-line therapy for the treatment of advanced pancreatic cancer. It also shows some modest success as second-line therapy in treating patients who have failed gemcitabine therapy.

# 3.10 Angiogenesis Inhibitors

Angiogenesis, the recruitment of new blood vessels, is critical for the growth of primary tumours above 1 to 2mm in diameter and is an essential component of the metastatic pathway. These vessels provide the principal route by which tumour cells exit the primary tumour site and enter the circulation. For many tumours, the vascular density can provide a prognostic indicator of metastatic potential, with highly vascular primary tumours having a higher incidence of metastasis than poorly vascular tumours. Tumour angiogenesis is regulated by the production of angiogenic stimulators including members of the fibroblast growth factor and vascular endothelial growth factor families. In addition, tumours may activate angiogenic inhibitors such as angiostatin and endostatin which can modulate angiogenesis both at the primary site and at downstream sites of metastasis. The potential use of these and other natural and synthetic angiogenic inhibitors as anticancer drugs is currently under intense investigation and has recently been reviewed by Zetter<sup>[185]</sup> and Harris.<sup>[186]</sup> Whether

these and similar drugs will cause tumour regression has not yet been studied in patients. These approaches for advanced disease should be more successful when applied early in an adjuvant situation.

# 4. Conclusion

It should be apparent from this review of past and current approaches to pancreatic cancer that any progress against this disease will have to come from a combination of advances on two fronts. First, disease detection must be improved. Methods of screening for early disease will need to be developed in concert with the identification of 'at risk' individuals. Second, a multi-modality approach designed to attack different aspects of tumour cell biology will need to be integrated into a treatment regimen that has resectional surgery as its foundation. In this regard, the use of neoadjuvant therapy prior to surgery, and adjuvant therapy following surgery, should play increasingly important roles in the management of pancreatic cancer.

# **Acknowledgements**

L. Rosenberg is a Senior Clinician-Scientist supported by the Fonds de la Recherche en Santé du Québec (FRSQ).

#### References

- Landis S, Taylor M, Bolden S, et al. Cancer statistics, 1998. CA Cancer J Clin 1998; 48: 6-30
- Wingo PA, Tong T, Bolden S. Cancer statistics 1995. CA Cancer J Clin 1995; 45: 8-30
- 3. Gudjonsson B. Cancer of the pancreas. Cancer 1987; 60: 2284-303
- Tsuchiya R, Noda T, Harada N, et al. Collective review of small carcinomas of the pancreas. Ann Surg 1986; 203: 77-81
- DiMagno EP, Reber HA, Tempero MA. AGA technical review on the epidemiology, diagnosis and treatment of pancreatic ductal adenocarcinoma. Gastroenterology 1998; 117:1464-84
- Kelly DM, Benjamin IS. Pancreatic carcinoma. Ann Oncol 1995; 6: 19-28
- Williamson RCN. Pancreatic cancer: the greatest oncologic challenge. BMJ 1988; 296: 445-6
- Gordis L, Gold EB. Epidemiology of pancreatic cancer. World J Surg 1984; 8: 808-21
- Douglass H. Adjuvant therapy for pancreatic cancer. World J Surg 1995; 19: 170-4
- American Cancer Society. Cancer facts and figures- 1991. Atlanta (GA): American Cancer Society, 1991
- Connolly M, Dawson P, Michelassi F, et al. Survival in 1001 patients with carcinoma of the pancreas. Ann Surg 1987; 206: 366-73
- Singh S, Longmire W, Reber H. Surgical palliation for pancreatic cancer: The UCLA experience. Ann Surg 1990; 212: 132-9

