

Pharmacological Neuroprotection for Glaucoma

Glyn Chidlow,^{1,2} John P.M. Wood^{1,2} and Robert J. Casson^{1,2}

1 Ophthalmic Research Laboratories, South Australian Institute of Ophthalmology, Hanson Institute, Adelaide, South Australia, Australia

2 University of Adelaide, Adelaide, South Australia, Australia

Contents

Abstract	726
1. Epidemiology and Definitions	726
2. Outline of Relevant Pathology	728
3. Pathogenesis of Glaucoma	728
3.1 The Mechanical Theory	728
3.2 The Vascular Hypothesis	729
3.3 Autoimmunity and Heat Shock Proteins	731
4. Laboratory Models of Glaucoma	731
5. Criteria for Evaluating Neuroprotection	732
6. Targets for Neuroprotection	733
6.1 Glutamate Receptors	733
6.1.1 Is There Evidence for Increased Glutamate in Glaucoma?	733
6.1.2 NMDA Receptor Antagonists	734
6.1.3 How are NMDA Receptor Antagonists Neuroprotective in Experimental Glaucoma?	735
6.1.4 Summary	735
6.2 Boosting Protective Autoimmunity	735
6.2.1 Glatiramer Acetate	736
6.2.2 Summary	736
6.3 Neurotrophin Deprivation	736
6.3.1 Obstructed Axonal Transport	737
6.3.2 Neurotrophin Deprivation	737
6.3.3 Brain-Derived Neurotrophic Factor	737
6.3.4 Other Neurotrophic Factors	737
6.3.5 Summary	738
6.4 Nitric Oxide Synthesis	738
6.4.1 Aminoguanidine	738
6.4.2 Summary	739
6.5 Voltage-Gated Sodium Channels	739
6.5.1 Phenytoin	739
6.5.2 Summary	740
6.6 Voltage-Gated Calcium Channels	740
6.7 Oxidative Stress	740
6.7.1 α -Tocopherol	741
6.7.2 <i>Ginkgo biloba</i>	741
6.7.3 Other Free Radical Scavengers	742
6.7.4 Summary	742
6.8 Heat Shock Proteins	742
6.8.1 Geranylgeranylacetone	743
6.8.2 Summary	743
6.9 Retinal Ganglion Cell Death and Apoptosis	743
6.9.1 Evidence for Apoptosis in Glaucoma	743

6.9.2	Apoptotic Pathways in Glaucoma	744
6.9.3	Pharmacological Intervention in Apoptosis	745
6.9.4	Summary	746
7.	Neuroprotective Properties of Drugs Already Used as Ocular Hypotensives	746
7.1	α_2 -Adrenoceptor Antagonists	747
7.1.1	Brimonidine	747
7.2	β -Adrenoceptor Antagonists (β -Blockers)	748
7.2.1	Betaxolol	748
7.2.2	Timolol	748
7.2.3	Metipranolol	749
7.3	Prostaglandin Derivatives	749
7.4	Carbonic Anhydrase Inhibitors	750
7.5	Summary	750
8.	Conclusions	750

Abstract

Glaucoma represents a group of neurodegenerative diseases characterised by structural damage to the optic nerve and slow, progressive death of retinal ganglion cells (RGCs). Elevated intraocular pressure is traditionally considered to be the most important risk factor for glaucoma, and treatment options for the disease have hitherto been limited to its reduction. However, visual field loss and RGC death continue to occur in patients with well controlled intraocular pressures and, thus, a consensus has recently emerged that additional treatment strategies are needed.

One such strategy is pharmacological neuroprotection, which in the context of glaucoma, refers to the situation in which a drug is deployed to interact with neuronal or glial elements within the retina/optic nerve head and thereby facilitate the survival of RGCs. The advent of animal models of chronic glaucoma has enhanced our understanding of many of the pathological processes occurring in glaucoma and, in doing so, described logical targets for pharmacological intervention. Such targets, which have been manipulated with varying degrees of success in relevant animal paradigms include glutamate receptors, autoimmune elements, neurotrophin deprivation, nitric oxide synthesis, oxidative stress products, sodium and calcium channels, heat shock proteins and apoptotic pathways.

With exciting data now emerging from many research laboratories, it is obvious that pharmacological neuroprotection for glaucoma without doubt represents an exciting development in the search for a treatment modality for this debilitating disease.

1. Epidemiology and Definitions

Glaucomatous optic neuropathy (glaucoma) represents a group of neurodegenerative diseases characterised by structural damage to the optic nerve (ON) and slow, progressive death of retinal ganglion cells (RGCs). It is the second most common cause of blindness worldwide.^[1] Risk factors for glaucoma include an elevated intraocular pressure (IOP), increasing age, family history, race and myopia. Historically, IOP was included in the definition of the

disease; subtypes of glaucoma were classified according to whether the IOP was elevated or not, whether the aqueous outflow channels appeared to be open or closed, and whether or not a cause for an elevated IOP, if present, could be detected. Although IOP has now been removed from the definition and the pathogenesis of the disease is poorly understood, there is convincing evidence that glaucoma is an IOP-sensitive optic neuropathy,^[2-5] and currently, the only clinically proven treatment is to reduce the IOP. This can be achieved either by

medical or surgical means.^[3-5] IOP reduction halts the progression of the disease in the majority of patients, but there remain a recalcitrant group who continue to lose vision despite very low IOPs.^[4] Furthermore, IOP reduction to the idealised levels can often not be achieved. Thus, a consensus has emerged proposing that additional treatment strategies are needed.

The term 'neuroprotection' is a relatively recent addition to the ophthalmological lexicon. It refers to physiological protection of undamaged, and rescue of damaged, RGCs and their axons embedded in a hostile environment. Most frequently the term is used in a pharmacological context, in which case a drug may be described as a neuroprotectant or as having neuroprotective properties. In ophthalmic research, the concept of neuroprotection has been regularly applied to diseases of the retina and ON, and in particular to glaucoma. Although any treatment strategy that preserves RGCs in glaucoma could be described as neuroprotective, most researchers limit use of this term to a drug that directly interacts with neuronal or glial elements within the retina/optic nerve head (ONH), thereby facilitating the survival of RGCs.^[6-8] Many of the concepts *à propos* neuroprotection in glaucoma have been derived from the study of other neurodegenerative diseases. For example, there is some support for the idea that a proportion of RGCs in glaucoma may be lost via secondary degeneration.^[9,10] Secondary degeneration is a phenomenon described in neurodegenerative diseases of the CNS in which neurons that are spared a primary insult are damaged by toxic products released from injured neighbouring neurons and glia. Thus, secondary degeneration is evident in a penumbra or zone that surrounds the ischaemic/traumatic lesion in affected brain tissue. A similar scenario is envisaged in glaucoma in which dying RGCs have the capacity to compromise the health of adjacent neurons.

A related concept to neuroprotection, and one that is sometimes referred to as 'vasoprotection',^[11] is the capacity of drugs to improve blood flow to the ONH. Numerous studies, utilising a variety of techniques, have demonstrated that, on average, perfusion to the ONH is decreased in glaucoma patients.^[12] Administration of drugs, such as calcium

channel antagonists, that counteract vascular insufficiency at the ONH via direct vasodilatory effects on blood vessels represents a potential therapeutic strategy for glaucoma; furthermore, some drugs may improve blood flow to the ONH and also have neuroprotective properties. Detailed discussion of the effect of drugs on ONH perfusion is, however, largely beyond the scope of this review and readers are directed to the comprehensive review by Costa and colleagues.^[13]

All of the drugs that are currently used for the treatment of glaucoma lower IOP. Some of these agents also have laboratory-proven neuroprotective properties;^[14,15] in some cases there is tantalising clinical evidence to support the laboratory findings. A large-scale, prospective, randomised study is currently evaluating the neuroprotective potential of one topical agent, brimonidine, for the treatment of normal tension glaucoma.^[16] Furthermore, there is evidence that drugs which have no effect on IOP may be neuroprotective in glaucoma. A 5-year, prospective, phase III clinical trial is currently underway investigating the effectiveness of the NMDA receptor antagonist memantine in open-angle glaucoma, while the vaccine glatiramer acetate, which displays neuroprotective properties in animal models of glaucoma, is expected to enter phase II trials in the near future.

Effective neuroprotection for glaucoma would offer a new treatment strategy and have a profound clinical impact. This review identifies potential targets for neuroprotection in glaucoma, discusses the evidence in favour of each target being a valid point for intervention, and highlights pharmacological classes of compounds that might prove useful as therapeutic agents. The search engine PubMed was primarily used to identify relevant references using numerous key words, such as neuroprotection, glaucoma, ganglion cell, retina, optic nerve and ocular hypertension in combination with keywords relevant to each individual section of the article. Papers for those references, as well as pertinent review articles and original publications cited by others, were evaluated for inclusion in this article. The final search data for inclusion of references within the article was 31 December 2006.

2. Outline of Relevant Pathology

The most striking clinical feature of glaucoma is the characteristic changes at the ONH, and there is a consensus that the primary site of injury in glaucoma is at this site. There are several different patterns of glaucoma, as distinguished by the ophthalmoscopic appearance; however, they all share a common feature: loss of RGC axons at the ONH. There are a number of different types of RGC; in particular, the parvo and magnocellular RGCs, subserving different visual functions. Evidence had accrued suggesting that the magnocellular RGCs with their larger somas and axons were preferentially affected in glaucoma;^[17,18] however, there is now considerable evidence against this concept.^[19,20]

The gradual loss of axons at the ONH produces characteristic changes to vision. Initially there is a generalised depression of retinal sensitivity followed by localised scotomata in a nerve-fibre bundle type distribution. Contrast sensitivity and motion detection are also affected early. There is evidence that one-third of RGCs can be lost before detectable field loss occurs,^[21] indicating considerable redundancy in the system; however, as axons continue to degenerate, the field loss continues: unchecked, blindness ensues.

An understanding of the early pathological changes that precede obvious axonal loss at the ONH has been impeded by the lack of available human tissue for study; however, some progress has been made; particularly as animal models have improved. In early glaucoma, there is a compression of the lamina cribrosa, followed by a backward bowing of the lamina cribrosa superiorly and inferiorly.^[22] Microvascular changes in the ONH have been described as an early feature,^[23] but other studies have indicated that capillaries are lost in proportion to the neural tissue.^[24] Axons at the superior and inferior poles are usually preferentially lost prior to generalised thinning of the neuroretinal rim (cupping of the disc).^[18,22] This has been attributed to anatomical differences in the lamina pores, with larger pores at the poles and less intervening supportive tissue.^[18,22] Changes to elastin and extracellular matrix have been reported,^[25,26] suggesting that loss of lamina cribrosa integrity contributes to the axonopathy. Activated astrocytes and production of matrix metal-

loproteinases have also been reported,^[27,28] and loss of glial tissue is a late feature.^[22]

In addition to the loss of axons at the ONH, there is a loss of RGC somata in the retina. This is a characteristic pathological feature. In fact, retinal pathology appears to be limited to this feature.^[29-31] There is no convincing evidence that other retinal cells are affected. Moreover, at least some of the RGC somata undergo apoptotic death;^[30,32] however, the precise cause for the apoptosis remains obscure. Furthermore, there is evidence that RGCs in glaucomatous eyes are on a pathological continuum. A number of RGCs are in a transitory 'sick' phase with altered morphology, including shrinkage of their dendritic tree.^[33,34] This pathological feature has very important clinical ramifications, because it offers a target population for potential neuroprotectants. Hence, current neuroprotective strategies are aimed at healthy and/or sick RGCs. Neuroregeneration of RGCs is generally considered to be a separate concept.

3. Pathogenesis of Glaucoma

The exact nature and cause of the demise of RGCs in glaucoma is still essentially unknown, although a multitude of cellular and tissue events, pathways and mediators that could play important roles in the process have been hypothesised.^[12,19,30,35-46] Moreover, although glaucoma has traditionally been associated with ocular mechanical strain arising from an elevation of IOP, this is now known not to be exclusively the case, with vascular, autoimmune and excitotoxic components among those influences also having been hypothesised to play roles.

3.1 The Mechanical Theory

It is believed that the major destructive effect of elevated IOP is a deformation in the lamina cribrosa leading to a misalignment of inter-lamellae pores and, thus, kinking and distortion of axon bundles.^[25,26,30,44,47-49] Morphologically this presents as retrograde depression of the lamina cribrosa, disorganisation of its structure and a decreased density of connective tissue in both inferior and superior poles of the ONH.^[50] This explains why different individuals exhibit damage at a range of IOP levels: varia-

tions in mechanical or elastic properties of lamina cribrosa components determine the susceptibility of the ONH to deviations in IOP. Damage to RGCs can also occur indirectly through mechanical compression of sensitive microvessels (hypoxia) or via the toxic influence of substances released from stretched (stressed) local glia including cytokines or trophic factors such as tumour necrosis factor- α (TNF α), nitric oxide (NO), glutamate or aspartate, D-serine or potassium ions.^[40,51-54] It is also possible that the compromised ONH area becomes infiltrated with blood-borne peptides, antibodies or other potentially harmful factors from the relatively permeable choroidal circulation.^[38]

One of the major consequences of mechanical compression to RGC axons is thought to be a reduction or blockade of target-derived trophic support, which is provided under physiological conditions by intra-axonal retrograde transport of brain-derived neurotrophic factor (BDNF) or other trophic factors from the lateral geniculate body.^[30,55-57] In this situation, the affected RGC experiences a similar fate to those RGCs that die off during retinal development: lack of appropriate target-derived trophic support causes cells to die in an apoptotic manner. The appearance of RGC apoptosis after both experimental glaucoma^[57-60] and ON transection,^[58,61,62] along with the demonstrable protective effect of exogenous BDNF or other neurotrophins in paradigms of axonal injury,^[63-69] lends good support to this theory.

3.2 The Vascular Hypothesis

There is considerable evidence for blood flow incompetence at the ONH in glaucoma patients (see review by Flammer et al.^[12]), with a clear correlation between retinal damage and vascular risk factors such as systemic hypotension, low perfusion pressure, increased local resistance and vasospasm. It has been proposed that the ONH and/or retina undergo chronic hypoglycaemic, hypoxic or ischaemic injury, as a result of compromised local blood flow.^[11,12,41,43,55] Vascular disturbances have long been known to have profound effects on brain neurons (see reviews by Nagahiro et al.^[70] and Siesjö^[71,72]) and a large body of evidence now exists also linking ischaemia or hypoxia with RGC death in experimental models.^[73]

The major consequence of blood flow compromise to a tissue and to its component cells stems from a reduction of nutrients and substrates for energy production.^[70,72] When a cell is deprived of glucose or oxygen a fall in cellular adenosine triphosphate (ATP) production will result. Without ATP, there will be a failure in many vital cellular systems, such as membrane pumps and channels, biosynthetic pathways, intracellular signalling pathways and maintenance of cellular and organellar (e.g. mitochondrial) integrity.^[73]

The initial outcome of ATP synthesis failure in neuronal tissue pathology is a failure of the plasma membrane-based, ATP-driven sodium-potassium pump.^[70,72] The resultant slow elevation in membrane potential increases the likelihood of depolarisation and also release of the voltage-dependent Mg²⁺ block on the NMDA-type glutamate receptor channel.^[74,75] This leads to membrane destabilisation and opening of multiple ligand- and voltage-gated ion channels with a net elevation in intracellular levels of sodium, calcium and chloride ions, an uncontrolled osmotic influx of associated water molecules causing cell swelling and necrotic lysis, action potential generation, intracellular acidosis and extracellular accumulation of neurotransmitters, either by synaptic release or by reversal of transmitter carriers. An excessive influx of ions, particularly calcium in the cell body and sodium in the axon region, leads to a loss of control for many cellular regulatory systems and an activation of enzymes that may contribute to cell death or degradation, e.g. lipases, NO synthases, kinases, endonucleases, proteases, phosphatases and caspases.^[70,76] The net result of the energy production failure in a cell is, therefore, an irreversible commitment to death via a series of self-reinforcing events. Secondary death, often by apoptosis, is also a natural consequence of tissue energy failure.^[70,76] In neuronal tissues it is believed that secondary death processes, involving cells initially unaffected by an energy production deficit, arise because of the increased extracellular levels of neurotransmitters (e.g. glutamate) and other factors such as TNF α , NO, endothelin, D-serine or potassium ions. Elevated neurotransmitter levels will necessarily activate appropriate local receptors and in the absence of energy-dependent transmitter removal, this process will continue in an uncon-

trolled manner. In such an instance, the receptor profile of neighbouring neurons becomes extremely important; if a cell expresses a preponderance of inhibitory receptors (e.g. for transmitters such as GABA or glycine), then it may not be unduly affected by this process. However, if expression of excitatory receptors predominates (particularly those for glutamate), then the cell can effectively be 'excited to death' in the process of excitotoxicity.^[7,40]

Excitotoxic neuronal injury involves a self-reinforcing cascade of events stemming from persistent activation of ionotropic glutamate receptors.^[77] It leads to a loss in cellular ionic homeostasis, membrane depolarisation, build-up in intracellular calcium and sodium levels and, ultimately, cell death, in a manner similar to that detailed for ischaemia. Calcium-induced cellular demise is triggered more efficiently when influx occurs through certain channels rather than others: influx through the NMDA-type glutamate receptor is believed to be particularly efficient at causing cell death.^[77] Excitotoxicity during glaucoma could theoretically arise from both mechanical and vascular insults, but it is mainly considered to be of consequence during hypoxia/ischaemia.^[37,39] Hence, if hypoxia/ischaemia plays a role in the pathogenesis of glaucoma, as is widely believed, then excitotoxicity will be a major component. Nevertheless, the precise molecular mechanisms by which excitotoxicity may act as a destructive component of 'optic nerve head' rather than 'retinal' ischaemia remain unclear (see section 6.1.3).

A pathological set of events that is considered central to the vascular hypothesis, but that is also implicated in the mechanical theorem, is damage caused by oxidative stress. All cells maintain a defence against oxidative damage. This comprises a series of enzymatic (e.g. catalase, superoxide dismutase) and non-enzymatic (e.g. ascorbate, α -tocopherol) antioxidants to detoxify oxidative free radical species arising as a result of normal cellular respiration (e.g. superoxide, hydroxyl, singlet oxygen). When energy levels are depleted or stressful events numerous, as is predicted to occur in glaucoma, this system will not operate at full capacity, meaning that there will be an intracellular build-up of damaging radical species. Such species have detrimental consequences for a cell by attacking

macromolecular constituents and causing both structural and functional damage. Furthermore, in the event of circulatory restoration following an ischaemic episode, oxygen will flood the tissue, further overloading the capacity of a cell to detoxify radicals. This phenomenon, termed 'reperfusion injury', is a well known consequence of re-establishing a compromised blood supply,^[78] and has been described in the retina following restoration of the blood supply after experimental ischaemia. Oxidative free radicals are not only produced in the retina after ischaemia-reperfusion but as a consequence of other injurious situations, for example, excitotoxic glutamate receptor stimulation, and oxidative stress may well contribute to tissue damage and cellular death in both the retina and lamina cribrosa during glaucoma (see section 6.7).

One free radical that has received much attention is NO. NO is a gaseous free radical species that acts as a potent physiological vasodilator as well as a messenger molecule within the CNS. It is synthesised by nitric oxide synthase (NOS) from arginine and oxygen. Three distinct isoforms of NOS have been identified: neuronal NOS (NOS-1; nNOS) and endothelial NOS (NOS-3; eNOS) are constitutively expressed by a variety of neurons and by endothelial cells of blood vessels, respectively, while inducible NOS (NOS-2; iNOS) is not typically found under normal physiological conditions, but is induced by certain, usually toxic, stimuli. NO has the advantage of not requiring synapses to pass from one neuron to another, and it is this ease of passage that underlies its detrimental influence: a rapid intracellular and intercellular diffusion combined with the ready combination with superoxide radicals to produce peroxynitrite lends this compound a potentially lethal character. Increased levels of NO production in the retina have been described as a direct consequence of pathological situations such as ischaemia-reperfusion,^[79,80] inflammation^[81] and excitotoxicity.^[82] For example, stimulation of NMDA receptors activates NOS-1 and causes transcriptional upregulation of NOS-2, leading to large increases in the cellular NO level.^[83] Importantly, NOS-1 and NOS-3 have been shown to be upregulated, and NOS-2 to be expressed, in astrocytes and microglia in the ONH of primary open-angle glaucoma (POAG) patients and experimental glaucomatous

rats, and these findings in particular have implicated NO in the pathogenesis of glaucoma (see section 6.4).^[84-89]

3.3 Autoimmunity and Heat Shock Proteins

Recent studies have suggested that the immune system may play one of two roles in glaucoma pathogenesis.^[38] The first is to actually cause RGC damage by stimulation of an autoimmune response subsequent to expression of autoantibodies or by a mimicked response to a sensitising antigen (e.g. the immunodominant heat shock proteins [HSPs]). The second is to provide a means of surveillance for RGC damage stimulated by other stresses (e.g. hypoxia, elevated pressure, raised glutamate levels, excessive TNF α or NO release). In the latter scenario, stressed RGCs produce antigens that will stimulate an immune response at the site of damage, thus protecting other RGCs from a similar destruction.^[38]

Evidence for a link between HSP reactivity and glaucoma pathogenesis has arisen from several studies that show that HSPs have an increased expression in the eyes of glaucoma patients,^[90,91] in animals subjected to chronic hypertension,^[92,93] and in stressed neurons and astrocytes in culture.^[94] Some glaucoma patients additionally possess an elevated titre of serum antibodies to HSP27.^[90] Furthermore, addition of HSP27 antibody itself can induce widespread apoptotic death of cultured retinal cells including RGCs.^[94] Other candidate antibodies for involvement that are elevated in the serum of glaucoma patients include those to glycosaminoglycan and glutathione *S*-transferase (see Tezel et al.^[38] for a review).

