

Interferon- β Therapy in Multiple Sclerosis

Evidence for a Clinically Relevant Dose Response

Douglas S. Goodin

Department of Neurology, University of California, San Francisco, California, USA

Abstract

There have been considerable advances made recently in the treatment of multiple sclerosis (MS). In particular, interferon (IFN) β has been demonstrated in several independent, multicentre clinical trials to lower unequivocally the biological activity of this illness. The results of these trials have been remarkably consistent, demonstrating a reduction in both disease activity and cumulative disability, using a combination of clinical and magnetic resonance imaging outcome measures. Nevertheless, the importance of the total weekly IFN β dose in the clinical management of individual patients has been controversial.

However, there is considerable information available regarding the effect of IFN β dose on the various biochemical and clinical markers that are affected by IFN β , which is derived both from pre-clinical studies and multicentre clinical trials. On balance, convincing evidence is provided to support the notion that there is a clinically relevant dose-response in the use of IFN β to treat patients with relapsing/remitting MS. However, many of the clinical trials of IFN β in MS have confounded the potential effects of dose with the possible effects of frequency of IFN β administration. As a result, it is possible that the apparent dose-response observed in these clinical trials may be due, in part, to the more frequent dose administration schedule rather than the total weekly dose.

Interferon (IFN) β , a naturally occurring immunomodulatory glycoprotein, was the first clinically available agent demonstrated to be effective in modifying the disease course in patients with multiple sclerosis (MS). Two forms of IFN β – IFN β -1a (AvonexTM¹, Rebif[®]¹) and IFN β -1b (Betaseron[®]¹, Betaferon[®]¹) – have been assessed in clinical trials. Both forms have been shown to reduce the frequency and severity of relapses, and to slow the progression of neurological disability.^[1-10] Magnetic resonance imaging (MRI) studies have also dem-

onstrated that IFN β reduces the number of active lesions and slows the increase in total MRI lesion volume over time.^[1-10] Different regimens for each form of IFN β have already been approved for the treatment of MS by the US Food and Drug Administration (FDA) and European regulatory bodies. Nevertheless, uncertainties remain about the optimal clinical application of these agents in the treatment of patients with MS. In particular, the optimal dose of IFN β is uncertain, as is the optimal time to start therapy, and the optimal route of administration.

With regard to initiating therapy, most authorities favour early treatment and this practice is, in

¹ Use of a tradename is for identification purposes only and does not imply endorsement.

fact, the position of recent consensus statements.^[11,12] In addition, the recently presented trials of IFN β -1a in patients at high risk of developing MS have shown that early treatment significantly slows the subsequent rate of conversion to clinically definite MS.^[13,14] There is also now evidence showing that the inflammatory demyelination of MS is accompanied by early axonal injury,^[15,16] which precedes and, presumably, underlies subsequent neurological disability.^[17] This kind of evidence provides considerable empirical support for the practice of initiating disease-modifying therapies as early as possible during the course of the disease, with the aim of limiting both irreversible tissue damage and permanent disability.

However, there is less consensus on the optimal dosage and route of administration of IFN β .^[18-20] Nonetheless, there is substantial information available regarding the effect of IFN β dose on various biochemical and clinical markers. This evidence derives both from preclinical studies (*in vitro* and in animals) as well as from large multicentre clinical trials. It is the purpose of this manuscript to review these diverse pharmacological, clinical and MRI findings as they relate to the use of IFN β in clinical practice.

1. Immunomodulatory Effects of Interferon (IFN) β

The cause of MS is unknown, although considerable evidence indicates that activated T cells, auto-reactive to self antigens such as myelin basic protein (MBP) or myelin oligodendrocyte glycoprotein (MOG), proliferate, cross the blood-brain barrier and, under the influence of proinflammatory cytokines and cellular adhesion molecules, enter the central nervous system (CNS) [for reviews see^[21,22]]. Other mononuclear cells such as macrophages and, to a lesser extent, B cells are also present in active MS lesions. Within the CNS, in concert with resident cells such as astrocytes and microglia, these mononuclear cells produce inflammation and, thereby, damage both to the myelin and to the oligodendrocytes, resulting in irreversible axonal transection.^[23]

The mechanism by which IFN β exerts its disease-modifying effects in MS is also unknown but could potentially be through one or more of a number of immunomodulatory mechanisms.^[24]

1.1 Inhibition of Gelatinase Secretion and T Cell Migration

Migration of activated T cells across the blood-brain barrier appears to be a key step in the pathophysiology of MS.^[25] An important part of this process is the release by T cells of matrix metalloproteinases (MMPs) in response to stimulation by the pro-inflammatory cytokine interleukin (IL)-2. MMPs cleave type IV collagen of the extracellular matrix (a component of the blood-brain barrier), thereby facilitating the transendothelial migration of T cells into the CNS.^[21,22]

Leppert et al.^[26] demonstrated that pretreatment of T cells with IFN β -1b inhibited IL-2-dependent secretion of MMP-2 and MMP-9, and reduced MMP-dependent migration across an artificial basement membrane by up to 90%, without significantly affecting normal cell locomotion. IFN β also down-regulated expression of the IL-2 cell surface receptor and the affinity of IL-2 for the T cell surface, indicating a potential further mechanism for the therapeutic effect of IFN β . Importantly, both the effect on MMP secretion and T cell migration were dependent on the dose of IFN β .^[26] IFN β has also been shown to dose-dependently inhibit activated leucocyte transmigration through an activated human brain microvascular endothelial cell (HB-MVEC) monolayer.^[27] Prestimulation of HB-MVEC with tumour necrosis factor- α (TNF α) and IFN γ significantly promoted transepithelial migration of activated leucocytes.^[27] Again, this migration is inhibited by IFN β -1a in a dose-dependent manner, mediated through inhibition of TNF α , IL-1 and MMP-9 production.^[27]

1.2 Inhibition of IFN γ -Induced Major Histocompatibility Complex Class II Expression

Vascular endothelial cells play a central role in T cell trafficking into the CNS. Under the influence of pro-inflammatory cytokines such as IFN γ and

TNF α , vascular endothelial cells express both major histocompatibility complex (MHC) class I and II molecules and adhesion molecules. These molecules help to activate and adhere leucocytes, and to facilitate their migration across the endothelium. Miller and colleagues^[28] showed that addition of different doses of IFN β to human vascular endothelial cells incubated with IFN γ led to a down-regulation of IFN γ -induced class II molecule expression in a dose-dependent manner.

