

EFFICIENT SYNTHESIS OF 3-FORMYL-1,2,4-TRIAZOLE NUCLEOSIDE
USING DIETHOXYACETONITRILE AS A SYNTHON[†]

Teiichi Murakami, Masami Otsuka, Susumu Kobayashi, and Masaji
Ohno*

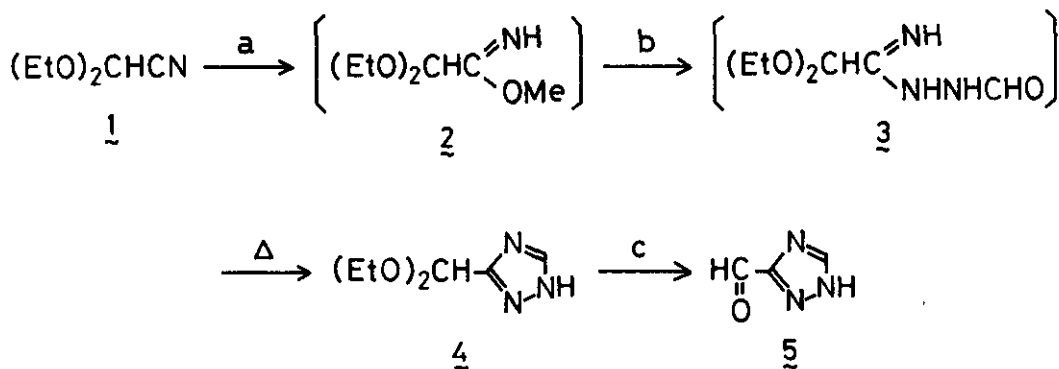
Faculty of Pharmaceutical Sciences, University of Tokyo,
Bunkyo-ku, Tokyo 113, Japan

Abstract — Diethoxyacetonitrile has been shown to be an efficient and versatile synthon for the synthesis of 1,2,4-triazole heterocycle (5), and 1- β -D-ribofuranosyl-1,2,4-triazole-3-carboxaldehyde (10) has been synthesized regiospecifically for the nitrogens and stereospecifically for the ribosylation.

Certain azole nucleosides of both natural and synthetic origin have shown significant biological activity.¹ Among these are the C-nucleoside antibiotic pyrazomycin² and the synthetic nucleoside ribavirin.³ Furthermore, it was recently disclosed that some nucleosides contain a formyl group in the base moiety. Nikkomycin X contains a 4-formyl-4-imidazolin-2-one moiety⁴ and polyoxin N contains 3-formyl-4-hydroxypyrazole.⁵ The objective of the investigation described herein was to develop a new methodology for the conversion of diethoxyacetonitrile to heterocyclic moiety of nucleosides structurally related to these biologically active compounds. Diethoxyacetonitrile is easily prepared from triethyl orthoformate and hydrogen cyanide⁶ or triethyl orthoformate and acetyl cyanide⁷ or ethyl diethoxyacetate by treatment with ammonia followed by POCl_3 .⁸ However, no systematic investigation has been carried out for the preparation of heterocyclic compounds using diethoxyacetonitrile as a synthon. Thus, we became interested in the selective conversion of the cyano group into various heterocyclic compounds, especially 1,2,4-triazole nucleoside of increasing importance.

[†] Dedicated to Professor Dr. Tetsuji Kametani, the Pharmaceutical Institute, Tohoku University, in commemoration of his retirement from the University.

Chart 1



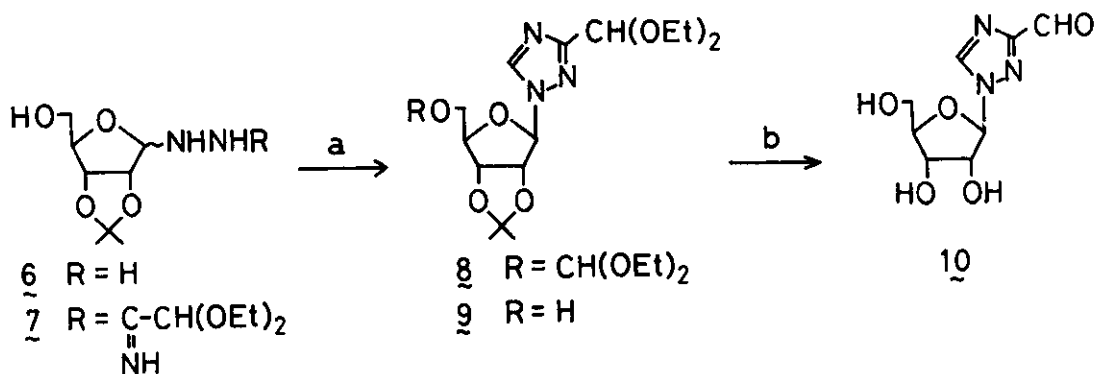
a, NaOMe / MeOH ; b, H₂NNHCHO ; c, 1N-HCl / Me₂CO

