

CONSOLARINE, A NOVEL NORDITERPENOID ALKALOID FROM *CONSOLIDA ARMENIACA*

Ali H. Meriçli,^a Filiz Meriçli,^a Ayhan Ulubelen,^{a,b} Haridutt K. Desai,^c Balawant S. Joshi,^c S. William Pelletier*,^{c,d} Seçkin Özden,^e and Mustafa Küçükislamoglu^f

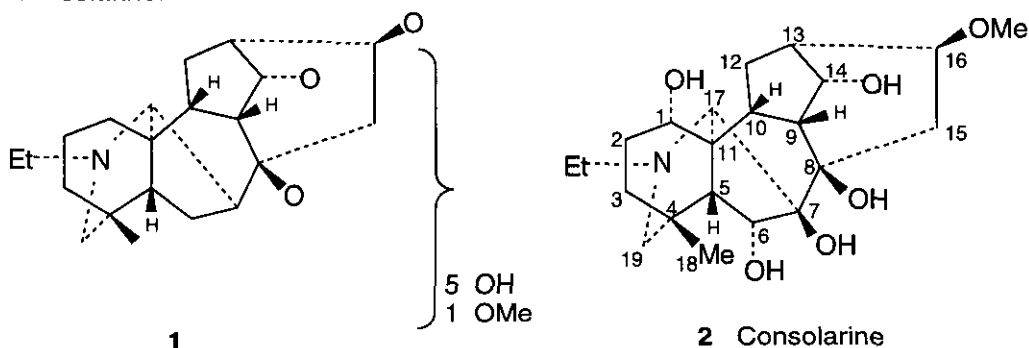
^aFaculty of Pharmacy, University of Istanbul, 34452, Istanbul, Turkey, ^bTUBITAK, Marmara Research Center, Department of Chemistry, P. O. Box 21, 41470, Gebze, Kocaeli, Turkey, ^cInstitute for Natural Products Research, ^dInstitute for Natural Products Research and Department of Chemistry, The University of Georgia, Athens, Georgia 30602-2556, U. S. A., ^eFaculty of Pharmacy, University of Ankara, Tandogan 06100, Ankara, Turkey, and ^fFaculty of Education, Technical University of Karadeniz, Trabzon, Turkey

(Dedicated to Professor Koji Nakanishi on the occasion of his 75th Birthday)

Abstract—From the aerial parts of *Consolida armeniaca*, (Stapf. Ex Huth.) Schröd. a new norditerpenoid alkaloid named *consolarine* has been isolated along with the known alkaloids ajadelphinine, gigactonine, and lycotone. The structure of *consolarine* (**2**) was established on the basis of ¹H, ¹³C, DEPT, homonuclear ¹H COSY, HETCOR, NOESY, and COLOC NMR spectral studies.

Except for the isolation of four acylated anthocyanin pigments from the flowers of *Consolida armeniaca* (Stapf. Ex Huth.) Schröd.,¹ no phytochemical work appears to have been carried out earlier on this plant. In continuation of our studies on the alkaloids of Turkish *Delphinium*, and *Consolida* species,²⁻⁹ an investigation of the aerial parts of *C. armeniaca* led to the isolation of a novel norditerpenoid alkaloid *consolarine*. The crude alkaloid isolated from *C. armeniaca* at pH 10 was purified on an Al₂O₃ column by VLC and six fractions (A-F) were collected. By chromatographic separation of the third fraction on an Al₂O₃ rotor, an amorphous homogeneous alkaloid designated as *consolarine* was isolated. The EIMS and the HRMS indicated the molecular ion at m/z 409 suggesting the formula C₂₂H₃₅NO₆ for the alkaloid. The NMR spectra showed that the alkaloid contains an *N*-ethyl group (δ_c 13.6, q; δ_H 1.12, 3H, t, J = 7.2 Hz; δ_c 50.9 t; δ_H 2.92, 2.99, 2H, AB J_{gem} = 11.0 Hz) and a methoxyl group (δ_c 57.0, q; δ_H 3.36, 3H, s) accounting for three carbons. Biogenetic