Such data are convincing, but it is not clear at present whether HSP expression exists as cause or effect in RGC axon injury. Caprioli,^[95] for example, has suggested that these proteins may just be expressed as part of an endogenous retinal protective response to stress (see section 6.8). Careful temporal studies may detail whether the increased HSP or HSP-antibody expression precedes all other signs of injury, thus indicating that these proteins may play a central role in stimulating an autoimmune attack on RGCs.

4. Laboratory Models of Glaucoma

The use of animal models to investigate glaucoma has been the subject of some excellent recent reviews.^[44,96-99] As a consequence, the topic is covered relatively briefly in this review. A number of different *in vitro* and *in vivo* models of RGC death have been used to study neuroprotection for glaucoma. Given that the aetiology and pathogenesis of glaucoma remain ill-defined, it seems reasonable to suggest that any potential neuroprotectant should be tested in multiple experimental models first. It follows that the more paradigms in which the drug shows activity, the more likely it is that the compound will be useful in the clinical setting. Nevertheless, the goal of a model is to resemble the disease as closely as possible, thus making extrapolation from bench to bedside more credible.

Tissue cultures and single cell type cultures provide a direct and rapid means of comparing different compounds. They are ideal for determining the potency and efficacy of a drug, and allow direct cellular accessibility and microenvironmental control, as well as efficient comparison between many experimental conditions or potential therapeutic compounds; however, they generally have less relevance to clinical glaucoma than *in vivo* models because of the unique complexity of the retina, the importance of the intact ON for RGC survival, the need to measure physiological parameters, the interaction between the anterior and posterior chambers of the eye, and the differing intra- and extra-cellular milieu of particular ocular tissues and cell layers, which cannot be accurately replicated *in vitro*.

A variety of insults can cause RGC loss *in vivo*, including models of retinal ischaemia-reperfusion, ON transection/crush and excitotoxic injury. There is evidence that ischaemia, mechanical injury and excitotoxicity all play a role in glaucoma pathogenesis; hence, successful neuroprotectants using these models can to some extent be extrapolated to glaucoma; however, the models do not closely mimic the clinical situation. Conversely, primate laser models of glaucoma probably exhibit the closest comparison to human glaucoma,^[100] but the high monetary and welfare costs associated often limit their use to what is essentially an immediately preclinical setting.

In recent years, a number of rodent models of ocular hypertension and glaucoma have been developed (see Morrison et al.^[44]). These rat models produce elevated IOP for several weeks, which results in glaucomatous-like changes at the ONH and a loss of RGCs. The models differ primarily in the means by which the IOP is raised. Elevated IOP can be achieved by cauterising two or three of the episcleral vessels,^[101,102] by scarring Schlemm's canal with hypertonic saline,^[60] or by lasering the trabecular meshwork.^[103,104] They are thought to represent a significant improvement upon ischaemia-reperfusion and mechanical injury models, and studies performed using these paradigms form the core of this review; however, more work is needed to determine their true similarity to glaucoma and whether any one model is superior to the others. In addition to rat models, mutant mice, such as the DBA/2J variant, have been developed, which spontaneously develop a form of secondary glaucoma.^[105,106] Mice offer the distinct advantage that their genetic background can be altered.

5. Criteria for Evaluating Neuroprotection

In order to evaluate whether any specific pharmacological agent has potential for use in the treatment of glaucoma, Wheeler et al.^[107] have set out four criteria that should be met by the drug in question. A prospective neuroprotective agent should (i) have (a) specific target(s) in the retina/optic nerve; (ii) increase RGC survival in laboratory models of glaucoma; (iii) reach the target at a neuroprotective concentration after clinical dosing; and (iv) display neuroprotective activity in human trials.

In the following sections, potential targets for neuroprotectants in glaucoma are identified. For each target, the evidence in favour of it being a valid point for intervention is discussed. Focus is then on pharmacological agents that have been tested in animal models of glaucoma or in the clinic, using the criteria listed above as a general framework for discussion (table I).

Table I. Selected studies detailing drugs tested in animal models of chronic ocular hypertension which promote survival of retinal ganglion cells (RGCs) via mechanisms that are independent of intraocular pressure

Drug	Potential Strategy	Species	Model	Reference
MK-801	NMDA receptor antagonism	Rat	Cauterisation	108
		Rat	Hypertonic saline	109
Memantine	NMDA receptor antagonism	Rat	Lasered	110
		Mouse	DBA/2J	111
		Primate	Lasered	112-114
Glatiramer acetate	Boosting autoimmunity	Rat	Lasered	115-117
BDNF	Neurotrophin deprivation	Rat	Lasered	67
			Cauterisation	63,118
CNTF	Neurotrophin deprivation	Rat	Lasered	119
Aminoguanidine	Nitric oxide inhibition	Rat	Cauterisation	120
		Rat	Hypertonic saline ^a	86
Phenytoin	Sodium channel blockade	Rat	Cauterisation	121
<i>Ginkgo biloba</i>	Reducing oxidative stress	Rat	Cauterisation	122
S-PBN	Reducing oxidative stress	Rat	Cauterisation ^b	63
Geranylgeranyl-acetone	Induce heat shock protein 70	Rat	Lasered	123
FK506	Calcineurin inhibition	Rat	Hypertonic saline	124
BIRC4	Caspase-3 inhibition	Rat	Hypertonic saline	125,126
Brimonidine	Activation of α_2 -adrenoceptors	Rat	Lasered cauterisation	104,127

a No protection of RGCs was observed using this model.

b Enhanced protection afforded by BDNF, failed to improve RGC survival when administered alone.

BDNF = brain-derived neurotrophic factor; **BIRC4** = baculoviral; **CNTF** = ciliary neurotrophic factor; **IAP** = repeat-containing protein 4; **S-PBN** = N-*tert*-butyl-(2-sulphophenyl)-nitron.

6. Targets for Neuroprotection

6.1 Glutamate Receptors

The rationale behind the excitotoxic hypothesis of glaucoma is simple: during glaucoma, there is an increase in the level of the excitatory neurotransmitter glutamate in the retina that is toxic to RGCs. While there is substantive evidence that high doses of, or prolonged exposure to, glutamate kills RGCs via overactivation of ionotropic glutamate receptors,^[128-131] no consensus has been reached in answering the question: is the level of glutamate elevated during glaucoma?

6.1.1 Is There Evidence for Increased Glutamate in Glaucoma?

There has been much speculation as to the pathological processes that could result in an elevated glutamate level, but conclusive data supporting an elevated level have proved tantalisingly elusive. Essentially, at the cellular level, two mechanisms are most likely to account for a rise in glutamate: (i) an increased release of glutamate from dying RGCs or metabolically compromised glial cells; and (ii) a reduced clearance of glutamate as a result of inefficient uptake.

Initial studies suggested that the level of glutamate was elevated in glaucomatous eyes. Dreyer et al.^[132] reported that in patients with glaucoma and in monkeys with experimentally elevated IOP the concentration of glutamate in the vitreous body was at least twice as high as in control eyes. This finding was replicated in studies that analysed the vitreous humours of dogs with breed-related primary glaucoma^[133] and quail with a glaucoma-like disorder.^[134] However, recent examinations of the glutamate content of vitreous bodies from human patients^[135] and from monkeys^[112,136,137] and rats^[138] with experimental glaucoma have failed to repeat these findings; in all of these studies the level in glaucomatous eyes was normal. Despite these negative findings, the excitotoxicity theory has not been markedly weakened. A number of authors have pointed out that the vitreal glutamate level may not correspond to the concentration present at the synapses of the RGCs. It is also possible that the glutamate concentration is only sporadically elevated, for example during ONH ischaemic episodes.

Lately, researchers have turned their attentions to investigating whether retinal glutamate clearance mechanisms are compromised in experimental glaucoma. Using the lasered model of chronic hypertension, Martin et al.^[139] found a significant reduction in the level of the glutamate transporters GLAST (EAAT1) and GLT-1 (EAAT2) in the retina when analysed by Western blot. They speculated that if the observed reductions were associated with a concurrent decrease in glutamate uptake capability, there would be an increased likelihood of glutamate-mediated injury. Conversely, WoldeMussie et al.,^[140] using the same model of chronic hypertension as Martin et al.,^[139] showed a sustained increase in the level of GLAST expression in the retina, when analysed by immunohistochemistry and Western blotting. In this case, the authors suggested that the increase in GLAST was most likely to be a compensatory mechanism in response to an increase in the extracellular glutamate concentration. The underlying reason for the contradictory results of the two studies is unclear and neither result is supported by recent work from the laboratory of Pow,^[141] who showed no change in the level of GLAST in glaucomatous rats. Interestingly, however, they showed a vastly altered expression and distribution of a splice variant of GLT-1 in glaucomatous rats and humans, which they suggest may be indicative of an anomaly in glutamate homeostasis. Nevertheless, none of these studies determined whether alterations in glutamate transporter expression in experimental glaucoma correlate with a change in functional capacity.

This issue was recently addressed in a study performed by Hartwick et al.^[142] They reasoned that if experimental glaucoma causes a decrease in the functional capacity of retinal glutamate clearance mechanisms, then exogenously applied glutamate would stimulate RGC glutamate receptors at lower concentrations. They tested the hypothesis by assessing the effect of glutamate on RGC calcium dynamics in living retinal wholemounts prepared from rats with experimental glaucoma and from controls. To validate the technique, they initially confirmed the presence of glutamate clearance mechanisms in the intact retinal preparation. They achieved this by demonstrating that in retinal wholemounts prepared from normal eyes, very high

concentrations of glutamate were necessary to affect RGC calcium dynamics, but pharmacological inhibition of glutamate transporters resulted in an increased sensitivity to glutamate. Subsequently, they investigated the responses in rats with experimental glaucoma (the hypertonic saline model). The results showed no difference between glaucomatous or control retinas in their sensitivity to exogenously applied glutamate. The authors concluded that there was no evidence for a global defect in retinal glutamate clearance mechanisms in the hypertonic saline rat model of glaucoma.

The inevitable weakness in the study of Hartwick et al.^[142] is that the functional capacity of the retinal glutamate transporters *in vitro* may not be identical to their capacity *in vivo*. If, for example, the energy requirements of the Müller cells are compromised during experimental glaucoma then the transporters may not be operating efficiently. This would not be evident in the retinal wholemount when the tissue is superfused with balanced salt solution and oxygen.

6.1.2 NMDA Receptor Antagonists

RGCs express multiple subtypes of ionotropic and metabotropic glutamate receptors,^[143-145] yet excitotoxic loss of RGCs is thought to result primarily from glutamate interacting with the NMDA receptor subtype. Administration of NMDA receptor antagonists is an effective method of preventing RGC loss in various models of RGC death in which excitotoxicity is implicated, such as injection of glutamate or NMDA,^[131] in models of retinal and ON ischaemia,^[146-148] and after ON crush.^[110,149,150] Therefore, if NMDA receptor antagonists similarly reduce RGC loss in models of chronic hypertension, then a role for excitotoxicity in glaucoma could be considered. The first study to address this issue was by Chaudhary et al.,^[108] who elevated IOP in rats by cauterising the episcleral veins, and administered the prototypical, non-competitive NMDA receptor antagonist MK-801. They found a marked preservation of RGC numbers in animals treated with the drug. A similar effect was documented by Guo et al.,^[109] who used the hypertonic saline model of RGC injury. However, MK-801 is unsuitable for use clinically because it blocks normal glutamatergic neurotransmission and would, therefore, give rise to potentially severe adverse effects. Accordingly, oth-

er studies have made use of the uncompetitive NMDA receptor antagonist memantine.

Memantine

Memantine (1-amino-3,5-dimethyl-adamantane) is an NMDA receptor antagonist that has recently been shown to be clinically useful in the treatment of moderate to severe Alzheimer's disease.^[151]

Activity: Memantine is a highly effective neuroprotective agent in acute animal models of RGC death.^[110,131,146-150] In the lasered rat model of chronic hypertension, daily infusion of memantine (5–10 mg/kg) via a subcutaneous osmotic pump increased the number of surviving RGCs and preserved the compound action potential of the ON.^[110] In the DBA/2J mouse model of glaucoma, twice daily intraperitoneal injections of memantine (5 mg/kg) protected against loss of RGCs to a degree similar to that achieved by the IOP-lowering drug timolol.^[111] Most significantly, in monkeys with experimental glaucoma, daily oral administration with memantine (4 mg/kg) afforded protection against structural changes and functional loss that occurred in vehicle-treated animals.^[112-114]

Mode of action: Memantine is classified as an uncompetitive NMDA receptor antagonist. An uncompetitive antagonist is defined as an inhibitor the action of which is contingent upon prior activation of the receptor by the agonist: the degree of blockade for a given concentration of antagonist increases as the concentration of the agonist increases. Thus, memantine is inactive during normal glutamatergic synaptic activity, but is an increasingly effective blocker under prolonged/excessive activation of the receptors, which occurs in pathological situations.^[152]

Penetration: The oral administration regimen used in the primate study resulted in a vitreous memantine concentration of approximately 1 µmol/L at the time the animals were killed.^[112] Memantine is known to block NMDA-mediated death of cultured RGCs at this concentration.^[152]

Clinical trials: A large-scale, randomised, prospective, phase III clinical trial is currently in progress to determine the effect of memantine on visual field loss in glaucoma patients. The results are not expected until 2007.

6.1.3 How are NMDA Receptor Antagonists Neuroprotective in Experimental Glaucoma?

The ability of NMDA receptor antagonists to attenuate injury is easy to explain when the injury involves ischaemia/trauma to a neuronal cell body that expresses NMDA receptors; however, the mechanism by which NMDA receptor blockade could attenuate experimental glaucoma remains unclear. If glaucomatous pathology occurs principally at the level of the unmyelinated axons in the ONH, which are unresponsive to NMDA,^[153] how could NMDA receptor blockade protect RGCs? Several explanations for the success of NMDA receptor antagonists in this situation exist.

- NMDA receptor antagonists prevent secondary degeneration of RGCs. In this context, secondary degeneration is hypothesised to occur when uninjured RGCs are exposed to the released toxic cellular contents (i.e. glutamate) of neighbouring RGCs that have died as a result of the primary insult.
- NMDA receptor blockade reduces glutamatergic injury to ONH astrocytes and this prevents subsequent injury to axons.
- The models inadvertently produce a primary injury to the retina and it is this injury that is being attenuated by the NMDA blocker.
- NMDA receptors *are* present on unmyelinated ON axons.
- Blockade of NMDA receptors located on the extra-ocular, myelinated portion of the RGC axons^[154] aids survival of these cells.

6.1.4 Summary

The evidence in support of an involvement of excitotoxicity in glaucoma is, at present, somewhat inconsistent. The findings that glutamate levels are not elevated in the vitreous of glaucomatous eyes and that retinal glutamate transporters appear to function normally in eyes with experimental glaucoma argue against an involvement of glutamate in the pathogenesis, yet the neuroprotective properties of memantine in models of glaucoma are impressive. Neuroprotection-based human studies can provide invaluable insights into the pathogenesis of glaucoma. If memantine proves to be clinically useful, then it will not only have provided the first pressure-independent, neuroprotective treatment,

but will have also proved that excitotoxicity plays a role in the pathogenesis of the disease.

6.2 Boosting Protective Autoimmunity

T-cell-mediated autoimmunity is traditionally considered to be an attack on host components by activated T cells leading to pathological changes and ultimately self-destructive autoimmune disease; however, an impressive body of work generated by the group of Schwartz has shown that boosting T cells weakly recognising self antigens following injury to CNS tissue can, under the right circumstances, protect against further degeneration and play a role in recovery.^[155] They propose that boosting autoimmunity by vaccination is a realistic treatment option for glaucoma.^[156,157]

The first convincing evidence that stimulating the immune system aids the survival of injured RGCs was the finding that systemic injection of encephalitogenic T cells directed to myelin basic protein in rats that had undergone partial ON crush resulted in an accumulation of these T cells at the site of injury, and histological and functional preservation of RGCs.^[158,159] The protection afforded by this passive immunisation was, however, accompanied by transient symptoms of autoimmune disease. In a follow-up study, it was ascertained that active immunisation, achieved by injection of various myelin-associated proteins, also increased survival of RGCs after ON injury, and perhaps more importantly that cryptic (non-encephalitogenic) myelin-associated peptides were equally protective of RGCs, but induced no signs of autoimmune disease.^[160] In further studies, Schwartz and colleagues found evidence consistent with the idea that the neuroprotective effects they had achieved by experimental manipulation of the number of autoimmune T cells were merely the augmentation of an endogenous response that occurs in stressful situations, i.e. that protective autoimmunity is a physiological phenomenon that can be boosted in a manner that is analogous to preconditioning. The experimental results that allowed this conclusion to be reached were as follows:

- in rats devoid of mature T cells, increased numbers of RGCs died after ON crush compared with normal rats;^[161,162]

- giving a prior unrelated CNS injury to a group of ON crush rats resulted in the loss of fewer RGCs than in animals that underwent just the ON injury;^[161,162]
- transfer of splenocytes from rats with the unrelated CNS injury to normal rats protected against an ON crush injury.^[161]

Initiation of an endogenous, autoimmune T cell-mediated self-protective mechanism is not exclusive to axonal injuries, but also occurs after excitotoxicity^[115] and chronic elevation of IOP.^[163] Interestingly, immunisation targeted at myelin-associated proteins failed to prevent RGC death after these injuries,^[115,116] from which the authors concluded that the myelin antigen is not specific to a RGC body injury. Immunisation with self-peptides derived from retinal neurons did result in beneficial effects.^[164] In glaucoma, the site of injury includes the RGC body and axon, which immunologically can be considered to be distinct tissues. One way of circumventing the need for tissue specificity and protecting against both axonal and cell body injuries is to use a synthetic antigen such as glatiramer acetate, which cross-reacts with multiple self-antigens.

6.2.1 Glatiramer Acetate

Glatiramer acetate (Copolymer-1; Cop-1; Copaxone) is a synthetic, random oligopeptide composed of tyrosine, glutamate, lysine and alanine residues. It is a low-affinity antigen that cross-reacts with a wide range of self-reactive T cells. This property allows glatiramer acetate to mimic self-antigens at various sites of injury by evoking neuroprotective autoimmune responses.

Activity: Glatiramer acetate emulsified with Freund's adjuvant increased survival of RGCs after glutamate-induced excitotoxicity in mice,^[115] after an acute increase in IOP^[165] and following ON crush in mice^[115] and rats,^[166] but not, surprisingly, after partial or complete transection of the ON in rats.^[167] The reason for the lack of effect of glatiramer acetate in the latter models is unclear. In the lasered rat model of chronic hypertension, Schwartz and colleagues demonstrated that immunisation with glatiramer acetate plus adjuvant resulted in a highly significant preservation of RGCs, irrespective of whether the drug was administered at the time of laser cauterisation or 10 days later.^[115,116] In a fol-

low-up study,^[117] the same group extended their findings. They showed, firstly, that glatiramer acetate was an effective neuroprotectant, even when administered without adjuvant, and secondly, that glatiramer acetate imparted functional as well as histological protection to RGCs.

Mode of action: The available evidence indicates that glatiramer acetate induces neuroprotection of RGCs via recruitment of systemic T cells to the site of injury:

- administration of glatiramer acetate caused a transient accumulation of T cells and microglia/macrophages in the retina;^[117]
- transfer of glatiramer acetate-reactive T cells elicited the same protective response as glatiramer acetate itself;^[115,166]
- glatiramer acetate was ineffective in animals devoid of mature T cells.^[117]

The precise mechanisms by which T cells confer resistance to injury have not yet been elucidated, although the release of trophic factors and cytokines may well be involved.^[166]

Penetration: Unlike other neuroprotective compounds, glatiramer acetate is an immune-based therapy and penetration of significant amounts of the drug to the site of injury is unnecessary.

Clinical trials: Glatiramer acetate is a US FDA-approved drug for multiple sclerosis, which elicited positive effects when administered daily in clinical trials.^[168] The compound is expected to enter phase II trials for use in glaucoma in the near future.

6.2.2 Summary

Boosting autoimmunity is an exciting and novel approach to treat glaucoma, and results obtained by Schwartz and colleagues using glatiramer acetate in a rat model of chronic hypertension are encouraging. It is important that other research groups investigate whether boosting autoimmunity is neuroprotective in rodent and primate models of glaucoma.

6.3 Neurotrophin Deprivation

The neurotrophin deprivation hypothesis, as expounded by Quigley,^[30] is essentially an adaptation of the traditional mechanical theory: raised IOP and/or defects in the extracellular matrix cause(s) compression and displacement of the lamina cribrosa. The resultant damage to the ON axons leads to

blockage of retrograde delivery of neurotrophic survival factors to the cell bodies and the initiation of RGC apoptosis. This section considers the evidence for obstructed axoplasmic transport and compromised neurotrophic support of RGCs in glaucoma.

