NITROSOUREA DERIVATIVES OF 3',4'-DIDEMETHOXY-3',4'-DIOXO-4-DEOXYPODOPHYLLOTOXIN AND UREA DERIVATIVES OF 4'-Q-DEMETHYLPODOPHYLLOTOXIN AS POTENT INHIBITORS OF HUMAN DNA TOPOISOMERASE II¹,†

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Abstract — Nitrosourea derivatives of 3',4'-didemethoxy-3',4'-dioxo-4-deoxypodophyllotoxin and urea derivatives of 4'-Q-demethylpodophyllotoxin were synthesized and evaluated for their inhibitory activity against DNA topoisomerase II and KB cells. Although the 4β -N'-nitrosoureido compounds demonstrated good inhibitory activity against topoisomerase II, they were found to possess low activity for protein linked-DNA complex formation ability. On the other hand, the 4β -ureido compounds exhibited better or similar activity compared to 1, etoposide.

Etoposide (1; VP-16) and teniposide (2) are known to be potent chemotherapeutic agents for the treatment of small-cell lung carcinoma, testicular cancer, and malignant lymphoma. Since they inhibit the catalytic activity of DNA topoisomerase II by stabilizing a cleavable enzyme-DNA complex, in which DNA is cleaved and covalently linked to the enzyme, DNA topoisomerase II has been considered as a target enzyme of 1.2 We

[†] Dedicated to Dr. Arnold Brossi on the occasion of his 70th birthday.

have previously synthesized and evaluated 4β -arylamino analogs related to 1, such as 4β -arylamino-3',4'-Q-Q-didemethylepipodophyllotoxins, 4 - 5 and -3',4'-didemethoxy-3',4'-dioxo-4-deoxypodophyllotoxins, 6 as potent inhibitors of human DNA topoisomerase II and as antitumor agents. As an extention of our studies, we have synthesized new 4β -substituted nitrosourea derivatives, together with 4β -urea derivatives, aimed at developing agents for the treatment of brain tumors, since nitrosourea derivatives, such as N-N-bis-(2-chloroethyl)-N-nitrosourea (BCNU, Carmustine), N-(2-chloroethyl)-N-nitrosourea (CCNU, Lomustine), and nitrosoureido sucroses, are able to penetrate the blood-brain barrier, $^{7-9}$ and have been used clinically for the treatment of brain cancer.

Scheme 1

The nitrosourea derivatives were synthesized as shown in Scheme I. 4'-Q-Demethylepipodophyllotoxin (3) was treated with trifluoroacetic acid and NaN3 to yield the 4β -azido compound (4).⁵ When a ketone, such as acetone or methyl ethyl ketone, was used as the solvent, hydrogenation of 4 gave the corresponding 4β -alkylamino derivatives (e.g., 5 and 6). This could possibly occur through the cyclic intermediate (A) to yield the N-alkylaminopodophyllotoxin on reduction. Reduction of 4 in the absence of acetone or methyl ethyl ketone furnished the 4β -amino compound (7). Compound (7) was further treated with the appropriate isocyanate to furnish the corresponding ureido derivatives (8 – 12).

In order to prepare the nitrosourea derivatives, 8 and 9 were treated with NaNO2 in the presence of HCOOH. Unexpectedly, they yielded a red compound in each case (13 and 14, respectively). The ¹H-nmr spectrum of 13 exhibited signals due to H-1 [δ 4.45 (d, J=4 Hz)], H-2 [δ 3.26 (m)], H-3 [δ 3.68 (dd, J=4, 14 Hz)], H-4 [δ 5.46 (dd, J=4, 8 Hz)], H2-11 [δ 4.23 (1H, dd, J=2, 9 Hz) and 4.55 (1H, dd, J=1, 9 Hz)], H-5 and -8 [δ 6.71 and 7.00 (each s)], and the methylenedioxy group [δ 6.00 (s)] of an epipodophyllotoxin skeleton. It also showed a three-proton singlet at δ 3.17, assignable to the N'-methylureido group, suggesting that the nitroso group was introduced at the distal nitrogen of the 4β-N'-methylureido group. However, one of the two methoxy signals found in epipodophyllotoxin was absent [δ 3.76 (3H,s)], and the signals ascribable to H-2' and -6' were observed as a pair of doublets [δ 6.46 and 5.36 (each 1H, d, J=1.5 Hz)]. In addition, the observation of two carbonyl carbon signals (δ 175.0 and 179.1), along with an ester (δ 176.2) and ureido (δ 157.5) carbonyl signals, in the ¹³C-nmr spectrum of 13, suggested the presence of an ortho-benzoquinone ring. The ¹H-nmr of 14 was similar to that of 13, except for the presence of the signals arising from the N'-nitroso-N'-2"-chloroethylureido group. These spectral data indicated that oxidation of ring-E had taken place,