considerations and the molecular formula $C_{22}H_{35}NO_6$ indicated that consolarine is a norditerpenoid alkaloid. As there are no carbonyl functionalities, ether oxygens or methylenedioxy groups, the alkaloid should contain five hydroxyl groups and one methoxyl group. The 1H and ^{13}C NMR spectra (Table 1) indicated the presence of a tertiary methyl (δ_c 30.2 q; δ_H , 1.27, 3H, s) group. As no other functional groups are discernible in the IR or the NMR spectra, a partial structure (1) can be written for consolarine.



The quaternary carbon signals at δ 33.7, 47.5 and 77.5 can be readily assigned to C-4, C-11 and C-8, respectively.¹⁰ The fourth quaternary carbon signal at δ 82.8 remains to be assigned. This carbon bearing a hydroxyl group can be located at either C-5, C-7, C-9, C-10 or C-13. The positions C-5, and C-10 can be discounted as the adjacent carbons C-4 and C-11 would have shown downfield shifts of ~ 4-6 ppm from their normal positions as in bonvalotine and bonvalol,¹¹ having a C-5 OH; and deltamine and dictyocarpine^{12,13} having a C-10 OH groups, respectively. The carbon signal at δ 75.2 (δ_H 4.12, 1H, t, $J = 3.5$ Hz) is clearly assigned to C-14 bearing an α hydroxyl group, not having any substituents on the adjacent carbons C-9 and C-13. There are numerous examples in support of this argument.⁹ Hence, the tertiary hydroxyl group should be located on the remaining position at C-7, consistent with the chemical shift of this quaternary carbon at δ 82.8.

The problem of locating the two secondary hydroxyls thus remains. One of the hydroxyls is present at C-1 (δ_c 72.2 d; δ_H , 3.63, 1H, br s, $W_{1/2} = 4.5$ Hz). This proton shows a correlation with the protons of H-2 (δ_c 28.9 t; δ_H , 1.50, 1H, m) in the COSY spectrum and with H-12 in the NOESY spectra (Table 2). The H-2 proton (δ_H , 1.50) in turn shows a correlation with one of the H-3 protons (δ_c 34.6 t; δ_H , 1.50, 1.80, 2H, m) in the COSY. The H-1 proton shows correlation with C-10 (δ_c 43.4) and C-11 (δ_c 47.5) in the COLOC spectrum (Figure 1). The remaining hydroxyl group should therefore be located at C-6, in preference to C-12 or C-15, which are the only other methylene groups. The H-15 protons (δ_c , 36.2, t; δ_H , 1.90, 2.85, 2H, m) shows COSY and NOESY correlations with H-16 (δ_H , 3.30, 1H, m). The proton signal for C-15 (δ_c 36.2 t; δ_H , 1.90, 2.85, 2H, m) is correlated with C-8 (δ_c 77.5) in the COLOC. The H-16 proton shows a correlation with H-13 (δ_c , 39.9 d; δ_H , 2.25, 1H, d d) and a W-type coupling with H-14.

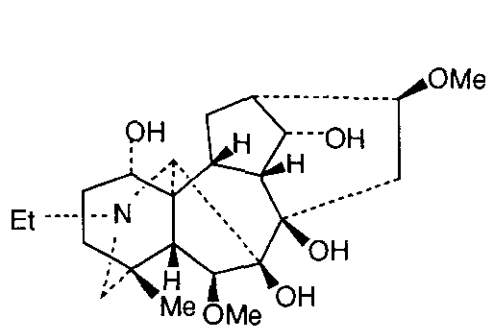
Table 1. ^1H and ^{13}C Chemical shifts assignments of consolarine (2) and dihydrogadesine (3) (in CDCl_3)

