

C(3)-ALKOXYCEPHEMS

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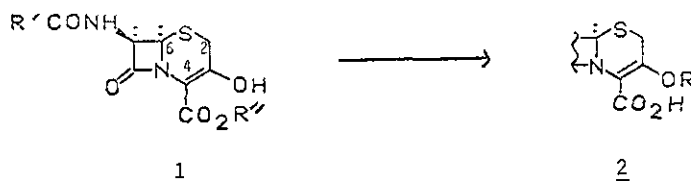
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Abstract - The C(3)-hydroxycephems 1 undergo the Mitsunobu reaction to give a series of heretofore unavailable C(3)-alkoxycephems 2.

Several research groups have reported the synthesis and activity of C(3)-alkoxy cephems 2, R being Me, Et, n-Bu, and CH₂Ph.^{1,2}

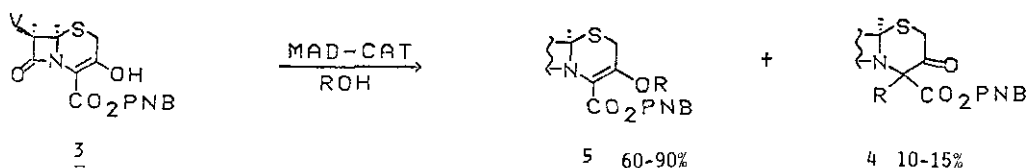


In general, these syntheses involve O-alkylation of the C(3)-hydroxycephem 1 with dimethyl sulfate, Meerwein's reagent, triazenes and the use of diazo derivatives.

We were interested in determining the *in vitro* and *in vivo* effect of lipophilic groups at the C(3)-position in the cephem molecule and needed a general, simple synthesis of C(3)-alkyl ethers.³ Use of a Williamson type ether synthesis was found not to be applicable, leading instead to alkylation at C(4) using either the cephem sulfide 1 or its sulfoxide.

The Mitsunobu reaction consists of an oxidative-reductive-dehydrative coupling in the presence of trivalent phosphorous and an azodicarboxylate, generally involving an inter or intramolecular dehydration between an alcohol and an acidic component.⁴ β -Keto esters have been reported to give low yields of the O-alkylated product. Mitsunobu, for example, reported the O-alkylation of ethyl acetoacetate under DEAD-CAT (diethyl azodicarboxylate-triphenylphosphine) conditions using n-propanol to give 18-37% O-alkylated product, depending on the reaction conditions.^{4,5}

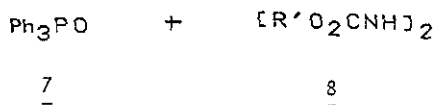
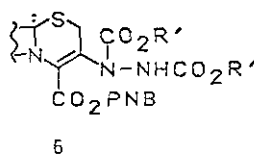
Application of the Mitsunobu reaction to the C(3)-hydroxycephem 3 using DEAD-CAT/MeOH gave, after silica chromatography, 60% C(3)-OMe, 5 (R=Me), identical to the product of 3 with diazomethane, and 10% C-alkylated product 4 (R=Me). The actual yield of the O-methylated product was somewhat higher, but we encountered problems separating the ether from the ethyl hydrazodicarboxylate by-product 8 (R'=Et). This problem was resolved by using dimethyl azodicarboxylate (MAD)⁶, whose by-product is moderately soluble in water⁷ and thus can be eliminated from concentrated ethyl acetate or methylene chloride by repeated (3-4 times) aqueous washings. In addition MAD appeared to be more reactive⁷ than DEAD and gave higher yields in certain cases. We thus reacted the enol 3 under MAD-CAT conditions (THF or CH₂Cl₂, r.t., 20 min.) with a series of alcohols (methanol to n-octanol) to give 60-90% of the O-alkylated product 5 plus 10-15% of the C-alkylated product 4.



