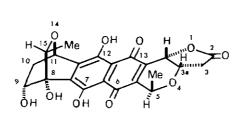
STUDIES ON THE TOTAL SYNTHESIS OF GRANATICIN: SYNTHESIS OF (\pm) -7-DEOXY-GRANATICIN 12-O-METHYL ETHER FROM CHRYSAZIN

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<u>Abstract</u> - The title compound (27) has been synthesized starting with chrysazin (1,8-dihydroxyanthraquinone) via 10-methoxy-8-(methoxy)methoxy-1-anthracenone (9). The strategy involves a stepwise and stereoselective construction of the oxabicycle and pyranolactone systems upon 9.

The naphthoquinone antibiotic granaticin (1), originally isolated from the culture of Streptomyces olivaceus in 1957¹ and since detected in a number of other actinomycetes, is active against Grampositive bacteria and protozoa and exhibits significant antitumor activity against P-388 lymphocytic leukemia in mice.² Of noteworthy in the structure 1, which had been determined by a combination of chemical degradation and X-ray analysis in 1968,³ is that granaticin can be regarded as a structural hybrid composed of sarubicin A (2)⁴ and nanaomycin D (3)⁵ in their C-glycoside derived oxabicycle⁶ and pyrano- γ -lactone systems, respectively. Our interest in this novel molecule has led us to embark on a program directed towards its total synthesis.



granaticin (1)

sarubicin A (2)

nanaomycin D(3)

The Scheme I illustrates the outline of two synthetic approaches to 1 that we envisioned based on our preliminary investigations. With regard to the preparation of the oxabicycle moiety, both routes (A & B) utilize the methodology that has been introduced by us for the total synthesis of 2.7 For the construction of the pyrano-Y-lactone moiety, on the other hand, the method of Kraus and Roth⁸ should meet our strict requirements: mildness of the reactions involved, which does not affect the preformed oxabicycle, and high stereoselectivity as improved by us.⁹ All other methods so far reported in nanaomycin synthesis, 10 which involve an acid-catalyzed epimerization of the benzylic C-Me, are not adaptable, since the oxabicycle would not survive under the reaction conditions. Here we report the first synthesis of the title compound having the complete skeleton of 1 by the route A.

Scheme I

Scheme II

Chrysazin (4), the starting material in this investigation, was first converted in three steps (75% yield) to triacetoxyanthracene (5) according to the procedures described in the literature. Extensive investigation on the partial hydrogenation of 5 has established the conditions leading to either tetrahydro compound (6) in 54% yield (5% Rh-Pd-C/AcOEt, 20 atm, room temp.) or its hydrogenolysis product 7 in 65% yield (5% Rh-C/AcOEt, 95 atm, 140°C). The compound 6 was transformed into diacetoxyanthracenone (8) in 80% yield by the following manipulation of the benzylic 1acetoxy group: i) trifluoroacetolysis (TFA, room temp., 1 h); ii) ethanolysis of the resulting trifluoroacetate (EtOH, Et₃N, 0°C, 2 h); iii) oxidation of the liberated 1-hydroxy function (PCC, 4A sieves, CH_2Cl_2 , O^OC to room temp.). The ketone 8 was also obtained from 7 in 63% yield by a regioselective benzylic oxidation with 3,5-dimethylpyrazole- CrO_3 complex. 12 It was then converted to 10-methoxy-8-(methoxy)methoxy-1-anthracenone (9) in 65% yield by a stepwise $\underline{0}$ -alkylation involving selective hydrolysis of the less hindered 8-acetate group: i) 1 eq 1.5% KOH-EtOH, 0°C; ii) MeOCH₂Cl, NaH, DMF; iii) 3% KOH-EtOH, room temp.; iv) Me₂SO₆, 6% KOH, CH₂Cl₂, n-Bu₆NBr). Alternatively, 9 was obtained from 7 in 55% yield by carrying out the stepwise O-alkylation first (formation of 10 in 80% yield) and then DDQ oxidation in MeOH 13 at room temperature (Scheme II). The structure of ${f 9}$ was confirmed by its conversion to ${f 11}$ (deprotection of the MOM group followed by ${f 0}$ ethylation), which was identical with an authentic sample prepared from 1-ethoxy-5-hydroxynaphthalene.14