- Ries LAG, Miller BA, Hankey BF, et al., editors. SEER cancer statistics review, 1973-1991: tables and graphs. NIH pub. no. 94-2789. Bethesda, MD: National Cancer Institute, 1994: 356-68
- Blackstock AW, Cox AD, Tepper JE. Treatment of pancreatic cancer: current limitations, future possibilities. Oncology 1996; 10: 301-30
- Flanders TY, Foulkes WD. Pancreatic adenocarcinoma: epidemiology and genetics. J Med Genet 1996; 33: 889-98
- Abruzzese JL. Pancreatic cancer: overview of current and future therapeutic approaches. Educational book of the American Society of Clinical Oncology, 33rd Annual Meeting, 1997: 65-70
- Rivenson A, Hoffman D, Prokopczyk B, et al. Induction of lung and exocrine pancreatic tumors in F344 rats by tobaccospecific and Areca-derived N-nitrosamines. Cancer Res 1988; 48: 6912-7
- Pour PM, Rivenson A. Induction of a mixed ductal-squamousislet cell carcinoma in a rat treated with a tobacco-specific carcinogen. Am J Pathol 1989; 134: 627-31
- Hoffman D, Rivenson A, Chung FL, et al. Nicotine-derived N-nitrosamines (TSNA) and their relevance in tobacco carcinogenesis. Crit Rev Toxicol 1991; 21: 305-11
- Ogawa T, Makino T, Mizumoto K, et al. Promoting effect of truncal vagotomy on pancreatic carcinogenesis initiated with N-nitrosobis-(2-oxopropyl) amine in Syrian golden hamsters. Carcinogenesis 1991; 12: 1227-30
- Corra S, Kazakoff K, Lawson TA, et al. Cholecystokinin inhibits DNA alkylation induced by N-nitrosobis (2-oxopropyl) amine (BOP) in hamster pancreas. Cancer Lett 1992; 62: 251-6
- 22. Hoffman D, Rivenson A, Abbi R, et al. A study of tobacco carcinogenesis: Effect of the fat content of the diet on the carcinogenic activity of 4 (methylnitros-amino)-1-(3-pyridyl)-1-butanone in F344 rats. Cancer Res 1993; 53: 2758-61
- Rosenberg L, Brown RA, Duguid WP. Development of experimental cancer in the head of the pancreas by surgical induction of tissue injury. Am J Surg 1984; 147: 146-51
- Silverman DT, Dunn JA, Hoover RN, et al. Cigarette smoking and pancreas cancer: a case-control study based on direct interviews. J Natl Cancer Inst 1994; 86: 1510-6
- La Vecchia C, Boyle P, Francesschi S, et al. Smoking and cancer with emphasis on Europe. Eur J Cancer 1991; 27: 94-104
- Yeo C, Cameron J, Lillemoe K, et al. Pancreaticoduodenectomy for cancer of the head of the pancreas. Ann Surg 1995; 221: 721-33
- Rothenberg ML. New developments in chemotherapy for patients with advanced pancreatic cancer. Oncology 1996; 10: 18-22
- Rothenberg ML, Abbruzzese JL, Moore M, et al. A rationale for expanding the endpoints for clinical trials in advanced pancreatic carcinoma. Cancer 1996; 78 (3 Suppl.): 627-32
- Andersen JR, Friss-Mollek A, Hancke S, et al. A controlled trial of combination chemotherapy with 5-FU and BCNU in pancreatic adenocarcinoma. Scand J Gastroenterol 1981; 16: 973
- Frey C, Twomey P, Keehn R, et al. Randomized study of 5-FU and CCNU in pancreatic cancer: report of the Veterans Administration Surgical Adjuvant Cancer Chemotherapy Study Group. Cancer 1981; 47: 27-32
- Mallinson CN, Rake MO, Cocking JB, et al. Chemotherapy in pancreatic cancer: results of a controlled, prospective, randomised, multicentre trial. BMJ 1980; 281: 1589-91
- Cullinan S, Moertel CG, Wieand HS, et al. A phase III trial on the therapy of advanced pancreatic cancer: evaluations of the

- Mallinson regimen and combined 5-fluorouracil, doxorubicin, and cisplatin. Cancer 1990; 65: 2207-12
- Kelsen D. The use of chemotherapy in the treatment of advanced gastric and pancreatic cancer. Semin Oncol 1994; 21: 58-66
- Bukowski RM. Role of chemotherapy in patients with adenocarcinoma of the pancreas. Adv Oncol 1995; 11: 25
- Carter SK. The integration of chemotherapy into a combined modality approach for cancer treatment. VI. Pancreatic adenocarcinoma. Cancer Treat Rev 1975; 3: 193
- Hubbard KP, Pazdur R, Ajani JA, et al. Phase II evaluation of iproplatin in patients with advanced gastric and pancreatic cancer. Am J Clin Oncol 1992; 15: 524-7
- Carlson RW, Doroshow JH, Odujinrin OO, et al. Trimetrexate in locally advanced or metastatic adenocarcinoma of the pancreas: a phase II study of the Northern California Oncology Group. Invest New Drugs 1990; 8: 387-9
- Casper ES, Schwartz GK, Johnson B, et al. Phase II trial of edatrexate in patients with advanced pancreatic cancer. Invest New Drugs 1992; 10: 313-6
- Casper ES, Schwartz GK, Kelsen DP. Phase II trial of fazarabine (arabinofuranosyl-5-azacytidine) in patients with advanced pancreatic adenocarcinoma. Invest New Drugs 1992; 10: 205-9
- Bukowski RM, Fleming TR, MacDonald JS, et al. Evaluation of combination chemotherapy and phase II agents in pancreatic adenocarcinoma: an Oncology Group study. Cancer 1993; 71: 322-5
- Linke K, Pazdur R, Abbruzzese J, et al. Phase II study of amonafide in advanced pancreatic adenocarcinoma. Invest New Drugs 1991; 9: 353-6
- 42. Hertel LW, Boder GB, Kroin JS, et al. Evaluation of the antitumor activity of gemcitabine (2',2'-difluoro-2'-deoxycytidine). Cancer Res 1990; 50: 4417-22
- Heinemann V, Hertel LW, Grindley GB, et al. Comparison of the cellular pharmacokinetics and toxicity of 2',2'-difluoro-2'-deoxycytidine and 1-β-D arabinofuranosylcytosine. Cancer 1988; 48: 4024-31
- Abbruzzese JL, Grunewald R, Weeks EA, et al. A phase I clinical, plasma and cellular pharmacology study of gemcitabine.
  J Clin Oncol 1991; 9: 491-8
- Casper ES, Green MR, Kelsen DP, et al. Phase II trial of gemcitabine (2',2'-difluoro-2'-deoxycytidine) in patients with adenocarcinoma of the pancreas. Invest New Drugs 1994; 12: 29-34
- 46. Burris III HA, Moore MJ, Andersen J, et al. Improvements in survival and clinical benefit with gemcitabine as first-line therapy for patients with advanced pancreas cancer: a randomized trial. J Clin Oncol 1997; 15: 2403-13
- Rothenberg ML, Moore MJ, Cripps MC, et al. A phase II trial of gemcitabine in patients with 5-FU-refractory pancreas cancer. Ann Oncol 1996; 7: 347-53
- 48. Gelber RD. Gemcitabine for pancreatic cancer: how hard to look for clinical benefit? An American perspective. Ann Oncol 1996; 7: 335-7
- Lawrence TS, Chang EY, Hahn T, et al. Radiosensitization of pancreatic cancer cells by 2',2'-difluoro-2'deoxycytidine. Int J Radiat Oncol Biol Phys 1996; 34: 867-72
- Hoffman JP, McGinn CJ, Szarka C, et al. A phase I study of preoperative gemcitabine with radiation therapy followed by postoperative gemcitabine for patients with localized, resectable pancreatic adenocarcinoma. Proc Am Soc Clin Oncol 1998; 17: 283