V = PhOCH₂CONH-

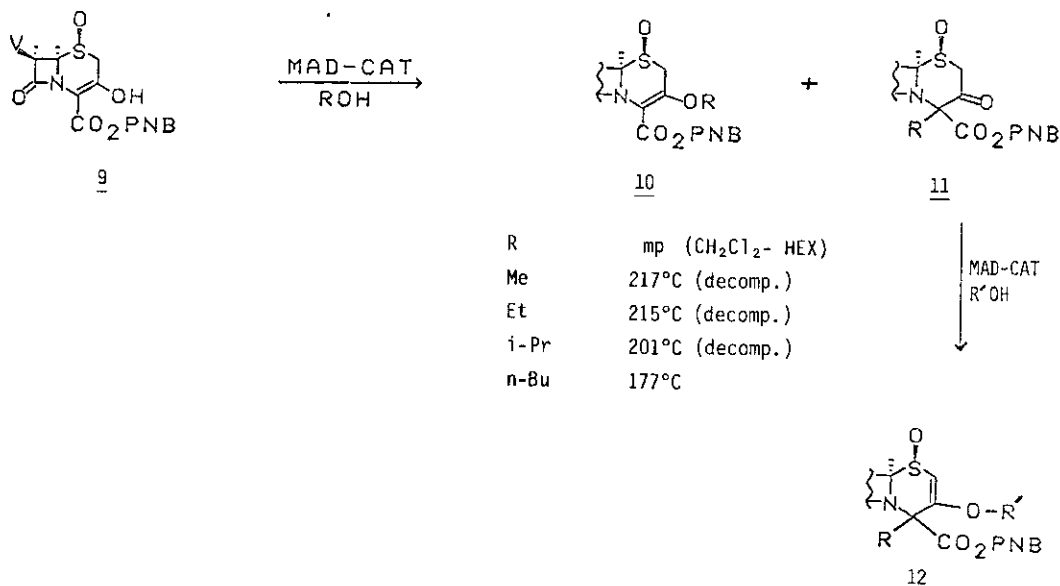
PNB = p-O₂NC₆H₄CH₂-

MAD-CAT = [MeO₂C-N=]₂/Ph₃P



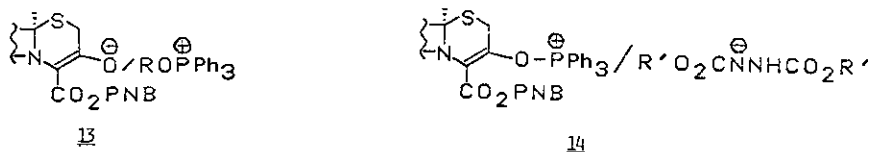
The C-alkylated product 4 was distinguishable from 5 by NMR (the chemical shift of the methylene alpha to the oxygen vs. carbon, and the difference in the methylene at C(2), i.e. the ketone 4 showed a broad AB, ca. 3.0 and 3.7 ppm, J = 14 HZ, while 5 showed a singlet at ca. 3.4 ppm) and by ketone carbonyl absorption at ca. 1729 cm⁻¹ in the i.r. spectrum.

The enol sulfoxide 9 under the MAD-CAT conditions also gave a mixture with a predominance of O-alkylation. This was determined by PBr₃ sulfoxide reduction followed by silica chromatography of the sulfides 4 and 5. The C-alkylated sulfoxide 11 further undergoes the Mitsunobu reaction to give the dialkylated Δ²-derivative 12.



The C(3)-hydrazino cephem derivative 6 (R'=Me) was observed when secondary alcohols were used or alternatively, when no alcohol was present.

