

# Current Management of Fungal Infections

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## Abstract

The management of superficial fungal infections differs significantly from the management of systemic fungal infections. Most superficial infections are treated with topical antifungal agents, the choice of agent being determined by the site and extent of the infection and by the causative organism, which is usually readily identifiable. One exception is onychomycosis, which usually requires treatment with systemically available antifungals; the accumulation of terbinafine and itraconazole in keratinous tissues makes them ideal agents for the treatment of onychomycosis. Oral candidiasis in immunocompromised patients also requires systemic treatment; oral fluconazole and itraconazole oral solution are highly effective in this setting.

Systemic fungal infections are difficult to diagnose and are usually managed with prophylaxis or empirical therapy. Fluconazole and itraconazole are widely used in chemoprophylaxis because of their favourable oral bioavailability and safety profiles. In empirical therapy, lipid-associated formulations of amphotericin-B and intravenous itraconazole are safer than, and at least as effective as, conventional amphotericin-B (the former gold standard). The high acquisition costs of the lipid-associated formulations of amphotericin-B have limited their use.

In humans, fungal infections can be classified into three broad groups: superficial, subcutaneous and systemic (see Garber<sup>[1]</sup>). Superficial infections of keratinised tissues such as nails and hair are usually by dermatophytes, whereas mucous membranes are most frequently infected by *Candida* spp. A variety of organisms cause subcutaneous infections, which are usually acquired by traumatic inoculation. The most serious life-threatening fungal infections are systemic. In some geographical locations systemic fungal infections are endemic, such as histoplasmosis in the Mississippi valley. However, immunocompromised patients worldwide are at risk of systemic infection from commensal and ubiquitous

species, primarily *Candida* spp. and *Aspergillus* spp.

The recent increase in the number of patients at risk of systemic fungal infections (patients with HIV, cancer patients receiving intensive chemotherapy)<sup>[2-4]</sup> has highlighted the need for effective therapy of fungal infections. Some superficial fungal infections can be treated or prevented with topical antifungal agents but others require systemic treatment with oral antifungals. Systemic infections are treated with orally and intravenously administered agents. Here we discuss the use and potential of the three classes of antifungal agents used most frequently – polyenes, azoles and allylamines (table I).

Table I. Selected properties of antifungal agents

Drug	Spectrum of activity	Properties/formulations
<b>Polyene macrolides</b>		
Amphotericin-B	<i>Candida</i> spp., <i>Aspergillus</i> spp. and other filamentous fungi, life-threatening infections with the endemic fungi <i>Coccidioides immitis</i> , <i>Histoplasma capsulatum</i> and <i>Blastomyces dermatitidis</i> Reports of resistance in non- <i>albicans Candida</i> spp.	Delivered topically or systemically, most frequently used in IV form for treatment of systemic fungal infections; oral tablets used mainly to treat thrush and gastrointestinal candidiasis; conventional formulations are nephrotoxic; lipid-associated formulations have reduced nephrotoxicity
Nystatin	<i>Candida</i> spp. and <i>Aspergillus</i> spp.	Oral formulation used for topical treatment of oropharyngeal candidiasis
<b>Allylamines</b>		
Terbinafine	<i>Candida</i> spp., <i>Aspergillus</i> spp., dermatophytes and dimorphic fungi	Topical and oral formulations used in treatment of cutaneous and nail dermatophyte infections; lipophilic and accumulates in skin, hair, sebum and nail plate
<b>Imidazoles</b>		
Ketoconazole	<i>Candida</i> spp., all dermatophytes, <i>Malassezia furfur</i> , <i>B. dermatitidis</i> , <i>C. immitis</i> , <i>H. capsulatum</i> , <i>Paracoccidioides brasiliensis</i> and <i>Phialophora</i> spp. NOT <i>Aspergillus</i> spp.	Oral formulations previously used in treatment of systemic infections; largely supplanted by other azole antifungals
Miconazole	<i>C. albicans</i> , <i>Pseudallescheria boydii</i> , <i>M. furfur</i> and all common dermatophytes NOT <i>Aspergillus</i> spp.	Available as topical cream, oral gel and IV formulation; parenteral form rarely used, because of toxicity
<b>Triazoles</b>		
Fluconazole	<i>Cryptococcus</i> spp., and many <i>Candida</i> spp., including <i>C. albicans</i> NOT <i>Aspergillus</i> spp. or some non- <i>albicans Candida</i> spp., e.g. <i>C. glabrata</i> and <i>C. krusei</i> Reports of resistance	Water-soluble oral capsules (or tablets), suspension or IV formulation used extensively in prophylaxis; treatment of choice for mucocutaneous candidiasis in neutropenic patients and one of the preferred treatments for oral candidiasis in patients with HIV infection
Itraconazole	<i>Candida</i> spp. <i>Aspergillus</i> spp., <i>B. dermatitidis</i> , <i>H. capsulatum</i> , <i>Cryptococcus</i> spp., <i>C. immitis</i> , <i>Trichophyton</i> spp., <i>Sporothrix schenckii</i> , <i>Penicillium marneffei</i> and <i>P. brasiliensis</i> Reports of resistance in <i>Aspergillus fumigatus</i>	Oral capsules taken with food; oral (taken before food) and IV solution have improved bioavailability; drug of choice for some rare fungal infections; valuable for prophylaxis and treatment of systemic fungal infections; lipophilic and accumulates in keratinous tissue

IV = intravenous.

1. Antifungal Agents

1.1 Polyenes

The most frequently used polyene macrolides are amphotericin-B and nystatin. These drugs act by binding to ergosterol in cell membranes (fig. 1), increasing permeability, disrupting metabolism and causing cell death.<sup>[5]</sup> Both amphotericin-B and nystatin have a broad spectrum of activity against most species of *Candida* and *Aspergillus*.

Amphotericin-B has been available for more than 40 years and exists in a variety of formulations; however, because the oral bioavailability of amphotericin-B is less than 5%,<sup>[6]</sup> the oral formu-

lations are essentially topical agents for the treatment or prevention of oral or intestinal colonisation. Intravenous formulations of amphotericin-B are used in the management of systemic fungal infections.

Nystatin also has low oral bioavailability and is generally used only as a topical treatment for thrush or in preventing colonisation with *Candida albicans* in the gut.<sup>[7]</sup>

1.2 Allylamines

The allylamine terbinafine inhibits squalene epoxidase (an enzyme involved in ergosterol synthesis; fig. 1), which results in ergosterol depletion,

membrane disruption and cell death.<sup>[8]</sup> Terbinafine has activity against dermatophytes, low activity against *Aspergillus* spp. and other filamentous fungi but limited activity against *Candida* spp. The drug is registered for topical and oral treatment of cutaneous and nail dermatophyte infections<sup>[9]</sup> and, because terbinafine is lipophilic and keratinophilic, it accumulates in sebum, hair and nails. Terbinafine has been used alone or in combination with amphotericin-B or azoles for the treatment of invasive fungal infections<sup>[10]</sup> but no controlled studies have been published.

### 1.3 Azoles

The azole antifungals are synthetic compounds that inhibit the fungal cytochrome P450 (CYP) 14 $\alpha$ -demethylase (fig. 1);<sup>[11]</sup> this results in a depletion of ergosterol and a loss of membrane integrity and activity. The azoles are divided into imidazoles and triazoles on the basis of their chemical structure and clinical effectiveness.