6.3.1 Obstructed Axonal Transport

The first data in support of the theory came from Anderson and Hendrickson in a 1974 study in which they showed that acute elevation of IOP in monkeys obstructed rapid orthograde axoplasmic transport.^[169] Subsequent experimental studies in monkeys and rats have confirmed and extended these findings:

- retrograde as well as orthograde axonal transport is blocked by acutely elevated IOP;^[170-172] the extent of axonal transport obstruction appears to be proportional to the magnitude and duration of the increase in IOP;^[170-175]
- axonal transport obstruction is evident in eyes with mild, chronic elevations in IOP;^[60,176]
- blockade of axonal transport can be reversed in some axons if the IOP is lowered and the duration of hypertension is relatively short-lived;^[170,176]
- there is an accumulation of dynein, a protein fundamental to retrograde axonal transport, in the ONH of rodents with chronic IOP elevation.^[177]

Histological analysis of human glaucomatous ONHs has revealed an accumulation of mitochondria and vesicles within axons where they cross the lamina cribrosa.^[49,178] These findings, although consistent with blockade of axonal transport at the ONH, are not conclusive. An alternative explanation has recently been postulated, which suggests that this increased mitochondrial density exists to serve the higher energy requirements necessary for electrical conduction in unmyelinated axons in the prelamellar and lamellar ON.^[179]

6.3.2 Neurotrophin Deprivation

The neurotrophin family consists of four members: BDNF, nerve growth factor, neurotrophin-3 and neurotrophin-4/5. To date, only a few studies have examined whether experimental glaucoma affects delivery of neurotrophins to, or expression of neurotrophins in, the retina. In rats with acutely elevated IOP, Quigley and co-workers found retrograde axonal transport of radioactive BDNF to the retina to be substantially inhibited.^[56,57] The results

indicated that during glaucoma RGCs may become depleted of BDNF. In two related studies, the chronology of neurotrophin changes after elevation of IOP were studied in models of chronic hypertension. Using the hypertonic saline model, Johnson et al.^[59] found results consistent with a decrease of neurotrophin delivery; they showed a gradual depletion of BDNF and neurotrophin-4/5 in the proximal ON and in the superior region of the inner retina. In contrast, Rudzinski et al.,^[180] using the episcleral vein cauterisation model, found a slow increase in BDNF protein expression in the retina over 4 weeks, no change in the level of neurotrophin-3, and a transient upregulation of nerve growth factor at 1 week, which had returned to normal by 4 weeks; these results do not imply a diminished delivery of neurotrophins. In these latter studies, differentiation between neurotrophins retrogradely transported from the superior colliculus and neurotrophins synthesised locally was not possible. New studies are needed to definitively address this issue.

6.3.3 Brain-Derived Neurotrophic Factor

BDNF enhanced survival of RGCs after ON transection,^[64,65] ON crush^[68] and excitotoxic injury;^[181] however, the effects after ON injuries were only transient. BDNF also promoted survival of RGCs in the episcleral vein cauterised-^[63,118] and lasered-^[67] rat models of chronic hypertension. Multiple intravitreal injections were needed to show positive effects in the former model, while in the latter model sustained delivery of BDNF was achieved by gene therapy. The effectiveness of BDNF in the episcleral cauterisation model was increased by co-administration of a free radical scavenger.^[63] This was also the case after ON transection,^[182] and it appears that the neuroprotective potential of BDNF, particularly at higher doses, is limited by adverse effects as a consequence of excessive free radical formation caused by BDNF itself.^[63,182]

6.3.4 Other Neurotrophic Factors

In models of ON crush or transection, RGC death can be delayed by a variety of other neurotrophins and trophic factors, including nerve growth factor, neurotrophin-4/5, glial cell-derived neurotrophic factor, ciliary neurotrophic factor (CNTF) and fibroblast growth factor-2.^[183,184] Of these compounds, only CNTF has thus far been tested in a

model of chronic hypertension. In the lasered rat model, a single intravitreal injection of CNTF enhanced survival of RGCs for up to 4 weeks and resulted in an increase in phosphorylated STAT3.^[119] It is not known whether activation of the JAK-STAT signalling pathway accounted for the observed neuroprotection.

6.3.5 Summary

Although there is no direct evidence from clinical studies that reduced delivery of neurotrophins is a contributory factor in the apoptotic death of RGCs in glaucoma, work performed in animal models does support the hypothesis. The majority of investigators have thus far concentrated their efforts on BDNF and the results are promising, but a number of questions remain unanswered. Are BDNF and its receptor TrkB downregulated in glaucomatous retinas? Does BDNF only afford transient neuroprotective effects in models of chronic glaucoma as is the case after ON transection? Will delivery of BDNF exacerbate any free radical-mediated injury in glaucoma? Does sustained delivery of BDNF lead to downregulation of TrkB and loss of trophic responsiveness? Will it be necessary to deliver BDNF and TrkB to achieve long-lasting neuroprotection? Is BDNF the most suitable candidate of the nerve growth factors/neurotrophins/neurotrophic factors to be used for neuroprotection? What is the most appropriate route of delivery for nerve growth factors?

6.4 Nitric Oxide Synthesis

It has been suggested that excessive NO, released by reactive astrocytes and microglia in the ONH, causes damage to ON axons at the level of the lamina cribrosa.^[83] Furthermore, it has been hypothesised that pharmacological inhibition of NOS-2 represents a novel strategy for the treatment of glaucoma.^[87] The available data is reviewed in this section.

The first evidence supporting the involvement of NO in the pathogenesis of POAG came from studies that localised the NOS isoforms in the ONH of POAG patients and healthy individuals. Neufeld et al.^[84] showed that NOS-1 and NOS-3 were upregulated in POAG, and NOS-2, which was absent in the normal ONH, was expressed in the ONH of POAG

patients. In follow-up studies, it was ascertained that reactive astrocytes and some microglia were the cells responsible for expressing NOS-2 in the ONH.^[52,89] These findings were strengthened by the results of a genetic analysis of glaucoma patients and controls, which showed an association between NOS-2 and POAG.^[185] A rationale for the expression of NOS-2 in the ONH of POAG patients was provided by the finding that when human ONH astrocytes were placed in culture and subjected to elevated hydrostatic pressure they synthesised large amounts of NOS-2.^[88] Experiments performed using the episcleral vein cauterisation model of chronic hypertension supported the association between NOS-2 expression and ONH pathophysiology. Shareef et al.^[85] showed that in rats with elevated IOP there was an expression of NOS-2 in ONH astrocytes, while Neufeld et al.^[120] demonstrated that an inhibitor of NOS-2, aminoguanidine, increased survival of RGCs compared with controls (see section 6.4.1).

The involvement of NO in ONH injury and RGC death was, however, recently questioned in a detailed paper by Pang et al.^[86] They found no evidence for the expression of NOS-2 by astrocytes in the ONH of POAG patients or in the ONH of rats with chronic ocular hypertension. Moreover, they documented no protection of RGCs by aminoguanidine.

6.4.1 Aminoguanidine

Activity: Aminoguanidine, when administered orally to rats (2 g/L), afforded protection to RGCs in the episcleral vein cauterised model of chronic hypertension.^[120] However, when administered in an identical fashion to rats injected with hypertonic saline to elevate IOP, it yielded no protection.^[86]

Mode of action: Aminoguanidine is a relatively potent inhibitor of NOS-2 (the concentration that produced 50% inhibition [IC₅₀] was 31 µmol/L), but it is also an inhibitor of NOS-1, with an affinity less than one order of magnitude lower. The drug is not specific for NOS; it is also a prototype α,β-dicarbonyl scavenging agent that inhibits formation of advanced glycosylation end products (IC₅₀ ≈ 200 µmol/L), and has antioxidant activity – at 100 µmol/L it decreased lipid peroxidation and the formation

of reactive oxygen species in retinal Müller cells (for a review, see Thornalley^[186]).

Penetration: The concentration of aminoguanidine in the retina following oral administration of 1 g/L to rats has been measured at approximately 50 $\mu\text{mol/L}$; ^[187] the concentration in the studies highlighted above is likely to be closer to 100 $\mu\text{mol/L}$, given that twice the dose was used. As such, it is doubtful that aminoguanidine functioned solely as an inhibitor of NOS-2 in the models of chronic hypertension.

6.4.2 Summary

The concept that reactive astrocytes/microglia release NO, which is toxic to ONs is an attractive one for two reasons: firstly, it agrees with the widely accepted viewpoint that the primary insult in POAG occurs at the level of the ONH; secondly, it offers a relatively straightforward means of treatment. However, to date, the evidence is ambiguous. It is not yet clear whether NOS-2 is expressed by cells within the ONH of glaucoma patients, and there is no consensus between the results that have been obtained from two different rodent models of ocular hypertension. As a further complication, truly isoform-specific NOS inhibitors are not yet available, which is problematic when all three isoforms of NOS are found in various cell types in the retina and at least two of the isoforms are present in the ONH region. Since there is no agreement as to which of the rodent models for glaucoma most closely simulates POAG, the most pressing need is for the role of NOS-2 to be explored in other laboratory paradigms of glaucoma, such as the lasered model of chronic hypertension.

6.5 Voltage-Gated Sodium Channels

The rationale for using Na⁺ channel antagonists as therapeutic agents in glaucoma stems from the knowledge that preventing Na⁺ influx through voltage-gated Na⁺ channels is an effective means of inhibiting axonal degeneration and preserving function in the CNS during periods of physiological stress, irrespective of whether the stress is caused by limited oxygen and nutrient supply, mechanical/compression injury or NO toxicity.^[153,188] Although there is little direct evidence that excessive influx of Na⁺ into ON axons contributes to the death of RGCs

in glaucoma, the hypothesis is logical for the following reasons:

- each of the physiological stresses referred to above is implicated in the pathogenesis of POAG;
- the ONH is particularly vulnerable to insults involving Na⁺ influx;
- experimental studies show that Na⁺ channel blockers can protect the ON from various insults.

The evidence for the involvement of ischaemia, mechanical compression and NO in the pathogenesis of POAG is discussed earlier (sections 3.1, 3.2 and 6.4), but the latter two statements are worth expanding upon.

The ONH is particularly vulnerable to injuries involving Na⁺ influx because of the very high density of Na⁺ channels, and in particular the Na_v 1.6 subtype, which is present in the unmyelinated prelaminar and laminar ON.^[179] Na_v 1.6, which produces a persistent, use-dependent, non-inactivating sodium current, is probably important for propagation and maintenance of action potentials;^[189] however, in pathological situations, these same properties are thought to trigger an uncontrolled influx of Na⁺ into the axoplasm, causing reversal of the Na⁺-Ca²⁺ exchanger, reversal of glutamate transport and, ultimately, irreversible Ca²⁺-mediated axonal injury.^[153]

In the isolated ON, Na⁺ channel antagonists have been demonstrated to preserve axonal function against injury caused by NO,^[188] anoxia,^[190,191] and the more damaging combination of oxygen and glucose deprivation.^[192] Blockade of voltage-gated Na⁺ channels has also been shown, *in vivo*, to protect against degeneration of the ON in models of inflammation,^[193] ON compression and in a chronic hypertensive glaucoma model (see section 6.5.1).

6.5.1 Phenytoin

Phenytoin (5,5-diphenylhydantoin) is an effective and commonly prescribed anticonvulsant used for the treatment of epilepsy.

Activity: Systemic administration of phenytoin reduced RGC death following partial ON crush^[149] and in the episcleral vein cauterised model of chronic hypertension.^[121] Phenytoin, at concentrations lower than those used clinically for the treatment of

epilepsy, has also been shown to protect the isolated rat ON from anoxia-induced injury.^[194]

Mode of action: Phenytoin binds to voltage-gated Na⁺ channels in the neuronal cell membrane and inhibits Na⁺ influx. It is especially effective in reducing neuronal excitability in neurons undergoing repetitive firing. In addition to its well documented effects on Na⁺ ion conductance, phenytoin has also been shown to interact with Ca²⁺ channels, ionotropic glutamate receptors, sigma binding sites and glatiramer acetateBAA receptors.^[195]

Penetration: Systemic administration of phenytoin to rats has been shown to affect the a- and b-waves of the electroretinogram,^[196] indicating that the drug crosses the blood-retinal barrier, but the exact concentrations reached after short- or long-term administration are not known.

Clinical trials: In a small-scale, short-term study of glaucoma patients, oral phenytoin produced a beneficial effect on visual fields.^[197]

6.5.2 Summary

The involvement of voltage-gated Na⁺ channels in the pathogenesis of acute ON injury, such as that caused by ischaemia, mechanical trauma and inflammation, is unequivocal; their involvement in the chronic optic neuropathy of POAG is at present uncertain. One of the difficulties of using Na⁺ channel antagonists as neuroprotectants in POAG is finding a dose of drug that will reduce persistent, damaging Na⁺ currents but have no effect on normal ion conductance. Phenytoin is considered to be one such drug, yet systemic administration of clinical doses to rats does lead to electrophysiological changes in the visual system,^[196] which may be related to its effects on other transmitter systems. In contrast, flunarizine and the β -adrenoceptor antagonist betaxolol (see section 7.2 for more detail), which both inhibit Na⁺ influx, have no adverse effects on the visual system, although whether either drug reaches the ONH at a sufficient concentration to reduce Na⁺ influx is unknown.

6.6 Voltage-Gated Calcium Channels

Calcium overload plays a central role in ischaemia- and excitotoxicity-induced death of white and grey matter, but the mechanisms by which Ca²⁺ is elevated in white and grey matter differ markedly.

This is exemplified by the retina/ON. In the retina, drugs that inhibit Ca²⁺ entry into cells through voltage-gated Ca²⁺ channels (termed Ca²⁺ channel antagonists) attenuate RGC death caused by ischaemia and excitotoxicity.^[73] Conversely, in the ON, Ca²⁺ channel antagonists are relatively ineffective against anoxic or ischaemic injuries; reversal of the Na⁺/Ca²⁺ exchanger rather than activation of voltage-gated Ca²⁺ channels is thought to be the route for Ca²⁺ entry into the axoplasm.^[153] Therefore, it has been hypothesised that Ca²⁺ channel antagonists would be effective as neuroprotectants in glaucoma if ionic fluxes in RGC bodies/dendrites are adversely affected, for example by hypoxic/ischaemic episodes or by elevated glutamate, but not if RGCs die only as a consequence of axonal injury.^[198]

Activity: To date, the role of retinal Ca²⁺ influx and the potential effectiveness of Ca²⁺ channel antagonists in chronic hypertensive models of RGC death have not been explored, although the results of these studies would not be straightforward to interpret since Ca²⁺ channel antagonists reduce IOP^[199-202] and increase ONH blood flow^[203,204] in laboratory animals.

Clinical trials: A number of studies have examined the effects of Ca²⁺ channel antagonists on ONH perfusion and visual parameters. The combined results suggest that some members of this class of drugs may increase ONH blood flow and improve vision in normotensive glaucoma patients.^[13,205] Because of their effects on blood flow, the mechanism of action of Ca²⁺ channel antagonists in normotensive glaucoma is thought to be primarily due to vasoprotection rather than neuroprotection. However, all of the studies performed thus far are short-term, and there is a real need for long-term clinical outcome data from randomised, placebo-controlled studies.

6.7 Oxidative Stress

It is hypothesised that oxidative stress plays a role in RGC death in POAG by causing damage at three discrete sites in the eye: the trabecular meshwork, the ONH and the retina. The involvement of oxidative stress in trabecular meshwork pathophysiology and obstruction of aqueous outflow is beyond the scope of this review and readers are directed to recent articles by Veach^[206] and Izotti et al.^[207] The

current review is concerned with the evidence supporting a role for oxidative stress in ON and RGC pathophysiology, and the ability of antioxidants to act as neuroprotectants to these tissues. As stated earlier (section 3.2), free radicals produced by activated astrocytes and by mitochondria within the ON, perhaps following ischaemia-reperfusion, are thought to contribute to extracellular matrix remodelling and atrophy in the lamina cribrosa, while in the retina free radicals are implicated in the death of RGCs caused by toxic mediators such as glutamate.

Because of the difficulties in analysing the ON or retina from POAG patients, direct evidence for an involvement of oxidative stress in ON atrophy and RGC death is sparse; however, there is an increasing body of indirect evidence that does support the hypothesis. The first strand of evidence comes from studies that have shown POAG patients to have reduced antioxidant defences. Birich et al.^[208] demonstrated that patients with POAG exhibited a higher concentration of serum lipid peroxidation products, Ferreira et al.^[209] conducted a prospective case control study and reported a lower total antioxidant capacity in aqueous humour from POAG patients than from controls, while Gherghel et al.^[210] measured total and reduced levels of the water-soluble antioxidant glutathione in the blood of untreated POAG patients and controls, and revealed that glaucoma patients exhibited lower levels of circulating glutathione. These studies suggest a general compromise of antioxidant defences in POAG patients, but which is not specific to the ON or retina.

The second strand of evidence concerns the detection of NOS-2 and the 27 kDa HSP in the ONH of glaucomatous eyes, and the elevated levels of iron-regulating genes in the retina of POAG patients. NOS-2, which is expressed by ONH astrocytes in glaucomatous eyes but is not present in healthy eyes,^[89] is a source of the free radical NO. Similarly, the redox chaperone HSP27, which serves as a cytoplasmic 'antioxidant',^[211] is present at higher levels in the ONH of glaucoma patients than healthy individuals.^[91] Transferrin, ceruloplasmin and ferritin regulate the availability of active iron as part of the antioxidant system and increased levels of these proteins are indicative of oxidative stress; all three

proteins have been detected at elevated levels in glaucomatous retinas relative to controls.^[212]

The third line of evidence comes from laboratory models of glaucoma. In both the episcleral vein cauterisation and the hyaluronic models of chronic hypertension, significant increases in the levels of reactive oxygen species and lipid peroxides have been detected in hypertensive eyes together with changes in the activities of antioxidant enzymes.^[213,214]

6.7.1 α -Tocopherol

α -Tocopherol (vitamin E) is quantitatively the major lipid-soluble antioxidant present in cell and mitochondrial membranes of the CNS and is present in high amounts within the rod photoreceptor outer segments and the retinal pigment epithelium. It acts as a powerful chain-breaking agent by scavenging peroxy radicals.

Activity: α -Tocopherol is an effective neuroprotectant in models of retinal ischaemia/reperfusion,^[215,216] but has not, to date, been tested in the chronic hypertensive models.

Clinical trials: In a short-term, nonblind, uncontrolled study, glaucoma patients receiving α -tocopherol had a reduced concentration of serum lipid peroxidation products and an expanded visual field,^[217] while in a separate study, glaucoma patients taking a complex containing α -tocopherol for 3 months displayed improved visual fields.^[218] Conversely, the results of a prospective, large-scale, long-term, epidemiological study that analysed the relationship between dietary antioxidant intake and POAG found little evidence that intake of α -tocopherol reduced the risk of developing POAG.^[219]

6.7.2 *Ginkgo biloba*

EGB 761 is a standardised extract of the leaves of the Chinese tree *Ginkgo biloba*, which improves blood flow in the whole body, including the eye,^[220] and which has been suggested to be beneficial in the treatment of mild to moderate dementia. EGB 761 contains two major groups of substances, the flavone glycosides and the terpene lactones, with >60 bioactive compounds in total.

Activity: EGB 761 has been demonstrated to protect the retina against ischaemia/reperfusion injury induced by transient ligation of the central retinal artery^[221,222] and acute elevation of IOP.^[223] In addi-

tion, pre-and post-treatment of rats with EGb 761 increased survival of RGCs in the episcleral vein cauterised model of chronic hypertension.^[122] In combination with other herbs, EGB 761 enhanced survival of RGCs in a model of ON injury.^[224] In retinal cultures, EGB 761 afforded protection against glutamate neurotoxicity.^[225]

Mode of action: In addition to its well documented vasotropic effect, the constituents of EGB 761 have been shown to have multiple cellular effects, which may account for the neuroprotective properties of the extract. EGB 761 is a potent antioxidant that has been shown to scavenge free radicals and to inhibit lipid peroxidation in the rat retina.^[226-228] It is also a potent inhibitor of platelet-activating factor, and has been shown to interfere with the glutamatergic NMDA receptor, to inhibit NOS activity and to stabilise the redox state of cells.^[229] Other as yet incompletely characterised mechanisms may also be involved.

Penetration: EGB 761 is taken orally, but information regarding the subsequent concentrations of its constituents in the retina is lacking. Several pharmacokinetic studies have been performed on EGb 761 and results indicate that extensive metabolism takes place following oral intake with bioavailability reaching <35%;^[230] thus, improved formulations are probably needed.

Clinical trials: In a small-scale, prospective, randomised, double-blind, crossover trial, EGb 761 administration effected an improvement in pre-existing visual field damage in some individuals with normal tension glaucoma.^[231]

6.7.3 Other Free Radical Scavengers

Aside from EGB 761, only one other study has assessed the effect of a free radical scavenger in a rat model of chronic hypertension. Ko et al.^[63] showed that systemic administration of the nonspecific free radical scavenger *N-tert-butyl*-(2-sulphophenyl)-nitron in the episcleral vein cauterisation model did not significantly improve survival of RGCs, although it enhanced the protection afforded by BDNF.