- Wolff RA, Evans DB, Gravel DM, et al. Phase I trial of gemcitabine combined with radiation for the treatment of locally advanced pancreatic adenocarcinoma. Proc Am Soc Clin Oncol 1998; 17: 283
- Abbruzzese JL, Levin B, Ajani JA, et al. A phase I trial of recombinant human interferon-gamma and recombinant human tumor necrosis factor in patients with gastrointestinal cancer. Cancer Res 1989; 49: 4057-61
- 53. Abbruzzese JL, Levin B, Ajani JA, et al. A pilot phase II trial of recombinant human interferon-gamma and recombinant human tumor necrosis factor in patients with gastrointestinal malignancies: results of a trial terminated by excessive toxicity. J Biol Response Modif 1992; 9: 522-7
- Andren-Sanderg A, Borg S, Dawiskiba I, et al. Estrogen receptors and estrogen binding protein in pancreatic cancer. Digestion 1982; 25: 12
- Berz C, Hollander C, Miller B. Endocrine responsive pancreatic carcinoma steroid binding and cytotoxicity studies in human tumor cell lines. Cancer Res 1986; 46: 2276-81
- Greenway B, Iqbal MJ, Johnson PJ, et al. Oestrogen receptor proteins in malignant and fetal pancreas. BMJ 1981; 283: 751-3
- Satake K, Yoshimoto T, Mukai R, et al. Estrogen receptors in 7,12-dimethylbenz (a) anthracene (DMBA) induced pancreatic carcinoma in rats and in human pancreatic carcinoma. Clin Oncol 1982; 8: 49-54
- Sandberg AA, Rosenthal HE. Steroid receptors in exocrine glands: the pancreas and prostate. J Steroid Biochem 1979; 11: 293-9
- 59. Pousette A, Carlstrom K, Skoldefors H, et al. Purification and partial characterization of a  $17\beta$ -estradiol-binding macromolecule in the human pancreas. Cancer Res 1982; 42: 633-7
- Theve NO, Pousette A, Carlstrom K. Adenocarcinoma of the pancreas – a hormone sensitive tumor? A preliminary report on Nolvadex treatment. Clin Oncol 1983; 9: 193-7
- Wong A, Chan A. Survival benefit of tamoxifen therapy in adenocarcinoma of pancreas. A case-control study. Cancer 1993; 71: 2200-3
- Lewin MJM. The somatostatin receptors in the GI tract. Annu Rev Physiol 1992; 54: 455-69
- Lamberts SWJ, Krenning EP, Reubi JC. The role of somatostatin and its analogs in the diagnosis and treatment of tumors. Endocr Rev 1991; 12: 450-8
- Arnold R, Benning R, Neuhaus R, et al. Gastroenteropancreatic endocrine tumors: effect of sandostatin on tumor growth. Digestion 1993; 54 Suppl. 1: 72-5
- Schally AV. Oncological applications of somatostatin analogues. Cancer Res 1988; 48: 6977-85
- Johnson LR. Effects of gastrointestinal hormones on pancreatic growth. Cancer 1981; 47: 1640-5
- 67. Comaru-Schally M, Schally AV. LH-RH agonists as adjuncts to somatostatin analogs in the treatment of pancreatic cancer. In: Lunefield B, Vickery B, editors. International symposium on Gn-RH analogues in cancer and human reproduction. Boston: Kluwer Academic Publishers, 1990: 203-10
- Konturek SJ, Bilski J, Jaworek J, et al. Comparison of somatostatin and its highly potent hexa- and octapeptide analogs on exocrine and endocrine pancreatic secretion. Proc Soc Exp Biol Med 1988; 187: 241-9
- Stoscheck CM, King Jr LE. Role of epidermal growth factor in carcinogenesis. Cancer Res 1986; 46: 1030-7
- Goustin AS, Leof EB, Shipley GS, et al. Growth factors and cancer. Cancer Res 1986; 46: 1015-29

