Institute, National Institutes of Health, U.S. Public Health Service, Bethesda, MD. The authors are grateful to Dr. Herbert Lazarus, formerly of the Dana-Farber Cancer Institute and now at the University of Miami School of Medicine, and Dr. John M. Idriss of these laboratories for the cell culture determinations and also to Linda B. Anderson for assistance with the in vivo antitumor assays. Generous supplies of bulk daunorubicin hydrochloride for this work were kindly provided by Farmitalia-Carlo Erba,

Milan, Italy, through the assistance of Adria Laboratories, Inc., Columbus, OH.

Registry No. 2·HCl, 23541-50-6; 7, 26295-55-6; 8, 65026-79-1; 9, 77270-18-9; 10, 97191-43-0; 11, 97191-44-1; 12, 80787-28-6; 13, 80787-29-7; 14, 97191-45-2; 15, 97191-46-3; 16, 97191-47-4; monosodium malonate, 2922-55-6; monosodium succinate, 2922-54-5; monosodium glutarate, 3343-88-2; monosodium adipate, 18996-34-4; monosodium pimelate, 6142-21-8; monosodium azelate, 17356-30-8; monosodium sebacate, 19455-73-3.

Synthesis and Antimetastatic Properties of Stereoisomeric Tricyclic Bis(dioxopiperazines) in the Lewis Lung Carcinoma Model¹

Donald T. Witiak,*† Raghunathan V. Nair,† and Franz A. Schmid‡

Division of Medicinal Chemistry and Pharmacognosy, College of Pharmacy, The Ohio State University, Columbus, Ohio 43210, and Laboratory of Experimental Antitumor Testing, Sloan-Kettering Institute for Cancer Research, Walker Laboratory, Rye, New York 10580. Received November 14, 1985

Synthesis of trans- and cis-tetrahydrodipyrazino[1,2-a:1',2'-d]pyrazine-1,3,7,9(2H,4H,8H,10H)-tetrone analogues 10 and 11 belonging to the bis(dioxopiperazine) class of antitumor agents and their bis(morpholinomethyl) derivatives 12 and 13 are described with use of 2,5-dimethylpyrazine as the starting material. Synthetic studies utilizing 3,6-disubstituted 2,5-dioxopiperazine precursors are included. Evaluation of 10–13 in the Lewis Lung carcinoma model indicated the bis(morpholinomethyl) analogue cis-13 to be antimetastatic, whereas the trans isomer 12 was toxic at a similar dose effecting a decrease in the life span of treated mice. The parent bis(dioxopiperazines) 10 and 11 were ineffective as antitumor or antimetastatic drugs.

Chemical, biochemical, and pharmacological properties of antitumor bis(dioxopiperazines) recently have been reviewed.² Cyclic analogues 4–9 of 1 and 2 exhibited stereoselective effects in various tumor models.^{3–6} In the solid state a cis "face to face" conformation of the dioxopiperazine rings were observed in antimetastatic racemic 2 and cis-5.^{7,8} However, such a conformation seems not to be essential for activity since tricyclic bis(dioxopiperazine) trans-6 exhibited antimetastatic properties in the B16-F10 melanoma model.^{5,9}

Recent reports from China¹⁰ have indicated the bis-(morpholinomethyl) derivative of 1 (namely, 3) to be effective in various human malignancies. Although such compounds are predictably unstable and undergo hydrolysis to the parent dioxopiperazines, morpholinomethyl-N groups may impart antineoplastic properties to a molecule owing to possible alkylating activities not unlike those proposed¹¹ for certain hydroxymethyl-N metabolites of therapeutically useful drugs. Comparative analysis of morpholinomethyl derivatives 8 and 9 with the respective parent imides 6 and 7 in a postoperative Lewis Lung (LL) carcinoma model revealed morpholinomethyl cis-syn-trans isomer 9 to be more effective as an inhibitor of metastasis than the other three analogues (6-8).12 Additionally, the order of decreasing activity was 9 > 8 > 7 > 6 when assessed in terms of survival or antimetastatic data. The increased activity observed for the morpholinomethyl derivatives over their respective parents was attributed12 either to increased solubility and drug delivery (prodrug) or to an intrinsic antitumor activity of the morpholinomethyl-N functionality possibly reflecting macromolecular alkylation. In this report we describe the synthesis of analogues 10-13 and biological evaluation of these compounds in the LL carcinoma model using the postoperative protocol. Such studies are of interest since previous investigations of conformationally constrained analogue pairs

(4, 5 and 6, 7) suggested that a "cisoid" relationship of imide functionalities was necessary for antimetastatic activity. 4-6 Clearly, interatomic distances between groups in 10 and 11 are markedly different than those found in 6 and 7. Consequently, selective antitumor activity observed for morpholinomethyl analogues would provide support for intrinsic antitumor activity of the morpholinomethyl-N functionality.