With a ready access to the dialkoxyanthracenone 9 being established, construction of 5,6-naphtho-2-oxabicyclo[2.2.2] octene system was initiated by transformation into the allyl alcohol 13 (Scheme III), which was performed by three steps in 77% overall yield as follows: i) reaction with $CH_2=C(OMe)$ Li followed by acid workup to give the α -ketol 12 (87%), ii) $NaBH_4$ reduction of 12 in iso-PrOH, iii) trifluoroacetic acid-catalyzed dehydration of the crude diol in CH_2Cl_2 (4 eq TFA, 0.14 M) at room temperature (88% yield from 12). cis-Dibydroxylation of 13 by a catalytic osmylation (0.1 eq OSO_4 , 6 eq Me_3NO , aqueous tert-BuOH, OCC) produced a diastereomeric mixture of triols, 14 and its 1'-epimer, which were separable on their di-Q-acetates by silica gel chromatography to provide 15 (83%) and its 1'-epimer (2.5%). The triol 14, 15 which was obtained as a gelatinous mass by hydrolysis of its acetate 15, was subjected to NBS mediated cyclization (OCCC) in 81% chromatoghraphed yield.

Transformation of 16 into the acetonaphtoquinone 20, that is required for attachment of the pyranolactone functionality, was nicely achieved as follows. Treatment of the carbonate 17 with 4 eq NBS in aqueous AcOH at 50° C for 30 min produced the bromoquinone 18^{15} in 78% yield. The position of the bromine atom as indicated in the structure (Scheme III) was assigned based on the

Scheme III

27, R= H

presumed reaction mechanism, which would be the same in principle as that suggested for the case of 1,5-diacetoxynaphthalene. Replacement of the bromine atom with acetyl group was then carried out by the following sequence of reactions via a lithio intermediate: i) reductive Q-methylation (aqueous $Na_2S_2O_4/AcOEt$; then $Me_2SO_4/K_2CO_3/acetone$, reflux, 70%); ii) exchange of the carbonate group for isopropylidene group (MeOH-KOH; then $CH_2=C(OMe)Me$, CSA, 92%); iii) acetylation (n-BuLi, -78°C; $CeCl_3$, $CeCl_3$,

Formation of the pyrano-Y-lactone system was now commenced by ${
m LiAlH}_{A}$ reduction of the ketone function in 21 (Et_2O , -50 to -30°C). The product, a diastereomeric mixture of the carbinols 22, was subjected to the deprotection-cyclization sequence according to a modification 9 of the Kraus' protocol as follows: i) Y-naphthylbutenolide formation by treatment with 1 eq TsOH in MeCN (room temp., ca. 8 min; quenching immediately after 22 had been consumed); ii) pyran-ring closure with 1 eq DBU in toluene-CH $_2$ Cl $_2$ (1:1) at -10 $^{\rm o}$ C for 10 min. There was obtained a ca. 4:4:1:1 mixture of four diastereomeric pyranolactones that possess the complete carbon skeleton of granaticin, in 55-60% overall yield from 21. These isomers could have been separated by MPLC (10y silica gel, AcOEt/hexane=2:1) in pure states. Assignment of the major two isomers to 23a,b (3a,5-trans) and the minor two to 24a,b (3a,5-cis), based on the known preference of the trans product in such annulation, 9 was supported by $^1\text{H-nmr}$ analysis (270 MHz), 15 diagnostic signals being those of C_{3a} -H (δ 4.77 for 23a,b; δ 4.40 for 24a,b) and C₅-H (δ 5.38 for 23a,b; δ 5.07 for 24a,b) on the pyranolactone group. Although 23a (less polar) and 23b (more polar) can be differentiated from each other by the resonances of of C_{15} -CH $_3$ in the oxabicycle (δ 0.81 and 0.87, respectively), the difference is too small to provide any information on their stereochemistry. The structures were, therefore, determined by X-ray analysis which was performed on the acetonide of 23b, 19

The compound 23a, which has the same stereochemistry as 1, could be oxidized with ceric ammonium nitrate 20 after protection of the 1,2-glycol system as acctonide or carbonate to furnish the corresponding naphthoquinones 25 or 26. Brief treatment of 25 with TsOH in MeCN afforded 27, (±)-7-deoxygranaticin 12-0-methyl ether. 15 At this point we encountered major difficulties in 0-demethylation of 27, the first and yet essential step for oxidation to the naphthazarin system in 1. Reaction of BC1 $_3$ with 26 occurred preferentially at the oxabicycle leading to a clean cleavage of the benzylic C_{11} -0 bond. Attempt for nucleophilic demethylation with MeSLi 21 resulted in the

destruction of the pyranolactone system. Thus, use of a chemoselectively removable protecting group for the phenolic C_{12} -OH should be considered to accomplish the total synthesis of 1. Investigations along this line are in progress in our laboratory.