| Carbon | δ (ppm) | | Proton | δ (ppm) | J (Hz) | |
|-------------------|--------------------------------|------|-----------------------|-------------------|-------------|---------------------------------------|
| (2) | (3) | (2) | | | | |
| 1 | 72.2 (<i>d</i>) [*] | 72.8 | 1b | 3.63 [#] | br <i>s</i> | $w_{1/2} = 4.5$ |
| 2 | 28.9 (<i>t</i>) | 29.3 | 2 | 1.50 | <i>m</i> | |
| 3 | 34.6 (<i>t</i>) | 32.2 | 3a, b | 1.50, 1.80 | <i>m</i> | |
| 4 | 33.7 (<i>s</i>) | 33.1 | 4 | - | - | |
| 5 | 50.1 (<i>d</i>) | 50.6 | 5 | 2.00 | <i>d</i> | $J_{5,6} = 6.7$ |
| 6 | 69.8 (<i>d</i>) | 91.0 | 6b | 4.49 | <i>d</i> | $J_{6,5} = 6.7$ |
| 7 | 82.8 (<i>s</i>) | 87.9 | 7 | - | - | |
| 8 | 77.5 (<i>s</i>) | 78.1 | 8 | - | - | |
| 9 | 46.8 (<i>d</i>) | 45.3 | 9 | 2.20 | <i>m</i> | |
| 10 | 43.4 (<i>d</i>) | 44.1 | 10 | 1.85 | <i>m</i> | |
| 11 | 47.5 (<i>s</i>) | 49.2 | 11 | - | - | |
| 12 | 29.6 (<i>t</i>) | 29.3 | 12a,b | 1.30, 1.70 | <i>m</i> | |
| 13 | 39.9 (<i>d</i>) | 39.4 | 13 | 2.25 | <i>d d</i> | $J_{13,12b} = 4.9$ |
| 14 | 75.2 (<i>d</i>) | 75.7 | 14 | 4.12 | <i>t</i> | $J_{14,9} = 3.5$ $J_{13,14} = 3.5$ |
| 15 | 36.2 (<i>t</i>) | 34.5 | 15a,b | 1.90, 2.85 | <i>m</i> | |
| 16 | 82.1 (<i>d</i>) | 82.1 | 16 | 3.30 | <i>m</i> | |
| 17 | 63.7 (<i>d</i>) | 65.7 | 17 | 2.80 | <i>s</i> | |
| 18 | 30.2 (<i>q</i>) | 27.7 | 18 | 1.27 | <i>s</i> | |
| 19 | 60.3 (<i>t</i>) | 60.8 | 19a,b | 2.36, 2.82 | AB | $J_{\text{gem}} = 11.0$ |
| N-CH ₂ | 50.9 (<i>t</i>) | 50.3 | N-CH ₂ a,b | 2.92, 2.99 | <i>m</i> | $J_{\text{vic}} = 7.2$ |
| CH ₃ | 13.6 (<i>q</i>) | 13.7 | CH ₃ | 1.12 | <i>s</i> | |
| C-6' | - | 58.0 | - | - | - | |
| C-16' | 57.0 (<i>q</i>) | 56.3 | OCH ₃ | 3.36 | <i>s</i> | |
| | | | 6-OH | 3.30 | br <i>m</i> | |

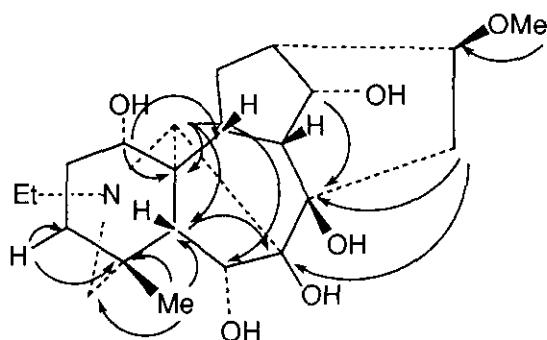
^{*} Multiplicity deduced by DEPT[#] Carbon showing long-range correlation with indicated protons deduced by HETCOR