- Korc M, Magnum BE. Recycling of epidermal growth factor in a human pancreatic carcinoma cell line. Proc Natl Acad Sci U S A 1985; 82: 6172-5
- Lamberts SWJ, Koper JW, Reubi JC. Potential role of somatostatin analogues in the treatment of cancer. Eur J Clin Invest 1987; 17: 281-7
- Pollak M, Constantino J, Polychronakos C, et al. Effect of tamoxifen on serum insulinlike growth factor I levels in stage 1 breast cancer patients. J Natl Cancer Inst 1990; 82: 1693-7
- Pollak M, Polychronakos C, Guyda H. Somatostatin analogue SMS 201-995 reduces serum IGF levels in patients with neoplasms potentially dependent on IGF-1. Anticancer Res 1989; 9: 889-91
- Huynh H, Pollak M. Enhancement of tamoxifen-induced suppression of insulin-like growth factor I gene expression and serum level by a somatostatin analogue. Biochem Biophys Res Commun 1994; 203 (1): 253-9
- Poston GJ, Townsend Jr CM, Rajaraman S, et al. Effect of somatostatin and tamoxifen on the growth of human pancreatic cancers in nude mice. Pancreas 1990; 5: 151-7
- 77. Klijn JGM, Hoff AM, Th AS, et al. Treatment of patients with metastatic pancreatic and gastrointestinal tumors with the somatostatin analogue Sandostatin: a phase II study including endocrine effects. Br J Cancer 1990; 62: 627-30
- Yamada Y, Post SR, Wang K, et al. Cloning and functional characterization of a family of human and mouse somatostatin receptors expressed in brain, gastrointestinal tract, and kidney. Proc Natl Acad Sci U S A 1992; 89: 251-5
- Yamada Y, Reisine T, Law S, et al. Somatosustin receptors, an expanding gene family cloning and functional characterization of human SSTR3, a protein coupled to adenylyl cyclase. Mol Endocrinol 1992; 6: 2136-42
- Xu Y, Song J, Bruno JF, et al. Molecular cloning and sequencing of a human somatostatin receptor, hSSTR4. Biochem Biophys Res Commun 1993; 193: 648-52
- Yamada Y, Kagimoto S, Kubota A, et al. Cloning, functional expression and pharmacological characterization of a fourth (hSSTR4) and fifth (hSSThS) human somatostatin receptor subtype. Biochem Biophys Res Commun 1993; 195: 844-52
- Bell GI, Reisine T. Molecular biology of somatostatin receptors. Trends Neurosci 1993; 16: 34-8
- Hoyar D, Bell GI, Berelowitz M, et al. Classification and nomenclature of somatostatin receptors. Trends Pharmacol Sci 1995; 16: 86-8
- 84. Buscail L, Delesque N, Esteve J-P, et al. Stimulation of tyrosine phosphatase and inhibition of cell proliferation by somatostatin analogues: mediation by human receptor subtypes SSTR1 and SSTR2. Proc Natl Acad Sci U S A 1994: 91: 2315-9
- Buscail L, Esteve J-P, Saint-Laurent N, et al. Inhibition of cell proliferation by the somatostatin analogue RC-160 is mediated by somatostatin receptor subtypes SSTR@ and SSSTR5 through different mechanisms. Proc Natl Acad Sci U S A 1995; 92: 1580-4
- Raynor K, Murphy WA, Coy DH, et al. Cloned somatostatin receptors: identification of subtype selective peptides and demonstration of high affinity linear peptides. Mol Pharmacol 1993; 43: 838-44
- O'Carroll A-M, Raynor K, Lolait SJ, et al. Characterization of cloned human somatostatin receptor SSTR5. Mol Pharmacol 1994; 46: 291-8
- Patel YC, Srikant CB. Subtype selectivity of peptide analogs for all five cloned human somatostatin receptors (hsstr 1-5). Endocrinology 1994; 135; 2814-7

