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Community-Associated Meticillin-Resistant Staphylococcus aureus Infections

Epidemiology, Recognition and Management

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Abstract

Meticillin-resistant *Staphylococcus aureus* (MRSA) is an important cause of infection, particularly in hospitalized patients and those with significant healthcare exposure. In recent years, epidemic community-associated MRSA

(CA-MRSA) infections occurring in patients without healthcare risk factors have become more frequent. The most common manifestation of CA-MRSA infection is skin and soft tissue infection, although necrotizing pneumonia, sepsis and osteoarticular infections can occur. CA-MRSA strains have become endemic in many communities and are genetically distinct from previously identified MRSA strains. CA-MRSA may be more capable colonizers of humans and more virulent than other S. aureus strains. Specific mechanisms of pathogenicity have not been elucidated, but several factors have been proposed as responsible for the virulence of CA-MRSA, including the Panton-Valentine leukocidin, phenol-soluble modulins and type I arginine catabolic mobile element. The movement of CA-MRSA strains into the nosocomial setting limits the utility of using clinical risk factors alone to designate community- or healthcare-associated status. Identification of unique genetic characteristics and genotyping are valuable tools for MRSA epidemiological studies. Although the optimum pharmacological therapy for CA-MRSA infections has not been determined, many CA-MRSA strains remain broadly susceptible to several non-β-lactam antibacterial agents. Empirical antibacterial therapy should include an MRSA-active agent, particularly in areas where CA-MRSA is endemic.

The widespread emergence of meticillinresistant *Staphylococcus aureus* (MRSA) in recent years highlights the impact of resistant bacterial infections on healthcare and society. MRSA has particularly been an important cause of infection in critically ill patients and in some settings is more common than meticillinsusceptible *S. aureus* (MSSA).^[1] *S. aureus* infections are estimated to have a significant cost to healthcare,^[2] and MRSA additionally has been an important cause of both morbidity and mortality in the US.^[3]

In addition to its role in nosocomial infections, MRSA has become a common cause of infections in healthy patient populations that lack traditional risk factors for MRSA exposure. The MRSA strains responsible for these infections have been found to be epidemiologically and genetically unique from nosocomial MRSA strains and are now designated as community-associated MRSA (CA-MRSA). CA-MRSA may be both more capable colonizers of humans and more virulent than other *S. aureus* strains as evidenced by the rapidity with which it has become endemic in many parts of the world. Indeed, CA-MRSA is the most frequent cause of cutaneous infections in some communities, as well as an important

cause of invasive infections such as pneumonia and sepsis. CA-MRSA strains continue to spread into new niches and are now common in some hospitals and responsible for causing nosocomial infections such as bacteraemia and surgical site infections.

This article discusses the epidemiology, unique genetic characteristics, virulence and pathogenesis of CA-MRSA. Pharmacological and nonpharmacological therapies for CA-MRSA are reviewed. The term 'community-associated' is used in this discussion, but is synonymous with the terms 'community-acquired' and 'communityonset', both of which are also present in the literature. Published literature from 1950 through to December 2008 was reviewed via the PubMed search engine using the primary search terms 'methicillin-resistant staphylococcus aureus', 'community-associated methicillin-resistant staphylococcus aureus', 'CA-MRSA' and 'HA-MRSA'. Individual antibacterials were also searched via PubMed and additional articles were obtained from the reference sections of source articles identified via PubMed. Finally, information on CA-MRSA management was also obtained from the US Centers for Disease Control and Prevention website (http://www.cdc.gov).

1. Epidemiology of Community-Associated Meticillin-Resistant Staphylococcus aureus (CA-MRSA)

MRSA was first identified in the early 1960s,^[4] became an increasingly common hospital pathogen in the 1980s and is now a major pathogen in most hospitals, particularly in the critical-care setting. In contrast to penicillin resistance, which is common in both nosocomial and community strains of S. aureus, MRSA was considered to be an exclusively nosocomial pathogen. Patients who developed MRSA infections typically had the following risk factors: (i) surgery, dialysis, hospitalization or residence in a long-term care facility within the prior year; (ii) indwelling percutaneous devices such as central venous catheters or feeding tubes; (iii) an MRSA infection identified more than 48 hours after hospital admission; or (iv) had previously had MRSA cultured. [5] MRSA cultured from patients with any of these risk factors are now clinically designated as healthcare-associated MRSA (HA-MRSA) to distinguish it from CA-MRSA, which is cultured from patients without any of these clinical risk factors. Before the widespread appearance of CA-MRSA strains, outbreaks of MRSA infections occurring in patients lacking the traditional risk factors were occasionally reported. [6,7] Phenotypically, HA-MRSA is often resistant to several antibacterial classes in addition to the β -lactam agents.

Early reports of CA-MRSA came from Western Australia. [8] Twenty-five MRSA isolates cultured between 1989 and 1991 were collected, with 18 representing infections and the remainder representing colonization. None of the patients had been hospitalized outside of the region in the previous 12 months. Of note, MRSA had not become established in the local hospitals and thus acquisition of MRSA in these patients was not felt to be nosocomial. Molecular genotyping revealed that the majority of these MRSA isolates were identical and were noted to be susceptible to chloramphenicol, gentamicin and trimethoprim, antibacterials to which nosocomial MRSA strains in Australia typically exhibited resistance.

By the late 1990s, reports of MRSA infections occurring in children in North America were

described in a Children's Hospital in Chicago, [9] as well as four cases of rapidly fatal infection in four children from the Midwestern US.[10] Subsequently, outbreaks were noted among incarcerated adults, [11-13] Alaska natives, [14] athletes [15-17] and military recruits.[18] Common to many of these outbreaks was the frequent presentation of patients with a soft-tissue infection, absence of traditional MRSA risk factors, genetic relatedness of the infecting MRSA strains and susceptibility of the bacteria to many non-β-lactam antibacterials. Given the epidemiological differences from nosocomial strains of MRSA, a distinction was made between CA- and HA-MRSA. Following recognition of epidemiologically related outbreaks of CA-MRSA infection, nonoutbreak infections have been reported from North America, Central and South America, Europe, Australia and Asia.[19-30]

1.1 Colonization

The prevalence of nasal colonization with MRSA appears to be increasing in the US. Based on data collected as part of the National Health and Nutrition Examination Survey, overall MRSA nasal colonization has increased from 0.9% (95% CI 0.5, 1.4) in 2001–2 to 1.5% (95% CI 1.2, 1.8) in 2002–4. [31,32] The proportion of MRSA strains identified genetically as CA-MRSA also increased from 8.1% (95% CI 1.1, 25.3) to 19.7% (95% CI 12.4, 28.8). It is possible these data may underestimate MRSA colonization, as other anatomical sites may also be important sites of colonization exclusive of the nares. [33-35]

Asymptomatic colonization with MRSA is a risk factor for subsequent infection with the colonizing strain. The risk of developing a subsequent infection may be increased if colonized with CA-MRSA versus MSSA. An observational study of 812 military recruits noted nasal colonization with CA-MRSA in 3% compared with 28% colonized with MSSA, but 38% of the CA-MRSA-colonized individuals developed softtissue infections compared with 3% of MSSA colonized individuals (relative risk 10.7; 95% CI 4.6, 25.2; p<0.01). Whether CA-MRSA strains are more likely to colonize and cause

subsequent infection than HA-MRSA or MSSA strains remains unanswered, although the endemicity and volume of CA-MRSA infections seem to suggest these strains may be more 'fit' and virulent than other *S. aureus* strains.

1.2 High-Risk Populations for CA-MRSA Infection

Various populations have been suggested as being at increased risk for developing CA-MRSA infections based on published reports of outbreaks in specific populations (table I). However, as CA-MRSA strains have become endemic in many parts of the world, it appears that almost any person can develop CA-MRSA infections. Nonetheless, certain populations may be at increased risk for infection because of inherent host factors or behaviours that enhance the transmission of CA-MRSA or the development of infection. Transmission of CA-MRSA between individuals is likely to be facilitated by crowded living conditions,[11-13,18] activities that involve skin-to-skin contact.[15-17] poor hygiene practices^[40] and sharing of contaminated household items.[16] In addition, men who have sex with men.[41] some indigenous populations[14,42] and individuals who have recently received antibacterials^[43] have been noted to have a higher incidence of CA-MRSA infection. Infection is more likely to occur if host defences against bacterial infections are diminished: shaving or skin abrasions, immature/compromised immune systems and loss of protective respiratory tract epithelium (as can occur following influenza infection). In communities where CA-MRSA is not yet endemic, outbreaks of infection in these previously recognized at-risk populations might herald future endemicity.

1.3 CA-MRSA Strains are Distinct from Healthcare-Associated MRSA

In addition to the epidemiological differences, CA- and HA-MRSA strains have been found to be both phenotypically and genotypically distinct (table II). Molecular typing methods have identified a small number of S. aureus strains that comprise the majority of CA-MRSA. Pulsedfield gel electrophoresis (PFGE), a commonly used method to determine bacterial strain relatedness, is an important epidemiological tool. A classification scheme categorizing isolates as USA100-USA1200 is used to describe MRSA by PFGE analysis. The most common CA-MRSA genotypes include USA300 or USA400,[44] but also include USA1000 and USA1100.[45] Common HA-MRSA genotypes include USA100, USA200 and USA500. [44] The Midwest strain MW2 that caused the epidemic infections in children^[10] is classified as USA400, and the pandemic clone of MRSA found throughout the US and Europe is classified as USA300. One particular strain of MRSA USA300, termed USA300-0114, is especially common in many outbreaks and surveillance studies.

Multi-locus sequence typing analysis is also useful for describing clones of CA-MRSA. Common CA-MRSA sequence types include ST1, which includes MW2, and the early Australian strains,^[8,10] ST30, ST59, ST80 and ST8, which includes the pandemic USA300 strain.^[44]

As noted in early reports of CA-MRSA outbreaks, the isolates exhibit susceptibility to many

Table I. Groups at risk for community-associated meticillin-resistant Staphylococcus aureus infection and associated risk factors

Population or group	Possible risk factors	References
Inmates/prisoners	Crowded living conditions, close contact	11-13
Men who have sex with men	Close contact	41
Athletes	Close contact, skin abrasions, sharing of equipment	15-17
Military recruits	Crowded living conditions, skin abrasions	18
Native Americans, Alaska natives	Close contact, crowded living conditions	14,42
Children	Close contact, skin abrasions	9,10
Methamphetamine users	Poor skin hygiene	40

Table II. Overview of general characteristics of community- and healthcare-associated meticillin-resistant *Staphylococcus aureus* (CA-MRSA and HA-MRSA)^[5,14,21,46-57]

Characteristic	CA-MRSA	HA-MRSA Nosocomial bacteraemia, pneumonia, wound infections	
Common manifestations	SSTI, necrotizing pneumonia		
Antibacterial susceptibility	Frequently susceptible to non- β -lactam antibacterials, low prevalence of iMLS $_{B}$ resistance	Broad resistance to non- β -lactam antibacterials, iMLS $_B$ resistance common	
SCCmec type	IV, V	I, II, III	
Accessory gene regulator type	agr III	agr I, II	
Genotype (PFGE)	USA300, USA400, USA1000 USA1100	USA100, USA200, USA500, USA600, USA800	
Sequence type (MLST)	ST1, ST8, ST30, ST59, ST80	ST5, ST36, ST45	
Virulence genes/factors	pvl, sea, seb, sec, seh and type I ACME common; higher expression of PSM; more rapid in vitro growth	pvl uncommon, type I ACME absent	

ACME = arginine catabolic mobile element; **iMLS**_B = inducible macrolide-lincosamide-streptogramin B; **MLST** = multi-locus sequence typing; **PFGE** = pulsed-field gel electrophoresis; **PSM** = α -type phenol-soluble modulins; **SCC** = staphylococcal cassette chromosome; **SSTI** = skin and soft tissue infection.

non-β-lactam antibacterials. In a prospective survey of MRSA in Minnesota, USA, CA-MRSA strains were significantly more susceptible than HA-MRSA strains to several antibacterials: ciprofloxacin (79% vs 16%), clindamycin (83% vs 21%), erythromycin (44% vs 9%) and gentamicin (94% vs 80%).^[5] In addition, CA-MRSA was more likely to be susceptible to the combination of ciprofloxacin, clindamycin, gentamicin and cotrimoxazole (trimethoprim/sulfamethoxazole) [odds ratio 5.88; 95% CI 4.86, 6.64]. Multidrugresistant MRSA USA300 (resistance to tetracycline, macrolides, lincosamides, streptogramin B and mupirocin) has been described in clinical isolates from San Francisco, CA, and Boston, MA, USA.[41,58,59] Multidrug resistance in many of these isolates is conferred by a conjugative plasmid, pUSA03, which has the potential to gain additional antibacterial resistance genes.^[58]

CA-MRSA strains have been noted to exhibit more rapid growth than HA-MRSA strains, with CA-MRSA mean doubling time of 28.79 minutes compared with 38.81 minutes for HA-MRSA strains (p<0.0001). [60] This more rapid growth has been hypothesized to allow CA-MRSA to outcompete slower growing strains of *S. aureus*.

