LEPIOTINS A AND B, NEW ALKALOIDS FROM THE MUSHROOMS, MACROLEPIOTA NEOMASTOIDEA AND CHLOROPHYLLUM MOLYBDITES

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Abstract—Two new alkaloids, lepiotins A and B were isolated from the poisonous mushrooms Macrolepiota neomastoidea and Chlorophyllum molybdites, which were classified to family Agaricaceae. These structures were determined on the basis of a spectroscopic analysis and a synthetic study.

Agaricaceous mushroom Chlorophyllum molybdites closely related to the genus Macrolepiota has similar form to M. neomastoidea. Both mushrooms C. molybdites and M. neomastoidea have been known to cause similar poisoning with severe gastrointestinal symptoms including intestinal irritant, vomiting and profuse diarrhea. Hypovolumetric shock from C. molybdites has also been reported. In the course of our program to find a new biologically active compound from Japanese mushroom (Basidiomycetes), we investigated the acute toxicity of the methanol extract on mice, and found temporary poisoning such as rising with both extracts from C. molybdites and M. neomastoidea. Attempts to find a toxic component led to the isolation of two new alkaloids, lepiotins A

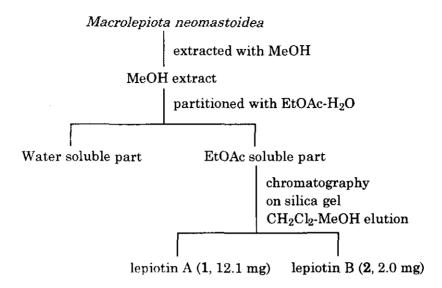


Chart 1. Isolation Procedure for Lepiotins A (1) and B (2) from M. neomastoidea

and B from the fruit bodies of M. neomastoidea and lepiotin B from C. molybdites. In this paper, we wish to report the isolation and the structural elucidation of lepiotins A (1) and B (2).

1) Lepiotins A (1) and B (2) from M. neomastoidea.

The fruit bodies of M. neomastoidea collected in Miyagi Prefecture were extracted with The methanol extract was partitioned between ethyl acetate and water. methanol. The ethyl acetate soluble part was concentrated in vacuo and the residue was repeatedly subjected to silica gel chromatography using dichloromethane-methanol mixed solvent system of increasing polarity to afford lepiotins A (1, 12.1 mg) and B (2, 2.0 mg). Lepiotin A (1) was obtained as a colorless oil. The molecular formula C₁₀H₁₁NO₃ was determined by the combination of HREI MS (m/z 193.0725, M+), ¹H and ¹³C NMR spectra. In the IR spectrum lepiotin A (1) showed the hydroxyl (3300) and carbonyl (1650 cm⁻¹) absorptions. Ferric chloride coloring test on TLC indicated the presence of phenol moiety. The ¹³C NMR spectrum of lepiotin A (1) showed ten signals due to a carbonyl carbon of amide or ester moiety ($\delta_{\rm C}$ 176.6), six sp² and three sp³ carbons as shown in Table 1. The ¹H and ¹³C NMR spectra of lepiotin A (1) exhibited signals for two sets of methine groups (δ_H/δ_C 6.85/116.8, 7.22/127.7) on 1,4-disubstituted phenol ring (partial structure P-A in Figure 1) The H-H COSY and a methine group (δ_H/δ_C 5.48/86.8) attached to two hetero atoms.

position	13C	·····	ıH
2	176.6	(s) ^b	
3	30.5	(t)	2.75-2.77 (1H, m, H _{3a})
			2.45-2.48 (1H, m, H _{3b})
4	29.2	(t)	2.45-2.48 (1H, m, H _{4a})
			2.02-2.05 (1H, m, H _{4b})
5	86.8	(d)	5.48 (1H, dd, <i>J</i> =4.5, 2.0, H ₅)
6	129.7	(a)	
7, 11	127.7	(d)	7.22 (2H, dd, <i>J</i> =8.0, 2.0)
8, 10	116.8	(d)	6.85 (2H, dd, <i>J</i> =8.0, 2.0)
9	156.9	(s)	

Table 1. ^{1}H and ^{13}C NMR Spectral Data for Lepiotin A (1)a

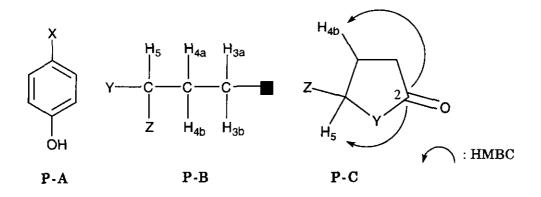


Figure 1. Partial Structures for Lepiotin (1)

^a CD₃OD, 300 MHz for ¹H, 75 MHz for ¹³C NMR. Coupling constants (J) were given in Hz.