- Upp JR, Olson D, Polson FJ, et al. Inhibition of growth of two human pancreatic adenocarcinomas in vivo by somatostatin analog SMS 201-995. Am J Surg 1988; 155: 29-35
- Liebow C, Reilly C, Serrano M, et al. Somatostatin analogs inhibit growth of pancreatic cancer by stimulating tyrosine phosphatase. Proc Natl Acad Sci U S A 1989; 86: 2003-7
- Fisher WE, Boros LG, O'Dorisio MS, et al. Somatostatin receptor status of pancreatic adenocarcinoma predicts response to somatostatin therapy in vitro and in vivo. Surg Forum 1995; 46: 137-40
- Fisher WE, Muscarella P, O'Dorisio TP, et al. Expression of the somatostatin receptor subtype-2 gene predicts response of human pancreatic cancer to octreotide. Surgery 1996; 120: 234-40
- Vidal C, Rauly I, Zeggari M, et al. Up-regulation of somatostatin receptors by epidermal growth factor and gastrin in pancreatic cells. Mol Pharmacol 1994; 45: 97-104
- 94. Lamberts SWJ, von Koetsveld P, Hofland LJ. The interrelationship between the antimitotic action of the somatostatin analog octreotide and that of cytostatic drugs and suramin. Int J Cancer 1991; 48: 938-41
- 95. Weckbecker G, Raulf F, Tolcsvai L, et al. Potentiation of the anti-proliferative effects of anti-cancer drugs by octreotide in vitro and in vivo. Digestion 1996; 57 Suppl. 1: 22-8
- 96. Radulovic S, Nagy A, Szoke B, et al. Cytotoxic analog of somatostatin containing methotrexate inhibits growth of MIA PaCa-2 human pancreatic xenografts in nude mice. Cancer Lett 1992; 62: 263-71
- Vennin PH, Peyret JP, Bonneterre J. Effect of the long-acting somatostatin analogue SMS 201-995 in advanced breast cancer. Anticancer Res 1989; 9: 153-6
- Guliana JM, Guillausseau PJ, Caron J, et al. Effects of shortterm subcutaneous administration of SMS 201-995 on calcitonin plasma levels in patients suffering from medullary thyroid carcinoma. Horm Metab Res 1989; 21: 584-6
- Kraenzlin ME, Ch'ng JC, Wood SM, et al. Long-term treatment of a VIPoma with somatostatin analogue resulting in remission of symptoms and possible shrinkage of metastases. Gastroenterology 1985; 88: 185-7
- Parmar H, Bogden A, Mollard M, et al. Somatostatin and somatostatin analogues in oncology. Cancer Treat Rev 1989; 16: 95-115
- Morisset J, Genik P, Lord A, et al. Effects of chronic administration of somatostatin on rat exocrine pancreas. Regul Pept 1982; 4: 49-58
- 102. Fekete M, Zalatnai A, Comura-Schally AM, et al. Membrane receptors for peptides in experimental and human pancreatic cancers. Pancreas 1989; 4: 521-8
- 103. Davies NM, Kapur P, Gillespie J, et al. Inhibitory effect of somatostatin analog RC-160 on EGF- and transforming growth factor alpha (TGF-α)-stimulated pancreatic cancer growth in vivo. Br J Cancer 1991; 64 Suppl. 15: 4
- 104. Schally AV, Srkalovic G, Szende B, et al. Anti-tumor effects of analogs of LH-RH and somatostatin: experimental and clinical studies. J Steroid Biochem Mol Biol 1990; 37: 1061-7
- 105. Szende B, Srkalovic G, Schally AV, et al. Inhibitory effects of analogs of luteinizing hormone-releasing hormone and somatostatin on pancreatic cancers in hamsters. Cancer 1990; 65: 2279-90
- 106. Paz-Bouza JR, Redding TW, Schally AV. Treatment of nitrosamine-induced pancreatic tumors in hamsters with analogues of somatostatin and luteinizing hormone-releasing hormone. Proc Natl Acad Sci U S A 1987; 84: 1112-6
- Poston GJ, Gillespie J, Guillou PJ. Biology of pancreatic cancer. Gut 1991; 32: 800-12