Genetic differences between CA- and HA-MRSA strains are numerous. Meticillin resistance is conferred by the *mec*A gene in staphylococci and is carried by the staphylococcal cassette chromosome (SCC).^[61] SCC is a mobile genetic element

that appears to allow the transfer of its associated genes, in this case the mecA gene, between staphylococci. Three SCCmecA types, I-III, had previously been described in MRSA.[62] SCCmec types I-III have several common features, including identical chromosomal integration site, conserved genetic organization around the mecA gene and the presence of cassette chromosome recombinase (ccr) genes, which allow horizontal transfer of SCCmec. Additional antibacterial resistance genes are common in SCCmec types II and III. SCCmec type I-III range in size from 34.4 to 66.9 kb. CA-MRSA strains contain the smaller SCCmec type IV. [63,64] or less commonly, SCCmec type V (20.9-24.3 kb and 28 kb, respectively). [65] SCCmec types I, IV and V do not typically carry additional antibacterial resistance genes with the exception of mecA.[60] Numerous variants of SCCmec have been identified in both CA- and HA-MRSA.[66]

1.4 Virulence of CA-MRSA

Virulence genes and toxins are present in CA-MRSA that are not common in other strains of MRSA. Baba and colleagues^[46] found 18 unique toxins, including the superantigen staphylococcal enterotoxin H (*seh*), in the MW2 strain. Clinical MRSA isolates from Minnesota revealed that several virulence genes were significantly more common in CA-MRSA than HA-MRSA:

Panton-Valentine leukocidin (PVL) gene, and genes for staphylococcal enterotoxins A, B, C and H.^[5] In addition, CA-MRSA strains are commonly accessory gene regulator (*agr*) type III, a regulatory operon that coordinates various virulence pathways whereas HA-MRSA are more likely to be *agr* I or II.^[5]

Numerous genetic differences have been proposed as a potential explanation for the perceived enhanced virulence of CA-MRSA. The precise mechanisms of pathogenicity in CA-MRSA have not been determined, and debate continues on the role of specific genes and toxins. Initial attention focused on the role of the PVL toxin. The PVL gene is uncommon in traditional nosocomial *S. aureus* strains, ^[5,47] thus it seemed reasonable to consider this as likely to be an important virulence determinant. PVL is a bicomponent toxin produced by the *lukPV* operon that results in pore formation in leukocytes. The PVL gene is common in many clinical CA-MRSA strains and uncommon in HA-MRSA. ^[5,14,21,46,48,49]

Animal studies have yielded conflicting results as to the importance of pvl. Deletion of the lukPV operon in MW2 and USA300 in a murine abscess model has not been shown to reduce abscess size or bacterial density when compared with the parent MRSA strain. Similar knockout experiments in a mouse bacteraemia model also did not show differences in lethality.[50] However, in a rabbit bacteraemia model, PVL may have a modest effect in the acute phase of infection that wanes as the disease progresses and does not appear to have an effect on bacterial gene regulatory networks.^[51] In a murine pneumonia model. PVL has been found both to be an important contributor to CA-MRSA virulence in one experiment^[52] and non-contributory in other experiments.[53,54]

Another recently identified family of toxins in *S. aureus* that is differentially expressed in CA-and HA-MRSA strains are α-type phenol-soluble modulins (PSMs). PSMs have leukocidal activity and are expressed at higher levels in CA-MRSA. CA-MRSA strains that express PSMs have been shown to be more virulent than non-PSM MRSA strains in bacteraemia and abscess mouse models.^[55]

Genome sequencing of the pandemic clone USA300 revealed it has acquired the type I arginine catabolic mobile element (ACME), a pathogenicity island that may enhance bacterial fitness and survival, from *S. epidermidis*. [56,57] Type I ACME appears to be unique to the USA300 pandemic clone and is uncommon in other MRSA. Type I ACME is physically linked to SCC*mec* using the same *ccr* recombinases for mobilization and transfer. [57] Thus, the combination of antibacterial resistance conferred by SCC*mec* and the virulence of type I ACME may provide the survival advantage that has allowed USA300 to become a dominant clone of MRSA.

2. Manifestations of CA-MRSA Disease

2.1 Skin and Soft Tissue Infection

The spectrum of disease caused by CA-MRSA appears to be similar to that of other *S. aureus*, although certain manifestations are particularly common. Early reports of CA-MRSA infections described severe disease including sepsis, osteomyelitis and pneumonia. Subsequent reports established that the most common manifestation of CA-MRSA is skin and soft tissue infection (SSTI), particularly abscess or furunculosis. Many affected patients often report a history of a 'spider bite' that they believe subsequently became infected, a description that is usually describing the natural history of CA-MRSA SSTI and not an infected arthropod bite.

A surveillance study of three different communities in the US found that SSTI was the most common manifestation of CA-MRSA disease (77%), followed by wound infection (10%), urinary tract infection (4%), sinus infection (4%) and pneumonia (2%).^[25] Among patients presenting to an urban medical centre with community-onset SSTI due to *S. aureus*, CA-MRSA caused 63% (244 of 389 episodes) of infections compared with 28% (110 of 389 episodes) due to MSSA.^[67] Similarly, in a multicentre study, *S. aureus* was found to be the most common cause of SSTI among patients presenting to the emergency department (76%).^[27] The prevalence of MRSA was 59% overall and pulsed-field type USA300

CA-MRSA accounted for 97% of all MRSA. Patients with CA-MRSA SSTIs often experience recurrent infections.^[68,69]

Serious CA-MRSA soft-tissue infections have been noted to occur. In a retrospective review of MRSA wound cultures at an urban hospital in Los Angeles, CA, USA, 14 of 843 patients from the community were found to have necrotizing fasciitis, necrotizing myositis or both.^[70] All of the MRSA isolates were susceptible to clindamycin, cotrimoxazole and rifampin. Molecular typing of the isolates found all were USA300, SCCmec type IV and PVL gene-positive. All of the patients described survived, suggesting that CA-MRSA may be less virulent than other organisms that cause fasciitis. Additional reports of fasciitis and myositis have since been reported, although both conditions appear to be relatively uncommon manifestations of CA-MRSA disease. [71-75] Fournier's gangrene due to CA-MRSA (PVL gene-positive) has also been reported.^[76]

2.2 Pulmonary Infection

S. aureus is an important cause of nosocomial pneumonia and less commonly a cause of community-acquired pneumonia (CAP). CA-MRSA strains, particularly PVL gene-positive strains, are increasingly reported to cause CAP in both adults and children.[77-79] Francis and colleagues^[78] reported four patients who developed necrotizing pneumonia due to PVL gene-positive, USA300 CA-MRSA. The patients ranged in age from 20 to 52 years and presented with cavitary lung lesions and shock. Two patients had laboratory-confirmed influenza infection and two patients had an influenza-like prodrome without confirmatory influenza testing. One patient died, the remaining three had prolonged hospitalizations with complications including shock, cavitary pneumonia, empyema, pneumothoraces and gangrene. Gonzalez and colleagues^[79] described 47 children with CA-MRSA infection and abnormal pulmonary imaging. Common manifestations of pulmonary infection included pneumonia, empyema and necrotizing pneumonia. Co-infection with influenza or parainfluenza virus was noted in three patients who developed

severe necrotizing pneumonia. Metastatic pulmonary disease was present in 20 patients with osteomyelitis. Additional cases of CA-MRSA CAP associated with influenza have been reported, particularly in young, healthy patients and often presenting with severe disease. [80,81]

With the success of pneumococcal vaccines in reducing *Streptococcus pneumoniae* pneumonia, CA-MRSA is an important consideration as a cause of severe CAP, especially in the setting of concomitant influenza infection or as a metastatic complication of existing CA-MRSA infection. Empirical therapy for severe CAP should include antibacterials active against CA-MRSA.

2.3 Osteoarticular Infections

Acute, haematogenous CA-MRSA osteoarticular infections are more frequently reported in children^[79,82-85] than in adults.^[86,87] Osteoarticular infection may be complicated by subperiosteal abscess, myositis and bursitis. Metastatic infection including pneumonia and bacteraemia are frequently associated with osteoarticular infection. Interestingly, several reports have noted an association between CA-MRSA osteomyelitis and development of venous thrombosis.^[84,88]

2.4 Other Infections

A variety of other manifestations of CA-MRSA infection have been reported. Infective endocarditis due to SCC*mec* type IV^[89] and USA300 genotype MRSA has been described.^[90] Sepsis can complicate virtually any manifestation of CA-MRSA disease, particularly pneumonia and osteomyelitis.^[10,78,91] Sepsis presenting with purpura fulminans^[92] and associated with Waterhouse-Friderichsen syndrome^[93] have also been reported. Uncommon manifestations of CA-MRSA infection include meningitis,^[94] cavernous sinus thrombosis,^[95] puerperal mastitis^[96,97] and ophthalmic infections.^[98]

3. Defining CA-MRSA

The presence or absence of clinical risk factors for MRSA has been the basis of clinical definitions for CA- and HA-MRSA, and broad non-β-lactam

antibacterial susceptibility is generally believed to be consistent with CA-MRSA. Initially, CA-MRSA strains were almost exclusively found in the outpatient setting. More recently, CA-MRSA strains have been increasingly isolated from hospitalized patients^[96,99-105] and have caused infections that are generally considered to be healthcare related such as prosthetic joint infections^[106] and surgical site infections.^[107] Several studies suggest that community strains of MRSA are replacing or adding to the traditional nosocomial strains of MRSA in many populations.^[108-110]

Defining CA- and HA-MRSA on clinical risk factors alone is becoming less useful as these strains intermingle within the healthcare setting. Antibacterial susceptibility phenotypes may still be useful to distinguish between some CA- and HA-MRSA strains, although increasing antibacterial resistance has been noted in some CA-MRSA isolates.^[41] Epidemiological investigations may require molecular genotyping methods and identification of various marker genes (e.g. SCCmec type, virulence/toxin genes) in addition to clinical data to fully characterize MRSA isolates. Strict definitions for CA- and HA-MRSA should be used whenever possible, and could include molecular genotype assignment, SCCmec type, and the presence or absence of various genes such as PVL, type I ACME and agr type.

4. Treatment of CA-MRSA Infections

Optimum management of CA-MRSA infections has not been determined. Current strategies include a combination of pharmacological and non-pharmacological interventions. In patients with recurrent infections, attempts to decolonize CA-MRSA are also frequently attempted, although the long-term benefit of this strategy remains unclear.

Many CA-MRSA are susceptible to several non-β-lactam antibacterial classes, providing clinicians with several outpatient options (table III). Frequently, older antibacterials such as clindamycin, doxycycline and cotrimoxazole are prescribed for the treatment of CA-MRSA infections, despite a lack of specific MRSA indications for these drugs. For more serious infections,

vancomycin remains the standard of care for most patients who require intravenous therapy. Other alternatives include daptomycin, tigecycline and quinupristin/dalfopristin. In addition, linezolid is available in both oral and intravenous formulations. Ceftobiprole is currently under review for approval and may be another intravenous option in the near future. It is unclear if the newer antibacterials necessarily represent an efficacy advantage over older options and each has unique limitations to consider.

4.1 Tetracyclines

Tetracyclines, particularly doxycycline and minocycline, have been used for many years clinically as treatment of S. aureus infections, especially SSTIs. At present, there have been no well designed clinical trial data published describing the efficacy of tetracyclines for the treatment of MRSA infections. One recent case series describes 24 patients with serious tetracyclinesusceptible MRSA infections who were treated with either doxycycline or minocycline.[120] The types of infection in this series were complicated SSTI (16 patients), osteomyelitis or septic arthritis (five patients), urinary tract infection (two patients) and bacteraemia (one patient). Thirteen (54%) patients received doxycycline and clinical cure was achieved in 83% of patients. The authors reviewed published literature describing patients with S. aureus (both MSSA and MRSA) infections treated with tetracyclines and found 85 patients from nine studies. The overall cure rate in this group was 85%. Tetracyclines remain an option for the treatment of uncomplicated CA-MRSA SSTIs, but are generally not recommended for the treatment of invasive infections.

4.2 Clindamycin

The emergence of CA-MRSA has renewed the empirical use of clindamycin, particularly for SSTIs. Clindamycin is a lincosamide that inhibits protein synthesis by binding to the bacterial 50S ribosomal subunit and has activity against several clinically important Gram-positive bacteria, including staphylococci, streptococci and anaerobes.