 $^{^{\}rm b}$ q: methyl, t: methylene, d: methine, s: quaternary carbon.

Figure 2. Structures of Lepiotin A (1) and Structure A

correlations from H_5 to H_{4a} and H_{4b} as well as the one from H_{4b} to H_{3a} indicated a partial structure **P-B** as shown in Figure 1. The HMBC spectrum supported the 1,4-disubstituted phenol moiety (**P-A**) and the HMBC correlations from carbonyl carbon (2-C) to methine (H_5) and methylene (H_{4a}) hydrogens indicated a five-membered ring (partial structure **P-C**). Taking into account of hetero atoms in the molecular formula, a nitrogen and an oxygen atoms must be assigned to atoms X, Y and Z, two of those must be the same atom, in the partial structures **P-A** and **P-C**. Thus the partial structures **P-A** and **P-C** assembled to the structures (1) and (A) as shown in Figure 2. The 13 C chemical shift of C-6 at δ 129.7 indicated a nitrogen as a neighboring atom (structure 1) instead of an oxygen (structure A).

For the structural confirmation and the estimate of the biological activity, lepiotin A (1) was synthesized from 4-aminophenol (3). 4-Aminophenol (3) and succinic anhydride were dissolved in pyridine-toluene (1:1). After adding triethylamine, the reaction mixture was heated under reflux for 48 h to give the imide (4) in 75% yield. The imide (4) in dichloromethane-methanol (1:1) was reduced with NaBH₄ at -15- -10 °C for 1 h to give

Scheme 1.

Table 2. ¹ H and ¹³ C NMR	Spectral Data	for Lepiotin B (2)a
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position	¹³ C	$^{1}\mathrm{H}$		
2	175.0	(s) ^b		
3	29.7	(t)	2.68-2.82	(1H, m)
			2.44-2.55	(1H, m)
4	24.8	(t)	2.23-2.37	(1H, m)
			2.08-2.17	(1H, m)
5	92.9	(d)	5.20	(1H, dd, J=6.1, 1.1)
6	130.1	(s)		
7, 11	126.1	(d)	7.22	(2H, ddd, J =8.7, 3.1, 2.1)
8, 10	116.1	(d)	6.75	(2H, ddd, J =8.7, 3.1, 2.1)
9	154.7	(s)		
O-CH ₃	54.2	(p)	3.26	(3H, s)
			5.80	(1H, br)

^a CD₃OD, 300 MHz for ¹H, 75 MHz for ¹³C NMR. Coupling constants (J) were given in Hz.

aminal (1) in 62% yield, that was identical with lepiotin A (1).⁴ Lepiotin B (2) was isolated as a colorless oil. The molecular formula $C_{11}H_{13}NO_3$ was determined by the combination of HREI MS (m/z 207.0912, M+), ¹H and ¹³C NMR spectra. The latter two spectra were similar to those of lepiotin A (1) indicating two sets of methine groups (δ_H/δ_C 6.75/116.1, 7.22/126.1) on 1,4-disubstituted phenol ring and a methine group (δ_H/δ_C 5.20/92.9) attached to two hetero atoms, except for a methyl (δ_H/δ_C 3.26/54.2) in lepiotin B (2) as shown in the Figure 3. Analyses of the 2D NMR spectra (H-H COSY, HMQC and HMBC) of lepiotin B (2) indicated similar γ -lactam ring as that of lepiotin A (1). HMBC correlations from a carbonyl (δ_C 175.0) to a methine (δ_H 5.20), and a methine (δ_C 92.9) to a

 $^{^{\}rm b}$ q: methyl, t: methylene, d: methine, s: quaternary carbon.

methoxyl (δ_H 3.26) along with the molecular formula indicated that the hetero atom X must be a nitrogen. The gross structure of lepiotin B is thus represented by the structure (2) as shown in Figure 3.

Figure 3. Gross Structure of Lepiotin B (2)

2) Isolation of lepiotin B (2) from C. molybdites

The ethyl acetate soluble part derived from the fruit bodies of *C. molybdites* as the similar procedure in Chart 1 was repeatedly subjected to silica gel chromatography using hexane-ethyl acetate solvent system and purified by preparative HPLC [ODS, methanolwater (3:7)] to afford lepiotin B (2, 2.3 mg) which was identical with the one isolated from *M. neomastoidea*.