together with nitrosation, giving the <u>ortho</u>-benzoquinone of the nitrosourea derivatives. The structures of the reaction products were concluded to be represented by 13 and 14. Since the ring-E of 4'-demethylpodophyllotoxins is quite sensitive to oxidating reagents, a nitrosourea derivative of 4'-demethylpodophyllotoxin could not be prepared, although several nitrosation conditions were attempted.

Compound (5) was also treated with NaNO₂ and HCOOH to afford the 4β-N-nitroso-N-isopropyl derivative (15). Purification of 15 by silica gel chromatography (hexane-acetone) gave a yellow substance (16). The ¹H-nmr of 16 was similar to that of 13. A downfield shift of H-4 [δ 6.03 (1H, d, J=4 Hz)] compared with that [δ 3.92 (d, J=4 Hz)] of 5 indicated the existence of a N-nitroso group. An asymmetric signal pattern due to H-2' and -6' [δ 5.02 and 5.77 (each 1H, br s)] and a methoxy singlet [δ 3.82 (3H, s)], similar to those found in 13, suggested that again the ring-E was oxidized. However, an additional isolated methyl [δ 2.09 (3H, s)] and a methylene [δ 2.90 and 3.08 (each 1H, d, J=15 Hz)] in the ¹H-nmr spectrum, together with a carbonyl signal (δ 204.3) in the ¹³C-nmr spectrum, suggested the presence of an acetonyl group. In addition, the signals ascribable to C-4' and -5' were observed at δ 74.3 and 199.4 in the ¹³C-nmr spectrum, suggesting that the acetonyl group was attached at either C-4' or -5' through a carbon-to-carbon linkage. The location of the acetonyl group was concluded to be at C-4' by ¹H-¹³C long-range COSY, which showed a long-range correlation between the methylene proton signal of the acetonyl group and the C-3' carbon signal. Based on these spectral data, the structure of this reaction product was concluded to be represented by 16. The configuration of C-4' still remains to be determined. This acetonyl group is considered to be derived from acetone, and similar examples are described in the literature.¹⁰

Table I shows the inhibition of DNA topoisomerase II activity, percentage of protein-linked DNA complex formation, and cytotoxicity against the KB cell line of the nitrosourea and urea derivatives. The nitrosourea derivatives (13 and 14) demonstrated good inhibitory activity against topoisomerase II. However, they possess low activity for protein linked-DNA complex formation ability. These results are similar to those found with 3',4'-didemethoxy-3',4'-dioxo-4-deoxy(epi)podophyllotoxin.⁶ In addition, the acetone condensed compound (16) showed no activity, although it does contain a N-nitroso group. This suggests that the N-nitroso group alone is not sufficient for inhibitory activity against topoisomerase II and that orthobenzoquinone derivatives have decreased ability to cause protein-linked DNA complex formation. On the other hand, the N-ureido compounds (8 – 12) exhibited better or similar activity compared with etoposide, 1. The aryl and β -chloroethyl-N-ureido compounds (9 – 12) also displayed cytotoxicity comparable to that of 1.

Table I. Biological evaluation of nitrosourea derivatives of 3',4'-didemethoxy-3',4'-dioxo-4-deoxypodophyllotoxin and urea derivatives of 4'-O-demethylpodophyllotoxin

Compound	Cytotoxicity (ID50 KB, μΜ) ^a	Inhibition of DNA topoisomerase II activity (ID50, µM) ^b	Cellular protein-DNA complex formation (% 20 µM)
etoposide (1)	0.2	50	100
5	NT	50	109
6	NT	50	73
8	1.4	50	81
9	< 0.2	25	143
10	< 0.2	25	148
11	< 0.2	50	125
12	< 0.2	50	118
13	1.5	5	41
14	1.3	< 5	7
16	> 10	>100	1

^a ID50 is the concentration of drug which affords 50% of KB cell growth after a 3 day incubation.