- 108. Szende B, Zalatnai A, Schally AV. Programmed cell death (apoptosis) in pancreatic cancer of hamsters after administration of analogs of luteinizing hormone releasing hormone and somatostatin. Proc Natl Acad Sci U S A 1989; 86: 1643-7
- 109. Klijn JGM, Setyono-Han B, Bakker GH, et al. Effects of somatostatin analog (sandostatin) treatment in experimental and human cancer. In: Klijn JGM, Paridaens R, Foekens JA, editors. Hormonal manipulation of cancer: peptides, growth factors and new (anti) steroidal agents. EORTC monograph series, Vol. 19. New York (NY): Raven Press, 1987: 459-68
- Friess H, Buchler M, Beglinger Ch, et al. Low dose octreotide treatment is not effective in patients with advanced pancreatic cancer. Pancreas 1993; 8: 540-5
- 111. Frieß H, Buchler M, Ebert M, et al. Treatment of advanced pancreatic cancer with high dose octreotide. Int J Pancreatol 1993; 14: 290-1
- 112. Ebert M, Frieβ H, Beger H, et al. Role of octreotide in the treatment of pancreatic cancer. Digestion 1994; 55 Suppl. 1: 48-51
- 113. Scambia G, Benedetti BP, Baiochhi G, et al. Antiproliferative effects of somatostatin analog SMS 201-995 on three human breast cancer cell lines. J Cancer Res Clin Oncol 1988; 144: 106-8
- 114. Cascinu S, Del Ferro E, Catalano G. A randomized trial of octreotide vs best supportive care only in advanced cancer patients refractory to chemotherapy. Br J Cancer 1995; 71: 97-101
- Canobbio L, Boccardo F, Cannata D, et al. Treatment of advanced pancreatic cancer with the somatostatin analogue BIM 23014. Cancer 1992; 69: 648-50
- 116. Huguier M, Samama G, Testart J,, et al. Treatment of adenocarcinoma of the pancreas with somatostatin and gonadoliberin (luteinizing hormone-releasing hormone). Am J Surg 1992; 164: 348-53
- 117. Weckbecker G, Tolcsvai L, Stolz B, et al. Somatostatin analogue octreotide enhances the antineoplastic effects of tamoxifen and ovariectomy on 7,12-dimethylbenz(a)anthracene-induced rat mammary carcinomas. Cancer Res 54: 6334-7
- Rosenberg L, Barkun AN, Denis MH, et al. Low-dose octreotide and tamoxifen in the treatment of adenocarcinoma of the pancreas. Cancer 1995; 75: 23-8
- 119. Fazeny B, Baur M, Prohaska M, et al. Octreotide combined with goserelin in the therapy of advanced pancreatic cancer: results of a pilot study and review of the literature. J Cancer Res Clin Oncol 1997; 123: 45-52
- Corbishley TP, Iqbal MJ, Wilkinson ML, et al. Androgen receptor in human normal and malignant pancreatic tissue and cell lines. Cancer 1986; 57: 1992-5
- 121. Iqbal MJ, Greenway BA, Wilkinson WL, et al. Sex steroid enzymes, aromatase and 5α reductase in the pancreas: a comparison of normal adult, foetal and malignant tissue. Clin Sci 1983; 65: 71-5
- 122. Greenway BA, Duke D, Pym B, et al. The control of human pancreatic adenocarcinoma xenografts in nude mice by hormone therapy. Br J Surg 1982; 69: 595-7
- 123. Greenway BA. Effect of flutamide on survival in patients with pancreatic cancer: results of a prospective, randomised, double blind, placebo controlled trial. BMJ 1998; 316: 1935-8
- Ryan DP, Fuchs C. Chemotherapy for pancreatic cancer. Adv Oncol 1998; 14: 11-9
- Gibbs JB, Oliff A, Kohl NE. Farnesyltransferase inhibitors: ras research yields a potential cancer therapeutic. Cell 1994; 77: 175-8

- 126. Kawa S, Nikaido T, Aoki Y, et al. Arotinoid mofarotene (R040-8757) up-regulates p21 and p27 during growth inhibition of pancreatic cancer cell lines. Int J Cancer 1997; 72: 906-11
- Joshhi US, Dergham ST, Chen YQ, et al. Inhibition of pancreatic tumour cell growth in culture by p21WAF1 recombinant adenovirus. Pancreas 1998; 16: 107-13
- Dergham ST, Dugan MC, Joshi US, et al. The clinical significance of p21WAF1/CIP-1 and p53 expression in pancreatic adenocarcinoma. Cancer 1997; 80: 372-81
- Joshi US, Dergham ST, Chen YQ, et al. Inhibition of pancreatic tumor cell growth in culture by p21WAF1 recombinant adenovirus. Pancreas 1998; 16: 107-13
- 130. Mohammad RM, Dugan MC, Mohamed AN, et al. Establishment of human pancreatic tumor xenograft model: potential application for preclinical evaluation of novel therapeutic agents. Pancreas 1998; 16: 19-25
- Lee JH, Federoff HJ, Schoeniger LO, et al. G207, modified herpes simplex virus type 1, kills human pancreatic cancer cells in vitro. J Gastrointest Surg 1999; 3: 127-33
- Mineta T, Rabkin SD, Yazaki T, et al. Attenuated multi-mutated herpes simplex virus-1 for the treatment of malignant gliomas. Nat Med 1995; 1: 938-43
- 133. Yazaki T, Manz HJ, Rabkins SD, et al. Treatment of human malignant meningiomas by G207, a replication-competent multimutated herpes simplex virus 1. Cancer Res 1995; 55: 4752-6
- Link Jr CJ, Levy LP, McCann LZ, et al. Gene therapy for colon cancer with the herpes simplex thymidine kinase gene. J Surg Oncol 1997; 64: 289-94
- Yeo C. Pancreatic cancer: 1998 update. J Am Coll Surg 1998; 187: 429-42
- Clary BM, Coveney EC, Blazer DG, et al. Active immunotherapy of pancreatic cancer with tumor cells genetically engineered to secrete multiple cytokines. Surgery 1996; 120: 174-81
- 137. Simons JW, Jalfee EM, Weber C, et al. Bioactivity of human GM-CSF gene transduced autologous renal vaccines. Cancer Res 1997; 57: 1537-46
- Cheson BD, Phillips PH, Sanol M. National Cancer Institute. Clinical trials. Oncology 1997; 11: 81-90
- 139. Jaffee EM, Schutte M, Gossett J, et al. Development and characterization of cytokine-secreting pancreatic adenocarcinoma vaccine from primary tumors for use in clinical trials. Cancer J Sci Am 1998; 4: 194-203
- Friess H, Gassmann M, Buchler MW. Adjuvant therapy of pancreatic cancer using monoclonal antibodies and immune response modifiers. Int J Pancreatol 1997; 11: 43-52
- Sears HF, Steplewski Z, Herlyn D, et al. Effects of monoclonal antibody immunotherapy on patients with gastrointestinal adenocarcinoma. J Biol Resp Mod 1984; 3: 138-50
- 142. Adams DO, Hall T, Steplewski Z, et al. Tumors undergoing regression induced by monoclonal antibodies of the IgG2a isotype contain increased numbers of macrophages activated for a distinctive form of antibodycytolysis. Proc Natl Acad Sci U S A 1984: 81: 3506-10
- 143. Tempero MA, Pour PM, Uchida E, et al. Monoclonal antibody CO 17-IA and 1eukopheresis in immunotherapy of pancreatic cancer. Hybridoma 1986; 5 Suppl. 1: 133-8
- 144. Sindelar WF, Maher MM, Herlyn D, et al. Trial of therapy with monoclonal antibody 17-1A in pancreatic carcinoma: preliminary results. Hybridoma 1986; 5 Suppl. 1: 125-32
- 145. Tempero MA, Sivinski CL, Steplewski Z, et al. Phase II trial of gamma interferon and monoclonal antibody 17-lA in pancreatic cancer: biological and clinical effects. J Clin Oncol 1990; 8: 2019-26