Methyl diethoxyacetimidate (2) was prepared from diethoxyacetonitrile (1) in the presence of a catalytic amount of sodium methoxide in methanol at room temperature for 3 h under argon atmosphere, and the resulting solution was treated with equiv of formylhydrazine dissolved in methanol at room temperature for 12 h, depositing colorless crystals 3.⁹ Then, the reaction mixture was heated at reflux for 14 h. After neutralization with AcOH and removal of the solvent, a pale pink solid was obtained. It was purified by column chromatography on silica gel (Et₂O:CH₃COCH₃ = 8:1) and afforded a colorless crystalline 1,2,4-triazole-3-carboxaldehyde diethyl acetal (4) in 90% yield, mp 68-70°C (recrystallized from n-hexane-ether). The acetal 4 was hydrolyzed to 1,2,4-triazole-3-carboxaldehyde¹⁰ (5, colorless foam, 93% yield) by treatment with 1N HCl in acetone at reflux for 3 h followed by neutralization with NaHCO₃, workup and chromatography on silica gel (CH₂Cl₂:MeOH = 5:1). The aldehyde 5 can be purified by sublimation, showing yellow at 180°C with mp 200-202°C (dec.), and exhibits infrared carbonyl band (KBr) at 1694 cm⁻¹, and ¹H-NMR in CD₃OD shows hemiacetal proton at δ 5.72 and an aromatic proton at δ 8.20, but ¹H-NMR in DMSO-d₆ shows aldehyde proton at δ 9.98 and an aromatic proton at δ 8.78. The mass spectrum shows molecular ions corresponding to the aldehyde (M⁺, 97), and was not to be dimeric hemiaminals.¹¹

The present methodology essentially consists of a two-step reaction affording

1,2,4-triazole-3-carboxaldehyde in excellent overall yield (Chart 1), and can be applied to the synthesis of 3-substituted 1,2,4-triazole system. Although the most successful glycosylation is based on the acid-catalyzed fusion procedure¹² or trimethylsilyl derivatives,³ it is recognized that the isomeric distribution varies greatly with the substituents on the heterocycle. Therefore, hydrazino-ribose was used to obtain 3-formyl-1,2,4-triazole nucleoside regioselectively and to take advantage of diethoxyacetonitrile as an efficient and versatile synthon. The hydrazone 6 of 2,3-O-isopropylidene-D-ribose was prepared from 2,3-O-isopropylidene-D-ribose and hydrazine according to Schmidt's method¹³. (Chart 2) The

Chart 2



a. HC(OEt)₃ on 7 ; b. HCO₂H on 9

acetimidate 2 (1.5 equiv) prepared as described above was gradually added to a methanol solution of the hydrazone 6, and the reaction mixture was stirred at room temperature for 3 h. A yellowish syrup (7) was obtained upon workup and directly treated with triethyl orthoformate, and the resulting solution was heated at reflux (120°C) for 1.5 h. After workup and complete removal of the excess orthoformate, and orange-red syrup was obtained and confirmed to be mainly 8, spectroscopically (¹H-NMR). The primary alcohol of 9 was generated by treatment with ethanol by heating at reflux for 8 h, affording a deep-red syrup upon workup. It was subjected to column chromatography on silica gel (Et₂O → AcOEt) and afforded a colorless syrup 9 in 39% overall yield from 6, showing M⁺+1, 344, and satisfactory IR, ¹H- and ¹³C-NMR data. The diethoxyl and isopropylidene groups were hydrolyzed together by treatment with aqueous formic acid at 40°C for 10 h.

1-β-D-Ribofuranosyl-1,2,4-triazole-3-carboxaldehyde (10) was obtained in 68% yield as a colorless foam after workup, chromatography on silica gel (CH₂Cl₂:MeOH=5:1), and complete removal of methanol under reduced pressure.¹⁰ The nucleoside 10 was converted to the oxime¹⁰ (mp 151-153°C, M⁺+1 245). Evidence for the structure of the product 10 as β-configuration was obtained by comparison of the δ value (5.87 in CD₃OD/TMS) and the coupling constant J_{1',2'} (3.5 Hz) for the anomeric proton with those of the triazole carboxamide nucleosides of known structure.¹⁴ Formation of the α anomer corresponding to 8, 9 and 10 was not detected in this procedure.

Acknowledgement: The work was financially supported in part by Grants-in-aid for Special Project Research from the Ministry of Education, Science and Culture of Japan.

REFERENCES AND NOTES

1. J. T. Witkowski and R. K. Robins, 'Chemistry and Biology of Nucleosides and Nucleotides', Academic Press, New York, 1979, p. 267 and references therein.
2. R. J. Suhadolnik, 'Nucleoside Antibiotics', Wiley-Interscience, New York, 1970, p. 390.
3. J. T. Witkowski, R. K. Robins, R. W. Sidwell, and L. N. Simon, J. Med. Chem., 1972, 15, 1150.
4. H. Hagenmaier and A. Keckeisen, Liebigs Ann. Chem., 1979, 1994.
5. M. Uramoto, J. Uzawa, S. Suzuki, K. Isono, J. G. Liehr, and J. A. McCloskey, Nucleic Acid Research, special publication No. 5, 1979, 327.
6. J. G. Erickson, J. Am. Chem. Soc., 1951, 73, 1338.
7. H. Böhme and R. Neidlein, Liebigs Ann. Chem., 1962, 1859.
8. T. Shiba, K. Inami, and K. Sawada, Heterocycles, 1979, 13, special issue, 175.
9. Compounds 2 and 3 were treated for further reaction without isolation, but they were easily confirmed to have the assigned structures by ¹H-NMR.
10. Satisfactory elemental analysis, IR, ¹H- & ¹³C-NMR and MS data were obtained.
11. E. J. Borowne, Aust. J. Chem., 1971, 24, 393.
12. L. Goodman, 'Basic Principles in Nucleic Acid Chemistry, Vol. 1', Academic Press, New York, 1974 p. 102 and references therein.
13. R. R. Schmidt, J. Karg, and W. Guilliard, Angew. Chem., internat, edit., 1975, 14, 64, see also ref. 6.
14. S. R. Naik, J. T. Witkowski, and R. K. Robins, J. Heterocyclic Chem., 1974, 11, 57.

Received, 23th July, 1980