NT: Not tested

b Each compound was examined with five concentrations at 5, 10, 25, 50 and 100 μM.

The ID50 value was established based on the degree of inhibition at these five concentrations.

EXPERIMENTAL

Optical rotations were determined using a Rudolph Research Autopol III polarimeter. 1 H and 13 C nmr spectra were recorded on a Bruker AC-300 (300 and 75.5 MHz, respectively) spectrometer. Chemical shifts are presented in terms of δ (ppm) with tetramethylsilane as an internal standard. Elemental analyses were performed by Atlantic Microlab, Inc. (Norcross, GA).

Synthesis of N-alkylamino Compounds. A solution of 4β-azido-4'-demethyl-4-deoxypodophyllotoxin (4, 425 mg, 1 mmol) in a mixture of ethyl acetate (50 ml) and acetone or methyl ethyl ketone (20 ml) was treated with 10% Pd-C under H₂ atomosphere with stirring overnight. After filtration, the filtrate was concentrated to give a product, which was purified by silica gel chromatography [hexane-EtOAc (3:2)].

4β-N-Isopropylamino-4-deoxy-4'-demethylpodophyllotoxin (5). Yield 75%; Colorless needles; mp 214 °C. [α]_D²⁰ – 86.1° (\underline{c} =0.55, CHCl₃). Anal. Calcd for C₂4H₂7NO₇•1/2H₂O: C, 63.99; H, 6.26; N, 3.11. Found: C, 64.36; H, 6.07; N, 3.28. ¹H-Nmr (CDCl₃, 300 MHz): δ 6.79 (1H, s, H-5), 6.46 (1H, s, H-8), 6.29 (2H, s, H-2',6'), 5.96, 5.94 (each 1H, s, -O-CH₂-O-), 4.51 (1H, d, <u>I</u>=5 Hz, H-1), 4.26 (d, <u>J</u>=9 Hz, H-11), 3.92 (1H, d, <u>I</u>=4 Hz, H-4), 3.77 (6H, s, OCH₃), 3.26 (dd, <u>J</u>=5, 14 Hz, H-2), 2.83–2.65 (2H, m, H-3 and -1"), 1.21 and 1.05 (each 3H, d, <u>I</u>=6 Hz, 1"-CH₃).

4β-N-Isobutylamino-4-deoxy-4'-demethylpodophyllotoxin (6). Yield 86%; Colorless needles; mp 220 °C. Anal. Calcd for C25H29NO7: C, 65.92; H, 6.42; N, 3.08. Found: C, 65.64; H, 6.37; N, 3.04. ¹H-Nmr (CDCl₃, 300 MHz): δ 6.79 (1H, s, H-5), 6.46 (1H, s, H-8), 6.29 (2H, s, H-2',6'), 5.96, 5.94 (each 1H, s, -O-CH₂-O-), 5.39(1H, br s, NH), 4.51 (1H, d, I=5 Hz, H-1), 4.26 (2H, m, H-11), 3.97 (0.4H, d, I=4 Hz, H-4), 3.92 (0.6H, d, I=4 Hz, H-4), 3.77 (6H, s, OCH₃), 3.30 (3/5H, dd, I=5, 14 Hz, H-2), 3.23 (0.4H, dd, I=5, 14 Hz, H-2), 2.77 (1H, m, H-3), 2.61 (0.6H, m, H-1"), 2.45 (0.4H, m, H-1"), 1.70 (0.4H, m, H-2"), 1.38 (0.6H, m, H-2"), 1.28 (1.2H, d, I=6 Hz, 1"-CH₃), 1.01 (1.8H, d, I=6 Hz, 1"-CH₃), 0.93 (3H, t, I=8 Hz, 2"-CH₃). The ¹H-nmr examination showed that 6 is a mixture of diastereoisomers in a molar ration of 2:3.