- 146. Tempero MA, Haga Y, Sivinski C, et al. Immunotherapy with monoclonal antibody (MAB) in pancreatic adenocarcinoma. Int J Pancreatol 1991; 9: 125-34
- 147. Weiner LM, Haney F, Padavic-Shaller K, et al. Phase II multicenter evaluation of prolonged murine monoclonal antibody 17-IA therapy in pancreatic carcinoma. J Immunother 1993; 13: 110-6
- 148. Bosslet K, Kern HF, Kanzy FJ, et al. A monoclonal antibody with binding and inhibiting activity towards human pancreatic carcinoma cells. I: Immunohistological and immunochemical characterization of a murine monoclonal antibody selecting for well differentiated adenocarcinomas of the pancreas. Cancer Jmmunol Immunother 1986; 23: 185-91
- 149. Kubel R, Buchler M, Baczako K, et al. Immunohistochemistry in pancreatic cancer with new monoclonal antibodies. Lang Arch Chir 1987; 371: 243-52
- Montz R, Klapdor R, Rothe B, et al. Immunoscintigraphy and radioimmunotherapy in patients with pancreatic carcinoma. Nuklearmedizin 1986; 25: 239-44
- 151. Kern HF, Bosslet K, Mollenhatter J, et al. Monocyte-related functions expressed in cell lines established from human pancreatic adenocarcinoma. I. Comparative analysis of endocytotic activity, lysosomal enzyme secretion, and superoxide anion production. Pancreas 1987; 2: 212-21
- 152. Kern HF, Bosslet K, Sedlacek HH, et al. Monocyte-related functions expressed in cell lines established from human pancreatic adenocarcinoma. II. Inhibition of stimulated activity by monoclonal antibodies reacting with surface antigens on tumor cells. Pancreas 1988; 3: 2-10
- 153. Klapdor R, Lander S, Bahlo M, et al. Radioimmunotherapy of xenografts of human pancreatic carcinomas intravenous and intratumoral application of 131I-labeled monoclonal antibodies. Nuklearmedizin 1986; 25: 235-8
- 154. Buchler M, Kubel R, Malfertheiner P, et al. Immunotherapy of advanced pancreatic carcinoma with the monoclonal antibody BW 494. Dtsch Med Wochenschr 1988; 113: 374-80
- Buchler NI, Friess H, Malfertheiner P, et al. Studies of pancreatic cancer utilizing monoclonal antibodies. Int J Pancreatol 1990; 7: 151-7
- Buchler M, Friess H, Schultheiss KH, et al. A randomized controlled trial of adjuvant immunotherapy (murine monoclonal antibody 494i32) in resectable pancreatic cancer. Cancer 1991;
  58: 1507-12
- 157. Buchler M, Kubel R, Klapdor R, et al. Immunotherapy of pancreatic cancer with monoclonal antibody BW 494: results from a multicentric phase I-II trial. In: Beger HG, Buehler M, Schulz G, et al., editors. Cancer therapy. Berlin: Springer. 1989: 3241
- 158. Friess H, Buchler M, Schulz G, et al. Therapy of pancreatic carcinoma with the monoclonal antibody BW494/32: first clinical results. Immunitat Infekt 1989; 17: 2-26
- 159. Bosslet K, Keweloh HC, Hermentin P, et al. Percolation and binding of monoclonal antibody BW494 to pancreatic carcinoma tissues during high dose immunotherapy and consequences for future therapy modalities. Br J Cancer 1990; 10: 37-9
- 160. Goodman GE, Hellstrom I, Yelton D, et al. Phase I trial of chimeric (human-mouse) monoclonal antibody L6 in patients with non-small-cell lung. colon. and breast cancer. Cancer Immunol Immunother 1993: 36: 267-73
- 161. Kushner BH, Cheung NK. GM-CSF enhances 3F8 monoclonal antibody-dependent cellular cytotoxicity against human melanoma and neuroblastoma. Blood 1989; 73: 1936-41