Table III. Antibacterials for the treatment of community-associated meticillin-resistant Staphylococcus aureus infections

Agent	Usual dosages	Adverse effects	Comments
Ceftobiprole ^[111]	500 mg IV q8h	Nausea, diarrhoea, vomiting	Not yet widely available
Clindamycin ^[112]	300–600 mg PO q6h 300–900 mg IV q8h	Nausea, diarrhoea, <i>Clostridium</i> difficile-associated disease	iMLS _B resistance may reduce activity
Daptomycin ^[113]	4-6 mg/kg IV q12h	Rhabdomyolysis, myopathy	6 mg/kg IV indicated for bacteraemia; do not use to treat pneumonia
Doxycycline ^[114]	100 mg PO or IV q12h	Nausea, hypersensitivity to sunlight	Contraindicated in pregnancy, children aged <8 y; activity against GAS is unknown
Linezolid ^[115]	600 mg PO or IV q12h	Nausea, pancytopenia if used >2 wk, optic neuritis, lactic acidosis, potential for serotonin syndrome	Not recommended for the treatment of bacteraemia
Quinupristin/dalfopristin ^[116]	7.5 mg/kg IV q12h	Myalgias, arthralgias, infusion site inflammation	iMLS _B resistance may reduce activity
Tigecycline ^[117]	100 mg IV once, then 50 mg IV q12h	Nausea, vomiting	Treatment of bacteraemia not recommended because of low serum concentration
Cotrimoxazole ^[118] (trimethoprim/ sulfamethoxazole)	160 mg PO q12h 2.5 mg/kg IV q12h	Rash, including Stevens-Johnson syndrome, cytopenias	Activity against GAS is unknown; not for use in infants aged <2 mo or during third trimester of pregnancy
Vancomycin ^[119]	15–20 mg/kg q8 to q12h	'Redman' syndrome, nephrotoxicity at higher doses, neutropenia, thrombocytopenia	Requires monitoring of trough concentrations when treating serious infection

GAS = group A streptococci; iMLS_B = inducible macrolide-lincosamide-streptogramin B; IV = intravenous; PO = oral; qxh = every x hours.

Resistance to clindamycin can be constitutive or inducible, and is related to methylation of the ribosomal binding site. *S. aureus* with inducible macrolide-lincosamide-streptogramin B (iMLS_B) resistance will phenotypically be resistant to erythromycin and susceptible to clindamycin. Detection of iMLS_B resistance can be performed using the double-disk diffusion test (D-zone test), in which an erythromycin disk will induce clindamycin resistance (see figure 1).^[121] *S. aureus* that express iMLS_B have an increased rate of spontaneous mutation to constitutive resistance (erythromycin and clindamycin resistant), which may become apparent during therapy with clindamycin.

Prevalence of iMLS_B resistance appears to be more common in HA-MRSA than CA-MRSA strains. A single-centre characterized 308 MRSA isolates between 2004 and 2006. Inducible clin-damycin resistance was present in 8.4% of 102 CA-MRSA and 50% of 115 HA-MRSA isolates (p \leq 0.001). The prevalence of iMLS_B resistance in clinically defined CA-MRSA was noted to decrease in one community from 93% in

1999 to 7% in 2002.^[122] The decline in the prevalence of iMLS_B resistance was associated with the emergence of MRSA strains consistent with USA300 genotype and is likely to represent a clonal shift as USA300 CA-MRSA became endemic in that community.

Expression of iMLS_B resistance associated with clinical failure of clindamycin has been reported, as have cases of patients who were successfully treated with clindamycin despite infection with iMLS_B MRSA strains. [123-128] *In vitro* and *in vivo* experiments in a neutropenic mouse thigh abscess model suggest clindamycin activity against iMLS_B CA-MRSA strains is partially impacted by inoculum size, exerting only bacteriostatic activity and selection for resistant bacteria when higher inoculum of bacteria is present. [129] In non-inducible CA-MRSA, clindamycin maintained bactericidal activity at higher bacterial inoculum.

Evaluation for the presence of iMLS_B resistance by the D-zone test is recommended for all MRSA isolates exhibiting the erythromycin-resistant, clindamycin-susceptible phenotype. Alternatives to clindamycin should be considered



Fig. 1. Double-disk diffusion (D-zone) test for identification of inducible macrolide-lincosamide-streptogramin B (iMLS $_{\rm B}$) resistance. An erythromycin disc (E) and clindamycin disc (CC) are placed on sheep blood agar streaked with *Staphylococcus aureus*. Note the blunted zone of growth inhibition in the region between the two discs, forming a zone of clearing in the shape of the letter 'D'. This represents a positive D-zone test, indicating the presence of iMLS $_{\rm B}$ resistance.

for the treatment of MRSA infections that express iMLS_B, particularly if there is a large burden of infection, as in the setting of undrained abscesses or osteomyelitis.

4.3 Cotrimoxazole

Folate antagonists have been used for the treatment of staphylococcal infections for decades. Although largely replaced with β -lactam agents or vancomycin in the past, the emergence of CA-MRSA has led to renewed interest in the off-label use of folate antagonists for the treatment of *S. aureus* infections. The combination of trimethoprim and sulfamethoxazole blocks folate biosynthesis, necessary for thymidine biosynthesis, by inhibition of two different enzymes. Sulfonamides inhibit dihydropteroate synthase and are bacteriostatic; trimethoprim inhibits tetrahydrofolate reductase. The combination of trimethoprim and sulfamethoxazole is bactericidal. [130,131]

Resistance to cotrimoxazole can occur if there are mutations in both target enzymes and can be spread horizontally by plasmids encoding the altered genes encoding resistance. [132] Addition of exogenous thymidine can also reverse the antistaphylococcal effects of folate antagonists.

Injured tissue and inflammatory cells release DNA, which *S. aureus* thermonuclease utilizes to release thymidine.^[133,134] Susceptibility of CA-MRSA to cotrimoxazole remains high with 91.1% of isolates from the SENTRY Antimicrobial Surveillance Program susceptible.^[135]

Despite the widespread use of cotrimoxazole for MRSA infection, it does not have a specific US FDA approved indication for the treatment of S. aureus infections. Only one randomized, prospective trial studying the efficacy of cotrimoxazole has been published. In this study, cotrimoxazole was compared with vancomycin for the treatment of S. aureus (MSSA and MRSA) infections in injection drug users.[130] Infection was caused by MRSA in 46% of patients and 64% of patients were bacteraemic, including some with endocarditis, and SSTIs were noted in 32%. Overall efficacy was 86% (95% CI 76, 96) for cotrimoxazole and 98% (95% CI 94, 100) for vancomycin. No statistically significant differences were noted between the treatment groups in terms of duration of fever, bacteraemia or positive wound cultures in the MRSA infected patients. All treatment failures occurred in patients with MSSA infections.

More recent evaluations of the efficacy of cotrimoxazole for the treatment of SSTIs are more likely to have included CA-MRSA strains, although no study to date has specifically reported efficacy in CA-MRSA strains. One urban ambulatory clinic noted an increase in MRSA SSTIs starting in 2002 that subsequently became more common than MSSA SSTIs by 2005.[136] During the same period of time, empirical cotrimoxazole use and clinical resolution of S. aureus SSTIs was noted, although this may have been affected by a concurrent increase in the number of patients who had incision and drainage procedures of their infection, which some authors have suggested as possibly having a greater impact on resolution of uncomplicated SSTIs than antibacterial choice.

Despite the lack of efficacy data for the treatment of CA-MRSA specifically, cotrimoxazole has become a common empirical agent for patients with suspected MRSA SSTI as it is convenient and generally well tolerated. An important disadvantage of cotrimoxazole is its reduced activity against group A streptococci precluding its use if streptococcal infection is suspected, and an alternative agent such as clindamycin may be more appropriate for its broader Gram-positive spectrum of activity.

4.4 Vancomycin

Vancomycin has been the preferred therapy for serious MRSA infections and is generally well tolerated with a low incidence of adverse effects. Despite its long history as the 'gold standard' of MRSA therapy, some data suggest that glycopeptide susceptibility may be decreasing and clinical failures of vancomycin therapy are increasing. Based on a growing body of evidence suggesting reduced vancomycin efficacy in treating isolates with borderline susceptible minimum inhibitory concentrations (MICs), the vancomycin susceptibility breakpoints were reduced in 2006.^[137] The vancomycin breakpoints were lowered from ≤ 4 to $\leq 2 \mu g/mL$ for 'susceptible', from 8–16 to 4–8 µg/mL for 'intermediate' and from ≥32 to ≥16 µg/mL for 'resistant'. Vancomycin is not recommended for the treatment of vancomycin-resistant S. aureus or vancomycinintermediate S. aureus (VISA).

Evaluation of a large collection of *S. aureus* strains between 1997 and 2003 did not show increasing vancomycin MICs and isolates with vancomycin MIC >2 μg/mL were uncommon. [138] In contrast, evaluation of *S. aureus* from single centres have shown significant increases in vancomycin MICs over time. [139,140] It is important to note that, although the vancomycin MICs increased over time, they still remained within the susceptible range for *S. aureus*. The differences in changes in vancomycin MIC may be related to the local vancomycin usage, with higher use associated with increased vancomycin MICs.

The significance of high-susceptible range vancomycin MICs is unclear, although as the MIC increases, the frequency of heteroresistant VISA (hVISA) strains also increases.^[141] hVISA strains appear to be susceptible to vancomycin on routine susceptibility testing, but may harbour

subpopulations of VISA that may be selected for by vancomycin treatment.

Several studies suggest that vancomycin efficacy may be reduced when treating MRSA infections with vancomycin MICs in the highsusceptible range.[142-145] Two studies with different designs have noted vancomycin MIC of $1.5 \,\mu g/m L^{[145]}$ and $2.0 \,\mu g/m L^{[142]}$ are predictors of clinical failure and mortality, respectively. All of the published studies have important limitations including retrospective study design for many of the studies, absence of specific vancomycin administration information and trough concentrations, inclusion of study populations that may be biased towards failure by previous extensive vancomycin failure and absence of information regarding non-pharmacological treatment strategies (e.g. central venous catheter removal, surgical interventions) that could affect infection outcomes. Further prospective studies are needed to define the role of vancomycin in the treatment of high-susceptible MIC S. aureus.

Although vancomycin heteroresistant USA300 MRSA strains have been identified, most CA-MRSA strains remain highly susceptible.[146,147] Monitoring of vancomycin trough concentrations, treatment response and changes in vancomycin MIC should be performed in patients with invasive CA- and HA-MRSA infections. Highdose vancomycin therapy has been suggested by some (maintaining vancomycin target trough concentrations of 15-20 µg/mL), although nephrotoxicity may be more common in this dose administration strategy.[148] Rising vancomycin MICs may indicate the presence of hVISA strains and alternative therapy should be considered in these situations, especially if clinical response is unsatisfactory.

4.5 Daptomycin

Daptomycin is a cyclic lipopeptide that is rapidly bactericidal against almost all Grampositive cocci including MRSA. Although the precise mechanism of action has not been elucidated, daptomycin is hypothesized to cause membrane depolarization via calcium-dependent insertion into the bacterial membrane. [149]

Evaluation of large collections of MRSA indicated all MRSA tested were susceptible to daptomycin with a MIC at which 90% of bacteria are inhibited (MIC₉₀) of 0.5 μg/mL.^[150,151] Decreasing susceptibility to daptomycin has been reported to occur in clinical S. aureus isolates both in patients with and without prior exposure to daptomycin.[152-156] hVISA may have an increased daptomycin MIC value, probably as a consequence of a thickened cell wall. Although vancomycin and daptomycin have different mechanisms of action, both mechanisms are directly related to the bacterial cell wall and the increased thickness may provide a physical barrier that limits the drug binding and activity. Decreasing susceptibility to daptomycin was reported in a USA300, PVL-positive CA-MRSA isolate from a patient with a ortic valve endocarditis.[157]

Daptomycin is approved for the treatment of complicated SSTIs (including MRSA), *S. aureus* (MSSA and MRSA) bacteraemia and right-sided endocarditis based on prospective, randomized clinical trials using vancomycin or antistaphylococcal penicillins as comparators. [158,159] In both the SSTI and bacteraemia trials, daptomycin was found to be noninferior to the comparator regimen. In the bacteraemia trial, low success rates were noted among patients with left-sided endocarditis in both the daptomycin and standard therapy groups (11.1% vs 22.2%, respectively).

Observational studies have described the efficacy of daptomycin for a variety of infections including osteomyelitis, [160,161] septic arthritis [162] and prosthetic joint infections [163] with varying degrees of success. At present, no prospective clinical trials have been performed to thoroughly evaluate the efficacy of daptomycin for the treatment of osteoarticular infections.

Daptomycin is not recommended for the treatment of pneumonia. In a phase III clinical trial for the treatment of CAP, daptomycin failed to achieve non-inferiority to ceftriaxone (79% vs 87% efficacy). [164] *In vitro* experiments have shown that pulmonary surfactant interacts directly with daptomycin and inhibits its antibacterial activity. Daptomycin irreversibly inserts into surfactant lipid aggregates. [164] Surfactant is not likely to be present in quantities to affect

daptomycin activity in pulmonary abscesses and thus daptomycin may still be active in patients with tricuspid endocarditis complicated by pulmonary abscesses.