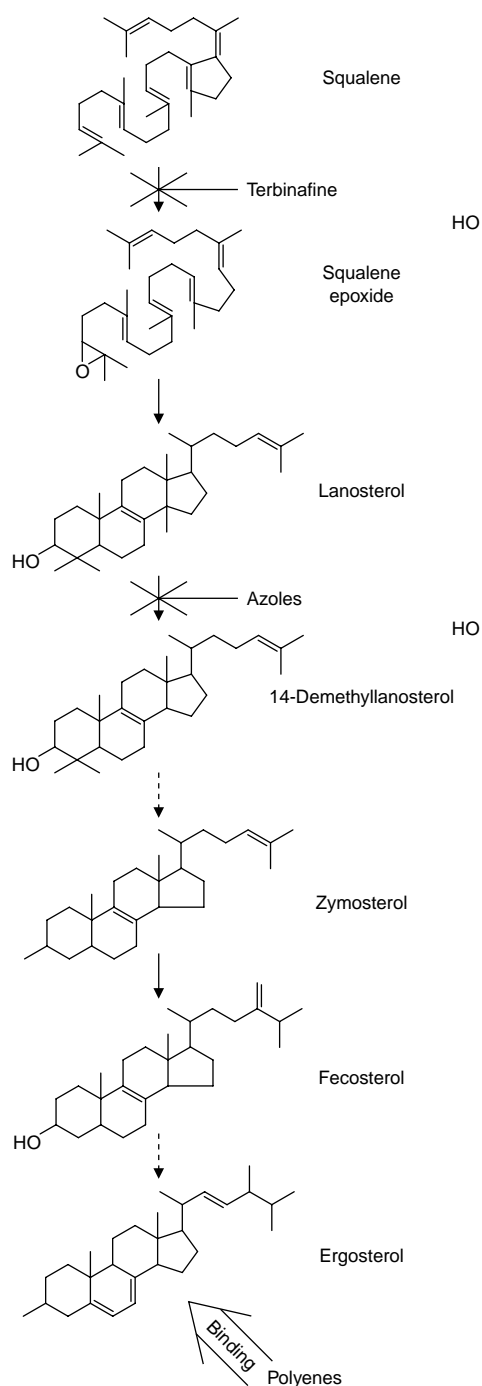
#### 1.3.1 Imidazoles

Miconazole has potent antifungal activity but is inactive against *Aspergillus* spp., and its insolubility restricts its use. Ketoconazole has a spectrum of activity similar to that of miconazole. When introduced 20 years ago, ketoconazole was the first orally absorbable antifungal azole (bioavailability approximately 75%);<sup>[12]</sup> it is now also available as a topical cream. Ketoconazole is primarily used for the topical treatment of superficial yeast and dermatophyte infections.

Miconazole and ketoconazole have been largely supplanted by the triazoles in the treatment of serious fungal infections.

#### 1.3.2 Triazoles

Itraconazole and fluconazole are the only systemic triazole antifungal agents currently available. Itraconazole has a broad spectrum of antifungal activity against *Candida* spp., *Aspergillus* spp. and dermatophytes; it accumulates in keratinous tissue, such as hair and nails, and is widely used for the treatment of fungal nail infections. In addition, itraconazole is orally absorbed and accu-



**Fig. 1.** Ergosterol synthesis pathway and points of inhibition by antifungal agents.

mulates in organs that are frequent sites for systemic fungal infections (such as the spleen and lungs),<sup>[13]</sup> and can therefore be used to treat and prevent a variety of systemic fungal infections (see Boogaerts and Maertens<sup>[14]</sup>). The bioavailability and clinical efficacy of itraconazole for systemic infections have recently been improved with the development of oral solution and intravenous (IV) formulations (see De Beule and Van Gestel<sup>[15]</sup>),<sup>[16]</sup>

Fluconazole has a narrower spectrum of antifungal activity than itraconazole; it is inactive against *Aspergillus* spp. and some species of *Candida*. Despite this, fluconazole is still used widely for the prophylaxis of systemic fungal infections and the treatment of confirmed systemic *Candida* infections in immunocompromised patients. Fluconazole is also an effective treatment for cryptococcal meningitis in patients with AIDS,<sup>[17]</sup> and is effective in preventing relapse when given as secondary prophylaxis.<sup>[18]</sup> Fluconazole has oral bioavailability of approximately 90% and can also be used to treat mucosal and cutaneous candidiasis. Because, like itraconazole, fluconazole accumulates in fingernails and toenails, it can also be used to treat onychomycosis.<sup>[19,20]</sup>

#### 1.4 Other Agents

Griseofulvin acts by binding to microtubular proteins and inhibiting fungal cell mitosis. Griseofulvin is effective only against dermatophytes and is indicated for the treatment of dermatophytoses of the skin, hair or nails when other treatments are considered inappropriate.

Flucytosine is a synthetic fluorinated pyrimidine that disrupts DNA and RNA synthesis. It has a narrow spectrum of activity and is largely used in combination with amphotericin-B for the treatment of cryptococcal meningitis or deep candidiasis.<sup>[21]</sup>

## 2. Treatment of Superficial Fungal Infections

Superficial fungal infections often produce characteristic lesions, and a combination of clinical

observation and laboratory investigation usually provides an accurate diagnosis.

### 2.1 Dermatophytosis

Fungal infections of the skin, nails and hair are usually caused by dermatophytes. Tinea corporis, tinea cruris, tinea pedis and tinea manuum are managed with administration of topical imidazoles or allylamines for 2 to 4 weeks; generally, more than 80% of patients are cured.<sup>[22-24]</sup> However, when the infections are extensive or when topical treatment fails, an oral antifungal agent is required. Treatment for 1 week with itraconazole capsules 200 to 400 mg/day or for 2 to 4 weeks with terbinafine 250 mg/day is usually effective.<sup>[25-30]</sup> The choice of first-line treatment for tinea capitis is less well defined; oral itraconazole capsules or terbinafine may be preferred to topical imidazoles. In a recent small trial, week-12 cure rates were 86% and 78% with itraconazole capsules and terbinafine, respectively.<sup>[31]</sup>

Most cases of tinea unguium and other fungal infections of the nails (onychomycosis) can be treated effectively only with the oral administration of itraconazole, terbinafine or to a lesser extent fluconazole.<sup>[32]</sup> Terbinafine 250 mg/day is usually administered for 6 or 12 weeks for the treatment of fingernail or toenail onychomycosis, respectively. These regimens provide mycological cure rates of 88% in fingernails and 82% in toenails.<sup>[33-35]</sup>

Therapeutic concentrations of itraconazole can be detected in the nail 6 to 9 months after therapy is stopped.<sup>[36]</sup> This has led to the development of pulse therapy, in which itraconazole capsules 400 mg/day are administered for only 1 week each month, repeated twice for fingernail onychomycosis and 3 times for toenail onychomycosis. The advantages of a pulse therapy regimen include reduced costs and improved compliance.<sup>[37]</sup> Comparative trials of continuous terbinafine therapy and itraconazole pulse therapy have given contradictory results.<sup>[38-40]</sup>

## 2.2 Superficial Candidiasis

*Candida* nail infections require oral treatment:<sup>[41]</sup> a 6-week course of itraconazole capsules 200 to 400 mg/day or 3 cycles of pulse therapy (each cycle consists of 1 week of drug therapy followed by 3 weeks with no therapy) with itraconazole capsules 400 mg/day are effective (mycological and clinical cure rates of more than 90%).<sup>[41-43]</sup> Other superficial *Candida* infections, such as vaginal, penile and cutaneous candidiasis respond well to topical treatment with nystatin or an imidazole, or oral treatment with itraconazole or fluconazole.