Synthesis of 4β -N-Ureido Compounds. A suspension of 7 (400 mg, 1 mmol) in benzene (50 ml) was added to a solution containing the appropriate isocyanate (0.20 mmol) in benzene (2 ml). The solution was stirred at room temperature overnight. The reaction mixture was concentrated to give a product, which was purified by crystallization (from hexane-EtOH) or silica gel chromatography [benzene-EtOAc (4:1)].

4β-(N'-Methylureido)-4-deoxy-4'- \underline{O} -demethylpodophyllotoxin (8). Yield 24%. Colorless crystals; mp 178 – 180 °C; $[\alpha]_D^{20}$ – 110.4° (\underline{c} =0.5, CHCl₃). Anal. Calcd for C₂₃H₂₄N₂O₈•H₂O: C, 58.22;

H, 5.52; N, 5.90. Found: C, 58.56; H, 5.46; N, 5.50. ¹H Nmr (CDCl₃, 300 MHz): δ 6.82 (1H, s, H-5), 6.49 (1H, s, H-8), 6.27 (2H, s, H-2',6'), 5.96 and 5.93 (each 1H, s, -O-CH₂-O-), 5.44 (1H, br s, OH), 5.11 (1H, dd, I=4, 6.5 Hz, H-4), 4.66 (1H, d, I=6.5 Hz, NH), 4.51 (2H, d, I=5 Hz, H-1 and NH), 4.38 (1H, dd, I=2, 9 Hz, H-11), 3.96 (1H, t, I=9 Hz, H-11), 3.76 (6H, s, OCH₃), 2.85–2.90 (2H, m, H-2 and 3), 2.82 (3H, d, I=5 Hz, NHCH₃).

4β-[N'-(2'''-Chloroethylureido)]-4-deoxy-4'-Q-demethylpodophyllotoxin (9). Yield 24%. Colorless crystals; mp 225 – 230 °C; $[\alpha]_D^{20}$ – 160.0° (c=0.1, CHCl3). Anal. Calcd for C24H25N2O8Cl*H2O: C, 55.12; H, 5.20; N, 5.36. Found: C, 55.11; H, 5.15; N, 4.78. ¹H Nmr (CDCl3, 300 MHz): δ 6.82 (1H, s, H-5), 6.48 (1H, s, H-8), 6.26 (2H, s, H-2',6'), 5.95 and 5.91 (each 1H, s, -O-CH2-O-), 5.45 (1H, br s, OH), 5.10 (1H, br s, NH), 5.10 (1H, d, I=4 Hz, H-4), 4.50 (1H, d, I=4 Hz, H-1), 4.38 (1H, t, I=9 Hz, H-11), 3.93 (1H, t, I=9 Hz, H-11), 3.75 (6H, s, OCH3), 3.78–3.50 (4H, m, CH2x2), 2.85–2.90 (2H, m, H-2 and 3).

4β-(N'-Phenylureido)-4-deoxy-4'-Q-demethylpodophyllotoxin (10). Yield 22%. Colorless crystals; mp 188 – 192 °C; $[α]_D^{20}$ – 86.1° (\underline{c} =0.55, CHCl₃). Anal. Calcd for C₂₈H₂₆N₂O₈•1/2H₂O: C, 63.75; H, 5.16; N, 5.31. Found: C, 63.44; H, 5.38; N, 5.23. ¹H Nmr (CDCl₃, 300 MHz): δ 7.00 (1H, br s, NH), 7.5–7.2 (5H in total, m, arom-H), 6.78 (1H, s, H-5), 6.40 (1H, s, H-8), 6.20 (2H, s, H-2',6'), 5.85 and 5.83 (each 1H, s, -O-CH₂-O-), 5.62 (1H, br s, OH), 5.08 (1H, d, <u>I</u>=4 Hz, H-4), 4.35 (1H, d, <u>I</u>=4 Hz, H-1), 4.25 (1H, t, <u>I</u>=9 Hz, H-11), 4.05 (1H, br s, NH), 3.86 (1H, t, <u>I</u>=9 Hz, H-11), 3.66 (6H, s, OCH₃), 2.68 (H, m, H-3), 2.65 (1H, dd, <u>I</u>=4, 14 Hz, H-2).