6.7.4 Summary

Free radical scavengers have proved effective retinal neuroprotectants in laboratory models of ischaemia/reperfusion and excitotoxicity; however,

the evidence to date in favour of their effectiveness in laboratory models of glaucoma, or more importantly in POAG patients, is at best tentative. Although recent studies have documented increased oxidative stress in the ONH and retina, only two free radical scavengers have been tested in models of chronic hypertension, and the beneficial effects of EGB 761 on RGC survival may not be due to its antioxidant properties. More basic and clinical research is needed into the role of oxidative stress in glaucoma.

6.8 Heat Shock Proteins

HSPs are ubiquitous, highly conserved molecular chaperones. They have multiple functions under normal conditions, such as facilitating proper assembly and intracellular folding of nascent polypeptides, and regulating protein degradation. HSPs are classified by their molecular weights (in kDa), comprising five main families: HSP110, HSP90, HSP70, HSP60 and the small HSPs. Induction of HSP expression occurs in response to a variety of stress situations, where it is believed to play an important role in the re-establishment of normal tissue function and in providing tolerance against further trauma. HSP70, often referred to as HSP72 owing to its apparent molecular weight, and HSP27, a small HSP, are of particular interest in the CNS because of their highly inducible nature in glia and neurons, and the established connection between their induction and subsequent neuroprotection from various injuries.^[232,233]

Recent studies have shown increased expression of HSPs in the eyes of glaucoma patients and animals with chronic hypertension. In glaucomatous human eyes, Tezel et al.^[91] found increased immunostaining for HSP60 and HSP27 in RGCs and the nerve fibre layer, together with increased labelling for HSP27 in astrocytes of the lamina cribrosa. A rationale for the latter finding was provided by the results of a study that showed cultured human ONH astrocytes exposed to elevated hydrostatic pressure increased their expression of HSP27.^[234] In monkeys with laser-induced glaucoma, Sakai et al.^[93] showed markedly enhanced labelling for HSP90, HSP60 and HSP27 in the RGC and nerve fibre layers, and moderately increased staining intensity for HSP70. In rats with chronic hypertension,

a robust and sustained increase in HSP27 was noted in RGCs,^[92] but only a moderate and transient increase in HSP70.^[95]

Increased expression of HSPs in the glaucomatous retina and ONH might indicate that these proteins form part of an endogenous defence mechanism in response to trauma. Support for this idea has been provided by the studies of Caprioli and colleagues^[95,123] who demonstrated that prior augmentation of HSP70 expression by RGCs, achieved by heat stress or pharmacological intervention, protected against RGC loss in rats caused by long-term elevation of IOP (see section 6.8.1). Since HSP27 is strongly upregulated in glaucomatous retinas, it would be interesting to determine if RGC loss in animals with chronic hypertension is accelerated in animals lacking the ability to synthesise this HSP.

6.8.1 Geranylgeranylacetone

Geranylgeranylacetone (GGA) is an anti-ulcer drug developed in Japan, which is used clinically for the treatment of gastric disorders. GGA has been shown to induce HSP70 synthesis in multiple tissues and to confer protection from various types of toxicity.

Activity: Repeated, systemic administration of GGA (200 mg/kg) reduced RGC loss and ON damage in the laser-induced model of chronic hypertension.^[123]

Mode of action: The authors demonstrated increased expression of HSP70 in RGCs after administration of GGA. The effects of GGA on HSP70 expression and RGC survival were prevented by inclusion of a recognised inhibitor of HSP expression, quercetin.

Clinical trials: GGA has not been tested in glaucoma patients. The compound is widely used as an oral antiulcer drug and has extremely low toxicity; thus, a clinical trial warrants consideration.

6.8.2 Summary

Overexpression of HSPs has been shown to afford protection to RGCs from injury caused by ischaemia-reperfusion, excitotoxicity, ON transection and experimental glaucoma, and pharmacological manipulation of HSP expression is an attractive therapeutic approach for use in glaucoma. Nevertheless, there are certain issues that need to be addressed if such a strategy is to be successful. Perhaps

the issue of most importance is the possibility that HSPs may contribute to, rather than retard, the progression of glaucoma. HSPs possess a number of immunological activities and their enhanced expression in RGCs of glaucomatous eyes may be a signal to the immune system of the location of stressed cells that need to be removed. The idea that HSPs may be immune targets was proposed by Wax and colleagues^[38] and arose from their findings of increased titres of HSP autoantibodies in the blood of glaucoma patients.^[235] Other issues to be addressed include delivery of HSPs to RGCs, and whether single or multiple HSPs should be upregulated in the tissue. There are many ways to increase HSP levels in a tissue, including physiological, pharmacological and genetic induction of HSP expression, and direct administration of the proteins. Treatment for a chronic, neurodegenerative disease such as glaucoma based on HSP induction will require a suitable delivery mechanism that takes into consideration the site of damage, the time course of the injury and the specificity of the response desired. Although studies to date have tended to concentrate on HSP70, evidence from the CNS suggests that HSP27 may have an equally potent and more wide-ranging protective effect.^[232] With regard to the retina, Kretz et al.^[236] recently showed that overexpression of HSP27 enhanced RGC survival following ON transection.

6.9 Retinal Ganglion Cell Death and Apoptosis

Apoptosis has been described as the opposite of necrosis (accidental, unregulated cell death) and is a genetically regulated cellular death process reliant upon a distinct temporal sequence of events such as mobilisation and post-translational activation of enzymes (see figure 1 and the excellent review by Hengartner^[237] for a summary of the basic pathways). During apoptosis, cellular constituents are internally degraded meaning that there is condensation and shrinkage of the nucleus and cytoplasm, but no associated tissue inflammation.

6.9.1 Evidence for Apoptosis in Glaucoma

Studies conducted over the last decade have suggested that apoptosis plays a major role in the process of RGC death. Cells dying by apoptosis were initially identified in the axotomised rat retina^[58,62]

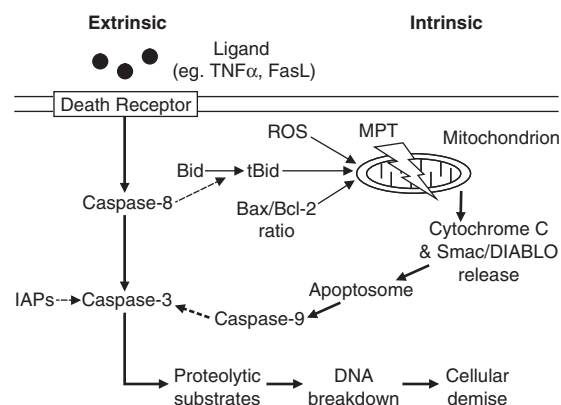


Fig. 1. Simplified scheme of events underlying the process of apoptotic cell death. Two major pathways have been described: the intrinsic pathway and the extrinsic pathway. The extrinsic pathway is stimulated by either the presence or absence of ligand-binding at cell surface receptors (e.g. FasR, tumour necrosis factor- α R [TNF α R]). This leads to the aggregation and activation of caspase-8 proforms and subsequent proteolytic cleavage and activation of caspase-3, which is instrumental in the dismantling of cellular structure and machinery. The intrinsic route to cell death, on the other hand, is stimulated by unknown events and results in a homeostatic imbalance in mitochondrial function, leading to mitochondrial permeability transition, and a release into the cytoplasm of apoptotic inducer proteins such as cytochrome C and Smac/DIABLO. This process also culminates in the activation of caspase-3. The roles of other caspase family members remain unclear. It is also likely that the cellular balance between pro-apoptotic Bax and anti-apoptotic Bcl-2, the cleavage by caspase-8 of pro-apoptotic Bid to truncated Bid (tBid) and the local levels of reactive oxygen species (ROS) contribute to stimulation of the mitochondrial-mediated intrinsic pathway. Major pathways to cell death are shown with thick arrows, speculative pathways in thin arrows and activation events by dotted arrows. **IAP** = inhibitor of apoptosis protein; **MPT** = mitochondrial permeability transition.

and in a monkey model of chronic IOP.^[238] Subsequent studies have confirmed the occurrence of apoptotic RGC death after axotomy,^[57,239-242] and similarly shown it to be a feature of central retinal artery occlusion,^[243] excitotoxicity^[244,245] and retinal ischaemia.^[246,247]

Apoptotic RGC death has been observed in rats with chronic ocular hypertension,^[58-60,248-253] as well as in quail with a glaucoma-like mutation.^[134] Furthermore, the quantities of TUNEL-positive cells in the RGC layer of human eyes with both POAG and secondary glaucoma have been shown to be significantly elevated with respect to control retinas.^[32,254] Apoptotic cells have also been detected by visualisation of apoptotic chromatin condensation in eyes

from patients diagnosed with normal pressure glaucoma^[255] and anterior ischaemic optic neuropathy.^[256]

6.9.2 Apoptotic Pathways in Glaucoma

Over recent years, intensive research has delineated many of the crucial signalling pathways involved in the process of apoptosis; figure 1 provides a summary of the major events that have been described in relation to RGC death in glaucoma models. It has become apparent that it is important to distinguish between the extrinsic, or receptor-activated pathways, and the intrinsic or mitochondrial-mediated events. Although triggers for the extrinsic pathway (e.g. TNF α , FasL, TNF-related apoptosis-inducing ligand [TRAIL]) have been well described and understood, the relationship between intracellular signals and mitochondria is not so clear.^[237] For example, it is apparent that the cellular balance between levels of pro-apoptotic Bax and anti-apoptotic Bcl-2 (and related family members) as well as the local availability of reactive oxygen species (ROS) play central roles in determining how and when mitochondria become stimulated to release their contents, initiating formation of the intrinsic pathway apoptosome (figure 1). However, the basis for mitochondrial opening, via the mitochondrial permeability transition (MPT), is unknown. Furthermore, there is an apparent redundancy in caspase-mediated substrate proteolysis, with many caspase enzymes appearing to perform very similar tasks. Regardless of these caveats, however, it is clear that there are certain key signalling mediators involved in RGC apoptosis (Bax/Bad, Bcl-2/Bcl-X_L, p53, caspases-1, 3, 8, 9, Bid and PKB/Akt) and these elements have been subject to increasing levels of scrutiny in order to determine their exact roles in the process.

A large volume of research information has been generated over the last decade concerning the involvement of intracellular mediators in experimental RGC death induced in models such as ON axotomy or acute elevation of IOP. However, information regarding the pathways involved in RGC death in experimental models of chronic IOP elevation is more limited. Activation of caspases contributes to cellular disassembly as a result of chronic ocular hypertension, with procaspases-3, 8 and 9 cleaved to

yield their active configurations.^[250,257-260] Furthermore, caspase-3-mediated proteolytic degradation of amyloid precursor protein (APP)^[258] and α -fodrin^[125] increases in rodent models of chronic ocular hypertension. There are also transient elevations in RGC expression of both Bax and Bcl-2, which almost completely revert to pre-hypertension levels after 2 weeks of IOP elevation.^[259] Bax is also involved in RGC death in the DBA/2J mouse model,^[261] as are increased dephosphorylation of pro-apoptotic Bad and cytoplasmic elevation of cytochrome C, both known to be associated with the induction of apoptosis.^[124] Bax is further elevated, along with Bak, p53, Fas, ICE and caspase-3 in RGCs^[262] in the mouse episcleral vein photocoagulation model^[263] and this is concurrent with a decrease in Bcl-2 expression.^[262] Moreover, expression of p53 gene family members *Gadd45a* and *Ei24* is also stimulated in RGC apoptosis after laser-induced chronic ocular hypertension in rats,^[264] although interestingly, this study also showed that anti-apoptotic gene expression of the caspase inhibitor *IAP-1* is simultaneously upregulated, perhaps as part of an endogenous neuroprotective mechanism.^[264] Finally, the PKB/Akt signalling pathway has been shown to have an involvement in RGC death in the episcleral vein model. PKB/Akt phosphorylation leads to its activation and this enzyme can subsequently promote cell survival by deactivating enzymes such as Bad, glycogen synthase kinase, caspase-3 and caspase-9, and influencing MAP kinase signalling pathways. Significantly, however, PKB/Akt undergoes dephosphorylation after episcleral vein cauterisation in the rat and this may prove to be a crucial early event in the initiation of RGC death in this model.^[250,265]

Two things are apparent from the studies mentioned in the previous paragraph. First, pro-apoptotic cellular mediators (e.g. Bax, caspase-3) are *always* shown to be activated/upregulated in chronic ocular hypertension models. Secondly, the role of anti-apoptotic components in relevant signalling pathways is not clear. In some cases, for example, Bcl-2 is shown to be decreased,^[262] which would be consistent with a downregulation event preceding initiation of death signalling. In other cases, Bcl-2 appears to be upregulated,^[259] and this may indicate a role for promotion of survival of some cells and

not others. Indeed, the role of anti-apoptotic signalling requires further investigation because knowledge of the function that these factors play in cell death may allow their manipulation in order to promote RGC survival.

6.9.3 Pharmacological Intervention in Apoptosis

Specific biomolecular triggers for apoptotic RGC death in glaucoma remain elusive; suggestions have included reduced essential trophic factor^[30,36] or cytokine^[266] supply to neurons, alterations in intracellular levels of calcium or reactive oxygen intermediates,^[267] or increased extracellular levels of certain neurotransmitters and neuromodulators.^[40] It follows that interventions such as trophic factor delivery, excitatory receptor antagonists and antioxidants should prevent either the initiation or the enactment of apoptotic pathways, thereby offering some degree of protection in glaucoma. These treatment modalities do indeed offer some protection to RGCs in animal models of glaucoma, but relatively little mechanistic information has been uncovered regarding whether they interact directly with specific elements of apoptotic pathways.

Direct manipulation of intracellular apoptotic pathways in RGCs, for example by gene therapy or by pharmacological intervention using pro-apoptotic enzyme modulators, offers a potential way in which glaucoma may be treated in the future. To date, the majority of ocular studies that have described pharmacological anti-apoptotic strategies have fallen into two categories. The first approach has been to manipulate the cellular levels of proteins involved in apoptotic pathways. Administration of the α_2 -adrenoceptor antagonist brimonidine (see section 7.1), for example, which protects RGCs in ocular hypertensive models, has been shown to activate the anti-apoptotic ERK and Akt pathways, thereby increasing the synthesis of Bcl-2 and Bcl-X_L.^[268] In cultured ganglion cells, lithium pretreatment also resulted in an upregulation of Bcl-2, and this was characterised to promote both cell survival and axon regeneration.^[269] Furthermore, treatment of ocular hypertensive rats with a calcineurin inhibitor prevented Bad (a proapoptotic molecule of the Bcl-2 family) dephosphorylation and cytochrome C release and this resulted in a marked increase in RGC survival.^[124]

The second approach has been to utilise caspase inhibitors to prevent RGC degradation; this has been the more widely utilised strategy. Intraocular injection of caspase-3 inhibitors has been demonstrated to increase the survival of RGCs after ON transection,^[240,270-273] high IOP-induced ischaemia,^[247] and ligation of the ophthalmic vessels.^[272] Blocking caspase-3 also prevented glutamate-induced RGC death in retinal explants.^[274] In rats with chronic ocular hypertension, inhibition of caspase-3 has been demonstrated to protect RGCs and to prevent cleavage of the neuronal cytoskeletal protein α -fodrin, a known target for caspase-3.^[125,126]

Caspase-3 is a pivotal and relatively late-stage component of multiple apoptotic cascades; hence its involvement in RGC death is to be expected. To date, the effectiveness of other classes of caspase inhibitors at preventing RGC death in ocular models of hypertension has not been investigated. Some success would be anticipated for two reasons: firstly, that several cell death pathways appear to be activated, and proapoptotic genes upregulated, in retinas with experimental glaucoma;^[250,260,264] and secondly, that inhibitors of other caspases have proved efficacious in alternative models of RGC injury. For example, inhibition of caspase-1 protected RGCs from NMDA,^[244,275] and ischaemia-induced^[247] apoptotic death, while caspase-8 inhibition aided survival of RGCs, *in vitro*, from simulated ischaemia, NMDA or HSP27 antibody application,^[94] as well as preventing RGC death, *in vivo*, as a result of axotomy.^[276]

6.9.4 Summary

The strong protective effects of such different caspase inhibitors indicate that different intracellular pathways are likely to contribute to RGC death. However, the actual pathways involved in the death of RGCs in glaucoma remain uncertain, meaning that, at present, it is impossible to comment on the validity of such experimental data for treatment of patients in the clinic. Moreover, the majority of investigations targeting the effect of caspase inhibition on paradigms of RGC death have employed intravitreal injections to apply the compounds. Obviously, in the case of human glaucoma this is unlikely to offer a valid therapeutic approach. Therefore, the potential of using caspase inhibitors

for human glaucoma treatment is, at present, also likely to be limited by potential pharmacokinetic constraints. Orally tolerated caspase inhibitor peptides have now been produced; for example, the Aventis compound pralnacasan (VX-740), which is a caspase-1 inhibitor that has recently been entered into a phase II clinical trial for patients with rheumatoid arthritis and other inflammatory diseases.^[277,278] More recently still, a series of oxamyl dipeptide compounds have been generated that have been optimised for pan-caspase inhibition, anti-apoptotic cellular activity and *in vivo* efficacy.^[279-281] On the basis of *in vitro* and *in vivo* experiments in murine models of liver injury, one inhibitor PF-03491390 [IDN-6556 (1)] has also been selected for two phase II clinical trials, evaluating its safety and efficacy for use in liver disease.^[282,283] Therefore, the potential exists to apply such compounds to the retina in the treatment of glaucoma.

In his excellent recent review, Nickells^[284] described the need for caution in interpretation and long-term significance of apoptotic findings in relation to glaucomatous RGC death. He astutely outlined that a number of points should be borne in mind when assessing the effects of anti-apoptotic strategies: first, that some events (e.g. mitochondrial dysfunction) are downstream in the overall apoptotic scheme and thus preventing these will not necessarily protect cells; second, that multiple pathways to cell death exist and shutting one or more of these down may, therefore, only lead to a circumvention of these specific processes; and third, that prevention of RGC apoptosis may leave behind a live but non-functioning cell. If these points are viewed in conjunction with the fact that not all RGC death is likely to be via classically defined apoptosis, then a note of caution must be applied to the possible anti-apoptosis strategy of treating RGC death in glaucoma.

7. Neuroprotective Properties of Drugs Already Used as Ocular Hypotensives

In recent years, a significant body of work has been conducted on the abilities of a range of drugs already used clinically as ocular hypotensives to act as retinal neuroprotectants. It is of course well documented that the best method of preventing RGC death in glaucoma or laboratory models of glaucoma

is to reduce the IOP; however, the concept of neuroprotection embraces the idea that a drug can protect RGCs independent of a reduction in IOP. If such a quality were to be demonstrated for a drug, this would enable it to be administered alongside a hypotensive agent to provide additional protection. The difficulty in determining the neuroprotective potential of drugs currently prescribed for the treatment of glaucoma in animal models of glaucoma is that the drugs are likely to lower IOP in the chronic hypertensive models, which makes it almost impossible to delineate whether any beneficial effect on RGC survival occurs via mechanisms that are independent of IOP. Bearing this in mind, in this review of the neuroprotective properties of hypotensive agents, particular attention has been paid to other models of RGC death, such as ischaemia/reperfusion, excitotoxicity and ON injury. As discussed previously (section 4), all of these paradigms of injury show some similarities to glaucoma, and information gained from them is, therefore, likely to have some relevance to the disease.

7.1 α_2 -Adrenoceptor Antagonists

7.1.1 *Brimonidine*

Brimonidine (Alphagan®)¹ is a highly selective α_2 -adrenoceptor agonist, which lowers IOP primarily by decreasing aqueous humour inflow. The evidence in favour of a neuroprotective action of brimonidine in the laboratory is convincing and has been covered in detail in a recent review article.^[14] Nevertheless, the data are worth summarising as the thoroughness of the work performed using this compound sets a benchmark for other drugs.

Activity: Systemic and topical administration of brimonidine resulted in protective effects in various acute and chronic rodent paradigms of RGC death, including ischaemia/reperfusion induced by acute elevation of IOP,^[285-287] ischaemia/reperfusion induced by transient ligation of the ophthalmic vessels,^[288-291] mechanical injury to the ON,^[292] and in the lasered^[104] and episcleral vein cauterised^[127] rat models of chronic ocular hypertension. Brimonidine also increased survival of retinal neurons in culture following exposure to the excitotoxin glutamate.^[293]

Target: α_2 -Adrenoceptors have been identified in the ganglion cell layer of the retina,^[107,294] and the protective effects of brimonidine in all of the models of injury are likely to be mediated via activation of these receptors since they can be blocked by co-administration of α_2 -adrenoceptor antagonists. Furthermore, indirect evidence for a receptor-mediated neuroprotective mode of action is provided by the knowledge that other α_2 -adrenoceptor agonists, such as clonidine and apraclonidine, are also neuroprotective in models of RGC death.^[295,296]

Mode of action: Activation of α_2 -adrenoceptors provides neuroprotection to RGCs via complex but seemingly independent mechanisms. Evidence is accumulating that α_2 -adrenoceptor activation leads to inhibition of pro-apoptotic pathways (see review by Tatton et al.^[268]), release of trophic factors, including BDNF^[297] and basic fibroblast growth factor,^[287] and inhibition of ischaemia-induced glutamate release.^[286]

Penetration: Following multiple, topical application of brimonidine 0.2% in patients for up to 2 weeks, the mean vitreous concentration of drug was determined to be 185 nmol/L, as measured by gas chromatography.^[298] Following multiple, topical administration of [¹⁴C]brimonidine 0.2% in rabbits and monkeys, non-metabolised radioactivity concentrations of 82 and 170 nmol/L, respectively, were detected in the vitreous humour.^[299] In the same study, the choroid/retina and ONH were found to contain even higher amounts of intact radioactivity. All of these levels of brimonidine are substantially higher than necessary to cause full activation of α_2 -adrenoceptors.