Oral candidiasis in immunocompetent patients can be controlled with topical imidazoles or a 2- to 3-week course of amphotericin-B oral suspension or nystatin oral suspension. However, in neutropenic cancer patients and patients with HIV infection, treatment with topical agents is associated with a high relapse rate and oral antifungals are preferred.<sup>[44]</sup> Oral fluconazole (100 to 200 mg/day for 2 weeks) has proven efficacy in treating oral candidiasis (48% of patients mycologically cured; 84% of patients clinically cured),<sup>[45]</sup> but fluconazole-resistant candidiasis is frequently reported during prolonged exposure in patients with HIV infection or chronic mucocutaneous candidiasis,<sup>[46-48]</sup> and fluconazole is ineffective against some species of *Candida*. Itraconazole capsules (200 mg/day for 2 to 4 weeks) are effective against oral and oesophageal candidiasis (40% of patients mycologically cured; 74% of patients clinically cured),<sup>[49]</sup> but absorption of the capsule formulation can be variable in patients with HIV and hypochlorhydria. Itraconazole oral solution has reliable absorption and greater bioavailability than the capsule,<sup>[50,51]</sup> and is at least as effective as fluconazole in immunocompromised patients with oral candidiasis (mycological cure rates of 88% and 77% for itraconazole oral solution and fluconazole, respectively; clinical response rate of 97% and 87% for itraconazole oral solution and fluconazole, respectively).<sup>[52]</sup> In addition, most patients with fluconazole-refractory oral candidiasis

respond to treatment with itraconazole oral solution (200 mg/day for 2 weeks).<sup>[53-55]</sup>

## 3. Management of Systemic Fungal Infections

Systemic fungal infections are life threatening and affect immunocompromised people, such as patients with HIV, bone marrow transplant recipients, patients with haematological malignancies and solid organ transplant recipients. Diagnosis of systemic fungal infections is difficult because symptoms are not specific and can be confused with those of bacterial infections, viral infections or the underlying condition, or with complications of treatment (see Garber<sup>[1]</sup>).

Although diagnostic techniques for systemic fungal infections are improving, prophylaxis and empirical treatment continue to play a key role in the management of systemic fungal infections. The poor prognosis associated with systemic fungal infections means that chemoprophylaxis is frequently administered to patients at high risk and empirical therapy is initiated without proof of infection – usually when the patient has persistent fever of unknown origin that is unresponsive to broad spectrum antibiotics. Both approaches can result in the prolonged administration of antifungal agents to high-risk patient groups.

### 3.1 Endemic Infections

Most endemic mycoses are treated with amphotericin-B or itraconazole (table II). Ketoconazole may also be effective for treating immunocompetent patients with non-life-threatening histoplasmosis, blastomycosis or paracoccidioidomycosis,<sup>[56]</sup> but fluconazole has relatively poor efficacy against endemic mycoses.

Itraconazole is recommended for the treatment of many endemic mycoses and as maintenance therapy in immunocompromised patients. Life-threatening histoplasmosis and blastomycosis may require intravenous treatment, such as with amphotericin-B.<sup>[57,58]</sup> However, oral itraconazole 100 to 400 mg/day for at least 6 months is the usual

**Table II.** First-choice therapies for endemic and other mycoses

Fungal infection	First-choice therapy
Blastomycosis	
Life-threatening and CNS	Amphotericin-B (1 mg/kg/day)
Mild or moderate	Itraconazole (≥200 mg/day)
Histoplasmosis	
Life-threatening and CNS	Amphotericin-B (1 mg/kg/day)
Mild or moderate	Itraconazole (≥200 mg/day)
Maintenance therapy in immunocompromised	Itraconazole (400 mg/day)
<i>Histoplasma duboisii</i> infection	Itraconazole (100-400 mg/day)
Paracoccidioidomycosis	Itraconazole (≥ 100 mg/day)
Coccidioidomycosis	
Life-threatening	Amphotericin-B (1-1.25 mg/kg/day)
Mild or moderate	Azoles (400 mg/day)
Maintenance therapy in immunocompromised	Azoles at recommended dose
Immunocompetent and non-meningeal	Itraconazole (400 mg/day; >1 year)
Immunocompetent and meningeal	Fluconazole (≥400 mg/day; life) or itraconazole (400 mg/day; life)
<i>Penicillium marneffei</i>	Amphotericin-B (≥0.6 mg/kg/day) then itraconazole (≥200 mg/day)
Sporotrichosis	
Lymphocutaneous	Itraconazole (200 mg/day)
Visceral	Itraconazole (400 mg/day)
Severe disseminated	Amphotericin-B (1 mg/kg/day)
Chromoblastomycosis	Itraconazole (100-200 mg/day)

CNS = central nervous system.

first-line treatment for infections caused by *Histoplasma duboisii*.<sup>[56]</sup>

Itraconazole capsules (≥200 mg/day for up to 6 months) are the treatment of choice for paracoccidioidomycosis and ketoconazole or amphotericin-B are the best alternatives.<sup>[56]</sup> The most effective therapeutic strategy for coccidioidomycosis in immunocompetent hosts is unclear: amphotericin-B, itraconazole capsules, ketoconazole and fluconazole have all been tested in clinical trials, with similar results.<sup>[56,59-61]</sup> However, a recent comparison of itraconazole capsules 400 mg/day with fluconazole 400 mg/day suggested that itraconazole may be more effective in the treatment of progressive nonmeningeal coccidioidomycosis, particularly skeletal infections.<sup>[62]</sup> In immuno-

compromised patients, amphotericin-B may be the first choice for treatment of coccidioidomycosis, although the potential benefit of itraconazole has not been tested.<sup>[56]</sup> In diffuse interstitial pneumonia, amphotericin-B is the most effective treatment, and in coccidioidal meningitis high dose fluconazole (400 to 1000 mg/day) has been recommended,<sup>[63]</sup> although itraconazole 400 mg/day may be as effective.<sup>[64]</sup>

In general, initial treatment with itraconazole capsules or amphotericin-B followed by itraconazole capsules are the recommended treatments for other rare endemic mycoses (table II).

3.2 Aspergillosis and Deep Candidiasis

The most frequently encountered systemic fungal infections of immunocompromised patients are those caused by species of *Candida* or *Aspergillus*. The various formulations of amphotericin-B and itraconazole are the most effective antifungal agents for the treatment and prevention of these systemic infections (table III). Fluconazole is effective only for the management of infections by susceptible species of *Candida*, such as *C. albicans*, and *C. parapsilosis*.

3.2.1 Chemoprophylaxis

Prophylaxis of systemic fungal infections is usually given when patients are considered to be at a particular risk of infection – for example, bone marrow transplant recipients. In addition, preemptive therapy is frequently given; this is antifungal prophylaxis in patients who have some evidence of infection but who do not meet the usual criteria for definitive diagnosis or for initiating empirical therapy.

Because of its poor oral absorption, amphotericin-B is an ineffective prophylactic agent for systemic fungal infection. The use of low dose IV amphotericin-B (<1 mg/kg/day) is associated with breakthrough infections,<sup>[65,66]</sup> and infusion-related adverse events and renal toxicity restrict the use of higher doses of amphotericin-B.