1.3 Inhibition of T Cell Proliferation and IFN γ Release

Proliferation and activation of auto-reactive T cells probably represent key events in the pathogenesis of MS. Noronha et al.^[29] demonstrated that IFN β significantly reduced *in vitro* mitogen-induced proliferation of T cells from both patients with MS and healthy individuals. Significantly, this reduction was dependent on the dose of IFN β in the concentration range 10 to 100 U/ml, and was observed irrespective of the mitogenic stimulus or the presence of IFN γ . Pre-activated T cells were down-regulated to similar extent. IFN β 1000 U/ml also down-regulated release of IFN γ from activated T cells obtained from patients with MS and healthy controls.

1.4 Increase in Interleukin-10 Expression

Inflammatory and autoimmune responses involve a complex interplay of mediators that either promote or inhibit immunological processes, and IFN β is known to exert inhibitory effects on immune promoters.^[24] For example, IL-10 is released by activated T cells and strongly inhibits cell-mediated immune responses.^[30-33] A study by Rudick et al.^[34] found that incubation of peripheral blood mononuclear cells (PBMC), *in vitro*, with IFN β -1a (varying in concentration between 0 and 1000 IU/ml) upregulated IL-10 mRNA levels in a dose-dependant manner.^[34] Serum IL-10 levels in healthy individuals and patients with MS were also increased after a single intramuscular injection of IFN β -1a and this effect was also dose-dependent at IFN β -1a doses of 6 and 12 MIU.^[34]

2. Effects of IFN β in Experimental Allergic Encephalomyelitis (EAE)

Experimental allergic encephalomyelitis (EAE) is a T cell-mediated autoimmune CNS disease and has been used extensively as an experimental model for MS (for reviews see^[35,36]).

2.1 Histological Impact of IFN in EAE

In a study by Abreu and colleagues,^[37] rats with a passively transferred form of EAE were treated with placebo or purified rat IFN at doses of 12 000, 40 000 or 120 000 U/kg. A clear dose response was observed on the number of lesions within the brain. The lowest IFN dose had no effect on the number of lesions (perivascular accumulation of mononuclear cells), but the 40 000 U/kg dose significantly reduced the number of these mononuclear cuffs and the 120 000 U/kg dose provided a further reduction, indicating a significant effect of IFN dose on the EAE disease process.

2.2 Effect of IFN β on Clinical Parameters in Mice with EAE

A later study by Yu et al.^[38] demonstrated that IFN β also exerts a therapeutic effect on the clinical manifestations of EAE. Mice with EAE received placebo or IFN β 5000 or 10000IU at the onset of clinical disease. Dose-dependent benefits were observed in several clinical parameters, including survival at 14 days, frequency of relapses, number of multiple relapses and time to first relapse. There was also a delay in the progression of disability to day 56. These clinical benefits were accompanied by a dose-dependent inhibition of *in vitro* proliferation of autoreactive T cells in the mice, as well as a reduction in inflammation and demyelination within the CNS.

3. Effects of IFN β on Non-Immunological Mediators

Although the mechanisms of action suggested for IFN β in the treatment of MS have emphasised its immunomodulatory effects, the influences of

IFN β on non-immunological processes may also be important.

3.1 Inhibition of Inducible Nitric Oxide Synthase

Nitric oxide generated by the inducible enzyme, nitric oxide synthase, has been implicated in damage to oligodendrocytes, contributing to the pathogenesis of MS.^[39] An *in vitro* study by Hua et al.^[40] showed that IFN β produced dose-dependent, selective and potent inhibition of IL-1 β /IFN γ -induced nitric oxide synthase expression in cultured human astrocytes. This observation suggests a potential mechanism by which IFN β may ameliorate inflammation and cytotoxicity within the CNS of patients with MS.

3.2 Promotion of Neurotrophin Nerve Growth Factor Release by Astrocytes

Growth factors released by astrocytes are important for oligodendrocyte development, maturation and survival; nerve growth factor (NGF) in particular stimulates adult porcine oligodendrocytes to extend processes and proliferate, and may also promote CNS remyelination.^[41] Boutros and colleagues^[42] showed that incubation of murine astrocytes in the presence of murine IFN β (concentration range 10 to 1000 U/ml) induced a dose-dependent release of NGF, up to 40 times that of untreated controls. If a similar effect occurs in humans, it may be one mechanism of IFN β action in MS. Interestingly, administration of NGF has also recently been shown to delay the onset and lessen the severity of EAE in marmosets.^[43] In this marmoset model, intrathecal administration of NGF seems to produce this anti-inflammatory effect by both the down-regulation of IFN γ and the up-regulation of IL-10 production by glial cells.^[43]

4. Pharmacodynamic, Pharmacological and Biological Effects of IFN β

IFN β is known to induce the expression of IFN-specific markers such as 2',5'-oligoadenylate synthetase (2',5'-OAS), neopterin, tryptophan, β_2 -

microglobulin and Mx protein.^[44-46] These markers reflect several biological activities of IFN β , such as MHC class I gene expression, antiviral and antiproliferative actions, and monocyte activation. These markers have been used to measure the pharmacodynamic response in clinical studies.^[47,48]

4.1 Dose Response: Pharmacodynamics After Single and Multiple Doses

Witt et al.^[49] examined the pharmacodynamic effects of IFN β -1b in 32 healthy individuals. Four groups of volunteers received 0.09, 0.9, 9 or 45MU ('old standard' MIUs – equivalent to the 'current standard' 0.018, 0.18, 1.8, and 8 MIU) as a single dose, then on 4 alternate days after a 7-day washout. Significant ($p < 0.02$) increases in β_2 -microglobulin and cellular 2',5'-OAS activity were observed at 24 hours for the 0.9MU dose. The 9 and 45MU doses were associated with significant increases in β_2 -microglobulin and cellular 2',5'-OAS, neopterin and tryptophan levels ($p < 0.01$). A clear dose response ($p < 0.01$) was evident for all markers over the IFN β -1b concentration range 0.09 to 45MU, and changes in all markers correlated with each other.