Clinical trials: Brimonidine has recently been shown to improve contrast sensitivity in glaucoma patients,^[300] and the weight of evidence in favour of brimonidine as a potential neuroprotective agent has led to a large, multicentre, prospective randomised study comparing the effect of brimonidine to timolol on visual field changes in patients with normal tension glaucoma.^[301]

1 The use of trade names is for product identification purposes only and does not imply endorsement.

7.2 β -Adrenoceptor Antagonists (β -Blockers)

7.2.1 Betaxolol

Betaxolol (Betoptic[®], Betoptic S[®]) is a relatively selective β_1 -adrenoceptor antagonist (β -blocker), which is nevertheless considered to lower IOP via inhibiting β_2 -adrenoceptors located in the non-pigmented ciliary epithelium, thereby decreasing aqueous humour inflow.

Activity: Racemic and levo-betaxolol have been demonstrated to reduce RGC death following ischaemia/reperfusion^[79,80,302-306] and excitotoxicity.^[303] Neuroprotection was evident irrespective of whether the drug was administered topically, intravitreally or systemically. Betaxolol also dramatically prevented RGC death and ON damage in the hypertonic saline rat model of chronic hypertension;^[251] however, since betaxolol reduced IOP significantly in treated eyes, it was not possible to distinguish between any direct neuroprotective effects and the effect due solely to lowering of IOP. *In vitro* studies support the case for a neuroprotective role for betaxolol in the retina, where it has been shown that betaxolol increases survival of retinal neurons in culture following excitotoxic and anoxic insults.^[293,305]

Target: Unlike brimonidine, the neuroprotective actions of betaxolol are not thought to be classically receptor-mediated. There is sparse evidence for the presence of β_1 -adrenoceptors in retinal/ON tissue, and although the β_2 subtype has been localised to ON astrocytes,^[307] it has not been identified in the retinal layers. Furthermore, studies have shown that betaxolol elicits a vastly superior neuroprotection to timolol or metoprolol,^[293,305,308] yet all three compounds have similar affinities for β_1 -adrenoceptors, and timolol a much higher affinity for the β_2 subtype. Instead, betaxolol is thought to increase RGC survival via direct inhibition of voltage-gated Ca^{2+} and Na^+ channels and glutamate currents in the retina/ON (see following paragraph).

Mode of action: There is considerable laboratory evidence to indicate that betaxolol can prevent excessive influx of Ca^{2+} and Na^+ ions into neurons, which occurs in pathological situations such as hypoxia, ischaemia and excitotoxicity.^[15] Electrophysiological experiments have revealed that betaxolol inhibits voltage-gated Ca^{2+} and Na^+ currents and

glutamate-gated currents in RGCs,^[309,310] while biochemical studies have shown the drug to inhibit NMDA- and/or glutamate-stimulated Ca^{2+} influx in whole retina and isolated RGCs,^[293,310-312] as well as Na^+ influx in rat brain.^[313] The inhibitory actions of betaxolol are most likely explained by the fact that the drug binds allosterically to L-type voltage-gated Ca^{2+} channels^[314] and voltage-gated Na^+ channels;^[313] however, it is also possible that betaxolol lowers the affinity of glutamate receptors for glutamate.^[312] Betaxolol has also been shown to upregulate expression of trophic factors in the retina^[304,315] and this too may play a role in the neuroprotection afforded by the drug.

Penetration: Studies in rabbits have shown that a significant portion of betaxolol reaches the retina when administered topically.^[303,316] These data indicate that local ocular and systemic routes both play a significant role in delivering betaxolol to the retina. It is difficult to estimate the concentration of betaxolol at the site of action in the retina/ON following topical application because betaxolol is highly lipophilic and will accumulate in membranes. Since the ion channels affected by betaxolol are located in cell membranes, the actual concentration of the drug at the site of action is likely to be noticeably higher than the concentration measured in the vitreous or retina as a whole.

Clinical trials: Although no large-scale clinical trials have been conducted, a number of independent, small-scale clinical trials have compared the effect of betaxolol with timolol on visual fields in patients with open-angle glaucoma.^[317-325]

The conclusion to be drawn from all of these studies is that betaxolol provides slightly better preservation of the visual field than timolol, despite its less efficacious effect on reducing IOP.

7.2.2 Timolol

Timolol (Timoptic[®]) is a non-selective β -blocker, which, like all β -blockers, lowers IOP by interacting with β_2 -adrenoceptors in the ciliary epithelium causing a reduced secretion of aqueous humour.

Activity: Scrutiny of the published literature relating to timolol reveals inconsistent results, wherein some studies show timolol to be moderately neuroprotective, yet others, using similar methodologies and models of injury, find little beneficial

effect. For example, Goto et al.^[326] showed that timolol attenuated glutamate toxicity in both mixed retinal and purified RGC cultures; conversely, Baptiste et al.^[293] found that timolol had no effect on neuronal survival in mixed retinal cultures treated with glutamate, and Kashiwagi et al.^[327] similarly found no effect of timolol on glutamate-mediated toxicity in purified RGC cultures. With regard to models of chronic ocular hypertension, it appears likely that the neuroprotective effectiveness of timolol is related to its ability to lower IOP rather than via any direct action on neuronal/glial components. Systemically administered timolol reduced IOP only slightly and failed to provide any neuroprotection,^[104] but topically applied timolol has been shown to reduce IOP efficaciously and counteract RGC loss.^[328,329] Timolol was protective in the DBA/2J mouse model of glaucoma,^[111] presumably via its ability to lower IOP in affected eyes; similarly in the episcleral vein cauterised model, timolol lowered IOP and caused a partial rescue of RGCs.^[330] In an acute model of ischaemia/reperfusion, Wood et al.^[305] found topically applied timolol to be only marginally neuroprotective against ischaemia/reperfusion injury, but Goto et al.,^[326] who used twice the dose of timolol, found the drug to be more effective. Timolol afforded no protection in an acute model of ON injury.^[331] The logical conclusion to be drawn from all of the results is that the compound is a weak neuroprotectant and needs to be used at high doses to exert direct neuroprotective effects. This conclusion is supported by studies that have analysed the mechanisms by which timolol might aid survival of RGCs.

Mode of action: Timolol does bind to voltage-gated Ca^{2+} and Na^{+} channels^[313,314] and reduce NMDA-stimulated Ca^{2+} influx,^[311,312] but with a very much lower affinity than betaxolol. This is manifest from electrophysiological studies where timolol did not display any detectable action on Ca^{2+} and Na^{+} currents.^[309,310] At present, other mechanisms of action of timolol are unknown. It does not act as a free radical scavenger^[332] and has not been documented to increase expression of trophic/survival factors.

Penetration: Timolol reaches the retina in significant amounts after topical application. Tan et al.^[333] suggest that absorption of timolol into the systemic

circulation plays a significant role in delivering timolol to the retina, while Goto et al.^[326] argue that periocular transscleral and/or vitreous routes are more important.

7.2.3 Metipranolol

Metipranolol (OptiPranolol®) is a non-selective β -blocker with an IOP-lowering efficacy similar to that of other non-selective β -blockers. It is less widely prescribed than other β -blockers, primarily because of early reports that it was associated with granulomatous uveitis. Topically applied metipranolol has been shown to partially attenuate the effects of ischaemia/reperfusion induced by acute elevation of IOP in the rat and anoxia-induced neuronal loss in rat retinal cultures,^[305] but the drug has not been tested in other paradigms of RGC death.

Mode of action: Metipranolol inhibits Na^{+} and Ca^{2+} influx into neurons in the same manner as betaxolol, albeit less potently.^[305] Interestingly, metipranolol, uniquely of the ophthalmic β -blockers, has also been shown to act as a potent antioxidant, reducing lipid peroxidation in brain homogenates^[332] and protecting the retina against damage caused by oxidants.^[334,335] Since free radical formation is implicated in the pathogenesis of glaucoma, it will be of interest to test the effect of metipranolol in other models of RGC death.

Penetration: Metipranolol is assumed to reach the posterior segment of the eye in rats since it elicits some protection when applied topically, but no penetration studies have been conducted in rats or humans.

No studies have been conducted to determine the neuroprotective properties of the other commonly prescribed β -blockers carteolol (Ocupress®/Teopitic®) or levobunolol (Betagan®).

7.3 Prostaglandin Derivatives

Latanoprost (Xalatan®) and travoprost (Travatan®), bimatoprost (Lumigan®) and unoprostone (Rescula®) are derivatives of prostaglandin $\text{F}_{2\alpha}$ ($\text{PGF}_{2\alpha}$) that reduce IOP by enhancing outflow of aqueous humor. Although the drugs are classified as $\text{PGF}_{2\alpha}$ analogues, a prostamide and a docosanoid, respectively, they are fundamentally alike in terms of structure and largely with respect to their mechanisms of action. In recent years, this new class of

drugs has replaced the topical β -blockers as the most commonly prescribed first-line treatment for glaucoma. Prostaglandins, including $\text{PGF}_{2\alpha}$, are well documented vasoconstrictors; they are also implicated in the pathogenesis of ischaemic and inflammatory injuries and can potentially influence many cell signalling pathways. However, to date, there is no evidence to indicate that the prostaglandin derivatives used to lower IOP are neurotoxic to the retina/ON. In fact, there are very limited data to suggest the reverse is true.

Activity: Intravitreal administration of latanoprost to rats has been documented to increase survival of RGCs after NMDA injection and ON transection, by an unknown mechanism.^[336] In a different study, systemically administered latanoprost partially prevented the accumulation of lactate induced by retinal ischaemia;^[337] unfortunately, no electrophysiological or histological data were recorded. The drug also attenuated glutamate and hypoxia-induced cell death in retinal cultures. The authors hypothesised that latanoprost may exert neuroprotective effects via a negative feedback on cyclo-oxygenase-2 activity.

Unoprostone has also been shown to protect the retina in a model of ischaemia.^[338] In this case, the mode of action of the drug is suggested to involve opening of maxi K^+ channels.

At present, no information is available with regard to travoprost or bimatoprost.

7.4 Carbonic Anhydrase Inhibitors

Dorzolamide (Trusopt®) and brinzolamide (Azopt®) are selective inhibitors of carbonic anhydrase isoenzyme II. Inhibition of this enzyme in cells of the ciliary epithelium causes a reduced formation of aqueous humour and a decrease in IOP.

Activity: Some studies have shown that carbonic anhydrase inhibitors increase choroidal and ONH blood flow via mechanisms that are independent of IOP,^[113,339] however, little evidence has been presented to date to indicate that these drugs can protect the retina in models of RGC death.

Dorzolamide failed to rescue RGCs in the DBA/2J mouse model of inherited glaucoma^[111] and, although the drug did afford protection in rat models of chronic hypertension,^[328,329] it also reduced IOP

in these eyes; the degree of damage/protection correlated with the level of IOP, indicating that the effect was entirely pressure-related and not caused by direct neuroprotection. Dorzolamide has not been tested in ischaemic, excitotoxic or ON injury models of RGC death.

7.5 Summary

Of the drugs that are currently prescribed to treat glaucoma, evidence to date indicates that brimonidine and betaxolol have significant neuroprotective properties. Small-scale clinical trials with betaxolol have shown that the drug performs slightly better than would be expected purely from its ability to lower IOP. The ongoing trial to identify whether brimonidine preserves visual fields more than timolol is of real importance for the future.

8. Conclusions

Despite the efforts of researchers to date, the specific site of the primary injury in glaucoma remains uncertain. The vast majority of clinical and experimental evidence indicates that the ONH is the primary pathological site, but there is limited evidence to suggest that RGC bodies may be primarily involved. The site of the primary injury has particular relevance to neuroprotection because different regions of the RGC may respond to different neuroprotective strategies. Indeed, it has been argued that a compartmentalised approach to neuroprotection in glaucoma may be the optimal strategy.^[46]

In recent years, much progress has been made in identifying potential pharmacological targets for glaucoma. This process has been aided considerably by the development of rodent models of chronic ocular hypertension, which appear to display many similarities to glaucoma. At present, pharmacological neuroprotection for glaucoma remains a concept rather than a reality. In the not too distant future, however, it is highly likely that administration of neuroprotectants will form an adjunct to IOP-lowering drugs in the treatment of this debilitating disease.

Acknowledgements

G. Chidlow and J.P.M. Wood would like to express their gratitude to Prof. Neville Osborne of Oxford University for his guidance, encouragement and friendship during the 12 years they spent in his laboratory. This research was supported by the Ophthalmic Research Institute of Australia (ORIA). R.J. Casson is a recipient of a Pfizer Neuroscience Award. The authors have no conflicts of interest directly relevant to the contents of this review. No sources of funding were used to assist in the preparation of this review.

References

- Quigley HA. Number of people with glaucoma worldwide. *Br J Ophthalmol* 1996; 80 (5): 389-93
- Kass MA, Heuer DK, Higginbotham EJ, et al. The Ocular Hypertension Treatment Study: a randomized trial determines that topical ocular hypotensive medication delays or prevents the onset of primary open-angle glaucoma. *Arch Ophthalmol* 2002; 120 (6): 701-13
- Collaborative Normal-Tension Glaucoma Study Group. Comparison of glaucomatous progression between untreated patients with normal-tension glaucoma and patients with therapeutically reduced intraocular pressures. *Am J Ophthalmol* 1998; 126 (4): 487-97
- The Advanced Glaucoma Intervention Study (AGIS). The relationship between control of intraocular pressure and visual field deterioration. The AGIS Investigators. *Am J Ophthalmol* 2000; 130 (6): 429-40
- Leskea MC, Heijl A, Hyman L, et al. Factors for progression and glaucoma treatment: the Early Manifest Glaucoma Trial. *Curr Opin Ophthalmol* 2004; 15 (2): 102-6
- Schwartz M, Belkin M, Yoles E, et al. Potential treatment modalities for glaucomatous neuropathy: neuroprotection and neuroregeneration. *J Glaucoma* 1996; 5 (6): 427-32
- Osborne NN, Chidlow G, Nash MS, et al. The potential of neuroprotection in glaucoma treatment. *Curr Opin Ophthalmol* 1999; 10 (2): 82-92
- Levin LA. Direct and indirect approaches to neuroprotective therapy of glaucomatous optic neuropathy. *Surv Ophthalmol* 1999; 43 Suppl. 1: S98-101
- Yoles E, Schwartz M. Degeneration of spared axons following partial white matter lesion: implications for optic nerve neuropathies. *Exp Neurol* 1998; 153 (1): 1-7
- Levkovitch-Verbin H, Quigley HA, Martin KR, et al. A model to study differences between primary and secondary degeneration of retinal ganglion cells in rats by partial optic nerve transection. *Invest Ophthalmol Vis Sci* 2003; 44 (8): 3388-93
- Harris A, Ciulla TA, Kagemann L, et al. Vasoprotection as neuroprotection for the optic nerve. *Eye* 2000; 14 (Pt 3B): 473-5
- Flammer J, Orgul S, Costa VP, et al. The impact of ocular blood flow in glaucoma. *Prog Retin Eye Res* 2002; 21 (4): 359-93
- Costa VP, Harris A, Stefansson E, et al. The effects of anti-glaucoma and systemic medications on ocular blood flow. *Prog Retin Eye Res* 2003; 22 (6): 769-805
- Wheeler L, WoldeMussie E, Lai R. Role of alpha-2 agonists in neuroprotection. *Surv Ophthalmol* 2003; 48 Suppl. 1: S47-51
- Osborne NN, Wood JP, Chidlow G. Invited review: neuroprotective properties of certain beta-adrenoceptor antagonists used for the treatment of glaucoma. *J Ocul Pharmacol Ther* 2005; 21 (3): 175-81
- Krupin T, Liebmann JM, Greenfield DS, et al. The Low-pressure Glaucoma Treatment Study (LoGTS) study design and baseline characteristics of enrolled patients. *Ophthalmology* 2005; 112 (3): 376-85
- Glovinsky Y, Quigley HA, Dunkelberger GR. Retinal ganglion cell loss is size dependent in experimental glaucoma. *Invest Ophthalmol Vis Sci* 1991; 32 (3): 484-91
- Quigley HA, Dunkelberger GR, Green WR. Chronic human glaucoma causing selectively greater loss of large optic nerve fibers. *Ophthalmology* 1988; 95 (3): 357-63
- Morgan JE, Uchida H, Caprioli J. Retinal ganglion cell death in experimental glaucoma. *Br J Ophthalmol* 2000; 84 (3): 303-10
- Morgan JE. Retinal ganglion cell shrinkage in glaucoma. *J Glaucoma* 2002; 11 (4): 365-70
- Kerrigan-Baumrind LA, Quigley HA, Pease ME, et al. Number of ganglion cells in glaucoma eyes compared with threshold visual field tests in the same persons. *Invest Ophthalmol Vis Sci* 2000; 41 (3): 741-8
- Quigley HA, Green WR. The histology of human glaucoma cupping and optic nerve damage: clinicopathologic correlation in 21 eyes. *Ophthalmology* 1979; 86 (10): 1803-30
- Zhao DY, Cioffi GA. Anterior optic nerve microvascular changes in human glaucomatous optic neuropathy. *Eye* 2000; 14 (Pt 3B): 445-9
- Quigley HA, Hohman RM, Addicks EM, et al. Blood vessels of the glaucomatous optic disc in experimental primate and human eyes. *Invest Ophthalmol Vis Sci* 1984; 25 (8): 918-31
- Quigley HA, Dorman-Pease ME, Brown AE. Quantitative study of collagen and elastin of the optic nerve head and sclera in human and experimental monkey glaucoma. *Curr Eye Res* 1991; 10 (9): 877-88
- Quigley HA, Brown A, Dorman-Pease ME. Alterations in elastin of the optic nerve head in human and experimental glaucoma. *Br J Ophthalmol* 1991; 75 (9): 552-7
- Agapova OA, Kaufman PL, Lucarelli MJ, et al. Differential expression of matrix metalloproteinases in monkey eyes with experimental glaucoma or optic nerve transection. *Brain Res* 2003; 967 (1-2): 132-43
- Agapova OA, Ricard CS, Salvador-Silva M, et al. Expression of matrix metalloproteinases and tissue inhibitors of metalloproteinases in human optic nerve head astrocytes. *Glia* 2001; 33 (3): 205-16
- Quigley HA. Ganglion cell death in glaucoma: pathology recapitulates ontogeny. *Aust N Z J Ophthalmol* 1995; 23 (2): 85-91
- Quigley HA. Neuronal death in glaucoma. *Prog Retin Eye Res* 1999; 18 (1): 39-57
- Kielczewski JL, Pease ME, Quigley HA. The effect of experimental glaucoma and optic nerve transection on amacrine cells in the rat retina. *Invest Ophthalmol Vis Sci* 2005; 46 (9): 3188-96
- Kerrigan LA, Zack DJ, Quigley HA, et al. TUNEL-positive ganglion cells in human primary open-angle glaucoma. *Arch Ophthalmol* 1997; 115 (8): 1031-5
- Jakobs TC, Libby RT, Ben Y, et al. Retinal ganglion cell degeneration is topological but not cell type specific in DBA/2J mice. *J Cell Biol* 2005; 171 (2): 313-25
- Weber AJ, Kaufman PL, Hubbard WC. Morphology of single ganglion cells in the glaucomatous primate retina. *Invest Ophthalmol Vis Sci* 1998; 39 (12): 2304-20
- Naskar R, Dreyer EB. New horizons in neuroprotection. *Surv Ophthalmol* 2001; 45 Suppl. 3: S250-5
- Nickells RW. Apoptosis of retinal ganglion cells in glaucoma: an update of the molecular pathways involved in cell death. *Surv Ophthalmol* 1999; 43 Suppl. 1: S151-61
- Vorwerk CK, Gorla MS, Dreyer EB. An experimental basis for implicating excitotoxicity in glaucomatous optic neuropathy. *Surv Ophthalmol* 1999; 43 Suppl. 1: S142-50
- Tezel G, Yang J, Wax MB. Heat shock proteins, immunity and glaucoma. *Brain Res Bull* 2004; 62 (6): 473-80