Daptomycin is generally well tolerated with a low incidence of renal dysfunction noted in the bacteraemia trial.[159] Elevation of creatine kinase (CK) with or without muscle pain has been noted in patients on daptomycin therapy. Among 534 patients with complicated SSTI receiving daptomycin 4 mg/kg, 0.2% reported myopathy. [158] In the bacteraemia trial (using daptomycin 6 mg/kg), elevation of CK to >500 IU/L was more common in the daptomycin arm than the standard therapy arm (9.5% vs 1.5%; p=0.02). CK levels normalized either with discontinuation of daptomycin or after completion of therapy. It is recommended that daptomycin-treated patients have CK levels monitored at baseline and weekly thereafter, and that the use of other drugs that may similarly increase CK or cause myopathy (e.g. HMG-CoA reductase inhibitors) is suspended. CK levels >1000 IU/L associated with unexplained myopathy or asymptomatic patients with CK levels ≥5 to 10 times the upper limit of normal (ULN) should have daptomycin discontinued.

4.6 Linezolid

Linezolid is a bacteriostatic, synthetic oxazolidinone that inhibits protein synthesis at the 50S ribosome and is active *in vitro* against *S. aureus* including MRSA, penicillin-resistant *S. pneumoniae* and vancomycin-resistant enterococci. Linezolid is available in both intravenous and oral formulations, and is approved in the US for the treatment of complicated SSTIs and nosocomial pneumonia caused by susceptible bacteria.

Patients with nosocomial pneumonia treated with linezolid were noted to have a significantly higher clinical cure rate than patients treated with vancomycin plus aztreonem in double-blind clinical trials (59% vs 36%, respectively; p<0.01). [165] Low cure rates in the vancomycin treatment arm suggest subtherapeutic vancomycin administration may have occurred as trough concentrations were not measured in these trials.

Linezolid is hypothesized to achieve higher lung tissue concentrations than vancomycin, [166-168] and thus may have an advantage over vancomycin for the treatment of pneumonia, although a recent clinical trial found that early microbiological response was similar in patients with MRSA ventilator-associated pneumonia who were treated with linezolid versus vancomycin. [169] Linezolid has not been evaluated specifically for the treatment of CA-MRSA necrotizing pneumonia, although it remains as an important option because it may reduce toxin production in *S. aureus* in addition to its antibacterial effects. [170]

Although a retrospective review of linezolid clinical trials shows patients with bacteraemia had similar outcomes to comparator drugs,[171] the role of linezolid for the treatment of S. aureus bacteraemia remains unclear. An open-label, randomized clinical trial comparing linezolid with vancomycin for the treatment of catheterassociated bloodstream infections was terminated early because of higher mortality in the linezolid arm.[172] The mortality difference was seen in patients with Gram-negative, mixed Gram-positive/Gram-negative infections, or no infection, and not in patients with Gram-positive infections alone. Post hoc analysis showed that treatment for infection with Gram-negative pathogens may have been inadequate in more than half of the deaths. Nonetheless, given the limited data available and bacteriostatic mode of action of linezolid, alternative agents should be considered for the treatment of bacteraemia.

The most common adverse reactions to linezolid include nausea, vomiting and diarrhoea. Significant limiting adverse effects include reversible thrombocytopenia, [173] serotonin toxicity, [174,175] peripheral neuropathy, [176] optic neuritis [177] and lactic acidosis. [178] Patients who receive >2 weeks of linezolid therapy should have weekly monitoring for myelosuppression. The ability of linezolid to cause serotonin syndrome is related to its weak non-selective inhibition of monoamine oxidase. Drugs with serotonergic activity (e.g. serotonin reuptake inhibitors, monoamine oxidase inhibitors, pethidine [meperidine], bupropion) should not be used concomitantly with linezolid.

4.7 Quinupristin/Dalfopristin

Quinupristin/dalfopristin is a combination streptogramin agent that is approved for the treatment of SSTIs due to MSSA and streptococci, and for the treatment of vancomycin-resistant Enterococcus faecium bacteraemia. Individually, quinupristin (a group B streptogramin) and dalfopristin (a group A streptogramin) are bacteriostatic, but the combination of quinupristin/dalfopristin is bactericidal against S. aureus via inhibition of protein synthesis by interfering with different components of 23S RNA in the 50S subunit of the bacterial ribosome. Resistance to quinupristin/dalfopristin is conferred by the MLS_R methylation mechanism previously described (section 4.2). Constitutive resistance confers resistance to quinupristin; however, synergy with streptogramin A agents may be retained, [179] although the activity may become bacteriostatic. Among S. aureus strains exhibiting iMLS_B resistance, quinupristin remains active, as it is not an inducer of the methylase.

Quinupristin/dalfopristin has been studied for the treatment of MRSA infections in an openlabel, emergency-use programme. [180] Patients with documented MRSA infections and who were either not responding to or intolerant of other MRSA active antibacterials were included in this study. Overall successful treatment was noted in 66.7% of patients who were clinically and bacteriologically evaluable, but this was largely driven by success in treating osteoarticular infections and SSTIs. Therapy failed in two patients with endocarditis. The MLS_B resistance phenotype (susceptible, constitutive or inducible resistance) did not appear to affect the response to therapy. Use of quinupristin/dalfopristin has been limited by its significant adverse effects including infusion site pain, arthralgias and myalgias, which can be severe enough to lead to discontinuation of therapy.[181]

4.8 Tigecycline

Tigecycline is a semisynthetic glycylcyline that has an expanded broad-spectrum antibacterial with activity against Gram-positive, Gramnegative, anaerobic and various atypical pathogens.

Although derived from minocycline, an altered 9-t-butylglycylamido side chain confers a broader spectrum of activity and allows tigecycline to overcome tetracycline resistance. Tigecycline has *in vitro* activity against several drug-resistant pathogens including MRSA, VISA, vancomycin-resistant enterococci and many extended-spectrum β-lactamase Gram-negative bacteria. Tigecycline has limited or no activity against *Pseudomonas* spp. and reduced activity against *Proteus mirabilis*.

Glycycyclines are bacteriostatic and exhibit activity by binding to bacterial 30S ribosomal subunit, preventing protein synthesis. Steric hindrance produced by the large substituent at position 9 allows tigecycline to overcome the major mechanisms of tetracycline resistance: active efflux of the drug out of the bacterial cell and ribosomal protection.[182] Tigecycline MIC values against MRSA are generally low, ranging from ≤0.06 to 2 µg/mL.[183] Among 1989 clinical isolates of CA-MRSA from North America, tigecycline was active against 98.2% of strains at the susceptibility breakpoint of $\leq 0.5 \,\mu\text{g/mL}$, the remaining 1.8% had an MIC of 1.0 µg/mL.[135] The majority of these strains were SCC*mec* type IV, PVL-positive MRSA (94.7%) and 88.4% were PFGE genotype USA300-0114.

The pharmacokinetic and pharmacodynamic properties of tigecycline have been well described elsewhere. [184] It is important to note that tigecycline is extensively distributed in tissues and there is a rapid decline in plasma concentration during the first 2 hours after administration. [184] Based on its relatively low plasma concentration, tigecycline should be used with caution in patients with suspected or proven bacteraemia.

Tigecycline is approved in the US for the treatment of complicated SSTI, including MRSA, and for complicated intra-abdominal infections (including MSSA but not MRSA infection). Tigecycline was noninferior to the combination of vancomycin and aztreonem for the treatment of complicated SSTIs in pooled data from two phase III, double-blind studies. Nausea and vomiting were the most common adverse effects reported in the clinical trials. Later evaluation of the MRSA strains from all of the tigecycline registrational studies found that 76 of 173 (44%)

isolates had at least one genetic feature associated with CA-MRSA.^[186]

4.9 Ceftobiprole

Ceftobiprole is a broad-spectrum cephalosporin that retains activity against MRSA by tightly binding to penicillin-binding protein 2a, the major determinant of meticillin resistance in staphylococci. Ceftobiprole is administered intravenously and appears to have a low potential to select for resistance. Prolonged serial passage in the presence of subinhibitory concentrations of ceftobiprole failed to select for clones with MIC values >4 times the parent strains, with a maximum MIC of 8 µg/mL in only one of ten strains tested. [187]

A randomized, double-blind trial comparing ceftobiprole with vancomycin plus ceftazidime for the treatment of complicated SSTI showed noninferiority of ceftobiprole monotherapy.[111] Infection due to MRSA was present in 22% and 18% of the ceftobiprole and comparator arms, respectively, with the majority of the remainder of infections due to MSSA, Enterobacteriaceae, P. aeruginosa and streptococci. Among MRSAinfected patients, PVL-positive strains were present in 92% and 84% of ceftobiprole and comparator arms, respectively. In vitro susceptibility testing of SCCmec type IV CA-MRSA isolates found all isolates were susceptible to ceftobiprole with MIC₅₀ and MIC₉₀ values of 1 and 2 µg/mL, respectively.[188] Ceftobiprole has been approved for the treatment of complicated SSTI including diabetic foot infections in Canada and Switzerland, and is currently under review for approval by regulatory authorities in the US, EU and Australia.

4.10 Other Agents and Combination Therapy

Many CA-MRSA strains show susceptibility to fluoroquinolone agents.^[5,104] Despite the *in vitro* susceptibility, fluoroquinolones are not recommended as therapy for suspected or proven MRSA infections because of concerns about the acquisition of resistance during therapy.^[189,190] Rifampin is active against many CA-MRSA isolates, but is not recommended as monotherapy

for the treatment of *S. aureus* infections because of the rapid emergence of resistance that can occur during therapy.^[191] Combination therapy of rifampin with other anti-staphylococcal agents, particularly tetracyclines and cotrimoxazole, has been used for the treatment of CA-MRSA, although there are limited data supporting this practice.^[192]

4.11 Investigational Agents

Several investigational agents for the treatment of MRSA infections have completed phase II or III studies and may potentially be available pending approval. Dalbavancin is a semisynthetic glycopeptide that has the unique characteristic of having a long half-life allowing for once-weekly administration.[193] The dose administration regimen is 1000 mg intravenously on day 1, then 500 mg intravenously on day 8. It inhibits cell-wall synthesis and exhibits concentration-dependent bactericidal activity. In a phase II study comparing dalbavancin to twice-daily linezolid therapy for 14 days, MRSA eradication was similar in the two groups (91% of dalbavancin recipients and 89% of linezolid recipients). However, clinical cure was not specifically reported.[194] An open-label, phase II study of catheter-related Gram-positive bacteria bloodstream infections showed dalbavancin was superior to vancomycin in curing infection (87% vs 50%; p < 0.05), although the number of patients with MRSA infection was small.[195]

Telavancin is a rapidly bactericidal lipoglycopeptide that exhibits activity through inhibition of cell-wall synthesis and membrane depolarization. [196] It has a long half-life of 7–9 hours allowing once-daily administration using 7.5–10 mg/kg/day. Telavancin has completed phase III studies with vancomycin as the comparator in patients with SSTI. [197] In these studies, MRSA caused a significant number of infections and most of these isolates were SCC*mec* type IV and PVL positive. [198] Telavancin therapy led to cure in 90.6% of patients versus 86.4% of vancomycin-treated patients. It is currently under review by regulatory authorities in the US.

Oritavancin is a semisynthetic glycopeptide that has completed phase III studies of efficacy in the treatment of complicated SSTI, although complete clinical details of these studies are not yet available. [199,200] Favourable outcomes were noted in the ortivancin-treated patients compared with those who received vancomycin followed by cephalexin. Oritavancin appears to exhibit a dual mode of action to inhibit cell-wall synthesis, [201] and has been shown to demonstrate activity against vancomycin-resistant staphylococci and enterococci. [202]

Other agents that are under study include the broad-spectrum cephalosporin ceftaroline and the diaminopyrimidine dihydrofolate reductase inhibitor iclaprim. Both agents have activity against MRSA and clinical studies are ongoing. [203-208]

4.12 Non-Pharmacological Therapy of CA-MRSA Infections

Data from epidemiological studies^[25,27,209] and randomized trials^[210,211] have noted cure of uncomplicated CA-MRSA SSTIs following incision and drainage despite the use of non-MRSA active antibacterial. Incision and drainage is recommended for any suspected uncomplicated cutaneous abscess and may be sufficient without systemic antibacterial therapy.^[212,213] Systemic antibacterial therapy is warranted in addition to incision and drainage if surrounding cellulitis or fever is present.^[212,213]

The importance of removing intravascular catheters or implanted prosthetic devices whenever possible cannot be overstated if these devices are potentially contaminated with *S. aureus*. Even in the presence of appropriate antibacterial therapy, infection can persist on artificial devices and lead to prolonged infection or treatment failure.