Similar results were reported recently by Stürzebecher et al.^[50] These authors contrasted the effects of IFN β -1a given subcutaneously or intramuscularly and IFN β -1b given subcutaneously to 75 healthy volunteers on the levels of human Mx protein, neopterin and 2',5'-OAS. Each patient in the study received a single dose, which was administered for IFN β -1a at doses of 1, 3, 6, 9 and 12 MIU and for IFN β -1b at doses of 2, 4, 8, 12 and 16 MIU. The production of all 3 markers was induced in a dose-dependent manner for both IFN β -1a and IFN β -1b. Moreover, although earlier authors had suggested that intramuscular administration produced greater IFN β activity than subcutaneous administration,^[47] this study found no differences in any of these biological effects between the 2 types of IFN β or between the different routes of administration. However, with regard to their effect on these biologic markers, intramuscular IFN β -1a 6 MIU was found to be approximately equal to sub-

cutaneous IFN β -1b 8 to 9 MIU.^[50] Importantly, even at the highest doses used in this study, these biological effects were not sustained for a week after a single injection.

4.2 Effect of Dose Frequency on Biological Effect Markers and Cytokine Release

Findings pertinent to the dosage schedule for IFN β were reported in a comparative study by Williams and Witt,^[51] in which healthy volunteers received either a single 6 MIU intramuscular dose of IFN β -1a (AvonexTM) or 8 MIU IFN β -1b (Betaseron[®]) subcutaneously every other day for 1 week. Biological response parameters reached similar maximum concentrations in both treatment groups. However, serum neopterin and β_2 -microglobulin levels remained significantly above baseline throughout the 7-day study among those receiving IFN β every other day, whereas this effect was sustained only to day 5 in the group given IFN β weekly. Furthermore, the overall induction of neopterin, β_2 -microglobulin and IL-10 levels, as assessed by area under the concentration-time curve (AUC), was significantly greater in the alternate-day group compared with the weekly ($p = 0.018$, $p = 0.031$, and $p = 0.014$, respectively). These data suggest that more frequent IFN β administration provides a more continuous biological response than once weekly administration.

In another study, 24 healthy individuals received IFN β -1a (Rebif[®]) subcutaneously at doses of 22 μ g once weekly, 66 μ g once weekly, 22 μ g 3 times a week or placebo for 4 weeks.^[52] Blood samples were obtained to assess pharmacodynamic responses; PBMCs from each sample were cultured and cytokine release measured with and without mitogenic stimuli. The immunomodulatory action of IFN β was found to be dose dependent. Moreover, the biological effect of the 66 μ g total weekly dose of IFN β was 2 to 3 times greater when divided into 3 injections compared with when it was administered as a single weekly injection, suggesting that more frequent administration may improve the biological effects of IFN β therapy.

A recent study investigated the levels of the anti-viral protein MxA following administration of the IFN β in 237 patients with clinically definite MS.^[53] 78 patients received IFN β -1b (Betaseron[®]) at a dosage of 8 MIU (250 μ g) every other day; 71 patients received IFN β -1a (Rebif[®]) at a dosage of 6 MIU (22 μ g) subcutaneously either weekly or 3 times weekly; and 21 patients received IFN β -1a (AvonexTM) at a dosage of 6 MIU (30 μ g) intramuscularly once weekly. Samples were drawn 1 to 2 days after IFN β administration, although, unfortunately, the time of MxA sampling was significantly delayed (by a half a day) in the Betaseron[®] group compared with the groups receiving other IFN β preparations. MxA stimulation in the untreated blood from 10 healthy volunteers was also studied. These authors reported that the levels of MxA were 2.29 ng/10⁵ peripheral blood lymphocytes (PBLs) in the Betaseron[®]-treated patients, 1.00 ng/10⁵ PBLs in the Rebif[®]-treated patients, and 0.57 ng/10⁵ PBLs in the AvonexTM-treated patients. *In vitro* stimulation of PBLs with all 3 of these agents resulted in a dose-dependant increase in MxA levels that was approximately equivalent (on an MIU for MIU basis) for each of the compounds. Although this trial seems somewhat poorly designed, these results, nevertheless, suggest that the biological activity of these different IFN β preparations are similar and that increasing the total weekly IFN β dose is associated with an increasing biological effect.

5. Clinical and Magnetic Resonance Imaging Evidence for a Dose Response in Patients with Multiple Sclerosis

Several multicentre clinical trials in patients with MS have demonstrated beneficial effects of IFN β on each of the 4 outcome measures in current clinical use, including clinical MS attack rate, clinical disability progression, MRI lesion activity and MRI lesion burden.^[1,5,7-9] Key results from the 4 principal phase III clinical trials of IFN β in relapsing-remitting MS (RRMS) are summarised in table I.

