SYNTHETIC STUDY OF USTILOXIN ANALOGS: BENZYLIC OXIDATION OF 13-MEMBERED CYCLIC PEPTIDE BY LEAD TETRAACETATE¹

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<u>Abstract</u>- The 13-membered cyclic peptides (5, 6), analogs of phomopsin-ustiloxin class of antibiotics, have been synthesized. Benzylic hydroxyl group was introduced in desired stereochemistry by lead tetraacetate oxidation followed by methanolysis.

There are a number of natural and synthetic compounds that interfere with microtubule function by binding to tubulin. Maytansine,² rhizoxin,³ dolastatin 10,⁴ phomopsin A(1)⁵ and ustiloxin A(2)⁶ are known to share the same binding site (RZX-MAYsite)^{7,8} on tubulin. However, their structural diversity remains it difficult to find their common structural elements to recognize the same binding site. Ustiloxin D(3),⁸ isolated as the minor component of ustiloxin family from the water extract of false smut balls, also exhibits potent antitubulin activity. The common structure found in 1, 2 and 3 demonstrates that 13-membered core structure is

Figure 1

responsible for anti-tubulin activity. Therefore, ustiloxin D was regarded as the promising candidate to elucidate the structural requirement for the RZX-MAYsite ligands.

We have been working in the synthesis of ustiloxin analogs to find the minimal structure responsible for anti-tubulin activity. In the previous report, synthesis of non-substituted 13-membered cyclic peptides (4a) and (4b) was described. However, both compounds did not inhibit the microtubule assembly even at higher concentration ($IC_{50} > 100 \mu$ M). Therefore, as the next challenge, we planned to synthesize more functionalized 13-membered cyclic peptides (5) and (6) to elucidate the minimal active structure as effective RZX-MAYsite ligand. Our synthetic strategy described below should make it possible not only to introduce a variety of functional groups into the 13-membered ring but also to modify the side chain easily.

Synthesis of **5** and **6** are achieved as shown in **Scheme 1** and **Scheme 2**. Starting L-DOPA derivative (**7**) was prepared from L-tyrosine by Boger's procedure.¹⁰ Williamson alkylation of **7** with tosylate (**8**) derived from D-serine, with Cs₂CO₃ as base, gave the corresponding aryl alkyl ether (**9**). Deprotection of *N*-Boc group of **10** with TFA gave the ammonium salt, which was condensed with *N*-Boc-L-valine to give the amide (**11**). Protection of primary hydroxy group as TBDPS (*tert*-butyldiphenylsilyl) ether followed by basic saponification afforded carboxylic acid (**12**). Macrolactamization of **12** was accomplished successfully according to the protocol developed by Schmidt.¹¹ An activation of **12** with DIPC (diisopropylcarbodiimide) and pentafluorophenol provided pentafluorophenyl ester (**13**), which was used without purification for the next

Scheme 1

Scheme 2

cyclization. Removal of *N*-Boc-protective group with TFA and successive macrolactamization in two phases system (CHCl₃-1M KHCO₃) under high dilution at 50°C gave 13-membered cyclic peptide (15) in 28% from 11 along with 34% of dimeric cyclic peptide (16). Removal of TBDPS group by tetra-*n*-butylammonium fluoride (TBAF), *N*-Cbz group and Bn group by hydrogenolysis gave 5. In order to introduce the oxygen function at benzylic carbon of 15, lead tetraacetate oxidation in AcOH was performed at 80°C to give acetate (17) as a single diastereomer in 30%.¹²⁻¹⁴ Removal of acetyl, TBDPS, *N*-Boc and Bn groups by K₂CO₃/MeOH, TBAF and hydrogenolysis gave 6 with hydroxyl group at benzylic center.

The absolute configuration of benzylic center of 6 was determined to be R, the same configuration as 3, by the comparison of its ¹H-NMR chemical shift and coupling constant with these of 1 and 3 (Figure 2).¹⁵ The benzylic and phenethylic protons of 6 appear at 4.43 and 3.17 ppm whereas these protons of 3 appear at 4.41 and 3.15 ppm, respectively. On the other hand, these protons of 1 appear at 5.03 and 4.07 ppm, respectively. Similarly, coupling constant between above protons are 3.9Hz for 1 and 7.6Hz for 3 and 6. The course of stereoselectivity is reasonably understood that the conjugate addition of acetic acid took place from the less hindered face of quinone methide transition state 18.