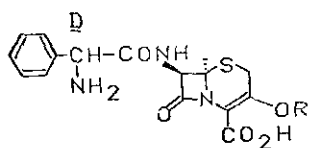
¹⁸O-Methanol studies⁸ show that the alkoxy phosphonium salt 13 derived from the alcohol is the intermediate for both O- and C-alkylation while the cephem alkoxy phosphonium salt 14 is believed to be the intermediate to the C(3)-hydrazinocephem 6.⁵



Use of trialkylphosphite⁴-MAD works equally well with the enol cephem. Thus the reaction of 3 with tri-n-butylphosphite-MAD gave 82% of O-alkylated n-Bu cephem 5, (R=n-Bu) while triethyl phosphite gave 77% of the O-ethyl derivative 5, (R=Et). PCl₅ cleavage⁸ of the side chain of 5 followed by acylation and deblocking led to the phenyl glycine acids 15. Alternatively, use of the benzhydryl-t-boc-phenyl glycine-C(3)-enol as the Mitsunobu substrate followed by deblocking (HCO₂H) gave the same acids.

MIC data (Table I) on these acids in general show that as the lipophilic character at C(3) increases the gram positive activity is enhanced with concomitant loss in gram negative activity. A similar trend was seen with the C(3)-alkylcephems.³

TABLE I



15

MIC ($\mu\text{g/ml}$)

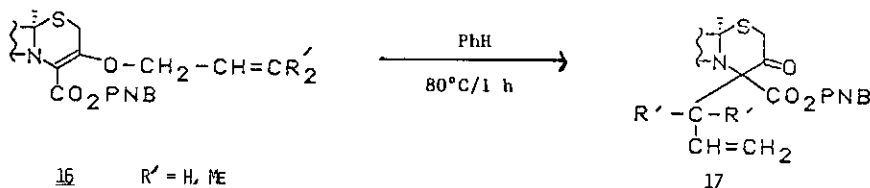
R	<u>S. aureus.</u>		<u>S. epidermidis</u>	<u>H. influenzae</u>	<u>E. coli</u>
			(amp ^s)		
	V41	S13E	222	CL	TEM
Me	128	64	8	8	8
Et	128	128	4	8	8
n-C ₃	64	128	1	8	16
i-C ₃	64	64	2	16	64
n-C ₄	16	16	1	8	64
n-C ₅	8	8	0.5	8	64
n-C ₆	16	32	0.5	8	64
n-C ₈	16	16	1	32	128

TABLE II

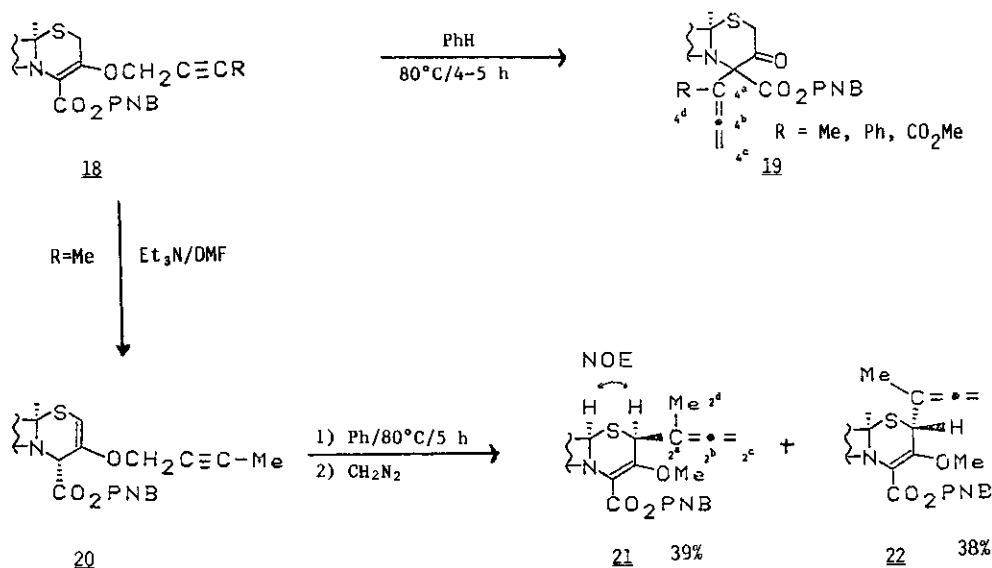
cephalexin	8	64	8	16	8
CH ₂ --Cl	8	8	0.125	8	64
CH ₂ -	32	32	1	8	32
CH ₂ -C \equiv CH	32	32	2	4	8
CH ₂ CO ₂ Me	128	128	4	8	8
CH ₂ CH ₂ SMe	128	128	4	16	64
CH ₂ CH ₂ -N-phthalimide	8	32	4	8	128
CH ₂ -	32	32	0.5	8	128
CH ₂ --Cl-purine.	128	128	4	32	128