4.13 Clinical Approach to Diagnosis and Therapy of CA-MRSA Infections

The ideal approach to management of CA-MRSA infection has not been determined, but current expert opinion suggests a combination of medical and surgical therapy based on the extent of infection. [214,215] CA-MRSA should be

considered as a cause of infection in patients presenting with disease consistent with frequently described manifestations, particularly SSTIs (especially if there is a description of 'spider bite'), fasciitis and myositis, necrotizing pneumonia and haematogenous osteoarticular infections. Culture data should be obtained whenever possible to guide antibacterial therapy, particularly if there is little response to empirical therapy. Incision and drainage of furuncles or carbuncles should be performed and may be sufficient therapy alone without antibacterial therapy for uncomplicated soft-tissue infections. Empirical antibacterial therapy with an MRSA-active agent is recommended if cellulitis is present in addition to abscess or if the patient is immunocompromised, has severe local infection or does not improve after incision and drainage. [215] Complicated SSTIs and suspected invasive CA-MRSA infection may require initial therapy with intravenous agents. The choice of antibacterial agent should be dictated by the type and severity of infection, patient factors (e.g. allergies, concomitant medications), potential drug adverse effects, cost and availability, and culture susceptibilities if available.

5. Decolonization of CA-MRSA

Decolonization of S. aureus carriage is often attempted to prevent recurrent CA-MRSA infection. Numerous strategies have been proposed, but the most common include the use of nasal mupirocin to eradicate nasal carriage, topical chlorhexidine gluconate to eradicate other cutaneous skin carriage and occasionally systemic antibacterial agents. The efficacy of these various strategies in preventing recurrent CA-MRSA infections has not been evaluated, but can be an adjunct to environmental hygiene and cleaning.[214] Colonization has been suggested to play a lesser role in the spread of CA-MRSA than person-to-person transmission and contaminated fomites, thus limiting the impact of decolonization.[216] Consultation with an infectious diseases expert is recommended when considering decolonization strategies for patients with recurrent CA-MRSA infections.

6. Conclusions

The rapid emergence and spread of CA-MRSA highlights the difficulty of treating drugresistant bacteria combined with an apparently more virulent strain. Initial reports of CA-MRSA infections represented focal outbreaks in specific populations, but current data indicate that CA-MRSA is now endemic in many regions of the world. In communities where CA-MRSA is endemic, empirical therapy for common manifestations of CA-MRSA disease, particularly SSTIs, should include MRSA active antibacterials. Clinical trials are needed to determine the optimal antibacterial therapy for CA-MRSA infections as well as the role of decolonization in preventing recurrent infections. In addition, further basic research into the virulence mechanisms of CA-MRSA strains may yield targets for nonpharmacological therapies.

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References

- National Nosocomial Infections Surveillance (NNIS) System Report data summary from January 1992 through June 2004 issued October 2004. Am J Infect Control 2004; 32: 470-85
- Noskin GA, Rubin RJ, Schentag JJ, et al. National trends in *Staphylococcus aureus* infection rates: impact on economic burden and mortality over a 6-year period (1998-2003). Clin Infect Dis 2007; 45: 1132-40
- Klevens RM, Morrison MA, Nadle J, et al. Invasive methicillin-resistant Staphylococcus aureus infections in the United States. JAMA 2007; 298: 1763-71
- Jevons MP, Coe AW, Parker MT. Methicillin resistance in staphylococci. Lancet 1963; I: 904-7
- Naimi TS, LeDell KH, Como-Sabetti K, et al. Comparison of community and health care-associated methicillinresistant *Staphylococcus aureus* infection. JAMA 2003; 290: 2976-84
- Levine DP, Cushing RD, Jui J, et al. Community-acquired methicillin-resistant Staphylococcus aureus endocarditis in the Detroit Medical Center. Ann Intern Med 1982; 97: 330-8
- Cafferkey MT, Hone R, Falkiner FR, et al. Gentamicin and methicillin-resistant *Staphylococcus aureus* in Dublin hospitals: clinical and laboratory studies. J Med Microbiol 1983; 16: 117-27

- Udo EE, Pearman JW, Grubb WB. Genetic analysis of community isolates of methicillin-resistant *Staphylo-coccus aureus* in Western Australia. J Hosp Infect 1993; 25: 97-108
- Herold BC, Immergluck LC, Maranan MC, et al. Community-acquired methicillin-resistant Staphylococcus aureus in children with no identified predisposing risk. JAMA 1998; 279: 593-8
- Centers for Disease Control and Prevention. Four pediatric deaths from community-acquired methicillin-resistant Staphylococcus aureus – Minnesota and North Dakota, 1997-1999. MMWR Morb Mortal Wkly Rep 1999; 48: 707-10
- Centers for Disease Control and Prevention. Methicillinresistant Staphylococcus aureus skin or soft tissue infections in a state prison – Mississippi, 2000. MMWR Morb Mortal Wkly Rep 2001; 50: 919-22
- Centers for Disease Control and Prevention. Outbreaks of community-acquired methicillin-resistant Staphylococcus aureus skin infections – Los Angeles County, California, 2002-2003 [letter]. MMWR Morb Mortal Wkly Rep 2003; 52: 88
- Centers for Disease Control and Prevention. Methicillinresistant Staphylococcus aureus infections in correctional facilities – Georgia, California, and Texas, 2001-2003. MMWR Morb Mortal Wkly Rep 2003; 52: 992-6
- Baggett HC, Hennessy TW, Leman RL, et al. Outbreak of community-onset methicillin-resistant *Staphylococcus aureus* skin infections in southwestern Alaska. Infect Control Hosp Epidemiol 2003; 24: 397-402
- Centers for Disease Control and Prevention. Methicillinresistant Staphylococcus aureus infections among competitive sports participants – Colorado, Indiana, Pennsylvania, and Los Angeles County, 2000-2003. MMWR Morb Mortal Wkly Rep 2003; 52: 793-5
- Begier EM, Frenette K, Barrett NL, et al. A high-morbidity outbreak of methicillin-resistant Staphylococcus aureus among players on a college football team, facilitated by cosmetic body shaving and turf burns. Clin Infect Dis 2004; 39: 1446-53
- Kazakova SV, Hageman JC, Matava M, et al. A clone of methicillin-resistant *Staphylococcus aureus* among professional football players. N Engl J Med 2005; 352: 468-75
- Campbell KM, Vaughn AF, Russell KL, et al. Risk factors for community-acquired methicillin-resistant *Staphylo-coccus aureus* infections in an outbreak of disease among military trainees in San Diego, California, in 2002. J Clin Microbiol 2004; 42: 4050-3
- Nimmo GR, Schooneveldt J, O'Kane G, et al. Community acquisition of gentamicin-sensitive methicillin-resistant Staphylococcus aureus in southeast Queensland, Australia. J Clin Microbiol 2000; 38: 3926-31
- Aires de Sousa M, Bartzavali C, Spiliopoulou I, et al. Two international methicillin-resistant *Staphylococcus aureus* clones endemic in a university hospital in Patras, Greece. J Clin Microbiol 2003; 41: 2027-32
- Vandenesch F, Naimi T, Enright MC, et al. Communityacquired methicillin-resistant *Staphylococcus aureus* carrying Panton-Valentine leukocidin genes: worldwide emergence. 2003; 9: 978-84

- Liassine N, Auckenthaler R, Descombes MC, et al. Community-acquired methicillin-resistant Staphylococcus aureus isolated in Switzerland contains the Panton-Valentine leukocidin or exfoliative toxin genes. J Clin Microbiol 2004; 42: 825-8
- Velazquez-Meza ME, Aires de Sousa M, Echaniz-Aviles G, et al. Surveillance of methicillin-resistant *Staphylococcus* aureus in a pediatric hospital in Mexico City during a 7-year period (1997-2003): clonal evolution and impact of infection control. J Clin Microbiol 2004; 42: 3877-80
- Ribeiro A, Dias C, Silva-Carvalho MC, et al. First report of community-acquired methicillin-resistant *Staphylo-coccus aureus* in South America. J Clin Microbiol 2005; 43: 1985-8
- Fridkin SK, Hageman JC, Morrison M, et al. Methicillinresistant *Staphylococcus aureus* disease in three communities. N Engl J Med 2005; 352: 1436-44
- Vourli S, Perimeni D, Makri A, et al. Community acquired MRSA infections in a paediatric population in Greece. Euro Surveill 2005; 10: 78-9
- Moran GJ, Krishnadasan A, Gorwitz RJ, et al. Methicillinresistant S. aureus infections among patients in the emergency department. N Engl J Med 2006; 355: 666-74
- Hsu LY, Koh YL, Chlebicka NL, et al. Establishment of ST30 as the predominant clonal type among communityassociated methicillin-resistant Staphylococcus aureus isolates in Singapore. J Clin Microbiol 2006; 44: 1090-3
- Huang YC, Su LH, Wu TL, et al. Changing molecular epidemiology of methicillin-resistant *Staphylococcus* aureus bloodstream isolates from a teaching hospital in Northern Taiwan. J Clin Microbiol 2006; 44: 2268-70
- Nimmo GR, Coombs GW, Parson JC, et al. Methicillinresistant Staphylococcus aureus in the Australian community: an evolving epidemic. Med J Aust 2006; 184: 374-5
- Kuehnert MJ, Kruszon-Moran D, Hill HA, et al. Prevalence of *Staphylococcus aureus* nasal colonization in the United States, 2001-2002. J Infect Dis 2006; 193: 172-9
- Gorwitz RJ, Kruszon-Moran D, McAllister SK, et al. Changes in the prevalence of nasal colonization with Staphylococcus aureus in the United States, 2001-2004. J Infect Dis 2008; 197: 1226-34
- 33. Eveillard M, de Lassence A, Lancien E, et al. Evaluation of a strategy of screening multiple anatomical sites for methicillin-resistant *Staphylococcus aureus* at admission to a teaching hospital. Infect Control Hosp Epidemiol 2006; 27: 181-4
- Mertz D, Frei R, Jaussi B, et al. Throat swabs are necessary to reliably detect carriers of *Staphylococcus aureus*. Clin Infect Dis 2007; 45: 475-7
- Buehlmann M, Frei R, Fenner L, et al. Highly effective regimen for decolonization of methicillin-resistant Staphylococcus aureus carriers. Infect Control Hosp Epidemiol 2008; 29: 510-6
- Roghmann MC, Siddiqui A, Plaisance K, et al. MRSA colonization and the risk of MRSA bacteraemia in hospitalized patients with chronic ulcers. J Hosp Infect 2001; 47: 98-103
- Huang SS, Platt R. Risk of methicillin-resistant Staphylococcus aureus infection after previous infection or colonization. Clin Infect Dis 2003; 36: 281-5

 Davis KA, Stewart JJ, Crouch HK, et al. Methicillinresistant Staphylococcus aureus (MRSA) nares colonization at hospital admission and its effect on subsequent MRSA infection. Clin Infect Dis 2004; 39: 776-82

- Ellis MW, Hospenthal DR, Dooley DP, et al. Natural history of community-acquired methicillin-resistant Staphylococcus aureus colonization and infection in soldiers. Clin Infect Dis 2004; 39: 971-9
- Cohen AL, Shuler C, McAllister S, et al. Methamphetamine use and methicillin-resistant *Staphylococcus aureus* skin infections. Emerg Infect Dis 2007; 13: 1707-13
- Diep BA, Chambers HF, Graber CJ, et al. Emergence of multidrug-resistant, community-associated, methicillinresistant *Staphylococcus aureus* clone USA300 in men who have sex with men. Ann Intern Med 2008; 148: 249-57
- Groom AV, Wolsey DH, Naimi TS, et al. Communityacquired methicillin-resistant *Staphylococcus aureus* in a rural American Indian community. JAMA 2001; 286: 1201-5
- Baggett HC, Hennessy TW, Rudolph K, et al. Communityonset methicillin-resistant Staphylococcus aureus associated with antibiotic use and the cytotoxin Panton-Valentine leukocidin during a furunculosis outbreak in rural Alaska. J Infect Dis 2004; 189: 1565-73
- McDougal LK, Steward CD, Killgore GE, et al. Pulsedfield gel electrophoresis of oxacillin-resistant *Staphylo-coccus aureus* isolates from the United States: establishing a national database. J Clin Microbiol 2003; 41: 5113-20
- Diep BA, Carleton HA, Chang RF, et al. Role of 34 virulence genes in the evolution of hospital- and communityassociated strains of methicillin-resistant *Staphylococcus* aureus. J Infect Dis 2006; 193: 1495-503
- Baba T, Takeuchi F, Kuroda M, et al. Genome and virulence determinants of high virulence community-acquired MRSA. Lancet 2002; 359: 1819-27
- 47. Lina G, Piémont Y, Godail-Gamt F, et al. Involvement of Panton-Valentine leukocidin-producing *Staphylococcus aureus* in primary skin infections and pneumonia. Clin Infect Dis 1999; 29: 1128-32
- Dufour P, Gillet Y, Bes M, et al. Community-acquired methicillin-resistant *Staphylococcus aureus* infections in France: emergence of a single clone that produces Panton-Valentine leukocidin. Clin Infect Dis 2002; 35: 819-24
- Diep BA, Sensabaugh GF, Somboona NS, et al. Widespread skin and soft-tissue infections due to two methicillin-resistant *Staphylococcus aureus* strains harboring the genes for Panton-Valentine leucocidin. J Clin Microbiol 2004; 42: 2080-4
- Voyich JM, Otto M, Mathema B, et al. Is Panton-Valentine Leukocidin the major virulence determinant in community-associated methicillin-resistant *Staphylococcus* aureus disease? J Infect Dis 2006; 194: 1761-70
- Diep BA, Palazzolo-Balance AM, Tattevin P, et al. Contribution of Panton-Valentine leukocidin in community-associated methicillin-resistant *Staphylococcus aureus* pathogenesis. PLoS One 2008; 3: e3198
- Labandeira-Rey M, Couzon F, Boisset S, et al. Staphylococcus aureus Panton Valentine leukocidin causes necrotizing pneumonia. Science 2007; 315: 1130-3
- Bubeck Wardenburg J, Bae T, Otto M, et al. Poring over the pores: α-hemolysin and Panton-Valentine leukocidin