Table 2. ^1H , ^1H correlations and nOe's of consolarine (2)

| Observed H | Correlations (COSY) | nOe's (NOESY) |
|-----------------------------------|---|--|
| H-1 β | H-2 | H-12 _a |
| H-2 | H-1 β , H-3 | - |
| H-3 _a | H-3 _b , CH ₃ -18 | N-CH ₂ CH ₃ |
| H-3 _b | H-3 _a | |
| H-5 | H-6 | H-6, CH ₃ -18 |
| H-6 | H-5 | H-5, H-9 |
| H-9 | H-10, H-13, H-14 | H-6 |
| H-10 | H-9, H-12 _b | - |
| H-12 _a | H-12 _b , H-13 | H-1 β |
| H-12 _b | H-10, H-12 _a | - |
| H-13 | H-9, H-12 _a , H-14, H-16 (W) | H-14, H-16 |
| H-14 | H-9, H-13, H-16 | H-13, H-16 |
| H-15 _a | H-15 _b , H-16 | H-15 _b |
| H-15 _b | H-15 _a | H-15 _a , H-16 |
| H-16 | H-13, H-14, H-15 _a , H-15 _b | H-13, H-14, H-15 _b , |
| H-17 | - | H-16 |
| CH ₃ -18 | H-3 _b | H-5 |
| H-19 _a | - | CH ₃ -18, H-19 _b |
| H-19 _b | - | CH ₃ -18, H-19 _a |
| N-CH ₂ CH ₃ | - | H-3 _a |
| N-CH ₂ CH ₃ | - | CH ₃ -18, N-CH ₂ CH ₃ |



3 Dihydrogadesine

Figure 1
COLOC Correlations of consolarine 2

Thus, the five hydroxyl groups of the alkaloid are located at C-1, C-6, C-7, C-8 and C-14 and the methoxyl is at C-16 in the partial structure (1). In most of the lycotoxine-type norditerpenoid alkaloids, the hydroxyl group at C-6 is in a β position and H-C-6 is α and the Dreiding model reveals the

H(5 β)-H(6 α) dihedral angle $\sim 90^\circ$. The chemical shift for H-6 normally appears at $\sim \delta$ 4.5, corresponding with 6- α H and $J_{5\beta,6\alpha} = \sim 0$ Hz. In the present alkaloid, the H-6 signal (δ_c 69.8 d; δ_H , 4.49, 1H, d, $J = 6.7$ Hz) clearly shows that the hydroxyl group is in an α -position. In the aconitine-type alkaloids, where the C-6 methoxyl is in an α -position, e.g., acoforesticine,¹⁴ acoforestinine,¹⁴ and flavaconidine,¹⁵ C-6- β H appears at $\sim \delta$ 4.1 as a doublet, showing a coupling of ~ 6 Hz with H-5. The coupling with H-7 is negligible as the dihedral angle is $\sim 90^\circ$. Consolarine indicates similarity to the C-6 α hydroxyl group of pubescenine (δ_c 70.8 d; δ_H , 4.45, 1H, d, $J = 6.8$ Hz, H-6 β),¹⁶ as against 6-*epi*-pubescenine where H-6 appears as a singlet showing no coupling with H-5.¹⁷ A Dreiding model shows that H-6 α hydroxyl group is in close proximity to the C-18 methyl group (δ_c 30.2 q; δ_H , 1.27, 3H, s). In a 1D difference nOe experiment, the C-18 methyl group showed an nOe to the C-6 α hydroxyl group which appears at δ 3.30. A plausible explanation for the downfield shifts of the C-18 methyl in the ^{13}C and ^1H spectra (δ_c 30.2; δ_H , 1.27), from the normal values (δ_c 25.0-26; δ_H , 0.8-1.1) lies in the deshielding influence of the 6- α -hydroxyl group on the C-18 methyl. On the basis of these data, structure (2) has been assigned to consolarine. Consolarine is the only example of a lycoctonine-type norditerpenoid alkaloid bearing a C-18 methyl and C-6 α OH groups. The structure of consolarine (2) resembles dihydrogadesine (3) (see Table 1 for ^{13}C NMR spectra) except for a β -methoxyl at C-6 instead of a C-6 α -hydroxyl group.¹⁸

By chromatographic separation of fractions (A) and (E) on an Al_2O_3 rotor, the known alkaloids ajadelphinine,¹⁹ lycoctonine¹⁰ and gigactonin²⁰ were isolated.