Table 1. Findings from 4 clinical trials of interferon- β (IFN β) in patients with relapsing remitting multiple sclerosis during years 1 and 2 by increasing IFN β dose. All comparisons are versus placebo in the same study^a

Study parameter	Betaseron [®] Trial ^[1-3]	Rebif [®] Trial ^[9]	Avonex [™] Trial ^[4,5]	Rebif [®] Trial ^[9]	Rebif [®] Trial ^[6,7]	Betaseron [®] Trial ^[1-3]	Rebif [®] Trial ^[6,7]
Dosage and administration information							
Total weekly dose of IFN β (MIU) ^b	5.6	6	6	12	18	28	36
weekly dose of IFN β (μ g)	175	22	30	44	66	875	132
Mean EDSS at entry	2.9	2.7	2.4	2.6	2.5	3.0	2.5
EDSS range at entry	0-5.5	0-5.0	1-3.5	0-5.0	0-5.0	0-5.5	0-5.0
Dose schedule (route)	qod (SC)	qw (SC)	qw (IM)	qw (SC)	tiw (SC)	qod (SC)	tiw (SC)
Measures of disease activity (%)							
Relapse rate, 1y	-15	0	-10 ^c	-19	-33****	-33****	-37****
Relapse rate, 2y (ITT)	-13*		-18*		-29**	-34***	-32****
Median time to first relapse	+18		+31		+70*	+86*	+113*
Relapse-free patients, 1y		0		-11	+59		+87
Relapse-free patients, 2y	+29		+42		+69*	+95**	+100**
Median MRI attack rate, 2y ^d	-67*		-33*		-67***	-83**	-78****
Measures of disease severity (%)							
Confirmed progression (1 EDSS point), 2y			-37*		-19*	-29	-30*
Final EDSS \geq 1 point more than baseline	0		-32			-31*	
Median change in MRI BOD, 2y	-5.1		-6.7		-12.1****	-17.3***	-14.7****

- a Percentage reductions (or increases) were calculated by dividing the reported rates in the treated group by the rates in the placebo group, except for MRI disease burden (BOD) which was calculated as the difference in % change between the treated and placebo groups.
- b Dosages of IFN β as expressed in millions of international units (MIU) of IFN β activity, as reported in the published papers. However, each company used a slightly different assay to measure IFN β activity and, therefore, the MIU scales are not directly comparable between the different publications. Approximate conversion formulas to reflect the different IFN β doses used are: (i) 6 MIU of Avonex[™] (30 μ g) is approximately equal to 9 MIU of Betaseron[®] (280 μ g). (ii) Avonex[™] and Rebif[®] are approximately equal in activity on a μ g basis.
- c Summary basis for US Food and Drug Administration approval.^[9,54]
- d MRI attack rate (activity) was measured differently in different trials. The Betaseron[®]^[1-3] and Rebif[®]^[6,7] trials include the number of new, recurrent and enlarging T2 lesions but not gadolinium (Gd) enhancement; The Avonex[™]^[4,5] trial used only the number of Gd enhancing lesions.

BOD = burden of disease; **EDSS** = Kurtzke Expanded Disability Status Scale; **IM** = intramuscular; **ITT** = intention to treat analysis; **MRI** = magnetic resonance imaging; **qod** = every other day; **qw** = once per week; **SC** = subcutaneous; **tiw** = 3 times per week.
* $p \leq 0.05$; ** $p \leq 0.01$; *** $p \leq 0.001$; **** $p \leq 0.0001$ compared with controls.

5.1 Dose-Finding Studies Using IFN β

One of the earliest clinical trials of IFN β in patients with MS was that of Knobler et al.^[54] in which 30 patients (5 groups) with RRMS were administered either placebo or IFN β -1b (Betaseron[®]) at doses of 0.8, 4, 8, or 16 MIU subcutaneously 3 times weekly for 24 weeks. This trial demonstrated a clear trend in favour of higher dosages of IFN β in the ability to reduce attack frequency over this short interval. Indeed, although the highest dosage

used (48 MIU per week) was poorly tolerated and several patients required dose reductions because of adverse effects, the observed attack rate at this IFN β dose was actually zero. A similar dose-effect also seems to be present using intramuscular IFN β -1a (Avonex[™]).^[47,55,56] Thus, following treatment with IFN β -1a, 6 MIU intramuscularly, the reported maximum concentration (C_{max}) for IFN activity was 34 U/ml and the AUC was 824 U/ml \square h.^[47] Following treatment with 12 MIU, these values were 44 and 1352, respectively,^[56] whereas fol-

lowing 15 MIU, they were 103 and 2242, respectively.^[55]

5.2 Phase III Clinical Trials of IFN β

The multicentre study of IFN β -1b (Betaseron®) in MS included 372 patients with RRMS who had scores on the extended disability status scale (EDSS) of 0 to 5.5.^[1,2] Patients were randomised to receive placebo, low dose (1.6 MIU; 50 μ g) or high dose (8 MIU; 250 μ g) IFN β -1b, subcutaneously, every other day for 2 years. Primary endpoints of the trial were relapse rate and proportion of patients remaining relapse free. Secondary outcomes included clinical disease progression, number of active MRI lesions (new, recurrent, or enlarged) and the total disease burden as measured by the volume of T2 white matter disease seen on MRI. After 2 years, compared with placebo, treatment with high dose IFN β -1b reduced both the clinical attack rate (–34%; $p < 0.0001$) and the MRI attack rate as measured by median number of T2 active lesions (–83%; $p < 0.009$). The median volume of T2 white matter disease seen on MRI (burden of disease) increased by 16.5% compared with baseline among placebo-treated patients, whereas with IFN β -1b treatment it decreased by 0.8% (i.e. a 17.3% difference compared with placebo; $p = 0.001$). The high dose also reduced the rate of confirmed 1-point EDSS progression but this was not significant (–29%; $p = 0.16$). In this trial, treatment with low dose IFN β -1b (5.6 MIU per week) was also better than placebo on some outcomes, although it was not as effective as the higher dose.