The superior safety profiles of the triazole antifungals have made them the most widely used drugs in the prophylaxis of systemic fungal infec-

**Table III.** Management options for systemic fungal infections

Approach	Options
Prophylaxis <sup>a</sup>	Itraconazole oral solution (5 mg/kg/day) Fluconazole capsules (50-400 mg/day)
Empirical treatment	Conventional amphotericin-B (0.6-1 mg/kg/day for 14 days or until resolution of symptoms) Amphotericin-B-liposomal (3 mg/kg/day for 14 days or until resolution of symptoms) ABLC (5 mg/kg/day for 14 days or until resolution of symptoms) ABCD (4 mg/kg/day for 14 days or until resolution of symptoms) IV itraconazole (400 mg/day for 2 days followed by 200 mg/day for 14 days or until resolution of symptoms)
Confirmed candidiasis	IV fluconazole (400 mg/day) for 3-7 days followed by capsules (400 mg/day until resolution of symptoms) <sup>c</sup> Conventional amphotericin-B (0.6-1 mg/kg/day until resolution of symptoms) ABLC (5 mg/kg/day until resolution of symptoms) IV itraconazole (400 mg/day until resolution of symptoms) <sup>b</sup>
Confirmed aspergillosis	Oral itraconazole (400 mg/day for 14 days or more) Conventional amphotericin-B (1-1.5 mg/kg/day for at least 14 days or until recovery of granulocytes and resolution of symptoms) Amphotericin-B-liposomal (3 mg/kg/day for at least 14 days or until recovery of granulocytes and resolution of symptoms) ABLC (5 mg/kg/day for at least 14 days or until recovery of granulocytes and resolution of symptoms) ABCD (4 mg/kg/day for at least 14 days or until recovery of granulocytes and resolution of symptoms) IV itraconazole (400 mg/day for 2 days followed by 200 mg/day for at least 14 days or until recovery of granulocytes and resolution of symptoms)

a Value only established in selected patient groups (bone marrow transplant patients, high risk liver transplant patients, selected high risk neutropenic patients with haematological malignancy, and those with previously documented invasive *Aspergillus* or other mould infection who are undergoing further courses of chemotherapy).

b No clinical data.

c Only for infections by *Candida* spp. shown to be susceptible to fluconazole.

**ABLC** = amphotericin-B-lipid-complex; **ABCD** = amphotericin-B-colloidal dispersion; **IV** = intravenous.

tions. Prophylaxis often requires long term anti-fungal administration and the ease of use of oral formulations makes them a preferred option. However, some patients at high risk have severely impaired swallowing or are unconscious or intubated and can only be given intravenous agents.

The high oral bioavailability of fluconazole and its availability in intravenous and oral formulations have led to the widespread use of this agent. Fluconazole has been shown to reduce the incidence of systemic *C. albicans* infections,<sup>[67,68]</sup> but it may be less effective in some high-risk patient groups, such as patients with acute leukaemia undergoing remission-induction therapy or intensive rescue therapy.<sup>[69,70]</sup> The most important weakness of fluconazole as a prophylactic agent is its narrow spectrum of activity; in some trials the

predominance of *Aspergillus* infections in fluconazole-treated patients has highlighted this weakness.<sup>[71]</sup> Similarly, fluconazole does not offer protection against non-*albicans Candida* infections; a concern surrounding the prophylactic use of fluconazole is the potential increase in fluconazole-resistant *Candida* spp. and increased colonisation by *C. krusei* and *C. glabrata*.<sup>[72-75]</sup>

The broad spectrum of activity and good safety profile of itraconazole make it an attractive choice for chemoprophylaxis. The capsule formulation is successful in preventing systemic fungal infections<sup>[76-78]</sup> and at a dosage of 200 mg/day is as effective as fluconazole (100 mg/day).<sup>[79]</sup> The absorption of the itraconazole capsule formulation may be variable in patients with damage to the intestinal epithelium or with reduced gastric acid-

ity.<sup>[80,81]</sup> An itraconazole oral solution, with its enhanced bioavailability, has overcome the limitations of the capsule formulation and is effective in the prophylaxis of systemic fungal infections in neutropenic patients.<sup>[82-85]</sup> In a placebo-controlled trial, itraconazole oral solution reduced the number of cases of invasive candidiasis but had no effect on the incidence of invasive aspergillosis.<sup>[86]</sup> However, in a comparative trial with oral fluconazole, itraconazole oral solution reduced the number of cases of aspergillosis.<sup>[87]</sup>

Clearly, many patients at risk of systemic fungal infections are receiving co-medications. Because the azole antifungals are metabolised by CYP3A, they can interact with other drugs metabolised by this enzyme. In solid organ transplant or bone marrow transplant recipients, the interaction between azoles and cyclosporin is the most significant,<sup>[88,89]</sup> but elevated plasma concentrations of cyclosporin can be managed by routine monitoring and reducing the dosage as necessary. In patients with HIV infection, itraconazole may interact with protease inhibitors (by inhibiting CYP3A)<sup>[90]</sup> and fluconazole may interact with the HIV reverse transcriptase inhibitor zidovudine (by inhibiting glucuronidation).<sup>[89,91,92]</sup> As with the co-administration of cyclosporin, blood concentration monitoring and dosage reduction may be advisable when azoles and some anti-HIV drugs are used concurrently. In patients with HIV, the HIV protease inhibitor saquinavir has low bioavailability, and it has been suggested that the addition of itraconazole may increase saquinavir plasma concentrations and ultimately improve anti-HIV efficacy.<sup>[93]</sup>

### 3.2.2 Empirical Treatment

Empirical treatment is usually initiated in immunocompromised patients with persistent fever of unknown origin (4 to 7 days) that is unresponsive to broad spectrum antibiotics or when pulmonary infiltrates or cavities are detected by computed tomography (CT) scan.

Amphotericin-B has been the first-choice treatment for systemic fungal infections for several years. A significant benefit (reduced death rate

from fungal infection and a decrease in clinically documented infection) is seen in patients receiving IV amphotericin-B.<sup>[94]</sup> The broad spectrum of activity of amphotericin-B has been the key to its success in empirical therapy, but the use of higher doses – which could be more effective – is prevented by infusion-related adverse events (nausea, fever and chills) and renal toxicity.

The triazole antifungal agents are the best alternative to amphotericin-B for empirical therapy. However, there have been no large-scale clinical trials comparing triazoles with amphotericin-B for empirical therapy. Both fluconazole and itraconazole capsules have shown equivalence to amphotericin-B in small-scale clinical trials,<sup>[95,96]</sup> although fluconazole is useful only in the treatment of confirmed deep candidiasis, and even then only when the isolate is shown to be susceptible to fluconazole. Confirmed aspergillosis has been treated with success with high doses of IV amphotericin-B<sup>[97,98]</sup> or oral itraconazole.<sup>[99,100]</sup> A combination of amphotericin-B and itraconazole capsules may improve outcomes and has been suggested as an alternative treatment strategy for aspergillosis,<sup>[101]</sup> despite some previous evidence of antagonism between these agents in animal studies.<sup>[102]</sup>

Many patients who require empirical therapy have already received antifungal prophylaxis. Until recently, empirical therapy with amphotericin-B was usually initiated if prophylaxis with azole antifungals failed; however, after failure of prophylaxis with an azole, empirical therapy with another azole or even a different formulation of the same azole may be at least as effective as using amphotericin-B. This has not been confirmed in clinical trials, but a recent report indicated that patients receiving previous prophylaxis (primarily fluconazole) benefited more from empirical therapy with itraconazole than with amphotericin-B.<sup>[103]</sup>

### 3.2.3 Advances in the Management of Systemic Fungal Infections

Despite the wide choice of antifungal agents available, death rates from systemic fungal infections are unacceptably high and improvements in management are still sought.



Several improvements in the diagnosis of systemic fungal infections have been made. Antigen detection methods for endemic fungal infections, such as histoplasmosis, have been developed; more recently, a double-sandwich enzyme-linked immunosorbent assay (ELISA) has been developed for detecting invasive *Aspergillus* infections. The sensitivity and specificity of the test were 92.6% and 95.4%, respectively, based on 71 autopsy-controlled patients.<sup>[104]</sup> The early use of CT scanning of the chest is also useful for the diagnosis of aspergillosis. The appearance of the distinctive 'halo' sign and later the 'air-crescent' sign on CT images can be highly suggestive of aspergillosis (or other fungal infections).<sup>[105]</sup> Other new diagnostic tools include biochemical methods and molecular biological techniques, such as polymerase chain reaction (PCR).<sup>[106]</sup> These tools may help to improve the diagnosis of fungal infections and thus the outcome of treatment; one group has reported reduced mortality among patients with aspergillosis when early CT scanning and antigen detection were used.<sup>[107]</sup>

Recent developments in antifungal therapy have produced useful re-formulations of the existing antifungal agents, including the lipid-associated formulations of amphotericin-B and cyclodextrin-associated formulations of itraconazole. In general, these formulations have been tested for the treatment of deep candidiasis and aspergillosis but not for the treatment of endemic mycoses.