EXPERIMENTAL

General Experimental Procedures.— IR spectra were recorded in CHCl_3 on a Perkin-Elmer Model 983 spectrophotometer. HRMS were determined on a VG Zap Spec instrument and Perkin-Elmer SCIEX AP1-1 mass spectrometer. NMR spectra including DEPT and 2D experiments, were recorded in CDCl_3 on a Bruker AC-250 and AC 300 spectrometers. The pulse sequences employed for the NMR experiments were those of the standard Bruker software. ^1H and ^{13}C NMR and COLOC spectra were determined on Bruker AC 200L instrument. Optical rotations were determined on Opt. Act Ltd AA-5 polarimeter. Chromatographic separations on a Chromatotron were carried out on rotors coated with 1 mm thick layers of Merck Al_2O_3 60 PF 254, 365 (EM 1104).

Plant Material.— The aerial parts of *Consolida armeniaca* were collected by one of the authors (M. K.) and identified by (A. H. M. and F. M.) (August 1996) near the Black sea between Gümüşhane-Bayburt 30 km from Bayburt at an elevation of 1650 m. A voucher specimen is deposited in the Herbarium of the Faculty of Pharmacy, University of Istanbul ISTE 72679.

Extraction of Crude Alkaloids.— Dried and powdered aerial parts of *C. armeniaca* (2.5 kg) were exhaustively extracted by percolation at rt with 95% EtOH. Evaporation (*in vacuo*) of the combined extracts gave a gummy residue which was dissolved in CH_2Cl_2 (500 mL) and extracted with 2% H_2SO_4 (200 mL x 10). The acidic extract was washed with CH_2Cl_2 (200 mL x 3) and then basified

to pH 10 with cold aq. 10% NaOH. Extractions with CH_2Cl_2 (250 mL x 5) and evaporation of the combined extracts in *vacuo* gave a crude mixture of alkaloids (2.5 g).

Purification of the Alkaloidal Mixture.— The crude alkaloidal mixture was chromatographed by VLC²¹ on an Al_2O_3 column. The eluting solvent was a gradient of hexane, EtOAc and MeOH and six fractions (A-F) were collected. These were separated on Al_2O_3 rotors of a Chromatotron. Fraction (A) was chromatographed on an Al_2O_3 rotor and gradient eluted with hexane, EtOAc, MeOH to afford ajadelphinine (30 mg) as an amorphous product. The identity was established by comparison of the TLC, ^1H and ^{13}C NMR spectra with an authentic sample. From the second fraction (B) (163 mg), consolarine (**2**) was obtained as an amorphous compound (103 mg), $[\alpha]_{\text{D}} + 0.65^\circ$ (c, 0.77, CHCl_3). EIMS: m/z 409 (M^+ , 75%), 394 (11), 376 (30), 353 (35), 145 (30), 122 (28), 58 (72). HRMS: Found, 409.2450; Calculated. for $\text{C}_{22}\text{H}_{35}\text{NO}_6$, 409.24643. IR (nujol): ν_{max} 3280, 2920, 2880, 1705, 1640, 1565, 1450, 1400, 1300, 1220, 1162, 1170, 1100 cm^{-1} . For ^1H and ^{13}C NMR spectra, see Table 1. The fraction (E) was chromatographed on an Al_2O_3 rotor to afford lycoc-tonine (190 mg) and gigactonine (25 mg). The alkaloids were identified by comparison of the TLC, ^1H and ^{13}C NMR spectra with those of authentic samples.

ACKNOWLEDGEMENTS

The authors thank NATO for a Collaborative Research Grant (CRG 931261). The Turkish authors also thank Istanbul University Research Fund for Grant No. UP 2-150197. Partial financial support provided by Grant HL 32562 from the National Institutes of Health is gratefully acknowledged.