This study extended for up to 5 years in some patients^[3] and the data demonstrated a persistent dose-dependent reduction in relapse rate (approximately 30% reduction in 8 MIU group compared with placebo in all 5 years). Furthermore, in the high dose group there was no significant increase in MRI disease burden compared to baseline, either yearly or for the entire duration of the study, whereas among patients receiving the low dose there was a significant increase each year ($p < 0.05$), with the exception of year 5.

The multicentre trial of IFN β -1a (Avonex™) included 301 patients with RRMS treated with placebo or intramuscular IFN β -1a 6 MIU per week (30 μ g per week).^[4] After 2 years, treatment with Avonex™ resulted in marginally significant reductions in the clinical attack rate (–18%; $p = 0.04$), the MRI attack rate as measured by the median number of gadolinium enhancing lesions (–33%; $p = 0.05$), and the rate of confirmed 1-point EDSS progression (–37%; $p = 0.02$) compared with placebo. The total volume of MRI T2 disease burden was also reduced but this was not significant (–6.7 %; $p = 0.36$). Because the design of this trial only included a single treatment arm, the question of the proper administration of IFN β -1a cannot be addressed from these results. Nevertheless, the generally greater effect sizes and the greater statistical significance of the Betaseron® trial compared with the Avonex™ trial, together with the fact that dose seemed to be an important consideration in the Betaseron® trial, suggests that dose may also be an important therapeutic consideration when using IFN β -1a in clinical practice.

In fact, this point is underscored by the results of the IFN β -1a (Rebif®) trial,^[6,7] in which a total of 560 patients with RRMS were treated for 2 years with placebo or IFN β -1a at doses of either 22 μ g (6 MIU) or 44 μ g (12 MIU) subcutaneously 3 times weekly. This trial is the first to report a statistically significant benefit on each of the 4 major outcome measures that are in current clinical use. Thus, compared with placebo, treatment with IFN β -1a 132 μ g per week (36 MIU per week) reduced the clinical relapse rate (–32%; $p < 0.005$), the MRI attack rate as measured by median number of T2 active lesions (–78%; $p < 0.0001$) and the MRI T2 volume of white matter disease (–14.7%; $p \leq 0.0001$; i.e. an increase of 10.9% *versus* baseline for placebo, compared with a decrease of 3.8% for IFN β -1a), and a reduction in the 1-point EDSS progression rate (–30%; $p < 0.05$). Treatment with IFN β -1a (66 μ g per week), subcutaneously was also highly effective on each of these outcome measures.

However, with the exception of the outcome of T2 active lesions, high dose IFN β -1a was not sta-

tistically better than the low dose on these clinical parameters at the 2-year time-point. Nevertheless, there was a trend favouring high dose on each of these other outcome measures.^[6,7] In addition, combined analysis of the results of the AvonexTM^[4] and Rebif[®]^[6,9] trials demonstrates a marked difference in efficacy (as measured by clinical relapse rate at 1 year) across the spectrum of administration from 22 to 132 µg weekly.^[9]

The IFNβ-1a (Rebif[®]) trial^[6] was continued for an additional 2 years.^[8] Placebo-treated patients were re-randomised in a blinded fashion to receive either IFNβ-1a 22 or 44 µg three times weekly. At the 4-year time-point a statistically significant dose response was apparent for selected clinical and MRI outcome measures.^[8] Thus, the high dose (132 µg per week) reduced the relapse rate during years 3 to 4, prolonged the time to second relapse and increased the proportion of relapse-free patients compared with the low dose of 66 µg per week ($p < 0.05$). Similarly, high dose IFNβ-1a was significantly better than low dose at reducing the MRI disease burden and T2 lesion activity ($p < 0.001$).

Recently, two head-to-head comparative trials have been reported in preliminary form.^[57,58] The first non-blind trial^[57] compared IFNβ-1b (Betaseron[®]; 28 MIU per week subcutaneously) to IFNβ-1a (AvonexTM; 30 µg per week intramuscularly) in 188 patients with RRMS over 2 years. After the first year of treatment, a significantly greater clinical benefit in the IFNβ-1b group (compared to the IFNβ-1a group) was reported for the clinical outcomes of relapse-free status and sustained progression, and for the MRI outcomes of new T2 lesions and gadolinium-enhancing lesions. The second was a randomised, single-blind, multicentre trial^[58] that compared high-dose IFNβ-1a (Rebif[®]; 132 µg per week subcutaneously) to low-dose IFNβ-1a (AvonexTM; 30 µg per week intramuscularly) in 677 patients with RRMS. After only 6 months of therapy, results for the high-dose group were significantly superior to the low-dose group on every clinical and MRI outcome measure related to attack rate examined in the trial, including the odds of

remaining attack-free, the attack rate, the time to first exacerbation, the odds of having no new T1 or T2 lesions, the total number of new lesions and the cumulative number of new active lesions.^[58] However, each of these trials varied both the total IFNβ dose and the frequency of IFNβ administration so that the relative contribution of each factor to the final results cannot be easily judged. Indeed, a recently presented comparative trial^[59] of IFNβ-1a (AvonexTM) at doses of 30 µg or 60 µg, intramuscularly, once weekly, found no significant difference on any outcome measure over 3 years between the two dosages studied. This result suggests that the frequency of IFNβ administration (in addition to total dose) may be an important therapeutic consideration.

6. Discussion

In summary, on the basis of several independent multicentre clinical trials, IFNβ therapy has demonstrated unequivocal efficacy in modifying the disease course in relapsing MS. Presumably, this therapeutic effect is mediated through one or more of a wide range of immunomodulatory and anti-inflammatory actions of IFNβ, although the actual mechanism of action remains to be elucidated. In addition, presumably the mechanism of action for IFNβ-1a and IFNβ-1b is the same and that the differences in outcome between the various clinical trials reflects both random error and dosage differences rather than a true difference in therapeutic effect between products.