Three lipid-associated formulations of amphotericin-B have been developed: amphotericin-B-liposomal, amphotericin-B-lipid-complex (ABLC) and amphotericin-B-colloidal dispersion (ABCD). These formulations are available for IV use and reduce the toxicity associated with amphotericin-B.

Amphotericin-B-liposomal (3 mg/kg/day) has efficacy equivalent to that of conventional amphotericin-B (1 mg/kg/day) and reduced toxicity.<sup>[108,109]</sup> Nephrotoxicity was significantly lower with amphotericin-B-liposomal (19%) than with conventional amphotericin-B (34%), but discontinuations because of toxic effects or lack of efficacy were similar between formulations.<sup>[109]</sup>

ABLC may also be effective for the treatment of confirmed or suspected systemic fungal infections and is generally well tolerated, except for infusion-related adverse events;<sup>[110,111]</sup> however, a high incidence of renal toxicity has been reported with ABLC.<sup>[112]</sup> This formulation has not been compared with conventional amphotericin-B in a randomised, double-blind trial, but, in a retrospective analysis, amphotericin-B-liposomal 1.9 mg/kg/day and ABLC 4.8 mg/kg/day were equally effective in treating systemic fungal infections and had similar safety profiles.<sup>[113]</sup>

Empirical therapy with ABCD 4 mg/kg/day and empirical therapy with conventional amphotericin-B 0.8 mg/kg/day were compared in a double-blind trial.<sup>[114]</sup> The results indicated that ABCD was as effective as conventional amphotericin-B, and was associated with less severe nephrotoxicity but more frequent infusion-related adverse events.

The use of the lipid-associated formulations of amphotericin-B is restricted by their high acquisition costs.<sup>[115-117]</sup> Although the costs of treating adverse events are lower with these formulations than with conventional amphotericin-B, it is not clear how significant these savings are.

Two new formulations of itraconazole have been developed using the solubilising excipient hydroxypropyl- $\beta$ -cyclodextrin. The itraconazole oral solution has reliable oral bioavailability in healthy individuals and in a variety of patient groups at risk of systemic fungal infections (see De Beule and Van Gestel<sup>[15]</sup>). In clinical trials comparing itraconazole oral solution with oral amphotericin-B or fluconazole suspension as prophylaxis in neutropenic patients, fewer superficial fungal infections, proven deep fungal infections, cases of aspergillosis and deaths with proven deep fungal infection were observed in the itraconazole oral solution groups.<sup>[87,118]</sup>

A new IV itraconazole formulation has also been developed, which provides an alternative formulation for those patients who cannot take oral medications. In addition, this IV itraconazole formulation achieves therapeutic plasma concentrations less than 48 hours after starting treatment,<sup>[119]</sup>

a feature that is of particular importance in the treatment of systemic fungal infections (suspected or confirmed). In a recent clinical trial, the IV itraconazole formulation (with subsequent therapy with itraconazole oral solution) was at least as effective as conventional amphotericin-B in the empirical treatment of systemic fungal infections and caused significantly fewer adverse events.<sup>[103]</sup> The IV itraconazole formulation may also be effective for first- or second-line treatment of invasive aspergillosis in a variety of patients, such as patients with AIDS, haematological malignancy or chronic granulomatous disease.<sup>[120]</sup> Unlike the lipid-associated formulations of amphotericin-B, the use of the new itraconazole formulations is cost effective.<sup>[121]</sup>

#### 4. Conclusion

The increasing number of antifungal agents, reformulations of existing agents and novel treatment strategies have all improved the management of fungal infections in recent years. Although high cure rates can be achieved in the treatment of many superficial infections, systemic fungal infections are still associated with high mortality. For several years, amphotericin-B was the most effective agent for the treatment and prevention of systemic fungal infections. However, the introduction of the triazoles – fluconazole and itraconazole – has challenged amphotericin-B as the gold standard. In particular, the triazoles have become the agents of choice in chemoprophylaxis; fluconazole has been widely used but the introduction of an itraconazole oral solution offers an agent with high bioavailability and a broader spectrum of activity than that of fluconazole. In the empirical treatment of systemic fungal infections, the lipid-associated formulations of amphotericin-B and the itraconazole IV formulation are at least as effective as conventional amphotericin-B and are less toxic. The high cost of lipid-associated formulations of amphotericin-B may make their use prohibitively expensive.

#### References

- Garber G. An overview of fungal infections. *Drugs* 2001; 61 Suppl. 1: 1-12
- Jantunen E, Ruutu P, Niskanen L, et al. Incidence and risk factors for invasive fungal infections in allogeneic BMT recipients. *Bone Marrow Transplant* 1997; 19: 801-8
- Tumbarello M, Tacconelli E, Pagano L, et al. Comparative analysis of prognostic indicators of aspergillosis in haematological malignancies and HIV infection. *J Infect* 1997; 34: 55-60
- Groll AH, Shah PM, Mentzel C, et al. Trends in the postmortem epidemiology of invasive fungal infections at a university hospital. *J Infect* 1996; 33: 23-32
- Bolard J. How do the polyene macrolide antibiotics affect the cellular membrane properties? *Biochim Biophys Acta* 1986; 864: 257-304
- Gallis HA, Drew RH, Pickard WW. Amphotericin B: 30 years of clinical experience. *Rev Infect Dis* 1990; 12: 308-29
- Schafer-Korting M, Blechschmidt J, Korting HC. Clinical use of oral nystatin in the prevention of systemic candidosis in patients at particular risk. *Mycoses* 1996; 39: 329-39
- Ryder NS. The mechanism of action of terbinafine. *Clin Exp Dermatol* 1989; 14: 98-100
- Balfour JA, Faulds D. Terbinafine: a review of its pharmacodynamic and pharmacokinetic properties, and therapeutic potential in superficial mycoses. *Drugs* 1992; 43: 259-84
- Harari S, Schiraldi G, de Juli E, et al. Relapsing *Aspergillus* bronchitis in a double lung transplant patient, successfully treated with a new oral antimycotic agent [letter]. *Chest* 1997; 111: 835-6
- Vanden Bossche H. Biochemical targets for antifungal azole derivatives: hypothesis on the mode of action. In: McGinnis MR, ed. *Current topics in medical mycology*. New York: Springer-Verlag, 1985: 313-51
- Como JA, Dismukes WE. Oral azole drugs as systemic antifungal therapy. *N Engl J Med* 1993; 330: 263-72
- Heykants J, Van Peer A, Van de Velde V, et al. The clinical pharmacokinetics of itraconazole: an overview. *Mycoses* 1989; 32 Suppl. 1: 67-87
- Boogaerts M, Maertens J. Clinical experience with itraconazole in systemic infections. *Drugs* 2001; 61 Suppl. 1: 39-47
- De Beule K, Van Gestel J. Pharmacology of itraconazole. *Drugs* 2001; 61 Suppl. 1: 27-37
- Hostetler JS, Hanson LH, Stevens DA. Effect of cyclodextrin on the pharmacology of antifungal oral azoles. *Antimicrob Agents Chemother* 1992; 36: 477-80
- Saag MS, Powderly WG, Cloud GA, et al. Comparison of amphotericin B with fluconazole in the treatment of acute AIDS-associated cryptococcal meningitis. The NIAID Mycoses Study Group and the AIDS Clinical Trials Group. *N Engl J Med* 1992; 326: 83-9
- Bozzette SA, Larsen RA, Chiu J, et al. A placebo-controlled trial of maintenance therapy with fluconazole after treatment of cryptococcal meningitis in the acquired immunodeficiency syndrome. California Collaborative Treatment Group. *N Engl J Med* 1991; 324: 580-4
- Wildfeuer A, Faergemann J, Laufen H, et al. Bioavailability of fluconazole in the skin after oral medication. *Mycoses* 1994; 37: 127-30
- Savin RC, Drake L, Babel D, et al. Pharmacokinetics of three once-weekly dosages of fluconazole (150, 300, or 450 mg) in distal subungual onychomycosis of the fingernail. *J Am Acad Dermatol* 1998; 38: S110-6
- Francis P, Walsh TJ. Evolving role of flucytosine in immunocompromised patients: new insights into safety, pharmacokinetics, and antifungal therapy. *Clin Infect Dis* 1992; 15: 1003-18