4β-[N'-(4'''-Chlorophenyl)ureido]-4-deoxy-4'-Q-demethylpodophyllotoxin (11). Yield 87%. Colorless crystals; mp 195 °C; $[\alpha]_D^{20}$ – 50.8° (α =0.69, CHCl3). Anal. Calcd for C28H25N2O8Cl*H2O: C, 58.90; H, 4.77; N, 4.91. Found: C, 59.13; H, 5.24; N, 5.37. H Nmr (CDCl3, 300 MHz): δ 8.05 (1H, br s, NH), 7.52 and 7.26 (each 2H, d, α =9 Hz, arom.-H), 6.95 (1H, s, H-5), 6.53 (1H, s, H-8), 6.40 (2H, s, H-2',6'), 6.27 (1H, d, α =7 Hz, NH), 5.99 (2H, s, -O-CH2-O-), 5.19 (1H, dd, α =4, 7 Hz, H-4), 4.56 (2H, d, α =4 Hz, H-1 and NH), 4.36 (1H, dd, α =2, 9 Hz, H-11), 4.04 (1H, t, α =9 Hz, H-11), 3.71 (6H, s, OCH3), 3.1–3.0 (2H, m, H-2 and -3).

4β-[N'-(4'''-Fluorophenyl)ureido]-4-deoxy-4'-Q-demethylpodophyllotoxin (12). Yield 43%. Colorless crystals; mp 185 °C; $[\alpha]_D^{20}$ – 99.6° (\underline{c} =0.6, CHCl3). Anal. Calcd for C₂₈H₂₅N₂O₈F•H₂O: C, 60.65; H, 4.91; N, 5.05. Found: C, 60.13; H, 5.14; N, 4.62. ¹H Nmr (CDCl3, 300 MHz): δ 7.25 (2H, dd, \underline{I} =5, 9 Hz, arom.-H), 7.00 (2H, t, \underline{I} =9 Hz, arom.-H), 6.83 (1H, s, H-5), 6.77 (1H, br s, NH), 6.48

(1H, s, H-8), 6.25 (2H, s, H-2',6'), 5.94 and 5.92 (each 1H, s, -O-CH₂-O-), 5.15 (1H, dd, <u>I</u>=4, 7 Hz, H-4), 5.01 (1H, br s, NH), 4.48 (1H, d, <u>I</u>=5 Hz, H-1), 4.40 (1H, dd, <u>I</u>=1, 9 Hz, H-11), 3.97 (1H, t, <u>I</u>=9 Hz, H-11), 3.73 (6H, s, OCH₃), 2.95 (1H, m, H-3), 2.77 (1H, dd, <u>I</u>=4, 14 Hz, H-2).

Synthesis of N-Nitroso Compounds. To a solution of 4β -N-ureido compounds (5, 8 or 9, 1 mmol) in a mixture of chloroform (50 ml) and formic acid (200 mg), ground sodium nitrite (150 mg, 2.2 mmol) was added and was stirred for 1 h at 0 °C. The reaction mixture was washed with water, dried over sodium sulfate, and concentrated to give a residue, which was purified by silica gel chromatography [CHCl3-EtOAc (9:1) or hexane-acetone (2:1)].

4β-(N'-Nitroso-N'-methylureido)-3',4'-didemethoxy-3',4'-dioxo-4-deoxypodophyllotoxin (13). Yield 40%. Brown powder; mp 175 °C (decomp.); $[\alpha]_D^{20} + 118.0^\circ$ (\underline{c} =0.1, CHCl3). Anal. Calcd for C₂₂H₁₉N₃O₉•1/2H₂O: C, 55.23; H, 4.21. Found: C, 55.67; H, 4.65. ¹H Nmr (CDCl₃, 300 MHz): δ 8.18 (1H, d, <u>I</u>=8 Hz, NH), 7.00 (1H, s, H-5), 6.71 (1H, s, H-8), 6.46 (1H, d, <u>I</u>=1.5 Hz, H-2'), 6.00 (2H, s, -O-CH₂-O-), 5.46 (1H, dd, <u>I</u>=4, 8 Hz, H-4), 5.36 (1H, d, <u>I</u>=1.5 Hz, H-6'), 4.45 (1H, d, <u>I</u>=4 Hz, H-1), 4.55 (1H, dd, <u>I</u>=1, 9 Hz, H-11), 4.23 (1H, dd, <u>I</u>=2, 9 Hz, H-11), 3.76 (3H, s, OCH₃), 3.68 (1H, dd, <u>I</u>=4, 14 Hz, H-2), 3.26 (1H, m, H-3), 3.17 (3H, d, <u>I</u>=5 Hz, NHCH₃).