Evidence now supports the value of early intervention with disease-modifying therapies in the hope of preventing the axonal injury that presumably precedes functional loss. As a result, it seems that the optimal therapy for MS should be that which provides maximum modulation of its pathogenic mechanisms and minimises damage to oligodendrocytes and axons, while, at the same time, produces the least adverse reactions. In the case of IFNβ, this would seem to translate into a recommendation that patients be given the highest dose that can be tolerated.

Importantly, evidence from clinical studies demonstrate a dose-response relationship for IFN β .^[1-3,6-9,57,58] The *in vitro* and animal data also support a dose response for many biological actions of IFN β that are of potential relevance to its clinical therapeutic effect.^[25-29,34,37,38,40,42] Moreover, pharmacodynamic studies indicate that the biological effects of IFN β are relatively short-lived, suggesting that the maximum clinical benefit of IFN β therapy may be achieved with administration that is more frequent than once weekly.^[47,49,50,52,56] Moreover, the clinical data suggest that, using the same dose schedule, higher doses are generally more effective than lower doses.^[1-3,6-9]

The concept of a dose response for IFN β in patients with MS is not surprising. A similar dose response has been reported in other conditions for which IFN therapy has been used, such as chronic^[60] and acute^[61] hepatitis, various cancers^[62-64] and anogenital warts.^[65]

However, as with any therapy, tolerability and the risk of toxicity place a limit on dose levels, and avoidance of adverse effects has been a significant concern in clinical trials with IFN β . As discussed in section 5.1, a pilot study with IFN β -1b reported that a dose of 16 MIU 3 times weekly was associated with unacceptable levels of adverse events including chills, fever and injection site reactions.^[55] However, more recently the prophylactic use of ibuprofen and paracetamol (acetaminophen) in trials with IFN β have proved useful in minimising unwanted effects. Indeed, in a recent comparison of IFN β -1b (Betaseron[®]) administered 3 times weekly versus IFN β -1a (Avonex[™]) once weekly, the incidence, duration and severity of adverse effects (mainly fever, chills and myalgia for 6 to 12 hours after first dose of either agent) did not differ between treatments.^[51] Moreover, the major IFN β clinical trials have generally found that treatment is well tolerated irrespective of dose.^[1-10]

Nevertheless, it seems that, at least for some of the outcomes, the therapeutic benefits of IFN β are approaching a plateau at the highest doses in current use.^[9] As a result, it seems unlikely that IFN β

(by itself) will eliminate the clinical activity of MS, even if substantially higher doses could be tolerated by patients.

7. Conclusion

There seems to be compelling evidence from multiple sources (including both clinical trials and pharmacodynamic data) to support the notion that there is a clinically relevant dose-response in the use of IFN β to treat patients with RRMS. Nevertheless, many of the clinical trials of IFN β in MS have confounded the potential effects of dose with the possible effects of frequency of IFN β administration. As a result, it is possible that a portion of the apparent dose-response observed in these clinical trials may be due to the more frequent dose administration schedule rather than the total weekly dose.

Acknowledgements

The author of this manuscript has participated (or is currently participating) in several industry-sponsored clinical therapeutic trials in multiple sclerosis. The sponsoring pharmaceutical companies for these trials have included (or do include): Biogen, Inc; Berlex Laboratories; Immunex Corp; Serono, Inc; and Teva Marion Partners. In addition, the author has lectured extensively at both medical conferences and in public on various aspects of the diagnosis and management of multiple sclerosis. In many cases these talks have been sponsored by non-restricted educational grants from one or another of each of the above-listed companies or by Athena Neurosciences.

References

1. The IFNB Multiple Sclerosis Study Group. Interferon beta-1b is effective in relapsing-remitting multiple sclerosis. I. Clinical results of a multicenter, randomised, double-blind, placebo-controlled trial. *Neurology* 1993; 43: 655-61
2. Paty DW, Li DKB, UBC MS/MRI Study Group, et al. Interferon beta-1b is effective in relapsing-remitting multiple sclerosis. II. MRI analysis results of a multicenter, randomised, double-blind, placebo-controlled trial. *Neurology* 1993; 43: 662-7
3. The IFNB Multiple Sclerosis Study Group and the University of British Columbia MS/MRI Analysis Group. Interferon beta-1b in the treatment of multiple sclerosis: final outcome of the randomised controlled trial. *Neurology* 1995; 45: 1277-85
4. Jacobs LD, Cookfair DL, Rudick RA, et al. Intramuscular interferon beta-1a for disease progression in relapsing multiple sclerosis. *Ann Neurol* 1996; 39: 285-94