We then turned our attention from alkyl ethers to C(3)-ethers having some sort of functional group in order to attempt to increase the antibacterial activity, for example, substituted benzyls, unsaturated groups (allyl and propargyl), esters, a serine protected ester, sulfides, imides, terpenes, and nucleosides (see Table II).

Stability problems were encountered with allyl ethers and these were shown to readily undergo the Claisen rearrangement ($80^{\circ}\text{C}/\text{PhH}/1\text{ h}$) to give a single product. Only one isomer was detected at C(4). The Claisen product **17** ($\text{R}'=\text{H}$) was identical (tlc, nmr) to the C-alkylated by-product of **3** using allyl alcohol.



Propargyl cephem ethers also undergo the Claisen rearrangement to give allenes **19**, but require longer reaction times ($4\text{--}5\text{ h}/80^{\circ}\text{C}$). Again only one isomer was detected at C(4). These are rather mild conditions for a propargyl Claisen rearrangement and we attributed this to the electronic effect of the β -carboalkoxy group, however, the Δ^2 -cephem propargyl ether **20** also rearranges under similar conditions to give a C(2)-mixture of allenes **21** and **22**. The stereochemistry at C(2) was determined by $\text{H}^2\text{--H}^6$ NOE studies and the presence of $\text{H}^2\text{--H}^7$ five bond coupling^{10,11} ($J=0.5\text{ Hz}$) in **22** which was absent in **21**.



In conclusion, we have shown the viability of the Mitsunobu reaction in preparing C(3)-alkoxycephems. Several of these derivatives show good *in vitro* antibacterial activity when compared to cephalixin.

EXPERIMENTAL

All melting points are uncorrected. The following instruments were used for obtaining the spectral data: ^1H nmr: Varian T-60, JEOL FX-90Q, Bruker WM-270 and 360; ^{13}C nmr: Bruker-270, JEOL FX-90Q; ir spectra: Perkin-Elmer 281; mass spectral data: Varian-m.a.t.-731.

(7 β)-3-Methoxy-7-[(phenoxyacetyl)amino]-3-cephem-4-carboxylic Acid, (4-nitrophenyl)-methyl Ester 5 (R=Me)

To a stirred solution of 3 (0.385 g, 1 mM) in 20 ml of THF was added 1 equiv. Ph_3P (0.262 g, 1 mM), 1 equiv. MeOH (0.032 g, 1 mM) followed by the addition of 1 equiv. (0.146 g) dimethyl azodicarboxylate using 5 ml of THF as washing. Allowed to stir at r.t. for 20 min. Evaporated to a low volume, added 30 ml of EtOAc and washed 4x 10 ml H_2O , 1x brine, dried (Na_2SO_4) and evaporated to dryness. Chromatographed on silica gel using a toluene-ethyl acetate gradient to give:

Frac. 14-17, 67 mg (13%) C-alkylated product 4 (R=Me) as a froth; m/e 499; ir v (CHCl_3) 1775 cm^{-1} (β -lactam), 1728 cm^{-1} (ketone); nmr (CDCl_3) δ 2.00 (s, 3, Me), 3.03, 3.70 (AB, J= 14 Hz, 2, C(2) protons), 4.63 (s, 2, PhOCH_2), 5.33-5.43 (BM, 4, H^6 H^7 , PNB).