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3H), 4.36 (dd, J = 9.1, 3.8 Hz, 1H), 4.50 (q, J = 6.6 Hz, 1H), 5.36, 5.39 (ABq, = 6.6 Hz, each 1H), 7.06 (d, J = 8.2 Hz, 1H), 7.38 (t, J = 8.2 Hz, 1H), 7.68 (d, = 8.2 Hz, 1H), 8.24 (s, 1H).

16:0.80 (d, J = 6.2 Hz, 3H), 1.49 (dt, J = 14.5, 1.4 Hz, 1H), 2.02 (br s, 1H, 0H), 2.86 (ddd, J = 14.5, 8.9, 3.8 Hz, 1H), 3.32 (br s, 1H, 0H), 3.53 (s, 3H), 3.89 (q, J = 6.2 Hz, 1H), 3.95 (s, 3H), 4.01 (dd, J = 8.9, 1.4 Hz, 1H), 5.26 (dd, J = 3.8, 1.4 Hz, 1H), 5.36 (s, 2H), 7.16 (d, J = 8.1 Hz, 1H), 7.42 (t, J = 8.1 Hz, 1H), 7.79 (d, J = 8.1 Hz, 1H), 8.21 (s, 1H).

18: 0.77 (d, J = 6.2 Hz, 3H), 1.68 (dd, J = 13.8, 5.9 Hz, 1H), 3.02 (ddd, J = 13.8, 9.5, 4.9 Hz, 1H), 3.95 (s, 3H), 4.40 (q, J = 6.2 Hz, 1H), 4.81 (dd, J = 9.5, 5.9 Hz, 1H), 5.45 (d, J = 4.9 Hz, 1H), 7.48 (s, 1H), 8.25 (s, 1H).

23a: 0.81 (d, J = 6.2 Hz, 3H), 1.59 (d, J = 6.6 Hz, 3H), 1.62 (d, J = 14.7 Hz, 1H), 2.73 (d, J = 17.5 Hz, 1H), 2.93.1 (br, 1H, 0H), 3.90 (s, 3H), 3.96 (s, 3H), 3.98 (q, J = 6.2 Hz, 1H), 4.00 (s, 3H), 4.15 (d, J = 8.8 Hz, 1H), 4.77 (dd, J = 4.8, 2.9 Hz, 1H), 5.33 (d, J = 3.7 Hz, 1H), 5.38 (q, J = 6.6 Hz, 1H), 5.64 (d, J = 2.9 Hz, 1H), 5.36 (q, J = 14.5 Hz, 1H), 1.65 (br s, 1H, 0H), 1.8 (br, 1H, 0H), 2.72 (d, J = 17.5 Hz, 1H), 2.90 (dd, J = 17.5, 4.8 Hz, 1H), 5.38 (q, J = 6.6 Hz, 1H), 5.64 (d, J = 2.9 Hz, 1H), 2.91 (ddd, J = 14.5, 8.8, 3.7 Hz, 1H), 5.38 (d, J = 6.2 Hz, 1H), 3.90 (dd, J = 17.5, 4.8 Hz, 1H), 3.90 (dd, J = 17.5, 4.8 Hz, 1H), 5.38 (q, J = 6.6 Hz, 1H), 5.36 (d, J = 17.5, 4.8 Hz, 1H), 5.37 (q, J = 6.6 Hz, 1H), 5.64 (d, J = 2.9 Hz, 1H), 5.97 (d, J = 14.5 Hz, 1H), 5.33 (d, J = 3.7 Hz, 1H), 5.38 (q, J = 6.6 Hz, 1H), 5.64 (d, J = 2.9 Hz, 1H), 5.97 (d, J = 14.5 Hz, 1H), 5.97 (d, J = 17.5, 4.8 Hz, 1H), 3.90 (dd, J = 17.5, 4.8 Hz, 1H), 3.90 (dd, J = 17.5, 4.8 Hz, 1H), 3.90 (dd, J = 17.5, 4.8 Hz, 1H), 3.91 (dd, J = 17.5, 4.8 Hz, 1H), 3.91 (dd, J = 17.5, 4.8 Hz, 1H), 3.91 (dd, J = 17.5, 4.8 Hz, 1H), 3.93 (dd, J = 17.5, 4.8 Hz, 1H

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