Lepiotins A (1) and B (2) are new alkaloids having a γ-lactam and a phenol rings. Lepiotin A (1) is optically inactive implying an epimeric mixture related to the anomeric equilibrium at C-5. Although lepiotin B (2) showed optical activity, the absolute configuration at C-5 was not clarified.

Methanol extracts of *M. neomastoidea* and *C. molybdites* exhibited transient toxicity that included rising at the dose of 100 mg/kg on mice. Synthetic lepiotin A (1), however, did not show any activity at less than 400 mg/kg. We are now investigating assay system itself because of low toxicity of the methanol extracts of both mushrooms on mice.

EXPERIMENT

UV spectra was recorded on a HITACHI U-3200 and IR spectra on JASCO A-100S and FT/IR 5300 spectrophotometer. ¹H and ¹³C NMR spectra were recorded on JEOL JNM GX500 (500 MHz for ¹H and 125 MHz for ¹³C) and VARIAN Gemini 2000 (300 MHz for ¹H and 75 MHz for ¹³C) spectrometers with TMS as an internal standard. Coupling

constants (J) were given in Hz. High resolution electron impact (HREI) MS spectra were recorded on a JEOL JMS DX303 spectrometer. Optical rotation was recorded on a JASCO DIP-370 spectrometer.

Isolation of lepiotins A (1) and B (2) from M. neomastoidea

The fruit bodies of M. neomastoidea (280 g) collected in Miyagi Prefecture were extracted three times with MeOH (3 x 400 mL) at rt for 30 h to give an extract (20 g) after being filtered and the solvent evaporated. This extract was partitioned between water and EtOAc. The organic layer was concentrated in vacuo and the residue (1.0 g) was repeatedly chromatographed on silica gel with mixtures of CH₂Cl₂ and MeOH of increasing polarity. Elution with CH₂Cl₂-MeOH (9:1) gave lepiotins A (1, 12.1 mg) and B (2, 2.0 mg). Lepiotin A (1), colorless oil. UV λ (2-PrOH) nm (log ϵ): 281.8 (3.28), 230.4 (4.32), 203.0 (4.02).IR v (CHCl₃): 3300, 2340, 1690, 1520, 760 cm⁻¹. ¹H NMR (CD₃OD, 500 MHz): δ 2.02-2.05 (1H, m), 2.45-2.48 (2H, m), 2.75-2.77 (1H, m), 5.48 (1H, dd, J=4.5, 2.0), 6.85 (2H, dd, *J*=8.0, 2.0), 7.22 (2H, dd, *J*=8.0, 2.0). ¹³C NMR (CD₃OD, 125 MHz): δ 29.2 (t), 30.5 (t), 86.8 (d), 116.8 (d), 116.8 (s), 127.7 (d), 156.9 (s), 176.6 (s). HREI MS: m/z 193.0725 (M+), Calcd 193.0739 for C₁₀H₁₁NO₃.

Lepiotin B (2), colorless oil. [α]_D²² +3.75° (c 0.160, MeOH). UV λ (MeOH) nm (log ε): 279.0 (3.13), 232.2 (3.77), 202.4 (4.09). IR ν (CHCl₃): 3270, 2350, 1680, 1510, 1070 cm⁻¹. ¹H NMR (CDCl₃, 300 MHz): δ 2.08-2.17 (1H, m), 2.23-2.37 (1H, m), 2.44-2.55 (1H, m), 2.68-2.82 (1H, m), 3.26 (3H, s), 5.20 (1H, dd, J=6.1, 1.1), 6.75 (2H, ddd, J=8.7, 3.1, 2.1), 7.22 (2H, ddd, J=8.7, 3.1, 2.1). ¹³C NMR (CDCl₃, 75 MHz): δ 24.8 (t), 29.7 (t), 54.2 (q), 92.9 (d), 116.1 (d), 126.1 (d), 130.1 (s), 154.7(s), 175.0 (s). HREI MS: m/z 207.0912(M⁺), Calcd 207.0895 for C₁₁H₁₃NO₃.