Frac. 18-22, 356 mg (71%) O-alkylated product 5 (R=Me) as a white solid; m/e 499, ir v (CHCl_3) 1773 cm^{-1} (β -lactam); nmr (CDCl_3 - DMSO-d_6) δ 3.70 (s, 2, C(2) protons), 3.88 (s, 3, OMe), 4.63 (s, 2, PhOCH_2) 5.12 (d, J= 4 Hz, 1, H^6), 5.33 (BS, 2, PNB), 5.47 (d, d J=4, 8 Hz, 1, H^7), 8.97 (d, J=8 Hz, 1, NH).

(7 β)-3-(1-propenyloxy)-7-[(phenoxyacetyl)amino]-3-cephem-4-carboxylic Acid, (4-nitrophenyl)-methyl Ester 16 (R'=H)

To a solution of 0.800 g of the acetic acid solvate of 3 in 30 ml THF was added 1.1 equiv. (0.423 g) Ph_3P , 1.1 equiv. (0.094 g) of allyl alcohol followed by 1.1 equiv. (0.236 g) dimethyl azodicarboxylate. After 15 min at r.t. the mixture was worked (vide supra) and chromatographed on silica gel using a toluene-ethyl acetate gradient to give:

Frac. 24-27, 144 mg (19%) C-alkylated product 4 (R= $\text{CH}_2\text{CH}=\text{CH}_2$) [17, R'=H] as a froth; m/e 525; ir v (CHCl_3) 1775 cm^{-1} (β -lactam), 1728 cm^{-1} (ketone); nmr (CDCl_3) δ 3.00, 3.65 (AB, J= 15 Hz, 2, C(2) protons), 3.12 (m, 2, CH_2 -CH=CH $_2$), 4.60 (s, 2, PhOCH_2), 4.90-6.00 (m, 7, PNB, CH=CH $_2$, H^6 , H^7).

F 29-34, 525 mg (57%) O-alkylated product 5 ($R=CH_2CH=CH_2$) [16, $R'=H$] as a froth; m/e 525; $ir\ v$ ($CHCl_3$) $1775\ cm^{-1}$ (β -lactam), nmr ($CDCl_3$) δ 3.40 (s, 2, C(2) protons), 4.58 (s, 2, $PhOCH_2$), 5.10 (d $J = 4\ Hz$, 1, H_6), 5.37 (s, 2, PNB), 5.5-6.2 (m, 4, $H_6 + CH=CH_2$). A solution of 16 ($R'=H$) (578 mg) in 30 ml of PhH was gently refluxed 1 hour and evaporated to 17 ($R'=H$). TLC and nmr show complete conversion of 16 to 17.

(7 β)-3-(2-Butynyloxy)-7-[(phenoxyacetyl)amino]-3-cephem-4-carboxylic Acid, (4-nitrophenyl)-methyl Ester 18 ($R=Me$)

To a solution of 1.00 g of the acetic acid solvate of 3 in 40 ml of THF was added 1.1 equiv. (0.529 g) Ph_3P , 1.1 equiv. (0.141 g) 2-butyne-1-ol followed by 1.1 equiv. (0.295 g) dimethyl azodicarboxylate. After 15 min at r.t. the mixture was worked up (vide supra) and chromatographed on silica gel using a toluene-ethyl acetate gradient to give:

Frac. 21-23, 71 mg (7%) C-alkylated product 4 ($R=CH_2C\equiv CCH_3$) as a white froth; m/e 537; $ir\ v$ ($CHCl_3$) $1775\ cm^{-1}$ (β -lactam), $1729\ cm^{-1}$ (ketone); nmr ($CDCl_3$) δ 1.70 (m, 3, Me), 3.03, 3.52 (AB, $J=16\ Hz$, 2, C(2) protons), 3.32 (m, 2, $CH_2C\equiv CCH_3$), 4.58 (s, 2, $PhOCH_2$), 3.45 (m, 4, PNB, H^6 , H^7).