- 162. Senter PD. Activation of prodrugs by antibody-enzyme conjugates: a new approach to cancer therapy. FASEB J 1990: 4: 188-93
- 163. Svensson HP, Kadow JF, Vrudhula VM, et al. Monoclonal antibody-beta-lactamase conjugates for the activation of a cephalosporin mustard prodrug. Bioconjug Chem 1992; 3: 176-81
- 164. Senter PD, Wallace PM, Svensson HP, et al. Activation of prodrugs by antibody-enzyme conjugates. Adv Exp Med Biol 1991; 303: 97-105
- 165. Stalb F, Link KH, Mitchell MS. Immunomodulation of pancreatic cancer in vitro. Effect of monoclonal antibodies, GM-CSF and lymphokine-activated killer cells. Digestion 1992: 52: 124
- 166. Butera J, Malachovsky M, Rathore R, et al. Novel approaches in development for the treatment of pancreatic cancer. Front Biosci 1998; 3: E226-9
- 167. Nagakawa T, Konishi I, Higashino Y, et al. The spread and prognosis of carcinoma in the region of the pancreatic head. Jpn J Surg 1989; 19: 510-8
- 168. Nagakawa T, Kavahara M, Ueno K, et al. A clinicopathologic study on neural invasion in cancer of the pancreatic bead. Cancer 1992; 69: 930-5
- Mollenhauer J, Roether 1, Kern HF. Distribution of extracellular matrix proteins in pancreatic ductal adenocarcinoma and its influence on tumor cell proliferation in vitro. Pancreas 1987; 2: 14-24
- 170. Bramhall SR. The matrix metalloproteinases and their inhibitors in pancreatic cancer. Int J Pancreatol 1997; 21: 1-12
- 171. Hart IR, Goode NT, Wilson RE. Molecular aspects of the metastatic cascade. Biochem Biophys Acta 1989; 989: 65-84
- Liotta LA, Tryggvason K, Garbisa S, et al. Metastatic potential correlates with enzymatic degradation of basement membrane collagen. Nature 1980; 284: 67-8
- 173. Liotta LA, Stetler-Stevenson WG. Tumor invasion and metastasis: an imbalance of positive and negative regulation. Cancer Res 1991; 51 Suppl.: 5054-9s
- 174. Martinez-Hernandez A, Amenta PS. The basement membrane in pathology. Lab Invest 1983; 48: 650-77
- 175. Forster SJ, Talbot IC, Clayton DG, et al. Tumour basement membrane laminin in adenocarcinoma of rectum: an immunohistochemical study of biological and clinical significance. Int J Cancer 1986; 37: 813-7
- 176. Charpin C, Lissitzky JC, Jacquemier J, et al. Immunohistochemical detection of laminin in 98 human breast carcinomas: a light and electron microscopic study. Hum Pathol 1986; 17: 355-65
- 177. Lee CS, Montebello J, Georgiou T, et al. Distribution of type IV collagen in pancreatic adenocarcinoma and chronic pancreatitis. Int J Exp Pathol 1994; 75: 79-83
- 178. Wang Z-h, Manabe T, Ohshio C, et al. Immunohistochemical study of heparan sulfate proteoglycan in adenocarcinomas of the pancreas. Pancreas 1994; 9: 758-63
- 179. Cottam DW, Rees RC. Regulation of matrix metalloproteinases: their role in tumor invasion and metastasis (review). Int J Oncol 1993; 2: 861-72
- 180. Docherty AJ, Murphy C. The tissue metalloproteinase family and the inhibitor TIMP: a study using cDNAs and recombinant proteins. Ann Rheum Dis 1990; 49: 469-79
- 181. Brown T, Tangen C, Fleming T, et al. A phase II trial of taxol and granulocyte colony stimulating factor (G-CSF) in patients with adenocarcinoma of the pancreas. Proc Am Soc Clin Oncol 1993; 12: 200

- 182. Kaue SB. Docetaxel (taxotere) in the treatment of solid tumors other than breast and lung cancer. Semin Oncol 1995; 22(4): 30-3
- Rougier D, DeForin M, Ademis A, et al. Phase II study of taxotere in pancreatic adenocarcinoma. Proc Am Soc Clin Oncol 1994; 13: 200
- 184. Stehlin JS, Giovanella BC, Natelson EA, et al. A study of 9nitrocamptothecin (RFS-2000) in patients with advanced pancreatic cancer. Int J Oncol 1999; 14: 821-31
- 185. Zetter BR. Angiogenesis and tumor metastases. Ann Rev Med 1998; 49: 407-24
- Harris AL. Anti-angiogenesis therapy and strategies for integrating it with adjuvant therapy. Recent Results Cancer Res 1998; 152: 341-52

Correspondence and offprints: Dr *Lawrence Rosenberg*, Montreal General Hospital, 1650 Cedar Ave., L9-424, Montreal, Quebec, H3G 1A4, Canada.

E-mail: cxlw@musica.mcgill.ca