22. Piérard GE, Arrese JE, Piérard-Franchimont C. Treatment and prophylaxis of tinea infections. *Drugs* 1996; 52: 209-24
23. Evans EG. Tinea pedis: clinical experience and efficacy of short treatment. *Dermatology* 1997; 194 Suppl. 1: 3-6
24. Noble SL, Forbes RC, Stamm PL. Diagnosis and management of common tinea infections. *Am Fam Physician* 1998; 58: 163-74
25. Farag A, Taha M, Halim S. One-week therapy with oral terbinafine in cases of tinea cruris/corporis. *Br J Dermatol* 1994; 131: 684-6
26. Parent D, Decroix J, Heenen M. Clinical experience with short schedules of itraconazole in the treatment of tinea corporis and/or tinea cruris. *Dermatology* 1994; 189: 378-81
27. Gupta AK, De Doncker P, Heremans A, et al. Itraconazole for the treatment of tinea pedis: a dosage of 400 mg/day given for 1 week is similar in efficacy to 100 or 200 mg/day given for 2 to 4 weeks. *J Am Acad Dermatol* 1997; 36: 789-92
28. Boonk W, de Geer D, de Kreek E, et al. Itraconazole in the treatment of tinea corporis and tinea cruris: comparison of two treatment schedules. *Mycoses* 1998; 41: 509-14
29. Hickman JG. A double-blind, randomized, placebo-controlled evaluation of short-term treatment with oral itraconazole in patients with tinea versicolor. *J Am Acad Dermatol* 1996; 34: 785-7
30. Tausch I, Decroix J, Gwiedzinski Z, et al. Short-term itraconazole versus terbinafine in the treatment of tinea pedis or manus. *Int J Dermatol* 1998; 37: 140-2
31. Jahangir M, Hussain I, Ul Hasan M, et al. A double-blind, randomized, comparative trial of itraconazole versus terbinafine for 2 weeks in tinea capitis. *Br J Dermatol* 1998; 139: 672-4
32. Elewski B, Hay RJ. International summit on cutaneous antifungal therapy. 1994 Nov 11-13; Boston, Massachusetts, USA. *J Am Acad Dermatol* 1995; 33: 816-22
33. Goodfield MJD. Short-duration therapy with terbinafine for dermatophyte onychomycosis: a multicentre trial. *Br J Dermatol* 1992; 126 Suppl. 39: 33-5
34. van der Schroeff JG, Cirkel PK, Crijns MB, et al. A randomized treatment duration-finding study of terbinafine in onychomycosis. *Br J Dermatol* 1992; 126 Suppl. 39: 36-9
35. Gupta AK, Scher RK, De Doncker P. Current management of onychomycosis: an overview. *Dermatol Clin* 1997; 15: 121-35
36. Willemsen M, De Doncker P, Willems J, et al. Posttreatment itraconazole levels in the nail. New implications for treatment in onychomycosis. *J Am Acad Dermatol* 1992; 26: 731-5
37. Gupta AK. Pharmacoeconomic analysis of oral antifungal therapies used to treat dermatophyte onychomycosis of the toenails: a US analysis. *Pharmacoeconomics* 1998; 13: 243-56
38. Tosti A, Piraccini BM, Stinchi C, et al. Treatment of dermatophyte nail infections: an open randomized study comparing intermittent terbinafine therapy with continuous terbinafine treatment and intermittent itraconazole therapy. *J Am Acad Dermatol* 1996; 34: 595-600
39. Evans EG, Sigurgeirsson B. Double blind, randomised study of continuous terbinafine compared with intermittent itraconazole in treatment of toenail onychomycosis. The LION Study Group. *BMJ* 1999; 318: 1031-5
40. Degreef H, Del Palacio A, Mygind S, et al. Randomized double-blind comparison of short-term itraconazole and terbinafine therapy for toenail onychomycosis. *Acta Derm Venereol* 1999; 79: 221-3
41. Hay RJ, Clayton YM, Moore MK, et al. An evaluation of itraconazole in the management of onychomycosis. *Br J Dermatol* 1988; 119: 359-66
42. Kim J-A, Ahn K-J, Kim J-M, et al. Efficacy and tolerability of itraconazole in patients with fingernail onychomycosis: a 6-week pilot study. *Curr Ther Res* 1995; 56: 1066-75
43. Li WD, Ping WA, Yu LR, et al. Therapeutic efficacy of intermittent pulse therapy with itraconazole in onychomycosis: a Chinese multicenter trial. 54th Annual Meeting of the American Academy of Dermatology; 1996 Feb 10-25; Washington, DC, 1996
44. Smith DE, Midgley J, Allan M, et al. Itraconazole versus ketoconazole in the treatment of oral and oesophageal candidosis in patients infected with HIV. *AIDS* 1991; 5: 1367-71
45. Hay RJ. Overview of studies of fluconazole in oropharyngeal candidiasis. *Rev Infect Dis* 1990; 12 Suppl. 3: S334-S7
46. Bailly GG, Perry FM, Denning DW, et al. Fluconazole-resistant candidosis in an HIV cohort. *AIDS* 1994; 8: 787-92
47. Ruhnke M, Eigler A, Tennagen I, et al. Emergence of fluconazole-resistant strains of *Candida albicans* in patients with recurrent oropharyngeal candidosis and human immunodeficiency virus infection. *J Clin Microbiol* 1994; 32: 2092-8
48. Willocks L, Leen CLS, Brettell RP, et al. Fluconazole resistance in AIDS patients [letter]. *J Antimicrob Chemother* 1991; 28: 937-9
49. Smith D, Midgley J, Gazzard B. A randomised, double-blind study of itraconazole versus placebo in the treatment and prevention of oral or oesophageal candidosis in patients with HIV infection. *Int J Clin Prac* 1999; 53: 349-52
50. Cartledge JD, Midgely J, Gazzard BG. Itraconazole solution: higher serum drug concentrations and better clinical response rates than the capsule formulation in acquired immunodeficiency syndrome patients with candidosis. *J Clin Pathol* 1997; 50: 477-80
51. Prentice AG, Warnock DW. Itraconazole more bioavailable in solution [letter]. *Blood* 1996; 88: 3662-3
52. Graybill JR, Vazquez J, Darouiche RO, et al. Randomized trial of itraconazole oral solution for oropharyngeal candidiasis in HIV/AIDS patients. *Am J Med* 1998; 104: 33-9
53. Cartledge JD, Midgley J, Youle M, et al. Itraconazole cyclo-dextrin solution – effective treatment for HIV-related candidosis unresponsive to other azole therapy [letter]. *J Antimicrob Chemother* 1994; 33: 1071-3
54. Phillips P, Zemcov J, Mahmood W, et al. Itraconazole cyclo-dextrin solution for fluconazole-refractory oropharyngeal candidiasis in AIDS: correlation of clinical response with *in vitro* susceptibility. *AIDS* 1996; 10: 1369-76
55. Saag MS, Fessel WJ, Kauffman CA, et al. Treatment of fluconazole-refractory oropharyngeal candidiasis with itraconazole oral solution in HIV-positive patients. *AIDS Res Hum Retroviruses* 1999; 15: 1413-7
56. Lortholary O, Denning DW, Dupont B. Endemic mycoses: a treatment update. *J Antimicrob Chemother* 1999; 43: 321-31
57. Kauffman CA. Newer developments in therapy for endemic mycoses. *Clin Infect Dis* 1994; 19 Suppl. 1: S28-S32
58. Dismukes WE, Bradsher Jr RW, Cloud GC, et al. Itraconazole therapy for blastomycosis and histoplasmosis. *Am J Med* 1992; 93: 489-97
59. Robinson PA, Knirsch AK, Joseph JA. Fluconazole for life-threatening fungal infections in patients who cannot be treated with conventional antifungal agents. *Rev Infect Dis* 1990; 12 Suppl. 3: S349-63
60. Galgiani JN, Stevens DA, Graybill JR, et al. Ketoconazole therapy of progressive coccidioidomycosis. Comparison of 400- and 800-mg doses and observations at higher doses. *Am J Med* 1988; 84: 603-10