Frac. 31-70, 566 mg (57%) O-alkylated product 18 ($R=Me$) as a white froth; m/e 537; $ir\ v$ ($CHCl_3$) $2250\ cm^{-1}$ ($C\equiv C$), $1778\ cm^{-1}$ (β -lactam); nmr ($CDCl_3$) δ 1.87 (m, 3, Me), 3.50 (s, 2, C(2) protons), 4.60 (s, 2, $PhOCH_2$), 4.72 (m, 2, $OCH_2C\equiv CCH_3$), 5.10 (d, $J=4\ Hz$, 1, H^6), 5.33 (m, 2, PNB), 5.67 (d, d $J=4$, 8 Hz, 1, H^7). Crystallized from CH_2Cl_2 -hexanes to give white needles, m.p. 97-120°C (undergoing Claisen as indicated by tlc). Anal. Calcd for $C_{26}H_{23}N_3O_8S$: C, 58.09; H, 4.31; N, 7.82; Found: C, 57.89; H, 4.01; N, 7.54.

(7 β)-4-(1-Methyl-1,2-propadienyl)-3-oxy-7-[(phenoxyacetyl)amino]cephem-4-carboxylic Acid, (4-nitrophenyl) methyl Ester 19 ($R=Me$)

A solution of 18 ($R=Me$) (0.120 g) in 30 ml of PhH was gently refluxed for 3 h and evaporated to give 19 ($R=Me$) as a froth; m/z 537; $ir\ v$ ($CHCl_3$) $1950\ cm^{-1}$ (allene), $1778\ cm^{-1}$ (β -lactam), $1720\ cm^{-1}$ (ketone); $^1H\ nmr$ ($CDCl_3$) δ 1.82 (m, $J=3.0\ Hz$, 3, Me), 3.48 (s, 2, C(2) protons), 4.57 (s, 2, $PhOCH_2$), 5.01 (m, $J=3.0\ Hz$, 2 allene methylene), 5.27, 5.43 (AB, 2, PNB), 5.31 (d, $J=5.0\ Hz$, 1, H^6), 5.53 (d, d $J=5$, 8 Hz, 1, H^7); $^{13}C\ nmr$ ($CDCl_3$) δ C^2 33.63 t, C^3 191.87 s, C^4 73.58 s, C^{4a} 95.70 s, C^{4b} 207.59 s, C^{4c} 79.29 t, C^{4d} 14.98 q, C^6 57.79 d, C^7 58.51 d.

(4 α ,7 β)-3-(2-Butynyloxy)-7-[(phenoxyacetyl)amino]-2-cephem-4-carboxylic Acid, (4-nitrophenyl)-methyl Ester 20

1.725 g of 18 (R=Me) in 25 ml of DMF was treated at r.t. for 23 h with 3.0 equiv. (1.34 ml) Et₃N. EtOAc was then added and then washed 1X 1N HCl, 4X H₂O, 1X brine, dried (Na₂SO₄), evaporated to dryness and chromatographed on silica gel using toluene-ethyl acetate gradient to give a 0.546 g (32%) 20 as a white solid; ir ν (CHCl₃) 1774 cm⁻¹ (β -lactam); nmr (CDCl₃) δ 1.87 (m, 3, Me), 4.37 (m, 2, OCH₂C \equiv Me), 4.53 (s, 2, PhOCH₂), 5.00 (bs, 1, H⁴), 5.35 (m, 3, PNB, H⁶), 5.70 (d, d J=4, 9 Hz, 1, H⁷).

(2 α or 2 β ,7 β)-3-Methoxy-2-(1-methyl-1,2-propadienyl)-7-[(phenoxyacetyl)amino]-3-cephem-4-carboxylic Acid, (4-nitrophenyl)methyl Ester 21 and 22