5. Simon JH, Jacobs LD, Campion M, et al. Magnetic resonance studies of intramuscular interferon β -1a for relapsing multiple sclerosis. *Ann Neurol* 1996; 43: 79-87
6. PRISMS Study Group. Randomised double-blind placebo-controlled study of interferon β -1a in relapsing-remitting multiple sclerosis. *Lancet* 1998; 352: 1498-1504
7. Li DH, Paty DW, UBC MS/MRI Analysis Research Group, et al. Magnetic resonance imaging results of the PRISMS trial: a randomised, double-blind, placebo-controlled study of interferon- β 1a in relapsing-remitting multiple sclerosis. *Ann Neurol* 1999; 4: 197-206
8. PRISMS Study Group. PRISMS-4: long term efficacy of interferon β -1a in relapsing MS. *Neurology* 2001; 56: 1628-36
9. OWIMS Study Group. Evidence of interferon β -1a dose response in relapsing remitting MS. The OWIMS study. *Neurology* 1999; 53: 679-86
10. European Study Group on Interferon β -1b in Secondary Progressive MS. Placebo-controlled multicentre randomised trial of interferon β -1b in treatment of secondary progressive multiple sclerosis. *Lancet* 1998; 352: 1491-7
11. Oger J, Freedman M. Consensus statement on the Canadian MS clinical network on: the use of disease modifying agents in multiple sclerosis. *Can J Neurol Sci* 1999; 26: 274
12. National Multiple Sclerosis Society disease management consensus statement. The National Multiple Sclerosis Society, New York, NY, USA, 1999
13. Jacobs LD, Beck RW, Simon JH, et al. and the CHAMPS Study Group. Intramuscular interferon beta-1a therapy initiated during a first demyelinating event in multiple sclerosis. *N Eng J Med* 2000; 343: 898-904
14. Comi G, Filippi M, Barkhof F, et al. Effect of early interferon treatment on conversion to definite multiple sclerosis: a randomised study. *Lancet* 2001; 357: 1576-82
15. Ferguson B, Matyszak MK, Esiri MM, et al. Axonal damage in acute multiple sclerosis lesions. *Brain* 1997; 120: 393-9
16. Trapp BD, Peterson J, Ransohoff RM, et al. Axonal transection in the lesions of multiple sclerosis. *N Engl J Med* 1998; 338: 278-85
17. Trapp BD, Ransohoff R, Rudick R. Axonal pathology in multiple sclerosis: relationship to neurologic disability. *Curr Opin Neurol* 1999; 12: 295-302
18. Goodkin DE. Interferon therapy for multiple sclerosis. *Lancet* 1998; 352: 1486-7
19. Johnson KP, Panitch HJ, Herndon RM, et al. Interferon therapy for multiple sclerosis. *Lancet* 1999; 353: 494-8
20. Goodin DS. Perils and pitfalls in the interpretation of clinical trials. A reflection on the recent experience in multiple sclerosis. *Neuroepidemiology* 1999; 18: 53-63
21. Bar-Or A, Oliveira EML, Anderson DE, et al. Molecular pathogenesis of multiple sclerosis. *J Neuroimmunol* 1999; 100: 252-9
22. Conlon P, Oksenberg JR, Zhang J. The immunobiology of multiple sclerosis: an autoimmune disease of the central nervous system. *Neurobiol Dis* 6; 1999: 149-66
23. Trapp BD, Bo L, Mork S, et al. Pathogenesis of tissue injury in MS lesions. *J Neuroimmunol* 1999; 98: 49-56
24. Yong VW, Chabot S, Stuve O, et al. Interferon beta in the treatment of multiple sclerosis: mechanisms of action. *Neurology* 1998; 51: 682-9
25. Stuve O, Dooley NP, Uhm JH, et al. Interferon beta-1b decreases the migration of T lymphocytes *in vitro*: effects on matrix metalloproteinase-9. *Ann Neurol* 1996; 40 (6): 853-63
26. Leppert D, Waubant E, Burk MR, et al. Interferon beta-1b inhibits gelatinase secretion and *in vitro* migration of human T-cells: a possible mechanism for treatment efficacy in multiple sclerosis. *Ann Neurol* 1996; 40 (6): 846-852
27. Lou J, Gasche Y, Zheng L, et al. Interferon β inhibits activated leucocyte migration through human brain microvascular endothelial cell monolayer. *Lab Invest* 1999; 79: 1015-25
28. Miller A, Lanir N, Shapiro S, et al. Immunoregulatory effects of interferon- β and interacting cytokines on human vascular endothelial cells. Implications for multiple sclerosis and other autoimmune diseases. *J Neuroimmunol* 1996; 64: 151-61
29. Noronha A, Toscas A, Jensen MA. Interferon β decreases T-cell activation and interferon γ production in multiple sclerosis. *J Neuroimmunol* 1993; 46: 145-54
30. Mosmann TR, Moore KW. The role of IL-10 in crossregulation of TH1 and TH2 responses. *Immunol Today* 1991; 12: 49-53
31. Wang P, Wu P, Anthes JC, et al. Interleukin-10 inhibits interleukin-8 production in human neutrophils. *Blood* 1994; 83: 2678-83
32. Joyce DA, Gibbons DP, Green P, et al. Two inhibitors of pro-inflammatory cytokine release, interleukin-10 and interleukin-4, have contrasting effects on release of soluble p75 tumour necrosis factor receptor by cultured monocytes. *Eur J Immunol* 1994; 24: 2699-705
33. Itoh K, Inoue T, Ito K, et al. The interplay of interleukin-10 (IL-10) and interleukin-2 (IL-2) in humoral immune responses: IL-10 synergizes with IL-2 to enhance responses of human B lymphocytes in a mechanism which is different from upregulation of CD25 expression. *Cell Immunol* 1994; 157: 478-88
34. Rudick RA, Ransohoff RM, Peppler R, et al. Interferon beta induces interleukin-10 expression: relevance to multiple sclerosis. *Ann Neurol* 1996; 40: 618-27
35. Gold R, Hartung H-P, Toyka KV. Animal models for autoimmune demyelinating disorders of the nervous system. *Mol Med Today* 2000; 6: 88-91
36. Steinman L. Assessment of animal models for MS and demyelinating disease in the design of rational therapy. *Neuron* 1999; 24: 511-4
37. Abreu SL, Tondreau J, Levine S, Sowinski R. Inhibition of passive localised experimental allergic encephalomyelitis by interferon. *Int Archs Allerg Appl Immun* 1983; 72: 30-3
38. Yu M, Nishiyama A, Trapp BD, et al. Interferon- β inhibits progression of relapsing-remitting experimental autoimmune encephalomyelitis. *J Neuroimmunol* 1996; 64: 91-100
39. Merrill JE, Ignarro LJ, Sherman MP, et al. Microglial cell cytotoxicity of oligodendrocytes is mediated through nitric oxide. *J Immunol* 1993; 151: 2132-41
40. Hua LL, Liu JS, Brosnan CF, et al. Selective inhibition of human glial inducible nitric oxide synthase by interferon-beta: implications for multiple sclerosis. *Ann Neurol* 1998; 43 (3): 384-7
41. Althaus HH, Klöppner S, Schmidt-Schulz T, et al. Nerve growth factor induces proliferation and enhances fibre regeneration in oligodendrocytes isolated from adult pig brain. *Neurosci Lett* 1992; 135: 219-23
42. Boutros T, Croze E, Yong VW. Interferon- β is a potent promoter of nerve growth factor production by astrocytes. *J Neurochem* 1997; 69: 939-46
43. Villoslada P, Hauser SL, Bartke I, et al. Human nerve growth factor protects common marmosets against autoimmune encephalomyelitis by switching the balance of T helper cell type 1 and 2 cytokines within the central nervous system. *J Exp Med* 2000; 191: 1799-1806
44. Pestka S, Langer JA, Zoon KC, et al. IFNs and their actions. *Ann Rev Biochem* 1987; 56: 727-72