61. Tucker RM, Denning DW, Arathoon EG, et al. Itraconazole therapy for nonmeningeal coccidioidomycosis: clinical and laboratory observations. *J Am Acad Dermatol* 1990; 23: 593-601
62. Galgiani JN, Catanzaro A, Cloud GA, et al. Comparison of oral fluconazole and itraconazole for progressive, nonmeningeal coccidioidomycosis. *Ann Intern Med* 2000; 133 (9): 676-86
63. Galgiani JN, Ampel NM, Catanzaro A, et al. Practice guideline for the treatment of coccidioidomycosis. *Clin Infect Dis* 2000; 30: 658-61
64. Tucker RM, Denning DW, Dupont B, et al. Itraconazole therapy for chronic coccidioid meningitis. *Ann Intern Med* 1990; 112: 108-12
65. Schuler US, Haag C. Prophylaxis of fungal infections. *Mycoses* 1997; 40 Suppl. 2: 41-4
66. Lortholary O, Dupont B. Antifungal prophylaxis during neutropenia and immunodeficiency. *Clin Microbiol Rev* 1997; 10: 477-504
67. Goodman JL, Winston DJ, Greenfield RA, et al. A controlled trial of fluconazole to prevent fungal infections in patients undergoing bone marrow transplantation. *N Engl J Med* 1992; 326: 845-51
68. Slavin MA, Osborne B, Adams R, et al. Efficacy and safety of fluconazole prophylaxis for fungal infections after marrow transplantation – a prospective, randomized, double-blind study. *J Infect Dis* 1995; 171: 1545-52
69. Winston DJ, Chandrasekar PH, Lazarus HM, et al. Fluconazole prophylaxis of fungal infections in patients with acute leukemia. Results of a randomized placebo-controlled, double-blind, multicenter trial. *Ann Intern Med* 1993; 118: 495-503
70. Kern W, Behre G, Rudolf T, et al. Failure of fluconazole prophylaxis to reduce mortality or the requirement of systemic amphotericin B therapy during treatment for refractory acute myeloid leukemia: results of a prospective randomized phase III study. German AML Cooperative Group. *Cancer* 1998; 83: 291-301
71. Schaffner A, Schaffner M. Effect of prophylactic fluconazole on the frequency of fungal infections, amphotericin B use, and health care costs in patients undergoing intensive chemotherapy for hematologic neoplasias. *J Infect Dis* 1995; 172: 1035-41
72. Pfaller MA, Lockhart SR, Pujol C, et al. Hospital specificity, region specificity, and fluconazole resistance of *Candida albicans* bloodstream isolates. *J Clin Microbiol* 1998; 36: 1518-29
73. Hoppe JE, Klingebiel T, Niethammer D. Selection of *Candida glabrata* in pediatric bone marrow transplant recipients receiving fluconazole. *Pediatr Hematol Oncol* 1994; 11: 207-10
74. Wingard JR, Merz WG, Rinaldi MG, et al. Association of *Torulopsis glabrata* infections with fluconazole prophylaxis in neutropenic bone marrow transplant patients. *Antimicrob Agents Chemother* 1993; 37: 1847-9
75. Wingard JR. Importance of *Candida* species other than *C. albicans* as pathogens in oncology patients [review]. *Clin Infect Dis* 1995; 20: 115-25
76. Prentice AG, Bradford GR. Prophylaxis of fungal infections with itraconazole during remission-induction therapy. *Mycoses* 1989; 32 Suppl. 1: 96-102
77. Vreugdenhil G, Van Dijke BJ, Donnelly JP, et al. Efficacy of itraconazole in the prevention of fungal infections among neutropenic patients with hematologic malignancies and intensive chemotherapy. A double blind, placebo controlled study. *Leuk Lymphoma* 1993; 11 (5-6): 353-8
78. Glasmacher A, Molitor E, Hahn C, et al. Antifungal prophylaxis with itraconazole in neutropenic patients with acute leukemia. *Leukemia* 1998; 12: 1338-43
79. Huijgens PC, Simoons-Smit AM, van Loenen AC, et al. Fluconazole versus itraconazole for the prevention of fungal infections in haemato-oncology. *J Clin Pathol* 1999; 52: 376-80
80. Boogaerts MA, Verhoef GE, Zachee P, et al. Antifungal prophylaxis with itraconazole in prolonged neutropenia: correlation with plasma levels. *Mycoses* 1989; 32 Suppl. 1: 103-8
81. Glasmacher A, Hahn C, Leutner C, et al. Breakthrough invasive fungal infections in neutropenic patients after prophylaxis with itraconazole. *Mycoses* 1999; 42: 443-51
82. Prentice AG, Warnock DW, Johnson SA, et al. Multiple dose pharmacokinetics of an oral solution of itraconazole in autologous bone marrow transplant recipients. *J Antimicrob Chemother* 1994; 34: 247-52
83. Prentice AG, Warnock DW, Johnson SA, et al. Multiple dose pharmacokinetics of an oral solution of itraconazole in patients receiving chemotherapy for acute myeloid leukaemia. *J Antimicrob Chemother* 1995; 36: 657-63
84. Reynes J, Bazin C, Ajana F, et al. Pharmacokinetics of itraconazole (oral solution) in two groups of human immunodeficiency virus-infected adults with oral candidiasis. *Antimicrob Agents Chemother* 1997; 41: 2554-8
85. Michallet M, Persat F, Kranzhofer N, et al. Pharmacokinetics of itraconazole oral solution in allogeneic bone marrow transplant patients receiving total body irradiation. *Bone Marrow Transplant* 1998; 21: 1239-43
86. Menichetti F, Favero AD, Martino P, et al. Itraconazole oral solution as prophylaxis for fungal infections in neutropenic patients with hematologic malignancies: a randomized, placebo-controlled, double-blind, multicenter trial. *Clin Infect Dis* 1999; 28: 250-5
87. Morgenstern GR, Prentice AG, Prentice HG, et al. A randomized controlled trial of itraconazole versus fluconazole for the prevention of fungal infections in patients with hematological malignancies. *Br J Haematol* 1999; 105: 901-11
88. Schafer-Korting M. Pharmacokinetic optimisation of oral antifungal therapy. *Clin Pharmacokinet* 1993; 25: 329-41
89. Gupta AK, Katz HI, Shear NH. Drug interactions with itraconazole, fluconazole, and terbinafine and their management. *J Am Acad Dermatol* 1999; 41: 237-49
90. MacKenzie-Wood AR, Whitfield MJ, Ray JE. Itraconazole and HIV protease inhibitors: an important interaction. *Med J Aust* 1999; 170: 46-7
91. Brockmeyer NH, Tillmann I, Mertins L, et al. Pharmacokinetic interaction of fluconazole and zidovudine in HIV-positive patients. *Eur J Med Res* 1997; 2: 377-83
92. Trapnell CB, Klecker RW, Jamis-Dow C, et al. Glucuronidation of 3'-azido-3'-deoxythymidine (zidovudine) by human liver microsomes: relevance to clinical pharmacokinetic interactions with atovaquone, fluconazole, methadone, and valproic acid. *Antimicrob Agents Chemother* 1998; 42: 1592-6
93. Koks CH, van Heeswijk RP, Veldkamp AI, et al. Itraconazole as an alternative for ritonavir liquid formulation when combined with saquinavir [letter]. *AIDS* 2000; 14: 89-90
94. EORTC International Antimicrobial Therapy Cooperative Group. Empiric antifungal therapy in febrile granulocytopenic patients. *Am J Med* 1989; 86: 668-72
95. Anaissie EJ, Darouiche RO, Abi-Said D, et al. Management of invasive candidal infections: results of a prospective, randomized, multicenter study of fluconazole versus amphotericin B and review of the literature. *Clin Infect Dis* 1996; 23: 964-72