39. Casson RJ. Possible role of excitotoxicity in the pathogenesis of glaucoma. *Clin Experiment Ophthalmol* 2006; 34 (1): 54-63
40. Osborne NN, Wood JPM, Chidlow G, et al. Ganglion cell death in glaucoma: what do we really know? *British J Ophthalmol* 1999; 83: 980-6
41. Osborne NN, Ugarte M, Chao M, et al. Neuroprotection in relation to retinal ischemia and relevance to glaucoma. *Surv Ophthalmol* 1999; 43 Suppl. 1: S102-28
42. Farkas RH, Grosskreutz CL. Apoptosis, neuroprotection, and retinal ganglion cell death: an overview. *Int Ophthalmol Clin* 2001; 41 (1): 111-30
43. Kuehn MH, Fingert JH, Kwon YH. Retinal ganglion cell death in glaucoma: mechanisms and neuroprotective strategies. *Ophthalmol Clin North Am* 2005; 18 (3): 383-95
44. Morrison JC, Johnson EC, Cepurna W, et al. Understanding mechanisms of pressure-induced optic nerve damage. *Prog Retin Eye Res* 2005; 24 (2): 217-40
45. Bathija R. Optic nerve blood flow in glaucoma. *Clin Exp Optom* 2000; 83 (3): 180-4
46. Whitmore AV, Libby RT, John SW. Glaucoma. Thinking in new ways: a role for autonomous axonal self-destruction and other compartmentalised processes? *Prog Retin Eye Res* 2005; 24 (6): 639-62
47. Albon J, Purslow PP, Karwowski WS, et al. Age related compliance of the lamina cribrosa in human eyes. *Br J Ophthalmol* 2000; 84 (3): 318-23
48. Albon J, Karwowski WS, Avery N, et al. Changes in the collagenous matrix of the aging human lamina cribrosa. *Br J Ophthalmol* 1995; 79 (4): 368-75
49. Quigley HA, Addicks EM, Green WR, et al. Optic nerve damage in human glaucoma II: the site of injury and susceptibility to damage. *Arch Ophthalmol* 1981; 99 (4): 635-49
50. Ethier CR. Scleral biomechanics and glaucoma: a connection? *Can J Ophthalmol* 2006; 41: 9-11
51. Morgan JE. Optic nerve head structure in glaucoma: astrocytes as mediators of axonal damage. *Eye* 2000; 14 (Pt 3B): 437-44
52. Yuan L, Neufeld AH. Activated microglia in the human glaucomatous optic nerve head. *J Neurosci Res* 2001; 64 (5): 523-32
53. Tezel G, Hernandez MR, Wax MB. *In vitro* evaluation of reactive astrocyte migration, a component of tissue remodeling in glaucomatous optic nerve head. *Glia* 2001; 34 (3): 178-89
54. Neufeld AH, Liu B. Glaucomatous optic neuropathy: when glia misbehave. *Neuroscientist* 2003; 9 (6): 485-95
55. Nickells RW. Retinal ganglion cell death in glaucoma: the how, the why, and the maybe. *J Glaucoma* 1996; 5 (5): 345-56
56. Quigley HA, McKinnon SJ, Zack DJ, et al. Retrograde axonal transport of BDNF in retinal ganglion cells is blocked by acute IOP elevation in rats. *Invest Ophthalmol Vis Sci* 2000; 41 (11): 3460-6
57. Pease ME, McKinnon SJ, Quigley HA, et al. Obstructed axonal transport of BDNF and its receptor TrkB in experimental glaucoma. *Invest Ophthalmol Vis Sci* 2000; 41 (3): 764-74
58. Garcia-Valenzuela E, Shareef S, Walsh J, et al. Programmed cell death of retinal ganglion cells during experimental glaucoma. *Exp Eye Res* 1995; 61 (1): 33-44
59. Johnson EC, Deppmeier LM, Wentzien SK, et al. Chronology of optic nerve head and retinal responses to elevated intraocular pressure. *Invest Ophthalmol Vis Sci* 2000; 41 (2): 431-42
60. Morrison JC, Moore CG, Deppmeier LM, et al. A rat model of chronic pressure-induced optic nerve damage. *Exp Eye Res* 1997; 64 (1): 85-96
61. Zhang C, Tso MO. Characterization of activated retinal microglia following optic axotomy. *J Neurosci Res* 2003; 73 (6): 840-5
62. Berkelaar M, Clarke DB, Wang YC, et al. Axotomy results in delayed death and apoptosis of retinal ganglion cells in adult rats. *J Neurosci* 1994; 14 (7): 4368-74
63. Ko ML, Hu DN, Ritch R, et al. The combined effect of brain-derived neurotrophic factor and a free radical scavenger in experimental glaucoma. *Invest Ophthalmol Vis Sci* 2000; 41 (10): 2967-71
64. Peinado-Ramon P, Salvador M, Villegas-Perez MP, et al. Effects of axotomy and intraocular administration of NT-4, NT-3, and brain-derived neurotrophic factor on the survival of adult rat retinal ganglion cells: a quantitative *in vivo* study. *Invest Ophthalmol Vis Sci* 1996; 37 (4): 489-500
65. Mansour-Robaey S, Clarke DB, Wang YC, et al. Effects of ocular injury and administration of brain-derived neurotrophic factor on survival and regrowth of axotomized retinal ganglion cells. *Proc Natl Acad Sci U S A* 1994; 91 (5): 1632-6
66. Unoki K, LaVail MM. Protection of the rat retina from ischemic injury by brain-derived neurotrophic factor, ciliary neurotrophic factor, and basic fibroblast growth factor. *Invest Ophthalmol Vis Sci* 1994; 35 (3): 907-15
67. Martin KR, Quigley HA, Zack DJ, et al. Gene therapy with brain-derived neurotrophic factor as a protection: retinal ganglion cells in a rat glaucoma model. *Invest Ophthalmol Vis Sci* 2003; 44 (10): 4357-65
68. Chen H, Weber AJ. BDNF enhances retinal ganglion cell survival in cats with optic nerve damage. *Invest Ophthalmol Vis Sci* 2001; 42 (5): 966-74
69. Klocker N, Kermer P, Weishaupt JH, et al. Brain-derived neurotrophic factor-mediated neuroprotection of adult rat retinal ganglion cells *in vivo* does not exclusively depend on phosphatidylinositol-3'-kinase/protein kinase B signaling. *J Neurosci* 2000; 20 (18): 6962-7
70. Nagahiro S, Uno M, Sato K, et al. Pathophysiology and treatment of cerebral ischemia. *J Med Invest* 1998; 45 (1-4): 57-70
71. Siesjo BK. Pathophysiology and treatment of focal cerebral ischemia. Part II: mechanisms of damage and treatment. *J Neurosurg* 1992; 77 (3): 337-54
72. Siesjo BK. Pathophysiology and treatment of focal cerebral ischemia. Part I: pathophysiology. *J Neurosurg* 1992; 77 (2): 169-84
73. Osborne NN, Casson RJ, Wood JP, et al. Retinal ischemia: mechanisms of damage and potential therapeutic strategies. *Prog Retin Eye Res* 2004; 23 (1): 91-147
74. Henneberry RC, Novelli A, Cox JA, et al. Neurotoxicity at the N-methyl-D-aspartate receptor in energy-compromised neurons: an hypothesis for cell death in aging and disease. *Ann N Y Acad Sci* 1989; 568: 225-33
75. Novelli A, Reilly JA, Lysko PG, et al. Glutamate becomes neurotoxic via the N-methyl-D-aspartate receptor when intracellular energy levels are reduced. *Brain Res* 1988; 451 (1-2): 205-12
76. Kristian T, Siesjo BK. Calcium in ischemic cell death. *Stroke* 1998; 29 (3): 705-18
77. Arundine M, Tymianski M. Molecular mechanisms of calcium-dependent neurodegeneration in excitotoxicity. *Cell Calcium* 2003; 34 (4-5): 325-37
78. Kuroda S, Siesjo BK. Reperfusion damage following focal ischemia: pathophysiology and therapeutic windows. *Clin Neurosci* 1997; 4 (4): 199-212
79. Cheon EW, Park CH, Kang SS, et al. Nitric oxide synthase expression in the transient ischemic rat retina: neuroprotection of betaxolol. *Neurosci Lett* 2002; 330 (3): 265-9
80. Cheon EW, Park CH, Kang SS, et al. Change in endothelial nitric oxide synthase in the rat retina following transient ischemia. *Neuroreport* 2003; 14 (3): 329-33
81. Aslan M, Yucel I, Akar Y, et al. Nitrotyrosine formation and apoptosis in rat models of ocular injury. *Free Radic Res* 2006; 40 (2): 147-53
82. Blute TA, Lee MR, Eldred WD. Direct imaging of NMDA-stimulated nitric oxide production in the retina. *Vis Neurosci* 2000; 17 (4): 557-66

83. Neufeld AH. Nitric oxide: a potential mediator of retinal ganglion cell damage in glaucoma. *Surv Ophthalmol* 1999; 43 Suppl. 1: S129-35
84. Neufeld AH, Hernandez MR, Gonzalez M. Nitric oxide synthase in the human glaucomatous optic nerve head. *Arch Ophthalmol* 1997; 115 (4): 497-503
85. Shareef S, Sawada A, Neufeld AH. Isoforms of nitric oxide synthase in the optic nerves of rat eyes with chronic moderately elevated intraocular pressure. *Invest Ophthalmol Vis Sci* 1999; 40 (12): 2884-91
86. Pang IH, Johnson EC, Jia L, et al. Evaluation of inducible nitric oxide synthase in glaucomatous optic neuropathy and pressure-induced optic nerve damage. *Invest Ophthalmol Vis Sci* 2005; 46 (4): 1313-21
87. Neufeld AH. Pharmacologic neuroprotection with an inhibitor of nitric oxide synthase for the treatment of glaucoma. *Brain Res Bull* 2004; 62 (6): 455-9
88. Liu B, Neufeld AH. Nitric oxide synthase-2 in human optic nerve head astrocytes induced by elevated pressure *in vitro*. *Arch Ophthalmol* 2001; 119 (2): 240-5
89. Liu B, Neufeld AH. Expression of nitric oxide synthase-2 (NOS-2) in reactive astrocytes of the human glaucomatous optic nerve head. *Glia* 2000; 30 (2): 178-86
90. Tezel G, Seigel GM, Wax MB. Autoantibodies to small heat shock proteins in glaucoma. *Invest Ophthalmol Vis Sci* 1998; 39 (12): 2277-87
91. Tezel G, Hernandez R, Wax MB. Immunostaining of heat shock proteins in the retina and optic nerve head of normal and glaucomatous eyes. *Arch Ophthalmol* 2000; 118 (4): 511-8
92. Lu HB, Yuan YS, Li Y, et al. The expression of heat shock protein 27 in retinal ganglion cells in the rat glaucoma model [in Chinese]. *Zhonghua Yan Ke Za Zhi* 2005; 41 (6): 533-9
93. Sakai M, Sakai H, Nakamura Y, et al. Immunolocalization of heat shock proteins in the retina of normal monkey eyes and monkey eyes with laser-induced glaucoma. *Jpn J Ophthalmol* 2003; 47 (1): 42-52
94. Tezel G, Wax MB. Inhibition of caspase activity in retinal cell apoptosis induced by various stimuli *in vitro*. *Invest Ophthalmol Vis Sci* 1999; 40 (11): 2660-7
95. Park KH, Cozier F, Ong OC, et al. Induction of heat shock protein 72 protects retinal ganglion cells in a rat glaucoma model. *Invest Ophthalmol Vis Sci* 2001; 42 (7): 1522-30
96. Weinreb RN, Lindsey JD. The importance of models in glaucoma research. *J Glaucoma* 2005; 14 (4): 302-4
97. Goldblum D, Mittag T. Prospects for relevant glaucoma models with retinal ganglion cell damage in the rodent eye. *Vision Res* 2002; 42 (4): 471-8
98. Morrison JC. Elevated intraocular pressure and optic nerve injury models in the rat. *J Glaucoma* 2005; 14 (4): 315-7
99. Levin LA. Retinal ganglion cells and supporting elements in culture. *J Glaucoma* 2005; 14 (4): 305-7
100. Rasmussen CA, Kaufman PL. Primate glaucoma models. *J Glaucoma* 2005; 14 (4): 311-4
101. Shareef SR, Garcia-Valenzuela E, Salierno A, et al. Chronic ocular hypertension following episcleral venous occlusion in rats. *Exp Eye Res* 1995; 61 (3): 379-82
102. Laquis S, Chaudhary P, Sharma SC. The patterns of retinal ganglion cell death in hypertensive eyes. *Brain Res* 1998; 784 (1-2): 100-4
103. Levkovitch-Verbin H, Quigley HA, Martin KR, et al. Translimbal laser photocoagulation to the trabecular meshwork as a model of glaucoma in rats. *Invest Ophthalmol Vis Sci* 2002; 43 (2): 402-10
104. WoldeMussie E, Ruiz G, Wijono M, et al. Neuroprotection of retinal ganglion cells by brimonidine in rats with laser-induced chronic ocular hypertension. *Invest Ophthalmol Vis Sci* 2001; 42 (12): 2849-55
105. Lindsey JD, Weinreb RN. Elevated intraocular pressure and transgenic applications in the mouse. *J Glaucoma* 2005; 14 (4): 318-20
106. John SW. Mechanistic insights into glaucoma provided by experimental genetics the cogan lecture. *Invest Ophthalmol Vis Sci* 2005; 46 (8): 2649-61
107. Wheeler LA, Gil DW, WoldeMussie E. Role of alpha-2 adrenergic receptors in neuroprotection and glaucoma. *Surv Ophthalmol* 2001; 45 Suppl. 3: S290-4
108. Chaudhary P, Ahmed F, Sharma SC. MK801-a neuroprotectant in rat hypertensive eyes. *Brain Res* 1998; 792 (1): 154-8
109. Guo L, Salt TE, Maass A, et al. Assessment of neuroprotective effects of glutamate modulation on glaucoma-related retinal ganglion cell apoptosis *in vivo*. *Invest Ophthalmol Vis Sci* 2006; 47 (2): 626-33
110. WoldeMussie E, Yoles E, Schwartz M, et al. Neuroprotective effect of memantine in different retinal injury models in rats. *J Glaucoma* 2002; 11 (6): 474-80
111. Schuettauf F, Quinto K, Naskar R, et al. Effects of anti-glaucoma medications on ganglion cell survival: the DBA/2J mouse model. *Vision Res* 2002; 42 (20): 2333-7
112. Hare WA, WoldeMussie E, Lai RK, et al. Efficacy and safety of memantine treatment for reduction of changes associated with experimental glaucoma in monkey, I: functional measures. *Invest Ophthalmol Vis Sci* 2004; 45 (8): 2625-39
113. Hare WA, WoldeMussie E, Weinreb RN, et al. Efficacy and safety of memantine treatment for reduction of changes associated with experimental glaucoma in monkey, II: structural measures. *Invest Ophthalmol Vis Sci* 2004; 45 (8): 2640-51
114. Yucel YH, Gupta N, Zhang Q, et al. Memantine protects neurons from shrinkage in the lateral geniculate nucleus in experimental glaucoma. *Arch Ophthalmol* 2006; 124 (2): 217-25
115. Schori H, Kipnis J, Yoles E, et al. Vaccination for protection of retinal ganglion cells against death from glutamate cytotoxicity and ocular hypertension: implications for glaucoma. *Proc Natl Acad Sci U S A* 2001; 98 (6): 3398-403
116. Bakalash S, Kessler A, Mizrahi T, et al. Antigenic specificity of immunoprotective therapeutic vaccination for glaucoma. *Invest Ophthalmol Vis Sci* 2003; 44 (8): 3374-81
117. Bakalash S, Shlomo GB, Aloni E, et al. T-cell-based vaccination for morphological and functional neuroprotection in a rat model of chronically elevated intraocular pressure. *J Mol Med* 2005; 83 (11): 904-16
118. Ko ML, Hu DN, Ritch R, et al. Patterns of retinal ganglion cell survival after brain-derived neurotrophic factor administration in hypertensive eyes of rats. *Neurosci Lett* 2001; 305 (2): 139-42
119. Ji JZ, Elyaman W, Yip HK, et al. CNTF promotes survival of retinal ganglion cells after induction of ocular hypertension in rats: the possible involvement of STAT3 pathway. *Eur J Neurosci* 2004; 19 (2): 265-72
120. Neufeld AH, Sawada A, Becker B. Inhibition of nitric-oxide synthase 2 by aminoguanidine provides neuroprotection of retinal ganglion cells in a rat model of chronic glaucoma. *Proc Natl Acad Sci U S A* 1999; 96 (17): 9944-8
121. Hains BC, Waxman SG. Neuroprotection by sodium channel blockade with phenytoin in an experimental model of glaucoma. *Invest Ophthalmol Vis Sci* 2005; 46 (11): 4164-9
122. Hirooka K, Tokuda M, Miyamoto O, et al. The *Ginkgo biloba* extract (EGb 761) provides a neuroprotective effect on retinal ganglion cells in a rat model of chronic glaucoma. *Curr Eye Res* 2004; 28 (3): 153-7
123. Ishii Y, Kwong JM, Caprioli J. Retinal ganglion cell protection with geranylgeranylacetone, a heat shock protein inducer, in a rat glaucoma model. *Invest Ophthalmol Vis Sci* 2003; 44 (5): 1982-92
124. Huang W, Fileta JB, Dobberfuhr A, et al. Calcineurin cleavage is triggered by elevated intraocular pressure, and calcineurin

- inhibition blocks retinal ganglion cell death in experimental glaucoma. *Proc Natl Acad Sci U S A* 2005; 102 (34): 12242-7
125. Tahzib NG, Ransom NL, Reitsamer HA, et al. Alpha-fodrin is cleaved by caspase-3 in a chronic ocular hypertensive (COH) rat model of glaucoma. *Brain Res Bull* 2004; 62 (6): 491-5
 126. McKinnon SJ, Lehman DM, Tahzib NG, et al. Baculoviral IAP repeat-containing-4 protects optic nerve axons in a rat glaucoma model. *Mol Ther* 2002; 5 (6): 780-7
 127. Ahmed FA, Hegazy K, Chaudhary P, et al. Neuroprotective effect of alpha(2) agonist (brimonidine) on adult rat retinal ganglion cells after increased intraocular pressure. *Brain Res* 2001; 913 (2): 133-9
 128. Lucas DR, Newhouse JP. The toxic effect of sodium L-glutamate on the inner layers of the retina. *AMA Arch Ophthalmol* 1957; 58 (2): 193-201
 129. Sisk DR, Kuwabara T. Histologic changes in the inner retina of albino rats following intravitreal injection of monosodium L-glutamate. *Graefes Arch Clin Exp Ophthalmol* 1985; 223 (5): 250-8
 130. Siliprandi R, Lipartiti M, Fadda E, et al. Activation of the glutamate metabotropic receptor protects retina against N-methyl-D-aspartate toxicity. *Eur J Pharmacol* 1992; 219 (1): 173-4
 131. Vorwerk CK, Lipton SA, Zurakowski D, et al. Chronic low-dose glutamate is toxic to retinal ganglion cells: toxicity blocked by memantine. *Invest Ophthalmol Vis Sci* 1996; 37 (8): 1618-24
 132. Dreyer EB, Zurakowski D, Schumer RA, et al. Elevated glutamate levels in the vitreous body of humans and monkeys with glaucoma. *Arch Ophthalmol* 1996; 114 (3): 299-305
 133. Brooks DE, Garcia GA, Dreyer EB, et al. Vitreous body glutamate concentration in dogs with glaucoma. *Am J Vet Res* 1997; 58 (8): 864-7
 134. Dkhissi O, Chanut E, Wasowicz M, et al. Retinal TUNEL-positive cells and high glutamate levels in vitreous humor of mutant quail with a glaucoma-like disorder. *Invest Ophthalmol Vis Sci* 1999; 40 (5): 990-5
 135. Honkanen RA, Baruah S, Zimmerman MB, et al. Vitreous amino acid concentrations in patients with glaucoma undergoing vitrectomy. *Arch Ophthalmol* 2003; 121 (2): 183-8
 136. Carter-Dawson L, Crawford ML, Harwerth RS, et al. Vitreal glutamate concentration in monkeys with experimental glaucoma. *Invest Ophthalmol Vis Sci* 2002; 43 (8): 2633-7
 137. Wamsley S, Gabelt BT, Dahl DB, et al. Vitreous glutamate concentration and axon loss in monkeys with experimental glaucoma. *Arch Ophthalmol* 2005; 123 (1): 64-70
 138. Levkovitch-Verbin H, Martin KR, Quigley HA, et al. Measurement of amino acid levels in the vitreous humor of rats after chronic intraocular pressure elevation or optic nerve transection. *J Glaucoma* 2002; 11 (5): 396-405
 139. Martin KR, Levkovitch-Verbin H, Valenta D, et al. Retinal glutamate transporter changes in experimental glaucoma and after optic nerve transection in the rat. *Invest Ophthalmol Vis Sci* 2002; 43 (7): 2236-43
 140. WoldeMussie E, Wijono M, Ruiz G. Muller cell response to laser-induced increase in intraocular pressure in rats. *Glia* 2004; 47 (2): 109-19
 141. Sullivan RK, WoldeMussie E, Macnab L, et al. Evoked expression of the glutamate transporter GLT-1c in retinal ganglion cells in human glaucoma and in a rat model. *Invest Ophthalmol Vis Sci* 2006; 47 (9): 3853-9
 142. Hartwick AT, Zhang X, Chauhan BC, et al. Functional assessment of glutamate clearance mechanisms in a chronic rat glaucoma model using retinal ganglion cell calcium imaging. *J Neurochem* 2005; 94 (3): 794-807
 143. Hamassaki-Britto DE, Hermans-Borgmeyer I, Heinemann S, et al. Expression of glutamate receptor genes in the mammalian retina: the localization of GluR1 through GluR7 mRNAs. *J Neurosci* 1993; 13 (5): 1888-98
 144. Brandstatter JH, Hartveit E, Sassoe-Pognetto M, et al. Expression of NMDA and high-affinity kainate receptor subunit mRNAs in the adult rat retina. *Eur J Neurosci* 1994; 6 (7): 1100-12
 145. Hartveit E, Brandstatter JH, Enz R, et al. Expression of the mRNA of seven metabotropic glutamate receptors (mGluR1 to 7) in the rat retina. An in situ hybridization study on tissue sections and isolated cells. *Eur J Neurosci* 1995; 7 (7): 1472-83
 146. Lagreze WA, Knorle R, Bach M, et al. Memantine is neuroprotective in a rat model of pressure-induced retinal ischemia. *Invest Ophthalmol Vis Sci* 1998; 39 (6): 1063-6
 147. Osborne NN. Memantine reduces alterations to the mammalian retina, in situ, induced by ischemia. *Vis Neurosci* 1999; 16 (1): 45-52
 148. Kim TW, Kim DM, Park KH, et al. Neuroprotective effect of memantine in a rabbit model of optic nerve ischemia. *Korean J Ophthalmol* 2002; 16 (1): 1-7
 149. Naskar R, Quinto K, Romann I, et al. Phenytoin blocks retinal ganglion cell death after partial optic nerve crush. *Exp Eye Res* 2002; 74 (6): 747-52
 150. Vorwerk CK, Zurakowski D, McDermott LM, et al. Effects of axonal injury on ganglion cell survival and glutamate homeostasis. *Brain Res Bull* 2004; 62 (6): 485-90
 151. Areosa SA, Sherriff F, McShane R. Memantine for dementia. *Cochrane Database Syst Rev* 2005; (3): CD003154
 152. Lipton SA. Possible role for memantine in protecting retinal ganglion cells from glaucomatous damage. *Surv Ophthalmol* 2003; 48 Suppl. 1: S38-46
 153. Stys PK. White matter injury mechanisms. *Curr Mol Med* 2004; 4 (2): 113-30
 154. Micu I, Jiang Q, Coderre E, et al. NMDA receptors mediate calcium accumulation in myelin during chemical ischaemia. *Nature* 2006; 439 (7079): 988-92
 155. Schwartz M, Kipnis J. Protective autoimmunity and neuroprotection in inflammatory and noninflammatory neurodegenerative diseases. *J Neurol Sci* 2005; 233 (1-2): 163-6
 156. Schwartz M. Neurodegeneration and neuroprotection in glaucoma: development of a therapeutic neuroprotective vaccine: the Friedenwald lecture. *Invest Ophthalmol Vis Sci* 2003; 44 (4): 1407-11
 157. Schwartz M. Vaccination for glaucoma: dream or reality? *Brain Res Bull* 2004; 62 (6): 481-4
 158. Moalem G, Leibowitz-Amit R, Yoles E, et al. Autoimmune T cells protect neurons from secondary degeneration after central nervous system axotomy. *Nat Med* 1999; 5 (1): 49-55
 159. Moalem G, Yoles E, Leibowitz-Amit R, et al. Autoimmune T cells retard the loss of function in injured rat optic nerves. *J Neuroimmunol* 2000; 106 (1-2): 189-97
 160. Fisher J, Levkovitch-Verbin H, Schori H, et al. Vaccination for neuroprotection in the mouse optic nerve: implications for optic neuropathies. *J Neurosci* 2001; 21 (1): 136-42
 161. Yoles E, Hauben E, Palgi O, et al. Protective autoimmunity is a physiological response to CNS trauma. *J Neurosci* 2001; 21 (11): 3740-8
 162. Kipnis J, Yoles E, Schori H, et al. Neuronal survival after CNS insult is determined by a genetically encoded autoimmune response. *J Neurosci* 2001; 21 (13): 4564-71
 163. Bakalash S, Kipnis J, Yoles E, et al. Resistance of retinal ganglion cells to an increase in intraocular pressure is immune-dependent. *Invest Ophthalmol Vis Sci* 2002; 43 (8): 2648-53
 164. Mizrahi T, Hauben E, Schwartz M. The tissue-specific self-pathogen is the protective self-antigen: the case of uveitis. *J Immunol* 2002; 169 (10): 5971-7