45. Borden E, Paulnock D, Spear G, et al. Biological response modification in man: measurement of interferon induced proteins. In: Baron S, Dianzani F, et al., (editors). The interferon system: a current review. University of Texas, Austin (TX), 1986: 1-7
46. Liberati AM, Horisberger MA, Palmisano L, et al. Double-blind randomised phase I study on the clinical tolerance and biological effects of natural and recombinant human interferon beta. *J Interferon Res* 1992; 12: 329-36
47. Alam J, Goelz S, Rioux P, et al. Comparative pharmacokinetics and pharmacodynamics of two recombinant human interferon beta-1a (IFN β -1a) products administered intramuscularly in healthy male and female volunteers. *Pharmaceut Res* 1997; 14 (4): 546-9
48. Munafo A, Trincharad-Lugan I, Nguyen TXQ, et al. Comparative pharmacokinetics and pharmacodynamics of recombinant human interferon beta-1a after intramuscular and subcutaneous administration. *Eur J Neurol* 1998; 5: 187-93
49. Witt PL, Storer BE, Bryan GT, et al. Pharmacodynamics of biological response *in vivo* after single and multiple doses of interferon- β . *J Immunother* 1993; 13 (3): 191-200
50. Stürzebecher S, Maibauer R, Heuner A, et al. Pharmacodynamic comparison of single doses of IFN β 1a and IFN β 1b in healthy volunteers. *J Interferon Cytokine Res* 1999; 19: 1257-64
51. Williams GJ, Witt PL. Comparative study of the pharmacodynamic and pharmacologic effects of Betaseron[®] and Avonex[™]. *J Interferon Cytokine Res* 1998; 18: 967-75
52. Rothuizen LE, Buclin T, Spertini F, et al. Influence of interferon β -1a dose frequency on PBMC cytokine secretion and biological effect markers. *J Neuroimmunol* 1999; 99: 131-41
53. Deisenhammer F, Mayringer I, Harvey J, et al. A comparative study of the relative bioavailability of different interferon beta preparations. *Neurology* 2000; 54: 2055-60
54. Knobler RL, Greenstein JI, Johnson KP, et al. Systemic recombinant human interferon-beta treatment of relapsing-remitting multiple sclerosis: pilot study analysis and six-year follow-up. *J Interferon Res* 1993; 13: 333-40
55. Alam J, McAllister A, Scaramucci J, et al. Pharmacokinetics and pharmacodynamics of interferon beta-1a in healthy volunteers after intravenous, subcutaneous or intramuscular administration. *Clin Drug Invest* 1997; 14: 35-43
56. Biogen Inc., 1995. Summary basis of approval. FDA official document for license of interferon beta-1a (Avonex[™]). Available from URL: www.fda.gov/cber/products/ifnbio051796.htm
57. Durelli L, Ferrero T, Ghezzi G, et al. The independent comparison of interferon (INCOMIN) trial: a multicenter randomized trial comparing clinical and MRI efficacy of IFN beta-1a and beta-1b in multiple sclerosis [abstract]. *Neurology* 2001; 56 Suppl. 3: A148
58. Coyle P. Results of comparative efficacy trial using two formulations of interferon beta-1a in RRMS [abstract]. *J Neuro Sci* 2001; 187 Suppl. 2: S436
59. Clanet M, Kappos L, Radue EW, et al. Results of the European interferon beta-1a (Avonex) dose-comparison study. *J Neurol* 2001; 248 Suppl. 2: II/63.
60. Guan R, Yeoh KG, Yap I, et al. Subcutaneously administered recombinant human β -interferon in the treatment of chronic hepatitis B virus infection. *Aliment Pharmacol Ther* 1996; 10: 807-14
61. Takano S, Satomura Y, Omata M, and Japan Acute Hepatitis Cooperative Study Group. Effects of interferon beta on non-A, non-B acute hepatitis: a prospective, randomised, controlled-dose study. *Gastroenterology* 1994; 107: 805-11
62. Ravandi F, Estrov Z, Kurzrock R, et al. A phase I study of recombinant interferon- β in patients with advanced malignant disease. *Clin Cancer Res* 1999; 5: 3990-8
63. Borden EC, Rinehart JJ, Storer BE, et al. Biological and clinical effects of interferon β ser at two doses. *J Interferon Res* 1990; 10: 559-70
64. Fine HA, Wen PY, Robertson M, et al. A phase I trial of a new recombinant human β -interferon (BG9015) for the treatment of patients with recurrent gliomas. *Clin Cancer Res* 1997; 3: 381-7
65. Bonnez W, Oakes D, Bailey-Farchione A, et al. A randomised, double-blind trial of parenteral low dose versus high dose interferon- β in combination with cryotherapy for treatment of condyloma acuminatum. *Antiviral Research* 1997; 35: 41-52

Correspondence and offprints: Dr Douglas S. Goodin, Department of Neurology, Rm M-794, University of California, San Francisco, 505 Parnassus Ave, San Francisco, CA 94143-0114, USA.
E-mail: dsg@itsa.ucsf.edu