Isolation of lepiotin B (2) from C. molybdites

The fruit bodies of *C. molybdites* (186 g) collected in Shiga Prefecture were extracted three times with MeOH (3 x 300 mL) at rt for 30 h to give an extract (42 g) after being filtered and the solvent evaporated. This extract was partitioned between water and EtOAc. The organic layer was concentrated *in vacuo* and the residue (1.0 g) was chromatographed on silica gel (elution with hexane-EtOAc 1:1) and purified by preparative HPLC [ODS, MeOH-H₂O (3:7)] to yield lepiotin B (2, 2.3 mg) as a colorless oil. $[\alpha]_D^{22} + 3.75^{\circ}$ (c 0.160, MeOH), UV λ (MeOH) nm (log ϵ): 279.0 (3.13), 232.2 (3.77), 202.4 (4.09). IR ν (CHCl₃):

3270, 2350, 1680, 1510, 1070 cm⁻¹. ¹H NMR (CDCl₃, 300 MHz): δ 2.08-2.17 (1H, m), 2.23-2.37 (1H, m), 2.44-2.55 (1H, m), 2.68-2.82 (1H, m), 3.26 (3H, s), 5.20 (1H, dd, J=6.1, 1.1), 6.75 (2H, ddd, J=8.7, 3.1, 2.1), 7.22 (2H, ddd, J=8.7, 3.1, 2.1). ¹³C NMR (CDCl₃, 75 MHz): δ 24.8 (t), 29.7 (t), 54.2 (q), 92.9 (d), 116.1 (d), 126.1 (d), 130.1 (s), 154.7(s), 175.0 (s). HREI MS: m/z 207.0912 (M+), Calcd 207.0895 for C₁₁H₁₃NO₃.

N-(4-Hydroxyphenyl)succimide (3)

4-Aminophenol (3, 1.1 g, 10 mmol) and succinic anhydride (1.5 g, 15 mmol) were dissolved in pyridine-toluene (1:1, 100 mL). After triethylamine (250 μ L, 2.5 mmol) was added, the reaction mixture was heated under reflux for 48 h. The mixture was evaporated *in vacuo*, and the residue was chromatographed over silica gel (100 g). Elution with CH₂Cl₂-MeOH (9:1) afforded imide (4, 1.4 g, 75 %) as colorless columns, mp 260-265 °C (MeOH). IR v (CHCl₃): 3690, 1712 cm⁻¹. ¹H NMR (CDCl₃, 500 MHz): δ 1.50 (4H, s), 6.80 (2H, d, J=8.3), 7.15 (2H, d, J=8.3). ¹³C NMR (CDCl₃, 125 MHz): δ 29.0 (t), 117.0 (d), 125.0 (s), 129.5 (d), 127.7 (d), 159.0 (s), 180.0 (s). HREI MS: m/z 191.0574 (M⁺), Calcd 191.0583 for C₁₀H₉NO₃.

5-Hydroxy-N-(4-hydroxyphenyl)-2-pyrrolidinone (1)

To the imide (4, 1.5 g, 7.9 mmol) in CH_2Cl_2 -MeOH (1:1, 150 mL) was added NaBH₄ (5 g, 132 mmol) at -15--10 °C and the mixture was stirred for 1 h. The reaction mixture was evaporated *in vacuo* and the residue was chromatographed on silica gel (300 g). Elution with EtOAc afforded aminal (1, 933.7 mg, 62 %) as an oil. IR v (CHCl₃): 3300, 2340, 1690, 1520, 760 cm⁻¹. ¹H NMR (CDCl₃, 300 MHz): δ 2.02-2.05 (1H, m), 2.40-2.50 (2H, m), 2.70-2.80 (1H, m), 5.50 (1H, dd, J=5.0, 1.8), 6.80 (2H, d, J=9.0), 7.22 (2H, d, J=9.0). HREI MS: m/z 193.0719 (M⁺), Calcd 193.0721 for $C_{10}H_{11}NO_3$.

Acute toxicity test

Male ddy mice, 6 weeks old, were used in this study. The animals were housed in air conditioned room at rt $(22\pm1~^{\circ}\text{C})$ and with standard food and tap water available adlibitum. In this study five mice per group were used. The samples (100, 200, 400 mg/kg) were dissolved in 2 % gum Arabic solution and were injected intraperitoneally (ip) in mice. The appearance of mice was observed every an hour.

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REFERENCES

- 1. P. F. Lehmann and U. Khazan, Mycopathologia, 1992, 118, 3.
- 2. P. H. Stenklyft and W. L. Augenstein, J. Toxicol. Clin. Toxicol., 1990, 28, 159.
- T. Ohta, A. Takahashi, M. Matsuda, S. Kamo, T. Agatsuma, T. Endo, and S. Nozoe, Tetrahedron Lett., 1995, 36, 5223; M. Matsuda, T. Kobayashi, S. Nagao, T. Ohta, and S. Nozoe, Heterocycles, 1996, 43, 685, and references cited there in.
- 4. T. Ohta, S. Shiokawa, E. Iwashita, N. Sato, K. Sakurai, T. Ineyama, H. Izawa, K. Izawa, and S. Nozoe, *Heterocycles*, 1992, 33, 143.

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