REFERENCES

1. H. Saita, K. Toki, S. Ozden, and T. Honda, *Phytochemistry*, 1996, **41**, 1599.
2. A. Ulubelen, H. K. Desai, B. S. Joshi, B. P. Hart, S. W. Pelletier, A. H. Meriçli, F. Meriçli, and H. Ç. Özen, *J. Nat. Prod.*, 1996, **59**, 907.
3. A. Ulubelen, H. K. Desai, S. K. Srivastava, B. P. hart, J. C. Park, B. S. Joshi, S. W. Pelletier, A. H. Meriçli, F. Meriçli, and R. Ilarsan, *J. Nat. Prod.*, 1996, **59**, 360.
4. F. Meriçli, A. H. Meriçli, H. Becker, A. Ulubelen, S. Özden, N. Dürüst, and M. Tanker, *Phytochemistry*, 1996, **42**, 1249.
5. A. Ulubelen, H. K. Desai, B. S. Joshi, V. Venkateswarlu, S. W. Pelletier, A. H. Meriçli, F. Meriçli, and H. Ü. Özçelik, *J. Nat. Prod.*, 1995, **58**, 1555.
6. A. Ulubelen, A. H. Meriçli, and F. Meriçli, *Nat. Prod. Lett.*, 1994, **5**, 135.
7. A. Ulubelen, A. H. Meriçli, F. Meriçli, and R. Ilarslan, and W. Voelter, *Phytochemistry*, 1993, **34**, 1165.
8. A. Ulubelen, A. H. Meriçli, and F. Meriçli, *J. Nat. Prod.*, 1993, **56**, 780.
9. A. Ulubelen, A. H. Meriçli, F. Meriçli, F. Ilarslan, and S. A. Matlin, *Phytochemistry*, 1992, **31**, 3239.

10. S. W. Pelletier, N. V. Mody, B. S. Joshi, and L. C. Schramm, ^{13}C and proton NMR Assignments and Physical Constants of C_{19} -Diterpenoid Alkaloids, in *Alkaloids, Chemical and Biological Perspectives*, Vol. 2, Ed. S. W. Pelletier, Wiley, N. Y. 1983, pp. 206-462.
11. Q. P. Jiang and W. L. Sung, *Heterocycles*, 1984, **22**, 2429.
12. A. S. Narzullaev, M. S. Yunusov, and Y. S. Yunosov, *Khim. Prior. Soedin*, 1973, **46**, 443.
13. S. Pelletier, N. V. Mody, and O. D. Dailey, *Can. J. Chem.*, 1980, **58**, 1875.
14. S. W. Pelletier, B. S. Joshi, J. A. Glinski, H. P. Chokshi, S. Y. Chen, K. T. Bhandary, and K. T. Go, *Heterocycles*, 1987, **25**, 365.
15. Z. G. Chen, A. N. Lao, H. C. Wang, and S. H. Hong, *Heterocycles*, 1989, **29**, 997.
16. G. de la Fuente, R. D. Acosta, J. A. Gavin, R. H. Lugo, and P. G. Jones, *Tetrahedron Letters*, 1989, **29**, 2723.
17. Y. Bai, M. Benn, and W. Majak, *Heterocycles*, 1989, **29**, 1017.
18. A. G. Gonzalez, G. de la Fuente, and R. Diaz, *Phytochemistry*, 1982, **21**, 1781; A. G. Gonzalez, G. de la Fuente, M. Reina, and R. D. Acosta, *Heterocycles*, 1986, **24**, 1513.
19. S. W. Pelletier, S. Bhandaru, H. K. Desai, S. A. Ross, and H. M. Sayed, *J. Nat. Prod.*, 1992, **55**, 736.
20. S. Sakai, N. Shinma, S. Hasegawa, and T. Okamoto, *J. Pharm. Soc. (Japan)*, 1978, **98**, 1376.
21. S. W. Pelletier, H. P. Chokshi, and H. K. Desai, *J. Nat. Prod.*, 1986, **49**, 892.

Received, 22nd April, 1997