96. van't Wout JW, Novakova I, Verhagen CA, et al. The efficacy of itraconazole against systemic fungal infections in neutropenic patients: a randomised comparative study with amphotericin B. *J Infect* 1991; 22: 45-52
97. Burch PA, Karp JE, Merz WG, et al. Favorable outcome of invasive aspergillosis in patients with acute leukemia. *J Clin Oncol* 1987; 5: 1985-93
98. Denning DW, Stevens DA. Antifungal and surgical treatment of invasive aspergillosis: review of 2,121 published cases. *Rev Infect Dis* 1990; 12: 1147-202
99. Stevens DA, Lee JY. Analysis of compassionate use itraconazole therapy for invasive aspergillosis by the NIAID Mycoses Study Group criteria. *Arch Intern Med* 1997; 157: 1857-62
100. Lebeau B, Pelloux H, Pinel C, et al. Itraconazole in the treatment of aspergillosis: a study of 16 cases. *Mycoses* 1994; 37: 171-9
101. Popp AI, White MH, Quadri T, et al. Amphotericin B with and without itraconazole for invasive aspergillosis: a three-year retrospective study. *Int J Infect Dis* 1999; 3: 157-60
102. Schaffner A, Bohler A. Amphotericin B refractory aspergillosis after itraconazole: evidence for significant antagonism. *Mycoses* 1993; 36: 421-4
103. Boogaerts M, Garber G, Winston D, et al. Itraconazole (IT) compared with amphotericin B (AMB) as empirical therapy for persistent fever of unknown origin (FUO) in neutropenic patients (PTS). *Bone Marrow Transplant* 1999; 23 Suppl. 1: S111
104. Maertens J, Verhaegen J, Demuynck H, et al. Autopsy-controlled prospective evaluation of serial screening for circulating galactomannan by a sandwich enzyme-linked immunosorbent assay for hematological patients at risk for invasive aspergillosis. *J Clin Microbiol* 1999; 37: 3223-8
105. Blum U, Windfuhr M, Buitrago-Tellez C, et al. Invasive pulmonary aspergillosis. MRI, CT, and plain radiographic findings and their contribution for early diagnosis. *Chest* 1994; 106: 1156-61
106. Brenier-Pinchart MP, Pelloux H, Lebeau B, et al. Towards a molecular diagnosis of invasive aspergillosis? a review of the literature. *J Mycol Med* 1999; 9: 16-23
107. Caillot D, Casasnovas O, Bernard A, et al. Improved management of invasive pulmonary aspergillosis in neutropenic patients using early thoracic computed tomographic scan and surgery. *J Clin Oncol* 1997; 15: 139-47
108. Prentice HG, Hann IM, Herbrecht R, et al. A randomized comparison of liposomal versus conventional amphotericin B for the treatment of pyrexia of unknown origin in neutropenic patients. *Br J Haematol* 1997; 98: 711-8
109. Walsh TJ, Finberg RW, Arndt C, et al. Liposomal amphotericin B for empirical therapy in patients with persistent fever and neutropenia. *N Engl J Med* 1999; 340: 764-71
110. Rapp RP, Gubbins PO, Evans ME. Amphotericin B lipid complex. *Ann Pharmacother* 1997; 31: 1174-86
111. Walsh TJ, Hiemenz JW, Seibel NL, et al. Amphotericin B lipid complex for invasive fungal infections: analysis of safety and efficacy in 556 cases. *Clin Infect Dis* 1998; 26: 1383-96
112. Ringden O, Jonsson V, Hansen M, et al. Severe and common side-effects of amphotericin B lipid complex (Abelcet). *Bone Marrow Transplant* 1998; 22: 733-4
113. Clark AD, McKendrick S, Tansey PJ, et al. A comparative analysis of lipid-complexed and liposomal amphotericin B preparations in haematological oncology. [erratum in *Br J Haematol* 1998; 103: 215] *Br J Haematol* 1998; 103: 198-204
114. White MH, Bowden RA, Sandler ES, et al. Randomized, double-blind clinical trial of amphotericin B colloidal dispersion vs. amphotericin B in the empirical treatment of fever and neutropenia. *Clin Infect Dis* 1998; 27: 296-302
115. Boogaerts M, Tormans G, Maes E, et al. Cost-effectiveness analysis of Ambisome (AMB) vs amphotericin B (AMPHOB) in the empiric treatment of febrile neutropenia in adults and children. *Blood* 1996; 88 Suppl. 1: 501a
116. Coukell AJ, Brogden RN. Liposomal amphotericin B. Therapeutic use in the management of fungal infections and visceral leishmaniasis. *Drugs* 1998; 55: 585-612
117. Verweij PE, Bos JJ, Severens JL, et al. Cost-effectiveness of liposomal amphotericin B for the treatment of invasive fungal infections in neutropenic patients [abstract no. O-14]. 38th Interscience Conference on Antimicrobial Agents and Chemotherapy; 1998 Sep 25-27; San Diego, California, USA. Washington DC, USA: American Society for Microbiology, 1998: 609
118. Harousseau JL, Dekker AW, Stamatoullas-Bastard A, et al. Itraconazole oral solution for primary prophylaxis of fungal infections in patients with hematological malignancy and profound neutropenia: a randomized, double-blind, double-placebo, multicenter trial comparing itraconazole and amphotericin B. *Antimicrob Agents Chemother* 2000; 44: 1887-93
119. Vandewoude K, Vogelaers D, Decruyenaere J, et al. Concentrations in plasma and safety of 7 days of intravenous itraconazole followed by 2 weeks of oral itraconazole solution in patients in intensive care units. *Antimicrob Agents Chemother* 1997; 41: 2714-8
120. Caillot D, Bassaris H, Seifert WF, et al. Efficacy, safety and pharmacokinetics of intravenous (IV) followed by oral itraconazole (ITR) in patients (pts) with invasive pulmonary aspergillosis (IPA) [abstract no. 1646]. 39th Interscience Conference on Antimicrobial Agents and Chemotherapy; 1999 Sep 26-29; San Francisco, California, USA. Washington DC, USA: American Society for Microbiology, 1999: 575
121. Annemans L, Moeremans K, Milligan DW, et al. Economic evaluation of intravenous itraconazole in presumed systemic fungal infections. 39th Interscience Conference on Antimicrobial Agents and Chemotherapy [abstract no. 1867]. 39th Interscience Conference on Antimicrobial Agents and Chemotherapy; 1999 Sep 26-29; San Francisco, California, USA. Washington DC, USA: American Society for Microbiology, 1999: 743

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