20 (0.682 g) in 80 ml of PhH was gently refluxed for 5 h. Cooled to r.t. and evaporated to dryness. CH₂Cl₂ (50 ml) was added and treated at 5°C with excess CH₂N₂ for 8 min. It was then evaporated to dryness and chromatographed on silica using a toluene-ethyl acetate gradient to give: Frac. 17-21, 0.263 g (38%) 22 as a white froth; m/e 551; ir ν (CHCl₃) 1945 cm⁻¹ (allene), 1770 cm⁻¹ (β -lactam); ¹H nmr (CDCl₃) δ 1.88 (m, J_{2d-2c} = 3.05 Hz, 3, Me), 3.75 (s, 3, OMe), 3.99 (m, J_{H²-2c} = 2.44 Hz, J_{H²-H⁷} = 0.5 Hz, 1, H₂), 4.56 (s, 2, PhOCH₂), 4.85 (m, J_{2c-2d} = 3.05 Hz, J_{2c-H²} = 2.44 Hz, 11.06 Hz, 2, allene CH₂), 5.06 (d, J=4.88 Hz, 1, H⁶), 5.42, 5.30 (m, 2, PNB), 5.89 (d, d J=4.88, 9.16, 0.5 Hz, 1, H⁷). NOE's were observed between H²-2^d and H²-C(3)-OMe but not H²-H⁶; ¹³C nmr (CDCl₃) δ C² 43.15, C^{2a} 98.81, C^{2b} 207.02, C^{2c} 78.46, C^{2d} 17.24, C³ 160.81.

Frac. 22-27, 0.279 g (39%) 21 as a white froth; m/z 551; ir ν (CHCl₃) 1953 cm⁻¹ (allene), 1779 cm⁻¹ (β -lactam); ¹H nmr (CDCl₃) δ 1.90 (m, J_{2d-2c} = 2.4 Hz, 3, Me), 3.90 (s, 3, OMe), 4.06 (m, J_{H²-2c} = 3.0 Hz, 1, H²), 4.60 (s, 2, PhOCH₂), 4.98, 4.85 (m, J_{H²-2c} = 3.0, 3.7 Hz, 2, allene CH₂), 5.25, 5.42 (m, 2, PNB), 5.34 (d, J=3.7 Hz, 1, H⁶), 5.55 (d, d J= 3.7, 7.9, 1, H⁷). NOE's were observed between H²-H⁶, and H²-2^d; ¹³C nmr (CDCl₃) δ C² 45.23, C^{2a} 95.18, C^{2b} 204.67, C^{2c} 79.84, C^{2d} 16.74, C³ 160.75.

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REFERENCES AND NOTES

1. (a) Ciba Geigy, R. Scartazzini and H. Bickel, Brit. Pat., 1976, 1435111.
 (b) R. Scartazzini and H. Bickel, Heterocycles, 1977, 7, 1165.
2. (a) R. R. Chauvette and P. A. Pennington, J. Med. Chem., 1975, 18, 403.
 (b) S. Kukolja and R. R. Chauvette, "Chemistry and Biology of β -Lactam Antibiotics", eds. by R. B. Morin and M. Gorman, Academic Press, New York, 1982, Vol. 1, p. 93-198.
3. For a study of C(3)-alkyl cephem lipophiles see: D. O. Spry and A. R. Bhala, "C(3)-Alkyl-cephems", Heterocycles, 1985, 23, 1901.
4. O. Mitsunobu, Synthesis, 1981, 1.
5. T. Kurihara, M. Sugizaki, I. Kime, M. Wada, and O. Mitsunobu, Bull. Chem. Soc. Jap., 1981, 54, 2107.
6. We abbreviate dimethyl azodicarboxylate as MAD since DMAD is used for dimethyl acetylenedicarboxylate.
7. J. C. Kauer, Org. Syn., Col. Vol., 4, 1963, 411.
8. The reaction of 3 with 90% Me¹⁸OH under MAD-CAT conditions gave 61% Ph₃P¹⁸O (80% incorporation), 54% 5 (R=Me) and 13% 4 (R=Me). Neither 4 or 5 contained ¹⁸O.
9. B. Fechtig, H. Peter, H. Bickel, and E. Vischer, Helv. Chim. Acta, 1968, 51, 1108.
10. D. O. Spry, Tetrahedron Lett., 1972, 3717.
11. A. Yoshida, S. Oida, and E. Ohki, Chem. Pharm. Bull., 1975, 23, 2507.

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