- in *Staphylococcus aureus* pneumonia. Nat Med 2007; 13: 1405-6
- 54. Wardenburg JB, Palazzolo-Balance AM, Otto M, et al. Panton-Valentine is not a virulence determinant in murine models of community-associated methicillin-resistant Staphylococcus aureus disease. J Infect Dis 2008; 198: 1166-70
- Wang R, Braughton KR, Kretschmer D, et al. Identification of novel cytolytic peptides as key virulence determinants for community-associated MRSA. Nat Med 2007; 13: 1510-4
- Diep BA, Gill SR, Chang RF, et al. Complete genome sequence of USA300, an epidemic clone of communityacquired meticillin-resistant Staphylococcus aureus. Lancet 2006; 367: 731-9
- 57. Diep BA, Stone GC, Basuino L, et al. The arginine catabolic mobile element and staphylococcal chromosomal cassette mec linkage: convergence of virulence and resistance in the USA300 clone of methicillin-resistant Staphylococcus aureus. J Infect Dis 2008; 197: 1523-30
- Diep BA, Gill SR, Change RF, et al. Complete genome sequence of USA300, an epidemic clone of communityacquired meticillin-resistant *Staphylococcus aureus*. Lancet 2006; 367: 731-39
- Han LL, McDougal LK, Gorwitz RJ, et al. High frequencies of clindamycin and tetracycline resistance in methicillin-resistant Staphylococcus aureus pulsed-field type USA300 isolates collected at a Boston ambulatory health center. J Clin Microbiol 2007; 45: 1350-2
- Okuma K, Iwakawa K, Turnidge JD, et al. Dissemination of new methicillin-resistant *Staphylococcus aureus* clones in the community. J Clin Microbiol 2002; 40: 4289-94
- Katayama Y, Ito T, Hiramatsu K. A new class of genetic element, staphylococcus cassette chromosome *mec*, encodes methicillin resistance in *Staphylococcus aureus*. Antimicrob Agents Chemother 2000; 44: 1549-55
- Ito T, Katayama Y, Asada K, et al. Structural composition of three types of staphylococcal cassette chromosome mec integrated in the chromosome of methicillin-resistant Staphylococcus aureus. Antimicrob Agents Chemother 2002; 46: 1147-52
- 63. Ma XX, Ito T, Tiensasitorn C, et al. Novel type of staphylococcal chromosome *mec* identified in communityacquired methicillin-resistant *Staphylococcus aureus* strains. Antimicrob Agents Chemother 2002; 46: 1147-52
- Daum RS, Ito T, Hiramatsu K, et al. A novel methicillinresistance cassette in community-acquired methicillinresistant *Staphylococcus aureus* isolates of diverse genetic backgrounds. J Infect Dis 2002; 186: 1344-7
- Ito T, Ma XX, Takeuchi F, et al. Novel type V Staphylococcal cassette chromosome mec driven by a novel cassette chromosome recombinase, ccrC. Antimicrob Agents Chemother 2004; 48: 2637-51
- 66. Shore A, Rossney AS, Keane CT, et al. Seven novel variants of the staphylococcal chromosomal cassette mec in methicillin-resistant Staphylococcus aureus isolates from Ireland. Antimicrob Agents Chemother 2005; 49: 2070-83
- King MD, Humphrey BH, Want YF, et al. Emergence of community-acquired methicillin-resistant Staphylococcus aureus USA 300 clone as the predominant cause of skin

- and soft-tissue infections. Ann Intern Med 2006; 144: 309-17
- 68. Miller LG, Quan C, Shay A, et al. A prospective investigation of outcomes after hospital discharge for endemic, community-acquired methicillin-resistant and -susceptible *Staphylococcus aureus* skin infection. Clin Infect Dis 2007; 44: 483-92
- Graber CJ, Jacobson MA, Perdreau-Remington F, et al. Recurrence of skin and soft tissue infection caused by methicillin-resistant *Staphylococcus aureus* in a HIV primary care clinic. J Acquired Immune Defic Syndr 2008; 49: 231-3
- Miller LG, Perdreau-Remington F, Rieg G, et al. Necrotizing fasciitis caused by community-associated methicillin-resistant *Staphylococcus aureus* in Los Angeles. N Engl J Med 2005; 352: 1445-53
- Fowler A, Mackay A. Community-acquired methicillinresistant *Staphylococcus aureus* pyomyositis in an intravenous drug user. J Med Microbiol 2006; 55: 123-5
- Dehority W, Wang E, Vernon PS, et al. Communityassociated methicillin-resistant Staphylococcus aureus necrotizing fasciitis in a neonate. Pediatr Infect Dis J 2006; 25: 1080-1
- Pannaraj PS, Hulten KG, Gonzalez BE, et al. Infective pyomyositis and myositis in children in the era of community-acquired, methicillin-resistant *Staphylococcus* aureus infection. Clin Infect Dis 2006; 43: 953-60
- Sokolov KM, Kreye E, Miller LG, et al. Postpartum iliopsoas pyomyositis due to community-acquired methicillin-resistant *Staphylococcus aureus*. Obstet Gynel 2007; 110: 535-8
- Shedek BK, Nilles EJ. Community-associated methicillinresistant *Staphylococcus aureus* pyomyositis complicated by compartment syndrome in an immunocompetent young woman. Am J Emerg Med 2008; 26: 737.e3-4
- Burton MJ, Shah P, Swiatlo E. Community-acquired methicillin-resistant *Staphylococcus aureus* as a cause of Fournier's gangrene. Am J Med Sci 2008; 335: 327-8
- Gillet Y, Issartel B, Vanhems P, et al. Association between Staphylococcus aureus strains carrying gene for Panton-Valentine leukocidin and highly lethal necrotizing pneumonia in young immunocompetent patients. Lancet 2002; 359: 753-9
- Francis JS, Doherty MC, Lopatin U, et al. Severe community-onset pneumonia in healthy adults caused by methicillin-resistant *Staphylococcus aureus* carrying the Panton-Valentine leukocidin genes. Clin Infect Dis 2005; 40: 100-7
- Gonzalez BE, Hulten KG, Dishop MK, et al. Pulmonary manifestations in children with invasive communityacquired *Staphylococcus aureus* infection. Clin Infect Dis 2005; 41: 583-90
- Hageman JC, Uyeki TM, Francis JS, et al. Severe community-acquired pneumonia due to *Staphylococcus aureus*, 2003-04 influenza season. Emerg Infect Dis 2006; 12: 894-9
- Centers for Disease Control and Prevention. Severe methicillin-resistant Staphylococcus aureus communityacquired pneumonia associated with influenza – Louisiana and Georgia, December 200 – January 2007. MMWR Morb Mortal Wkly Rep 2007; 56: 325-9

- Martínez-Aguilar G, Hammerman WA, Mason EO, et al. Clindamycin treatment of invasive infections caused by community-acquired methicillin-resistant and methicillinsusceptible *Staphylococcus aureus* in children. Pediatr Infect Dis J 2003; 22: 593-8
- Arnold SR, Elias D, Buckingham SC, et al. Changing patterns of acute hematogenous osteomyelitis and septic arthritis. J Pediatr Orthop 2006; 26: 703-8
- Gonzalez BE, Teruya J, Mahoney DH, et al. Venous thrombosis associated with staphylococcal osteomyelitis in children. Pediatrics 2006; 117: 1673-9
- Gillet Y, Dohin B, Dumitrescu O, et al. Osteoarticular infections with *Staphylococcus aureus* secreting Panton-Valentine leucocidin. Arch Pediatr 2007; 14: S102-7
- Crum NF. The emergence of severe, community-acquired methicillin-resistant *Staphylococcus aureus* infections. Scand J Infect Dis 2005; 37: 651-6
- Seybold U, Talati NJ, Shah M, et al. Hematogenous osteomyelitis mimicking osteosarcoma due to community-associated methicillin-resistant *Staphylococcus aureus*. Infection 2007; 35: 190-3
- Nourse C, Starr M, Munckhof W. Community-acquired methicillin-resistant Staphylococcus aureus causes severe disseminated infection and deep venous thrombosis in children: literature review and recommendations for management. J Paediatr Child Health 2007; 43: 656-61
- Bahrain M, Vasiliades M, Wolff M, et al. Five cases of bacterial endocarditis after furunculosis and the ongoing saga of community-acquired methicillin-resistant *Staphy-lococcus aureus* infections. Scand J Infect Dis 2006; 38: 702-7
- Haque NZ, Davis SL, Manierski CL, et al. Infective endocarditis caused by USA300 methicillin-resistant Staphylococcus aureus (MRSA). Int J Antimicrob Agents 2007; 30: 72-7
- Gonzalez BE, Martinez-Aguilar G, Hulten KG, et al. Severe staphylococcal sepsis in adolescents in the era of community-acquired methicillin-resistant *Staphylococcus* aureus. Pediatrics 2005; 115: 642-8
- Kravitz GR, Dries DJ, Peterson ML, et al. Purpura fulminans due to *Staphylococcus aureus*. Clin Infect Dis 2005; 40: 941-7
- Adem PV, Montgomery CP, Husain AN, et al. Staphylococcus aureus sepsis and the Waterhouse-Friderichsen syndrome in children. N Engl J Med 2005; 353: 1245-51
- Valentine P, Parisi G, Monaco M, et al. An uncommon presentation for a severe invasive infection due to methicillinresistant *Staphylococcus aureus* clone USA300 in Italy: a case report. Ann Clin Microbiol Antimicrob 2008; 7: 11
- Munckhof WJ, Krishnan A, Kruger P, et al. Cavernous sinus thrombosis and meningitis from communityacquired methicillin-resistant Staphylococcus aureus infection. Intern Med J 2008; 38: 283-7
- Saiman L, O'Keefe M, Graham PL, et al. Hospital transmission of community-acquired methicillin-resistant Staphylococcus aureus among postpartum women. Clin Infect Dis 2003; 37: 1313-9
- 97. Stafford I, Hernandez J, Laibl V, et al. Communityacquired methicillin-resistant *Staphylococcus aureus*

- among patients with puerperal mastitis requiring hospitalization. Obstet Gynecol 2008; 112: 533-7
- Rutar T, Chamber HF, Crawford JB, et al. Ophthalmic manifestations of infections caused by the USA300 clone of community-associated methicillin-resistant *Staphylo*coccus aureus. Ophthalmology 2006; 113: 1455-62
- Healy CM, Hulten KG, Palazzi DL, et al. Emergence of new strains of methicillin-resistant *Staphylococcus aureus* in a neonatal intensive care unit. Clin Infect Dis 2004; 39: 1460-6
- De A Trindade P, Pacheco RL, Costa SF, et al. Prevalence of SCCmec Type IV in nosocomial bloodstream isolates of methicillin-resistant Staphylococcus aureus. J Clin Microbiol 2005; 43: 3435-7
- Regev-Yochay G, Rubinstein E, Barzilai A, et al. Methicillin-resistant *Staphylococcus aureus* in neonatal intensive care unit. Emerg Infect Dis 2005; 11: 453-56
- 102. Seybold U, Kourbatova EV, Johnson JG, et al. Emergence of community-associated methicillin-resistant *Staphylococcus aureus* USA300 genotype as a major cause of health-care-associated blood stream infections. Clin Infect Dis 2006; 42: 647-56
- 103. Davis SL, Rybak MJ, Amjad M, et al. Characteristics of patients with healthcare-associated infection due to SCCmec type IV methicillin-resistant Staphylococcus aureus. Infect Control Hosp Epidemiol 2006; 27: 1025-31
- 104. Laplante KL, Rybak MJ, Amjad M, et al. Antimicrobial susceptibility and staphylococcal chromosomal cassette mec type in community- and hospital-associated methicillin-resistant Staphylococcus aureus. Pharmacotherapy 2007; 27: 3-10
- 105. Benoit SR, Estivariz C, Mogdasy C, et al. Community strains of methicillin-resistant Staphylococcus aureus as potential cause of healthcare-associated infections, Uruguay, 2002-2004. Emerg Infect Dis 2008: 14: 1216-23
- 106. Kourbatova EV, Halvosa JS, King MD, et al. Emergence of community-associated methicillin-resistant Staphylococcus aureus USA 300 clone as a cause of health careassociated infections among patients with prosthetic joint infections. Am J Infect Control 2005; 33: 385-91
- 107. Patel M, Kumar RA, Stamm A, et al. USA300 genotype community-associated methicillin-resistant *Staphylo-coccus aureus* as a cause of surgical-site infections. J Clin Microbiol 2007; 45: 3431-3
- 108. Popovich KJ, Weinstein RA, Hota B. Are communityassociated methicillin-resistant Staphylococcus aureus (MRSA) strains replacing traditional nosocomial MRSA strains? Clin Infect Dis 2008; 46: 787-94
- 109. Patel M, Waites KB, Hoesley CJ, et al. Emergence of USA300 MRSA in a tertiary medical centre: implications for epidemiologic studies. J Hosp Infect 2008; 68: 208-13
- 110. Liu C, Graber CJ, Karr M, et al. A population-based study of the incidence and molecular epidemiology of methicillin-resistant *Staphylococcus aureus* disease in San Francisco, 2004-2005. Clin Infect Dis 2008; 46: 1637-46
- 111. Noel GJ, Bush K, Bagchi P, et al. A randomized, doubleblind trial comparing ceftobiprole medocaril with vancomycin plus ceftazidime for the treatment of patients with complicated skin and skin-structure infections. Clin Infect Dis 2008; 46: 647-55