Figure 2

Our successful synthesis of 5 and 6 would provided a basis of future synthesis of phomopsin-ustiloxin class of antibiotics. Further studies on the structure-activity relationship of ustiloxin by synthetic approach and chemical degradation of ustiloxin A are in progress.¹⁶

ACKNOWLEDGMENT

This work was supported in part by Grant-in-Aid for Scientific Research from the Ministry of Education, Science, Sports and Culture, Japan. We thank Dr. Naoko Morisaki for FABMS and HRFABMS measure-

ments, Mrs. Hiroko Hino for elemental analyses.

REFERENCES AND NOTES

- Dedicated to Professor Koji Nakanishi, Columbia University, on the occasion of his 75th birthday.
- 2. S. M. Kupchan, Y. Komoda, W. A. Court, G. J. Thomas, R. M. Smith, A. Karim, C. J. Gilmore, R. C. Haltwanger, and R. F. Bryan, *J. Am. Chem. Soc.*, 1972, **94**, 1354.
- 3. S. Iwasaki, H. Kobayashi, J. Furukawa, M. Namikoshi, S. Okuda, Z. Sato, I. Matsuda, and T. Noda, J. Antibiot., 1984, 37, 354.
- 4. G. R. Pettit, Y. Kamano, C. L. Herald, A. A. Tuinman, F. E. Boetter, H. Kizu, J. M. Schmidt, L. Baczynskyj, K. B. Tomer, and R. J. Bontems, J. Am. Chem. Soc., 1987, 109, 6883.
- 5. M. F. Mackay, A. V. Donkelaar, and C. C. J. Culvenor, J. Chem. Soc., Chem. Commun., 1986, 1219.
- 6. Y. Koiso, M. Natori, S. Iwasaki, S. Sato, R. Sonoda, Y. Fujita, H. Yaegashi, and Z. Sato, *Tetrahedron Lett.*, 1992, 33, 4157.
- 7. S. Iwasaki, Med. Res. Rev., 1993, 13, 183.
- 8. Y. Koiso, Y. Li, S. Iwasaki, K. Hanaoka, T. Kobayashi, R. Sonoda, Y. Fujita, H. Yaegashi, and Z. Sato, J. Antibiot., 1994, 47, 765.
- 9. R. Mutoh, R. Shirai, Y. Koiso, and S. Iwasaki, Heterocycles, 1995, 41, 9.
- 10. D. L. Boger and D. Yohannes, J. Org. Chem., 1987, 52, 5283.
- 11. U. Schmidt, H. Griesser, A. Lieberknecht, and J. Talbiersky, *Angew. Chem., Int. Ed. Engl.*, 1981, 20, 280. U. Schmidt, H. Griesser, A. Lieberknecht, and J. Talbiersky, *J. Org. Chem.*, 1982, 47, 3261.
- 12. O. Dimroth and R. Schweizer, Ber., 1923, 56, 1375.
- 13. W. S. Johnson, J. M. Anderson, and W. E. Shelberg, J. Am. Chem. Soc., 1944, 66, 218.
- 14. G. W. K. Cavill and D. H. Solomon, J. Chem. Soc., 1954, 3943.
- 15. H-NMR spectra of 6, phomopsin A and ustiloxin D were measured in d_ε-DMSO at rt.
- 16. An inhibitory activity of microtubule assembly was determined as described previously. M. Takahashi, S. Iwasaki, H. Kobayashi, S. Okuda, T. Murai, Y. Sato, T. Haraguchi-Hiraoka, and H. Nagano, *J. Antibiot.*, 1987, **40**, 66. Although ustiloxin D exhibited strong anti-tubulin activity ($IC_{50} = 3.6\mu M$), both **5** and **6** did not inhibit the microtubule assembly ($IC_{50} > 100 \mu M$).

Received, 12th May, 1997