4β-[N'-Nitroso-N'-(2"'chloroethylureido)]-3',4'-didemethoxy-3',4'-dioxo-4-deoxypodo-phyllotoxin (14). Yield 33%. Brown powder; mp 200 °C (decomp.); $[\alpha]_D^{20}$ + 126.1° (\underline{c} =0.1, CHCl₃). Anal. Calcd for C₂4H₂5N₂O₈Cl•H₂O: C, 55.12; H, 5.20; N, 5.36. Found: C, 55.11; H, 5.15; N, 4.78. ¹H Nmr (CDCl₃, 300 MHz): δ 7.11 (1H, d, \underline{I} =8 Hz, NH), 6.86 (1H, s, H-5), 6.48 (1H, s, H-8), 6.43 (1H, d, \underline{I} =1.5 Hz, H-2'), 5.21 (1H, d, \underline{I} =1.5 Hz, H-6'), 5.94 (2H, s, -O-CH₂-O-), 5.25 (1H, dd, \underline{I} =4, 7 Hz, H-4), 4.24 (1H, d, \underline{I} =5 Hz, H-1), 4.60 (1H, dd, \underline{I} =1, 9 Hz, H-11), 4.14 (2H, t, \underline{I} =6 Hz, H-2"), 4.10 (1H, t, \underline{I} =9 Hz, H-11), 3.75 (3H, s, OCH₃), 3.49 (2H, t, \underline{I} =6 Hz, H-1"), 3.15 (1H, dd, \underline{I} =2, 14 Hz, H-2), 3.00 (1H, m, H-3).

4β-(N-Nitroso-N-Isopropyl)-3',4'-didemethoxy-3'-oxo-4'-hydroxy-4'-acetonyl-4-deoxy-podophyllotoxin (16). Yield 33%. Yellow powder; mp 195 °C (decomp.); $[\alpha]_D^{20}$ – 56.9° (\underline{c} =0.47, CHCl3). Anal. Calcd for C26H28N2O9•1/2H2O: C, 59.87; H, 5.61; N, 5.37. Found: C, 60.28; H, 5.57; N, 5.33. ¹H Nmr (CDCl3, 300 MHz): δ 6.65 (1H, s, H-5), 6.59 (1H, s, H-8), 6.04 (1H, d, I=4 Hz, H-4), 6.05, 6.01 (each 1H, s, -O-CH2-O-), 5.77, 5.02 (each 1H, br s, H-2' and 6'), 4.50 (1H, dd, I=1, 9 Hz, H-11), 4.32 (1H, d, I=4 Hz, H-1), 3.82 (3H, s, OCH3), 3.63 (1H, m, H-1"), 3.23 (1H, t, I=9 Hz, H-11),

3.20 (1H, m, H-3), 3.08, 2.90 (each 1H, s, <u>I</u>=15 Hz, acetonyl-CH₂), 2.78 (1H, dd, <u>I</u>=5, 14 Hz, H-2), 2.09 (3H, s, acetonyl-CH₃), 1.54, 1.53 (each 3H, d, <u>I</u>=6.5 Hz, 1"-CH₃).

Biological Assay. Assays for the inhibition of human DNA topoisomerase II, for the production of cellular protein-linked DNA breaks, and for the cytotoxicity towards KB cells were carried out according to the procedures described previously. 11

ACKNOWLEDGEMENT

The authors thank Dr. Mike Fisher of the Cancer Research Center, UNC-Chapel Hill, for the KB cell culture assay. This work was supported by a grant from the American Cancer Society, No. DHP-13F (K. H. Lee).

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Received, 3rd February, 1994