- Cleocin (clindamycin) for injection [package insert].
 Bedford (OH): Ben Venue Laboratories, 2004
- Cubicin (daptomycin) for injection [package insert].
 Lexington (MA): Cubist Pharmaceuticals, 2007
- Vibramycin (doxycycline) [package insert]. New York: Pfizer Pharmaceuticals, 2007
- Zyvox (linezolid) [package insert]. New York: Pharmacia & Upjohn, Division of Pfizer, 2007
- Synercid (quinupristin and dalfopristin) for injection [package insert]. Bristol (TN): Monarch Pharmaceuticals, 2003
- Tygacil (tigecycline) for injection [package insert]. Philadelphia (PA): Wyeth Pharmaceuticals, 2006
- Bactrim (sulfamethoxazole and trimethoprim tablets) [package insert]. Nutley (NJ): Roche Pharmaceuticals, 2002
- 119. Rybak M, Lomaestro B, Rotschafer JC, et al. Therapeutic monitoring of vancomycin in adult patients: a consensus review of the American Society of Health-System Pharmacists, the Infectious Diseases Society of America, and the Society of Infectious Diseases Pharmacists. Am J Health-Syst Pharm 2009; 66: 82-98
- Ruhe JJ, Monson T, Bradsher RW, et al. Use of long-acting tetracyclines for methicillin-resistant *Staphylococcus aureus* infections: case series and review of the literature. Clin Infect Dis 2005; 40: 1429-34
- 121. Clinical and Laboratory Standards Institute/NCCLS. Methods for antimicrobial susceptibility testing for bacteria that grow aerobically. CLSI/NCCLS M100-S14. Wayne (PA): National Committee for Clinical Laboratory Standards, 2004
- 122. Chavez-Bueno S, Bozdogan B, Katz K, et al. Inducible clindamycin resistance and molecular epidemiologic trends of pediatric community-associated methicillinresistant *Staphylococcus aureus* in Dallas, Texas. Antimicrob Agents Chemother 2005; 49: 2283-88
- McGehee RF, Barrett FF, Finland M. Resistance of Staphylococcus aureus to lincomycin, clindamycin, and erythromycin. Antimicrob Agents Chemother 1968; 8: 392-7
- Rao GG. Should clindamycin be used in treatment of patients with infections caused by erythromycin-resistant staphylococci? [letter]. J Antimicrob Chemother 2000; 45: 715
- Drinkovic D, Fuller ER, Shore KP, et al. Clindamycin treatment of *Staphylococcus aureus* expressing inducible clindamycin resistance. J Antimicrob Chemother 2001; 48: 315-6
- Frank AL, Marcinak JF, Mahgat PD, et al. Clindamycin treatment of methicillin-resistant *Staphylococcus aureus* infections in children. Pediatr Infect Dis J 2002; 21: 530-34
- 127. Levin TP, Suh B, Axelrod P, et al. Potential clindamycin resistance in clindamycin-susceptible, erythromycin-resistant Staphylococcus aureus: report of a clinical failure. Antimicrob Agents Chemother 2005; 49: 1222-4
- 128. Siberry GK, Tekle T, Carroll K, et al. Failure of clindamycin treatment of methicillin-resistant *Staphylococcus aureus* expressing inducible clindamycin resistance in vitro. Clin Infect Dis 2003; 37: 1257-60
- LaPlante KL, Leonard SN, Andes DR, et al. Activities of clindamycin, daptomycin, doxycycline, linezolid,

- trimethoprim-sulfamethoxazole, and vancomycin against community-associated methicillin-resistant *Staphylococcus aureus* with inducible clindamycin resistance in murine thigh infection and in vitro pharmocodynamic models. Antimicrob Agents Chemother 2008; 52: 2156-62
- Markowitz N, Quinn EL, Saravolatz LD. Trimethoprimsulfamethoxazole compared with vancomycin for the treatment of *Staphylococcus aureus* infection. Ann Intern Med 1992; 117: 390-8
- Kaka AS, Rueda AM, Shelburn 3rd SA, et al. Bactericidal activity of oral agents against methicillin-resistant Staphylococcus aureus. J Antimicrob Chemother 2006; 58: 680-3
- Berg T, Firth N, Apisiridej S, et al. Complete nucleotide sequence of pSK41: evolution of staphylococcal conjugative plasmids. J Bacteriol 1998; 180: 4350-9
- Stokes A, Lacey RW. Effect of thymidine on activity of trimethoprim and sulphamethoxazole. J Clin Pathol 1978; 31: 165-71
- 134. Hamilton-Miller JM. Reversal of activity of trimethoprim against Gram-positive cocci by thymidine, thymine, and 'folates'. J Antimicrob Chemother 1988; 22: 35-9
- 135. Mendes RE, Sader HS, Deshpande L, et al. Antimicrobial activity of tigecycline against community-acquired methicillin-resistant Staphylococcus aureus isolates recovered from North American medical centers. Diagn Microbiol Infect Dis 2008; 60: 433-6
- Szumowski JD, Cohen DE, Kanaya F, et al. Treatment and outcomes of infections by methicillin-resistant *Staphylo-coccus aureus* at an ambulatory clinic. Antimicrob Agents Chemother 2007; 51: 423-28
- 137. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. CLSI approved standard M100-S16. Wayne (PA): Clinical and Laboratory Standards Institute, 2006
- 138. Jones RN. Microbiologic features of vancomycin in the 21st century: minimum inhibitory concentration creep, bactericidal/static activity, and applied breakpoints to predict clinical outcomes or detect resistant strains. Clin Infect Dis 2006; 42 Suppl. 1: S13-24
- 139. Wang G, Hindler JF, Ward KW, et al. Increased vancomycin MICs for *Staphylococcus aureus* clinical isolates from a university hospital during a 5-year period. J Clin Microbiol 2006; 44: 3883-6
- 140. Steinkraus G, White R, Friedrich L. Vancomycin MIC creep in non-vancomycin-intermediate Staphylococcus aureus (VISA), vancomycin-susceptible clinical methicillin-resistant S. aureus (MRSA) blood isolates from 2001-2005. J Antimicrob Chemother 2007; 60: 788-94
- 141. Rybak MJ, Leonard SN, Rossi KL, et al. Characterization of vancomycin-heteroresistant *Staphylococcus aureus* from the metropolitan area of Detroit, Michigan, over a 22year period (1986-2007). J Clin Microbiol 2008; 46: 2950-4
- 142. Moise-Broder PA, Sakoulas G, Eliopoulas J, et al. Accessory gene regulator group II polymorphism in methicillin-resistant *Staphylococcus aureus* is predictive of failure of vancomycin therapy. Clin Infect Dis 2004; 38: 1700-5
- 143. Sakoulas G, Moise-Broder PA, Schentag J, et al. Relationship of MIC and bactericidal activity to efficacy of vancomycin for treatment of methicillin-resistant Staphylococcus aureus bacteremia. J Clin Microbiol 2004; 42: 2398-402

- 144. Soriano A, Marco F, Martinez JA, et al. Influence of vancomycin minimum inhibitory concentration on the treatment of methicillin-resistant *Staphylococcus aureus* bacteremia. Clin Infect Dis 2008; 46: 193-200
- 145. Lodise TP, Graves J, Evans A, et al. Relationship between vancomycin MIC and failure among patients with methicillin-resistant *Staphylococcus aureus* bacteremia treated with vancomycin. Antimicrob Agents Chemother 2008; 52: 3315-20
- 146. Graber CJ, Wong MK, Carleton HA, et al. Intermediate vancomycin susceptibility in a community-associated MRSA clone. Emerg Infect Dis 2007; 13: 491-3
- 147. Chua T, Moore CL, Perri MB, et al. Molecular epidemiology of methicillin-resistant *Staphylococcus aureus* bloodstream isolates in urban Detroit. J Clin Microbiol 2008; 2345-52
- Hidayat LK, Hsu DI, Quist R, et al. High-dose vancomycin therapy for methicillin-resistant *Staphylococcus aureus* infections. Arch Intern Med 2006; 166: 2138-44
- Silverman JA, Perlmutter NG, Shapiro HM. Correlation of daptomycin bactericidal activity and membrane depolarization in *Staphylococcus aureus*. Antimicrob Agents Chemother 2003; 47: 2538-44
- Richter SS, Kealey DE, Murray CT, et al. The in vitro activity of daptomycin against *Staphylococcus aureus* and *Enterococcus* species. J Antimicrob Chemother 2003; 52: 123-7
- 151. Critchley IA, Draghi DC, Sahm DF, et al. Activity of daptomycin against susceptible and multidrug-resistant gram-positive pathogens collected in the SECURE study (Europe) during 2000-2001. J Antimicrob Chemother 2003; 51: 639-49
- Mangali A, Bica I, Snydman R, et al. Daptomycin-resistant methicillin-resistant *Staphylococcus aureus* bacteremia. Clin Infect Dis 2005; 40: 1058-60
- 153. Marty FM, Yeh WW, Wennersten CB, et al. Emergence of a clinical daptomycin-resistant *Staphylococcus aureus* isolate during treatment of methicillin-resistant *Staphylococcus aureus* bacteremia and osteomyelitis. J Clin Microbiol 2006; 44: 595-7
- 154. Pillai SK, Gold HS, Sakoulas G, et al. Daptomycin nonsusceptibility in *Staphylococcus aureus* with reduced vancomycin susceptibility is independent of alterations in MprF. Antimicrob Agents Chemother 2007; 51: 2223-5
- 155. Huang Y, Hsiao C, Liao C, et al. Bacteremia and infective endocarditis caused by a non-daptomycin-susceptible, vancomycin-intermediate, and methicillin-resistant Staphylococcus aureus strain in Taiwan. J Clin Microbiol 2008; 46: 1132-6
- Sakoulas G, Rose W, Rybak MJ, et al. Evaluation of endocarditis caused by methicillin-susceptible *Staphylo-coccus aureus* developing nonsusceptibility to daptomycin. J Clin Microbiol 2008; 46: 220-4
- Murthy MH, Olson ME, Wickert RW, et al. Daptomycin non-susceptible methicillin-resistant *Staphylococcus aureus* USA 300 isolate. J Med Microbiol 2008; 57: 1036-8
- Arbeit RD, Maki D, Tally FP, et al. The safety and efficacy of daptomycin for the treatment of complicated skin and skin-structure infections. Clin Infect Dis 2004; 38: 1673-81