165. Ben Simon GJ, Bakalash S, Aloni E, et al. A rat model for acute rise in intraocular pressure: immune modulation as a therapeutic strategy. *Am J Ophthalmol* 2006; 141 (6): 1105-11
166. Kipnis J, Yoles E, Porat Z, et al. T cell immunity to copolymer 1 confers neuroprotection on the damaged optic nerve: possible therapy for optic neuropathies. *Proc Natl Acad Sci U S A* 2000; 97 (13): 7446-51
167. Blair M, Pease ME, Hammond J, et al. Effect of glatiramer acetate on primary and secondary degeneration of retinal ganglion cells in the rat. *Invest Ophthalmol Vis Sci* 2005; 46 (3): 884-90
168. Simpson D, Noble S, Perry C. Glatiramer acetate: a review of its use in relapsing-remitting multiple sclerosis. *CNS Drugs* 2002; 16 (12): 825-50
169. Anderson DR, Hendrickson A. Effect of intraocular pressure on rapid axoplasmic transport in monkey optic nerve. *Invest Ophthalmol* 1974; 13 (10): 771-83
170. Minckler DS, Bunt AH, Johanson GW. Orthograde and retrograde axoplasmic transport during acute ocular hypertension in the monkey. *Invest Ophthalmol Vis Sci* 1977; 16 (5): 426-41
171. Johansson JO. Retrograde axoplasmic transport in rat optic nerve *in vivo*: what causes blockage at increased intraocular pressure? *Exp Eye Res* 1986; 43 (4): 653-60
172. Johansson JO. Inhibition and recovery of retrograde axoplasmic transport in rat optic nerve during and after elevated IOP *in vivo*. *Exp Eye Res* 1988; 46 (2): 223-7
173. Quigley H, Anderson DR. The dynamics and location of axonal transport blockade by acute intraocular pressure elevation in primate optic nerve. *Invest Ophthalmol* 1976; 15 (8): 606-16
174. Quigley HA, Anderson DR. Distribution of axonal transport blockade by acute intraocular pressure elevation in the primate optic nerve head. *Invest Ophthalmol Vis Sci* 1977; 16 (7): 640-4
175. Quigley HA, Guy J, Anderson DR. Blockade of rapid axonal transport: effect of intraocular pressure elevation in primate optic nerve. *Arch Ophthalmol* 1979; 97 (3): 525-31
176. Quigley HA, Addicks EM. Chronic experimental glaucoma in primates, II: effect of extended intraocular pressure elevation on optic nerve head and axonal transport. *Invest Ophthalmol Vis Sci* 1980; 19 (2): 137-52
177. Martin KR, Quigley HA, Valenta D, et al. Optic nerve dynein motor protein distribution changes with intraocular pressure elevation in a rat model of glaucoma. *Exp Eye Res* 2006; 83 (2): 255-62
178. Hollander H, Makarov F, Stefani FH, et al. Evidence of constriction of optic nerve axons at the lamina cribrosa in the normotensive eye in humans and other mammals. *Ophthalmic Res* 1995; 27 (5): 296-309
179. Barron MJ, Griffiths P, Turnbull DM, et al. The distributions of mitochondria and sodium channels reflect the specific energy requirements and conduction properties of the human optic nerve head. *Br J Ophthalmol* 2004; 88 (2): 286-90
180. Rudzinski M, Wong TP, Saragovi HU. Changes in retinal expression of neurotrophins and neurotrophin receptors induced by ocular hypertension. *J Neurobiol* 2004; 58 (3): 341-54
181. Kido N, Tanihara H, Honjo M, et al. Neuroprotective effects of brain-derived neurotrophic factor in eyes with NMDA-induced neuronal death. *Brain Res* 2000; 884 (1-2): 59-67
182. Klocker N, Cellerino A, Bahr M. Free radical scavenging and inhibition of nitric oxide synthase potentiates the neurotrophic effects of brain-derived neurotrophic factor on axotomized retinal ganglion cells *in vivo*. *J Neurosci* 1998; 18 (3): 1038-46
183. Chaum E. Retinal neuroprotection by growth factors: a mechanistic perspective. *J Cell Biochem* 2003; 88 (1): 57-75
184. Isenmann S, Kretz A, Cellerino A. Molecular determinants of retinal ganglion cell development, survival, and regeneration. *Prog Retin Eye Res* 2003; 22 (4): 483-543
185. Motalebipour M, Rada-Iglesias A, Jansson M, et al. The promoter of inducible nitric oxide synthase implicated in glaucoma based on genetic analysis and nuclear factor binding. *Mol Vis* 2005; 11: 950-7
186. Thornalley PJ. Use of aminoguanidine (Pimagedine) to prevent the formation of advanced glycation endproducts. *Arch Biochem Biophys* 2003; 419 (1): 31-40
187. Nyengaard JR, Chang K, Berhorst S, et al. Discordant effects of guanidines on renal structure and function and on regional vascular dysfunction and collagen changes in diabetic rats. *Diabetes* 1997; 46 (1): 94-106
188. Garthwaite G, Goodwin DA, Batchelor AM, et al. Nitric oxide toxicity in CNS white matter: an *in vitro* study using rat optic nerve. *Neuroscience* 2002; 109 (1): 145-55
189. Zhou W, Goldin AL. Use-dependent potentiation of the Nav1.6 sodium channel. *Biophys J* 2004; 87 (6): 3862-72
190. Stys PK, Waxman SG, Ransom BR. Ionic mechanisms of anoxic injury in mammalian CNS white matter: role of Na⁺ channels and Na⁺/Ca²⁺ exchanger. *J Neurosci* 1992; 12 (2): 430-9
191. Stys PK, Ransom BR, Waxman SG. Tertiary and quaternary local anesthetics protect CNS white matter from anoxic injury at concentrations that do not block excitability. *J Neurophysiol* 1992; 67 (1): 236-40
192. Garthwaite G, Brown G, Batchelor AM, et al. Mechanisms of ischaemic damage to central white matter axons: a quantitative histological analysis using rat optic nerve. *Neuroscience* 1999; 94 (4): 1219-30
193. Lo AC, Black JA, Waxman SG. Neuroprotection of axons with phenytoin in experimental allergic encephalomyelitis. *Neuroreport* 2002; 13 (15): 1909-12
194. Fern R, Ransom BR, Stys PK, et al. Pharmacological protection of CNS white matter during anoxia: actions of phenytoin, carbamazepine and diazepam. *J Pharmacol Exp Ther* 1993; 266 (3): 1549-55
195. Tunnicliff G. Basis of the antiseizure action of phenytoin. *Gen Pharmacol* 1996; 27 (7): 1091-7
196. Goto Y, Taniwaki T, Shigematsu J, et al. The long-term effects of antiepileptic drugs on the visual system in rats: electrophysiological and histopathological studies. *Clin Neurophysiol* 2003; 114 (8): 1395-402
197. Becker B, Stamper RL, Asseff C, et al. Effect of diphenylhydantoin on glaucomatous field loss: a preliminary report. *Trans Am Acad Ophthalmol Otolaryngol* 1972; 76 (2): 412-22
198. Osborne NN, Chidlow G, Wood JP, et al. Expectations in the treatment of retinal diseases: neuroprotection. *Curr Eye Res* 2001; 22 (5): 321-32
199. Santafe J, Martinez de Ibarreta MJ, Segarra J, et al. A long-lasting hypotensive effect of topical diltiazem on the intraocular pressure in conscious rabbits. *Naunyn Schmiedeberg Arch Pharmacol* 1997; 355 (5): 645-50
200. Siegner SW, Netland PA, Schroeder A, et al. Effect of calcium channel blockers alone and in combination with antiglaucoma medications on intraocular pressure in the primate eye. *J Glaucoma* 2000; 9 (4): 334-9
201. Osborne NN, Wood JP, Cupido A, et al. Topical flunarizine reduces IOP and protects the retina against ischemia-excitotoxicity. *Invest Ophthalmol Vis Sci* 2002; 43 (5): 1456-64
202. Campana G, Bucolo C, Murari G, et al. Ocular hypotensive action of topical flunarizine in the rabbit: role of sigma 1 recognition sites. *J Pharmacol Exp Ther* 2002; 303 (3): 1086-94
203. Tamaki Y, Araie M, Fukaya Y, et al. Effects of lomerizine, a calcium channel antagonist, on retinal and optic nerve head circulation in rabbits and humans. *Invest Ophthalmol Vis Sci* 2003; 44 (11): 4864-71
204. Ishii K, Fukaya Y, Araie M, et al. Topical administration of igandipine, a new water-soluble Ca²⁺ antagonist, increases

- ipsilateral optic nerve head circulation in rabbits and cynomolgus monkeys. *Curr Eye Res* 2004; 29 (1): 67-73
205. Orgul S, Zawinka C, Gugleta K, et al. Therapeutic strategies for normal-tension glaucoma. *Ophthalmologica* 2005; 219 (6): 317-23
 206. Veach J. Functional dichotomy: glutathione and vitamin E in homeostasis relevant to primary open-angle glaucoma. *Br J Nutr* 2004; 91 (6): 809-29
 207. Izzotti A, Bagnis A, Sacca SC. The role of oxidative stress in glaucoma. *Mutat Res* 2006; 612 (2): 105-14
 208. Birich TV, Birich TA, Marchenko LN, et al. Lipid peroxidation in the blood of primary glaucoma patients [in Russian]. *Vestn Oftalmol* 1986; 102 (1): 13-5
 209. Ferreira SM, Lerner SF, Brunzini R, et al. Oxidative stress markers in aqueous humor of glaucoma patients. *Am J Ophthalmol* 2004; 137 (1): 62-9
 210. Gherghel D, Griffiths HR, Hilton EJ, et al. Systemic reduction in glutathione levels occurs in patients with primary open-angle glaucoma. *Invest Ophthalmol Vis Sci* 2005; 46 (3): 877-83
 211. Papp E, Nardai G, Soti C, et al. Molecular chaperones, stress proteins and redox homeostasis. *Biofactors* 2003; 17 (1-4): 249-57
 212. Farkas RH, Chowers I, Hackam AS, et al. Increased expression of iron-regulating genes in monkey and human glaucoma. *Invest Ophthalmol Vis Sci* 2004; 45 (5): 1410-7
 213. Moreno MC, Campanelli J, Sande P, et al. Retinal oxidative stress induced by high intraocular pressure. *Free Radic Biol Med* 2004; 37 (6): 803-12
 214. Ko ML, Peng PH, Ma MC, et al. Dynamic changes in reactive oxygen species and antioxidant levels in retinas in experimental glaucoma. *Free Radic Biol Med* 2005; 39 (3): 365-73
 215. Aydemir O, Naziroglu M, Celebi S, et al. Antioxidant effects of alpha-, gamma- and succinate-tocopherols in guinea pig retina during ischemia-reperfusion injury. *Pathophysiology* 2004; 11 (3): 167-71
 216. Dilsiz N, Sahaboglu A, Yildiz MZ, et al. Protective effects of various antioxidants during ischemia-reperfusion in the rat retina. *Graefes Arch Clin Exp Ophthalmol* 2006; 244: 627-33
 217. Birich TV, Birich TA, Marchenko LN, et al. Vitamin E in the complex treatment of patients with primary glaucoma [in Russian]. *Vestn Oftalmol* 1986; 102 (2): 10-3
 218. Cellini M, Caramazza N, Mangiafico P, et al. Fatty acid use in glaucomatous optic neuropathy treatment. *Acta Ophthalmol Scand Suppl* 1998; (227): 41-2
 219. Kang JH, Pasquale LR, Willett W, et al. Antioxidant intake and primary open-angle glaucoma: a prospective study. *Am J Epidemiol* 2003; 158 (4): 337-46
 220. Chung HS, Harris A, Kristinsson JK, et al. *Ginkgo biloba* extract increases ocular blood flow velocity. *J Ocul Pharmacol Ther* 1999; 15 (3): 233-40
 221. Szabo ME, Droy-Lefaix MT, Doly M, et al. Ischemia and reperfusion-induced histologic changes in the rat retina: demonstration of a free radical-mediated mechanism. *Invest Ophthalmol Vis Sci* 1991; 32 (5): 1471-8
 222. Szabo ME, Droy-Lefaix MT, Doly M, et al. Free radical-mediated effects in reperfusion injury: a histologic study with superoxide dismutase and EGB 761 in rat retina. *Ophthalmic Res* 1991; 23 (4): 225-34
 223. Kim SY, Kwak JS, Shin JP, et al. The protection of the retina from ischemic injury by the free radical scavenger EGB 761 and zinc in the cat retina. *Ophthalmologica* 1998; 212 (4): 268-74
 224. Cheung ZH, So KF, Lu Q, et al. Enhanced survival and regeneration of axotomized retinal ganglion cells by a mixture of herbal extracts. *J Neurotrauma* 2002; 19 (3): 369-78
 225. Wang YS, Xu L, Ma K, et al. Protective effects of *Ginkgo biloba* extract 761 against glutamate-induced neurotoxicity in cultured retinal neuron. *Chin Med J (Engl)* 2005; 118 (11): 948-52
 226. Droy-Lefaix MT, Cluzel J, Menerath JM, et al. Antioxidant effect of a *Ginkgo biloba* extract (EGB 761) on the retina. *Int J Tissue React* 1995; 17 (3): 93-100
 227. Droy-Lefaix MT, Bonhomme B, Doly M. Protective effect of *Ginkgo biloba* extract (EGB 761) on free radical-induced changes in the electroretinogram of isolated rat retina. *Drugs Exp Clin Res* 1991; 17 (12): 571-4
 228. Szabo ME, Droy-Lefaix MT, Doly M. Direct measurement of free radicals in ischemic/reperfused diabetic rat retina. *Clin Neurosci* 1997; 4 (5): 240-5
 229. Ahlemeyer B, Kriegstein J. Neuroprotective effects of *Ginkgo biloba* extract. *Cell Mol Life Sci* 2003; 60 (9): 1779-92
 230. Cooper R. Gin(kgo) and tonic: with a twist! *J Altern Complement Med* 2003; 9 (5): 599-601
 231. Quaranta L, Bettelli S, Uva MG, et al. Effect of *Ginkgo biloba* extract on preexisting visual field damage in normal tension glaucoma. *Ophthalmology* 2003; 110 (2): 359-62
 232. Franklin TB, Krueger-Naug AM, Clarke DB, et al. The role of heat shock proteins Hsp70 and Hsp27 in cellular protection of the central nervous system. *Int J Hyperthermia* 2005; 21 (5): 379-92
 233. Richter-Landsberg C, Goldbaum O. Stress proteins in neural cells: functional roles in health and disease. *Cell Mol Life Sci* 2003; 60 (2): 337-49
 234. Salvador-Silva M, Ricard CS, Agapova OA, et al. Expression of small heat shock proteins and intermediate filaments in the human optic nerve head astrocytes exposed to elevated hydrostatic pressure *in vitro*. *J Neurosci Res* 2001; 66 (1): 59-73
 235. Wax MB, Tezel G, Kawase K, et al. Serum autoantibodies to heat shock proteins in glaucoma patients from Japan and the United States. *Ophthalmology* 2001; 108 (2): 296-302
 236. Kretz A, Schmeer C, Tausch S, et al. Simvastatin promotes heat shock protein 27 expression and Akt activation in the rat retina and protects axotomized retinal ganglion cells *in vivo*. *Neurobiol Dis* 2006; 21: 421-30
 237. Hengartner MO. The biochemistry of apoptosis. *Nature* 2000; 407 (6805): 770-6
 238. Quigley HA, Nickells RW, Kerrigan LA, et al. Retinal ganglion cell death in experimental glaucoma and after axotomy occurs by apoptosis. *Invest Ophthalmol Vis Sci* 1995; 36 (5): 774-86
 239. Kikuchi M, Tenneti L, Lipton SA. Role of p38 mitogen-activated protein kinase in axotomy-induced apoptosis of rat retinal ganglion cells. *J Neurosci* 2000; 20 (13): 5037-44
 240. Chaudhary P, Ahmed F, Quebada P, et al. Caspase inhibitors block the retinal ganglion cell death following optic nerve transection. *Brain Res Mol Brain Res* 1999; 67 (1): 36-45
 241. Watanabe M, Fukuda Y. Survival and axonal regeneration of retinal ganglion cells in adult cats. *Prog Retin Eye Res* 2002; 21 (6): 529-53
 242. Levkovitch-Verbin H, Quigley HA, Kerrigan-Baumrind LA, et al. Optic nerve transection in monkeys may result in secondary degeneration of retinal ganglion cells. *Invest Ophthalmol Vis Sci* 2001; 42 (5): 975-82
 243. Zhang Y, Cho CH, Atchaneeyasakul LO, et al. Activation of the mitochondrial apoptotic pathway in a rat model of central retinal artery occlusion. *Invest Ophthalmol Vis Sci* 2005; 46 (6): 2133-9
 244. Lam TT, Abler AS, Kwong JM, et al. N-methyl-D-aspartate (NMDA)-induced apoptosis in rat retina. *Invest Ophthalmol Vis Sci* 1999; 40 (10): 2391-7
 245. Laabich A, Cooper NG. Neuroprotective effect of AIP on N-methyl-D-aspartate-induced cell death in retinal neurons. *Brain Res Mol Brain Res* 2000; 85 (1-2): 32-40