- Fowler Jr VG, Boucher HW, Corey GR, et al. Daptomycin versus standard therapy for bacteremia and endocarditis caused by *Staphylococcus aureus*. N Engl J Med 2006; 355: 653-65
- 160. Lamp KC, Friedrick LV. Clinical experience with daptomycin for the treatment of osteomyelitis in patients with post-therapy follow-up. 46th Interscience Conference on Antimicrobial Agents and Chemotherapy; 2006 Sep 27-30; San Francisco (CA), L-1557
- Finney MS, Crank CW, Segreti J. Use of daptomycin to treat drug-resistant gram-positive bone and joint infections. Curr Med Res Opin 2005; 21: 1923-26
- 162. Forrest G, Donovan B, Lamp K, et al. Daptomycin use in patients with septic arthritis: post-marketing experience from CORE 2005. 46th Interscience Conference on Antimicrobial Agents and Chemotherapy; 2006 Sep 27-30; San Francisco (CA), A-1556
- Rao N, Regalla DM. Uncertain efficacy of daptomycin for prosthetic joint infections: a prospective case series. Clin Orthop Relat Res 2006; 451: 34-7
- 164. Silverman JA, Mortin LI, Vanpraagh AD, et al. Inhibition of daptomycin by pulmonary surfactant: in vitro modeling and clinical impact. J Infect Dis 2005; 191: 2149-52
- 165. Wunderink RG, Rello J, Cammarata SK, et al. Linezolid vs vancomycin: analysis of two double-blind studies of patients with methicillin-resistant *Staphylococcus aureus* nosocomial pneumonia. Chest 2003; 124: 1789-97
- Cruciani M, Gattie G, Lazzarini L, et al. Penetration of vancomycin into human lung tissue. J Antimicrob Chemother 1996; 38: 865-9
- Honeybourne D, Tobin C, Jevons G, et al. Intrapulmonary penetration of linezolid. J Antimicrob Chemother 2003; 51: 1431-4
- 168. Boselli E, Breilh D, Rimmelé T, et al. Pharmacokinetics and intrapulmonary concentrations of linezolid administered to critically ill patients with ventilator-associated pneumonia. Crit Care Med 2005; 33: 1529-33
- 169. Wunderink RG, Mendelson MH, Somero MS, et al. Early microbiological response to linezolid versus vancomycin in ventilator-associated pneumonia due to methicillinresistant Staphylococcus aureus. Chest 2008; 134: 1200-7
- 170. Stevens DL, Ma Y, Salmi DB, et al. Impact of antibiotics on expression of virulence-associated exotoxins genes in methicillin-sensitive and methicillin-resistant *Staphylo*coccus aureus. J Infect Dis 2007; 195: 202-11
- Shorr AF, Kunkel MJ, Kollef M. Linezolid versus vancomycin for *Staphylococcus aureus* bacteraemia: pooled analysis of randomized studies. J Antimicrob Chemother 2005; 56: 923-9
- 172. Wilcox MH, Tack KJ, Bouza E, et al. Complicated skin and skin-structure infections and catheter-related blood-stream infections: noninferiority of linezolid in a phase 3 study. Clin Infect Dis 2009; 48: 203-12
- 173. Wu VC, Wang YT, Wang CY, et al. High frequency of linezolid-associated thrombocytopenia and anemia among patients with end-stage renal disease. Clin Infect Dis 2006: 42: 66-72
- Lawrence KR, Adra M, Gillman PK. Serotonin toxicity associated with the use of linezolid: a review of postmarketing data. Clin Infect Dis 2006; 42: 1578-83

- 175. Taylor JJ, Wilson JW, Estes LL. Linezolid and serotonergic drug interactions: a retrospective survey. Clin Infect Dis 2006; 43: 180-7
- Lee E, Burger S, Shah J, et al. Linezolid-associated toxic optic neuropathy: a report of 2 cases. Clin Infect Dis 2003; 37: 1389-91
- 177. Frippiat F, Bergiers C, Michel C, et al. Severe bilateral optic neuritis associated with prolonged linezolid therapy. J Antimicrob Chemother 2004; 53: 1114-5
- Palenzuela L, Hahn NM, Nelson Jr RP, et al. Does linezolid cause lactic acidosis by inhibiting mitochondrial protein synthesis? Clin Infect Dis 2005; 40: e113-6
- Leclerq R, Courvalin P. Bacterial resistance to macrolides, lincosamide, and streptogramin antibiotics by target modification. Antimicrob Agents Chemother 1991; 35: 1267-72
- 180. Drew RH, Perfect JR, Srinath L, et al. Treatment of methicillin-resistant Staphylococcus aureus infections with quinupristin-dalfopristin in patients intolerant of or failing prior therapy. J Antimicrob Chemother 2000; 46: 775-84
- Olsen KM, Rebuck JA, Rupp ME. Arthralgias and myalgias related to quinupristin-dalfopristin administration. Clin Infect Dis 2001; 32: e83-6
- Projan SJ. Preclinical pharmacology of GAR-936, a novel glycylcycline antibacterial agent. Pharmacotherapy 2000; 20: 219S-23S
- Noskin GA. Tigecycline: a new glycylcycline for treatment of serious infections. Clin Infect Dis 2005; 41: S303-14
- Meagher AK, Ambrose PG, Grasela TH, et al. The pharmacokinetic and pharmocodynamic profile of tigecycline. Clin Infect Dis 2005; 41: S333-40
- 185. Ellis-Grosse EJ, Babinchak T, Dartois N, et al. The efficacy and safety of tigecycline in the treatment of skin and skinstructure infections: results of 2 double-blind phase 3 comparison studies with vancomycin-aztreonem. Clin Infect Dis 2005; 41: S341-53
- 186. McAleese F, Murphy E, Babinchak T, et al. Use of ribotyping to retrospectively identify methicillin-resistant Staphylococcus aureus isolates from phase 3 clinical trials for tigecycline that are genotypically related to community-associated isolates. Antimicrob Agents Chemother 2005; 49: 4521-9
- 187. Bogdanovich T, Ednie LM, Shapiro S, et al. Antistaphylococcal activity of ceftobiprole, a new broadspectrum cephalosporin. Antimicrob Agents Chemother 2005; 49: 4210-9
- Leonard SN, Cheung CM, Rybak MJ. Activities of ceftobiprole, linezolid, vancomycin, and daptomycin against community-associated and hospital-associated methicillin-resistant Staphylococcus aureus. Antimicrob Agents Chemother 2008; 52: 2974-6
- Blumberg HM, Rimland D, Carroll DJ, et al. Rapid development of ciprofloxacin resistance in methicillin-susceptible and -resistant *Staphylococcus aureus*. J Infect Dis 1991; 163: 1279-85
- Peterson LR, Quick JN, Jensen B, et al. Emergence of ciprofloxacin resistance in nosocomial methicillin-resistant Staphylococcus aureus isolates: resistance during

- ciprofloxacin plus rifampin therapy for methicillin-resistant *S. aureus* colonization. Arch Intern Med 1990; 150: 2151-5
- 191. Eng RHK, Smith SM, Buccini FJ, et al. Differences in ability of cell-wall antibiotics to suppress emergence of rifampicin resistance in *Staphylococcus aureus*. J Antimicrob Chemother 1985; 15: 201-7
- Iyer S, Jones DH. Community-acquired methicillinresistant Staphylococcus aureus skin infection: a retrospective analysis of clinical presentation and treatment of a local outbreak. J Am Acad Dermatol 2004; 50: 854-8
- Lin G, Credito K, Ednie LM, et al. Antistaphylococcal activity of dalbavancin, and experimental glycopeptide. Antimicrob Agents Chemother 2005; 49: 770-2
- 194. Jauregi LE, Babazadeh S, Seltzer E, et al. Randomized, double-blind comparison of once-weekly dalbavancin versus twice-daily linezolid therapy for the treatment of complicated skin and skin structure infections. Clin Infect Dis 2005; 41: 1407-15
- 195. Raad I, Darouiche R, Vazquez J, et al. Efficacy and safety of weekly dalbavancin therapy for catheter-related bloodstream infection caused by Gram-positive pathogens. Clin Infect Dis 2005; 40: 374-80
- Kanafani ZA. Telavancin: a new lipoglycopeptide with multiple mechanisms of action. Expert Rev Anti Infect Ther 2006; 4: 743-9
- 197. Stryjewski M, Chu VH, O'Riordan W, et al. Telavancin versus standard therapy for treatment of complicated skin and skin structure infections caused by Gram-positive bacteria: FAST 2 study. Antimicrob Agents Chemother 2006; 50: 862-7
- 198. Fowler Jr VG, Rude TH, Nelson CL, et al. Activity of telavancin against Staphylococcus aureus isolates carrying the Panton-Valentine leukocidin gene in the ATLAS studies [abstract 847]. In: Program and abstracts of the 17th European Congress of Clinical Microbiology and Infectious Diseases (Munich). Oxford: Blackwell Publishing, 2007
- 199. Giamarellou H, O'Riordon W, Harris H, et al. Phase 3 trial comparing 3-7 days of oritavancin vs. 10-14 days of vancomycin/cephalexin in the treatment of patients with complicated skin and skin structure infections (cSSSI) [abstract]. In: Program and abstracts of the 43rd Interscience Conference on Antimicrobial Agents and Chemotherapy (Chicago). Washington, DC: American Society for Microbiology, 2003
- 200. Wasilewski M, Dish D, McGill J, et al. Equivalence of shorter course of therapy with oritavancin vs. vancomycin/ cephalexin in complicated skin and skin structure infections (cSSSI) [abstract UL-18]. In: Program and abstracts of the 41st Interscience Conference on Antimicrobial Agents and Chemotherapy (Chicago). Washington, DC: American Society for Microbiology, 2001
- Kim SJ, Cegelski L, Stueber D, et al. Oritavancin exhibits dual mode of action to inhibit cell-wall biosynthesis in Staphylococcus aureus. J Mol Biol 2008; 14: 281-93
- Ward KE, Mersfelder TL, LaPlante KL. Oritavancin: an investigational glycopeptide antibiotic. Expert Opin Investig Drugs 2006; 15: 417-29
- Cerexa, Inc. Comparative study of seftaroline vs. vancomycin plus aztreonam in adult subjects with complicated

- skin infections [ClinicalTrials.gov identifier NCT 00423657]. US National Institutes of Health, Clinical Trials.gov [online]. Available from URL: http://www.clinicaltrials.gov [Accessed 2009 Apr 2]
- 204. Cerexa, Inc. Comparative study of ceftaroline vs. vancomycin plus aztreonam in adult subjects with complicated skin infections (cSSSI) [ClinicalTrials.gov identifier NCT00424190]. US National Institutes of Health, Clinical Trials.gov [online]. Available from URL: http://www. clinicaltrials.gov [Accessed 2009 Apr 2]
- 205. Cerexa, Inc. Efficacy and safety of ceftaroline versus linezolid in subjects with complicated skin and skin structure infections [ClinicalTrials.gov identifier NCT00633152]. US National Institutes of Health, ClinicalTrials.gov [online]. Available from URL: http://www.clinicaltrials.gov [Accessed 2009 Apr 2]
- 206. Arpida AG. Study of intravenous (I.V.) iclaprim versus linezolid in complicated skin and skin structure infections [cSSSI] (ASSIST-2) [ClinicalTrials.gov identifier NCT 00303550]. US National Institutes of Health, Clinical Trials.gov [online]. Available from URL: http://www.clinicaltrials.gov [Accessed 2009 Apr 2]
- 207. Arpida AG. Phase 3 safety and efficacy study of I.V. iclaprim v linezolid in cSSSI (ASSIST-1) [ClinicalTrials.gov identifier NCT00299520]. US National Institutes of Health, ClinicalTrials.gov [online]. Available from URL: http://www.clinicaltrials.gov [Accessed 2009 Apr 2]
- 208. Arpida AG. Clinical efficacy of intravenous iclaprim versus vancomycin in the treatment of hospital-acquired, ventilator-associated, or health-care-associated pneumonia [ClinicalTrials.gov identifier NCT00543608]. US National Institutes of Health, ClinicalTrials.gov [online]. Available from URL: http://www.clinicaltrials.gov [Accessed 2009 Apr 2]
- 209. Lee MC, Rios AM, Aten MF, et al. Management and outcome of children with skin and soft-tissue abscesses caused by community-acquired methicillin-resistant Staphylococcus aureus. Pediatr Infect Dis J 2004; 23: 123-7
- Giordano PA, Elston D, Akinlade BK, et al. Cefdinir vs cephalexin for mild to moderate uncomplicated skin and skin structure infections in adolescents and adults. Curr Med Res Opin 2006; 22: 2419-28
- 211. Rajendran PM, Young D, Maurer T, et al. Randomized, double-blind, placebo-controlled trial of cephalexin for treatment of uncomplicated skin abscesses in a population at risk for community-acquired methicillin-resistant Staphylococcus aureus infection. Antimicrob Agents Chemother 2007; 51: 4044-8
- 212. Stevens DL, Bisno AL, Chambers HF, et al. Practice guidelines for the diagnosis and management of skin and soft-tissue infections. Clin Infect Dis 2005; 41: 1373-406
- Chambers HF, Moellering Jr RC, Kamitsuka P. Management of skin and soft-tissue infection. N Engl J Med 2008; 359: 1063-7
- 214. Gorwitz RJ, Jernigan DB, Powers JH, et al. and Participants in the CDC Convened Experts' Meeting on Management of MRSA in the Community. Strategies for clinical management of MRSA in the community: summary of an experts' meeting. convened by the Centers for Disease Control and Prevention 2006 [online]. Available

- from URL: http://www.cdc.gov/ncidod/dhqp/pdf/ar/CAMRSA_ExpMtgStrategies.pdf [Accessed 2008 Dec 14]
- 215. Centers for Disease Control and Prevention. Outpatient management of skin and soft tissue infections in the era of community-associated MRSA [online]. Available from URL: http://www.cdc.gov/ncidod/dhqp/ar_mrsa_ca_skin. html [Accessed 2009 Feb 19]
- 216. Miller LG, Diep BA. Colonization, fomites, and virulence: rethinking the pathogenesis of community-associated

methicillin-resistant *Staphylococcus aureus* infection. Clin Infect Dis 2008; 46: 752-60

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