246. Zhang C, Rosenbaum DM, Shaikh AR, et al. Ischemic preconditioning attenuates apoptotic cell death in the rat retina. *Invest Ophthalmol Vis Sci* 2002; 43 (9): 3059-66
247. Katai N, Yoshimura N. Apoptotic retinal neuronal death by ischemia-reperfusion is executed by two distinct caspase family proteases. *Invest Ophthalmol Vis Sci* 1999; 40 (11): 2697-705
248. Guo L, Moss SE, Alexander RA, et al. Retinal ganglion cell apoptosis in glaucoma is related to intraocular pressure and IOP-induced effects on extracellular matrix. *Invest Ophthalmol Vis Sci* 2005; 46 (1): 175-82
249. Wang P, Jiang Y, Huang P, et al. Glutamate in experimental acute elevated intraocular pressure models of rabbits [in Chinese]. *Zhonghua Yan Ke Za Zhi* 2000; 36 (5): 378-80
250. Kim HS, Park CK. Retinal ganglion cell death is delayed by activation of retinal intrinsic cell survival program. *Brain Res* 2005; 1057 (1-2): 17-28
251. Morrison JC, Nylander KB, Lauer AK, et al. Glaucoma drops control intraocular pressure and protect optic nerves in a rat model of glaucoma. *Invest Ophthalmol Vis Sci* 1998; 39 (3): 526-31
252. Schlamp CL, Johnson EC, Li Y, et al. Changes in Thyl gene expression associated with damaged retinal ganglion cells. *Mol Vis* 2001; 7: 192-201
253. Levkovitch-Verbin H, Kaley-Landoy M, Hahot-Wilner Z, et al. Minocycline delays death of retinal ganglion cells in experimental glaucoma and after optic nerve transection. *Arch Ophthalmol* 2006; 124 (4): 520-6
254. Okisaka S, Murakami A, Mizukawa A, et al. Apoptosis in retinal ganglion cell decrease in human glaucomatous eyes. *Jpn J Ophthalmol* 1997; 41 (2): 84-8
255. Tatton NA, Tezel G, Insolia SA, et al. In situ detection of apoptosis in normal pressure glaucoma. a preliminary examination. *Surv Ophthalmol* 2001; 45 Suppl. 3: S268-72
256. Levin LA, Louhab A. Apoptosis of retinal ganglion cells in anterior ischemic optic neuropathy. *Arch Ophthalmol* 1996; 114 (4): 488-91
257. Hanninen VA, Pantcheva MB, Freeman EE, et al. Activation of caspase 9 in a rat model of experimental glaucoma. *Curr Eye Res* 2002; 25 (6): 389-95
258. McKinnon SJ, Lehman DM, Kerrigan-Baumrind LA, et al. Caspase activation and amyloid precursor protein cleavage in rat ocular hypertension. *Invest Ophthalmol Vis Sci* 2002; 43 (4): 1077-87
259. Wang X, Ng YK, Tay SS. Factors contributing to neuronal degeneration in retinas of experimental glaucomatous rats. *J Neurosci Res* 2005; 82 (5): 674-89
260. Huang W, Dobberfuhl A, Filippopoulos T, et al. Transcriptional up-regulation and activation of initiating caspases in experimental glaucoma. *Am J Pathol* 2005; 167 (3): 673-81
261. Libby RT, Li Y, Savinova OV, et al. Susceptibility to neurodegeneration in a glaucoma is modified by Bax gene dosage. *PLoS Genet* 2005; 1 (1): 17-26
262. Ji J, Chang P, Pennesi ME, et al. Effects of elevated intraocular pressure on mouse retinal ganglion cells. *Vision Res* 2005; 45 (2): 169-79
263. Gross RL, Ji J, Chang P, et al. A mouse model of elevated intraocular pressure: retina and optic nerve findings. *Trans Am Ophthalmol Soc* 2003; 101: 163-9; discussion 9-71
264. Levkovitch-Verbin H, Dardik R, Vander S, et al. Experimental glaucoma and optic nerve transection induce simultaneous upregulation of proapoptotic and prosurvival genes. *Invest Ophthalmol Vis Sci* 2006; 47 (6): 2491-7
265. Kanamori A, Nakamura M, Nakanishi Y, et al. Akt is activated via insulin/IGF-1 receptor in rat retina with episcleral vein cauterization. *Brain Res* 2004; 1022 (1-2): 195-204
266. Zhou X, Li F, Kong L, et al. Involvement of inflammation, degradation, and apoptosis in a mouse model of glaucoma. *J Biol Chem* 2005; 280 (35): 31240-8
267. Brookes PS, Yoon Y, Robotham JL, et al. Calcium, ATP, and ROS: a mitochondrial love-hate triangle. *Am J Physiol Cell Physiol* 2004; 287 (4): C817-33
268. Tatton W, Chen D, Chalmers-Redman R, et al. Hypothesis for a common basis for neuroprotection in glaucoma and Alzheimer's disease: anti-apoptosis by alpha-2-adrenergic receptor activation. *Surv Ophthalmol* 2003; 48 Suppl. 1: S25-37
269. Huang X, Wu DY, Chen G, et al. Support of retinal ganglion cell survival and axon regeneration by lithium through a Bcl-2-dependent mechanism. *Invest Ophthalmol Vis Sci* 2003; 44 (1): 347-54
270. Kermer P, Klocker N, Labes M, et al. Inhibition of CPP32-like proteases rescues axotomized retinal ganglion cells from secondary cell death *in vivo*. *J Neurosci* 1998; 18 (12): 4656-62
271. Kermer P, Klocker N, Labes M, et al. Activation of caspase-3 in axotomized rat retinal ganglion cells *in vivo*. *FEBS Lett* 1999; 453 (3): 361-4
272. Vidal-Sanz M, Lafuente M, Sobrado-Calvo P, et al. Death and neuroprotection of retinal ganglion cells after different types of injury. *Neurotox Res* 2000; 2 (2-3): 215-27
273. Kurimoto T, Miyoshi T, Suzuki A, et al. Apoptotic death of beta cells after optic nerve transection in adult cats. *J Neurosci* 2003; 23 (10): 4023-8
274. Chen TA, Yang F, Cole GM, et al. Inhibition of caspase-3-like activity reduces glutamate induced cell death in adult rat retina. *Brain Res* 2001; 904 (1): 177-88
275. Kwong JM, Lam TT. N-methyl-D-aspartate (NMDA) induced apoptosis in adult rabbit retinas. *Exp Eye Res* 2000; 71 (4): 437-44
276. Weishaupt JH, Diem R, Kermer P, et al. Contribution of caspase-8 to apoptosis of axotomized rat retinal ganglion cells *in vivo*. *Neurobiol Dis* 2003; 13 (2): 124-35
277. Siegmund B, Zeitz M, Pralnacasan (vertex pharmaceuticals). *IDrugs* 2003; 6 (2): 154-8
278. Rudolph K, Gerwin N, Verzijl N, et al. Pralnacasan, an inhibitor of interleukin-1beta converting enzyme, reduces joint damage in two murine models of osteoarthritis. *Osteoarthritis Cartilage* 2003; 11 (10): 738-46
279. Linton SD, Karanewsky DS, Ternansky RJ, et al. Acyl dipeptides as reversible caspase inhibitors, part 2: further optimization. *Bioorg Med Chem Lett* 2002; 12 (20): 2973-5
280. Linton SD, Karanewsky DS, Ternansky RJ, et al. Acyl dipeptides as reversible caspase inhibitors, part 1: initial lead optimization. *Bioorg Med Chem Lett* 2002; 12 (20): 2969-71
281. Linton SD, Aja T, Allegrini PR, et al. Oxamyl dipeptide caspase inhibitors developed for the treatment of stroke. *Bioorg Med Chem Lett* 2004; 14 (10): 2685-91
282. Linton SD. Caspase inhibitors: a pharmaceutical industry perspective. *Curr Top Med Chem* 2005; 5 (16): 1697-717
283. Linton SD, Aja T, Armstrong RA, et al. First-in-class pan caspase inhibitor developed for the treatment of liver disease. *J Med Chem* 2005; 48 (22): 6779-82
284. Nickells RW. The molecular biology of retinal ganglion cell death: caveats and controversies. *Brain Res Bull* 2004; 62 (6): 439-46
285. Wheeler LA, Lai R, WoldeMussie E. From the lab to the clinic: activation of an alpha-2 agonist pathway is neuroprotective in models of retinal and optic nerve injury. *Eur J Ophthalmol* 1999; 9 Suppl. 1: S17-21
286. Donello JE, Padillo EU, Webster ML, et al. Alpha(2)-Adrenoceptor agonists inhibit vitreal glutamate and aspartate accumulation and preserve retinal function after transient ischemia. *J Pharmacol Exp Ther* 2001; 296 (1): 216-23

287. Lai RK, Chun T, Hasson D, et al. Alpha-2 adrenoceptor agonist protects retinal function after acute retinal ischemic injury in the rat. *Vis Neurosci* 2002; 19 (2): 175-85
288. Lafuente MP, Villegas-Perez MP, Sobrado-Calvo P, et al. Neuroprotective effects of alpha(2)-selective adrenergic agonists against ischemia-induced retinal ganglion cell death. *Invest Ophthalmol Vis Sci* 2001; 42 (9): 2074-84
289. Lafuente MP, Villegas-Perez MP, Mayor S, et al. Neuroprotective effects of brimonidine against transient ischemia-induced retinal ganglion cell death: a dose response *in vivo* study. *Exp Eye Res* 2002; 74 (2): 181-9
290. Aviles-Trigueros M, Mayor-Torroglosa S, Garcia-Aviles A, et al. Transient ischemia of the retina results in massive degeneration of the retinotectal projection: long-term neuroprotection with brimonidine. *Exp Neurol* 2003; 184 (2): 767-77
291. Mayor-Torroglosa S, De la Villa P, Rodriguez ME, et al. Ischemia results 3 months later in altered ERG, degeneration of inner layers, and deafferented tectum: neuroprotection with brimonidine. *Invest Ophthalmol Vis Sci* 2005; 46 (10): 3825-35
292. Yoles E, Wheeler LA, Schwartz M. Alpha2-adrenoreceptor agonists are neuroprotective in a rat model of optic nerve degeneration. *Invest Ophthalmol Vis Sci* 1999; 40 (1): 65-73
293. Baptiste DC, Hartwick AT, Jollimore CA, et al. Comparison of the neuroprotective effects of adrenoceptor drugs in retinal cell culture and intact retina. *Invest Ophthalmol Vis Sci* 2002; 43 (8): 2666-76
294. Kalapesi FB, Coroneo MT, Hill MA. Human ganglion cells express the alpha-2 adrenergic receptor: relevance to neuroprotection. *Br J Ophthalmol* 2005; 89 (6): 758-63
295. Chao HM, Chidlow G, Melena J, et al. An investigation into the potential mechanisms underlying the neuroprotective effect of clonidine in the retina. *Brain Res* 2000; 877 (1): 47-57
296. Chao HM, Osborne NN. Topically applied clonidine protects the rat retina from ischaemia/reperfusion by stimulating alpha(2)-adrenoceptors and not by an action on imidazoline receptors. *Brain Res* 2001; 904 (1): 126-36
297. Gao H, Qiao X, Cantor LB, et al. Up-regulation of brain-derived neurotrophic factor expression by brimonidine in rat retinal ganglion cells. *Arch Ophthalmol* 2002; 120 (6): 797-803
298. Kent AR, Nussdorf JD, David R, et al. Vitreous concentration of topically applied brimonidine tartrate 0.2%. *Ophthalmology* 2001; 108 (4): 784-7
299. Acheampong AA, Shackleton M, John B, et al. Distribution of brimonidine into anterior and posterior tissues of monkey, rabbit, and rat eyes. *Drug Metab Dispos* 2002; 30 (4): 421-9
300. Evans DW, Hosking SL, Gherghel D, et al. Contrast sensitivity improves after brimonidine therapy in primary open angle glaucoma: a case for neuroprotection. *Br J Ophthalmol* 2003; 87 (12): 1463-5
301. Krupin T, Leibmann JM, Greenfield DS, et al. The Low-pressure Glaucoma Treatment Study (LoGTS) study design and baseline characteristics of enrolled patients. *Ophthalmology* 2005; 112 (3): 376-85
302. Osborne NN, Cazevielle C, Carvalho AL, et al. *In vivo* and *in vitro* experiments show that betaxolol is a retinal neuroprotective agent. *Brain Res* 1997; 751 (1): 113-23
303. Osborne NN, DeSantis L, Bae JH, et al. Topically applied betaxolol attenuates NMDA-induced toxicity to ganglion cells and the effects of ischaemia to the retina. *Exp Eye Res* 1999; 69 (3): 331-42
304. Wood JP, DeSantis L, Chao HM, et al. Topically applied betaxolol attenuates ischaemia-induced effects to the rat retina and stimulates BDNF mRNA. *Exp Eye Res* 2001; 72 (1): 79-86
305. Wood JP, Schmidt KG, Melena J, et al. The beta-adrenoceptor antagonists metipranolol and timolol are retinal neuroprotectants: comparison with betaxolol. *Exp Eye Res* 2003; 76 (4): 505-16
306. Cheon EW, Park CH, Kang SS, et al. Betaxolol attenuates retinal ischemia/reperfusion damage in the rat. *Neuroreport* 2003; 14 (15): 1913-7
307. Mantyh PW, Rogers SD, Allen CJ, et al. Beta 2-adrenergic receptors are expressed by glia *in vivo* in the normal and injured central nervous system in the rat, rabbit, and human. *J Neurosci* 1995; 15 (1 Pt 1): 152-64
308. Osborne NN, Wood JP, Chidlow G, et al. Effectiveness of levobetaxolol and timolol at blunting retinal ischaemia is related to their calcium and sodium blocking activities: relevance to glaucoma. *Brain Res Bull* 2004; 62 (6): 525-8
309. Gross RL, Hensley SH, Gao F, et al. Retinal ganglion cell dysfunction induced by hypoxia and glutamate: potential neuroprotective effects of beta-blockers. *Surv Ophthalmol* 1999; 43 Suppl. 1: S162-70
310. Hirooka K, Kelly ME, Baldrige WH, et al. Suppressive actions of betaxolol on ionic currents in retinal ganglion cells may explain its neuroprotective effects. *Exp Eye Res* 2000; 70 (5): 611-21
311. Melena J, Stanton D, Osborne NN. Comparative effects of antiglaucoma drugs on voltage-dependent calcium channels. *Graefes Arch Clin Exp Ophthalmol* 2001; 239 (7): 522-30
312. Zhang J, Wu SM, Gross RL. Effects of beta-adrenergic blockers on glutamate-induced calcium signals in adult mouse retinal ganglion cells. *Brain Res* 2003; 959 (1): 111-9
313. Chidlow G, Melena J, Osborne NN. Betaxolol, a beta(1)-adrenoceptor antagonist, reduces Na(+) influx into cortical synaptosomes by direct interaction with Na(+) channels: comparison with other beta-adrenoceptor antagonists. *Br J Pharmacol* 2000; 130 (4): 759-66
314. Melena J, Wood JP, Osborne NN. Betaxolol, a beta1-adrenoceptor antagonist, has an affinity for L-type Ca2+ channels. *Eur J Pharmacol* 1999; 378 (3): 317-22
315. Agarwal N, Martin E, Krishnamoorthy RR, et al. Levobetaxolol-induced up-regulation of retinal bFGF and CNTF mRNAs and preservation of retinal function against a photic-induced retinopathy. *Exp Eye Res* 2002; 74 (4): 445-53
316. DeSantis L, Dahlin D, Barnes G. A role for the calcium channel blocking-like action of betaxolol for providing therapeutic benefit to glaucoma patients. In: Drance SM, editor. *Update to glaucoma, ocular blood flow and drug treatment*. Amsterdam: Kugler, 1995: 137-43
317. Messmer C, Flammer J, Stumpf D. Influence of betaxolol and timolol on the visual fields of patients with glaucoma. *Am J Ophthalmol* 1991; 112 (6): 678-81
318. Collignon-Brach J. Long-term effect of ophthalmic beta-adrenoceptor antagonists on intraocular pressure and retinal sensitivity in primary open-angle glaucoma. *Curr Eye Res* 1992; 11 (1): 1-3
319. Collignon-Brach J. Longterm effect of topical beta-blockers on intraocular pressure and visual field sensitivity in ocular hypertension and chronic open-angle glaucoma. *Surv Ophthalmol* 1994; 38 Suppl.: S149-55
320. Kaiser HJ, Flammer J, Stumpf D, et al. Longterm visual field follow-up of glaucoma patients treated with beta-blockers. *Surv Ophthalmol* 1994; 38 Suppl.: S156-9
321. Drance SM. A comparison of the effects of betaxolol, timolol, and pilocarpine on visual function in patients with open-angle glaucoma. *J Glaucoma* 1998; 7 (4): 247-52
322. Vainio-Jylha E, Vuori ML. The favorable effect of topical betaxolol and timolol on glaucomatous visual fields: a 2-year follow-up study. *Graefes Arch Clin Exp Ophthalmol* 1999; 237 (2): 100-4
323. Araie M, Azuma I, Kitazawa Y. Influence of topical betaxolol and timolol on visual field in Japanese open-angle glaucoma patients. *Jpn J Ophthalmol* 2003; 47 (2): 199-207

324. Rainer G, Dorner GT, Garhofer G, et al. Changing antiglaucoma therapy from timolol to betaxolol: effect on ocular blood flow. *Ophthalmologica* 2003; 217 (4): 288-93
325. Miki H, Miki K. The effects on the intraocular pressure and visual field resulting from a switch in the treatment from timolol to betaxolol. *J Ocul Pharmacol Ther* 2004; 20 (6): 509-17
326. Goto W, Ota T, Morikawa N, et al. Protective effects of timolol against the neuronal damage induced by glutamate and ischemia in the rat retina. *Brain Res* 2002; 958 (1): 10-9
327. Kashiwagi K, Iizuka Y, Tsukahara S. Neuroprotective effects of nipradilol on purified cultured retinal ganglion cells. *J Glaucoma* 2002; 11 (3): 231-8
328. Seki M, Tanaka T, Matsuda H, et al. Topically administered timolol and dorzolamide reduce intraocular pressure and protect retinal ganglion cells in a rat experimental glaucoma model. *Br J Ophthalmol* 2005; 89 (4): 504-7
329. Sarup V, McEwan GC, Thompson C, et al. Dorzolamide and timolol saves retinal ganglion cells in glaucomatous adult rats. *J Ocul Pharmacol Ther* 2005; 21 (6): 454-62
330. Diaz F, Villena A, Moreno M, et al. Effects of a non-selective beta-blocker on adult rat anterograde axonal transport and retinal ganglion layer after increased intraocular pressure. *Histol Histopathol* 2005; 20 (4): 1077-84
331. Nakazawa T, Tomita H, Yamaguchi K, et al. Neuroprotective effect of nipradilol on axotomized rat retinal ganglion cells. *Curr Eye Res* 2002; 24 (2): 114-22
332. Melena J, Osborne NN. Metipranolol attenuates lipid peroxidation in rat brain: a comparative study with other antiglaucoma drugs. *Graefes Arch Clin Exp Ophthalmol* 2003; 241 (10): 827-33
333. Tan AY, LeVatte TL, Archibald ML, et al. Timolol concentrations in rat ocular tissues and plasma after topical and intraperitoneal dosing. *J Glaucoma* 2002; 11 (2): 134-42
334. Osborne NN, Wood JP. Metipranolol blunts nitric oxide-induced lipid peroxidation and death of retinal photoreceptors: a comparison with other anti-glaucoma drugs. *Invest Ophthalmol Vis Sci* 2004; 45 (10): 3787-95
335. Osborne NN, Wood JP. The beta-adrenergic receptor antagonist metipranolol blunts zinc-induced photoreceptor and RPE apoptosis. *Invest Ophthalmol Vis Sci* 2006; 47 (7): 3178-86
336. Kudo H, Nakazawa T, Shimura M, et al. Neuroprotective effect of latanoprost on rat retinal ganglion cells. *Graefes Arch Clin Exp Ophthalmol* 2006; 244 (8): 1003-9
337. Drago F, Valzelli S, Emmi I, et al. Latanoprost exerts neuroprotective activity *in vitro* and *in vivo*. *Exp Eye Res* 2001; 72 (4): 479-86
338. Melamed S. Neuroprotective properties of a synthetic docosanoid, unoprostone isopropyl: clinical benefits in the treatment of glaucoma. *Drugs Exp Clin Res* 2002; 28 (2-3): 63-73
339. Fuchsjager-Mayrl G, Wally B, Rainer G, et al. Effect of dorzolamide and timolol on ocular blood flow in patients with primary open angle glaucoma and ocular hypertension. *Br J Ophthalmol* 2005; 89 (10): 1293-7

Correspondence: Dr *Glyn Chidlow*, Ophthalmic Research Laboratories, South Australian Institute of Ophthalmology, Institute of Medical and Veterinary Services, Adelaide, SA 5000, Australia.
E-mail: glyn.chidlow@imvs.sa.gov.au