Chemistry. Our synthetic strategy utilized the analysis shown in Scheme I. Two pathways were explored for the synthesis of piperazines 20–22. One involved selective reduction of the amide functionalities in 3,6-disubstituted 2,5-dioxopiperazine precursors. Dioxopiperazine diester 23 of undefined stereochemistry was reported¹³ to have been formed on cooling a solution of ethyl aminomalonate

- (1) A portion of this work was presented to the Division of Medicinal Chemistry at the 187th National Meeting of the American Chemical Society in St. Louis, MO, April 8-13, 1984.
- Herman, E. H.; Witiak, D. T.; Hellmann, K.; Waravdekar, V. S. Adv. Pharmacol. Chemother. 1982, 19, 249-290.
- (3) Witiak, D. T.; Lee, H. J.; Hart, R. W.; Gibson, R. E. J. Med. Chem. 1977, 20, 630-635.
- (4) Witiak, D. T.; Lee, H. J.; Goldman, H. D.; Zwilling, B. S. J. Med. Chem. 1978, 21, 1194-1197.
- (5) Witiak, D. T.; Trivedi, B. K.; Campolito, L. B.; Zwilling, B. S.; Reiches, N. A. J. Med. Chem. 1981, 24, 1329-1332.
- (6) Zwilling, B. S.; Campolito, L. B.; Reiches, N. A.; George, T.; Witiak, D. T. Br. J. Cancer 1981, 44, 578-583.
- (7) Hempel, A.; Camerman, N.; Camerman, A. J. Am. Chem. Soc. 1982, 104, 3453-3456.
- (8) Hempel, A.; Camerman, N.; Camerman, A. J. Am. Chem. Soc. 1982, 104, 3456-3458.
- (9) Hempel, A.; Camerman, N.; Camerman, A. J. Am. Chem. Soc. 1983, 105, 2350-2354.
- (10) Ren, Y. F.; Shu, H. L. Zhang, T. M.; Chen, Z. Y.; Lin, C. Kexue Tongbao 1980, 25, 189.
- (11) Soloway, A. H.; Brumbaugh, R. J.; Witiak, D. T. J. Theor. Biol. 1983, 102, 361–373.
- (12) Witiak, D. T.; Trivedi, B. K.; Schmid, F. A. J. Med. Chem. 1985, 28, 1111.
- (13) Cheeseman, G. W. H. J. Chem. Soc. 1960, 242-247.

[†]The Ohio State University.

[‡]Sloan-Kettering Institute for Cancer Research.

in acetone. Later reports by Hino and Sato^{14,15} indicated that ethyl aminomalonate did not dimerize to 21 at elevated temperatures. Our results were in agreement with these later reports; ethyl aminomalonate prepared according to the method of Schipper and Day¹⁶ could not be dimerized under various conditions.

We therefore investigated the dioxopiperazinediol 24 as a possible precursor to the piperazinedicarboxylates 20-22. Geometric isomers of diol 24 were prepared according to the procedure of Rao and Ravindranath¹⁷ from commercially available serine methyl ester hydrochloride. Hydroxyl protection was necessary for further synthetic work since isomers of 24 are insoluble in most organic solvents. Attempts to prepare the THP ethers failed. Isomers of diacetates 25 were obtained from reaction of isomerically pure 24 with AcCl in HOAc, but such derivatization did not markedly improve solubility characteristics. Geometric assignments for 25 are tentative. Respective ¹³C and ¹H NMR spectra for the pure isomeric esters were nearly identical and thus not useful for defining stereochemistry.

Selective Borch reduction¹⁸ of cis-25 was not successful; reaction of cis-25 with triethyloxonium tetrafluoroborate

Scheme I

Scheme Π^a

H₃C
$$\stackrel{N}{\longrightarrow}$$
 CH₃ $\stackrel{A}{\longrightarrow}$ HO₂C $\stackrel{N}{\longrightarrow}$ HO₂C $\stackrel{\longrightarrow}$ HO₂C $\stackrel{N}{\longrightarrow}$ HO₂C $\stackrel{N}{\longrightarrow}$ HO₂C $\stackrel{N}{\longrightarrow}$ HO₂C $\stackrel{$

^a $a = SeO_2$, pyridine-water, reflux; b = KOH, H_2O , Pd/C, H_2 , 40-42 psi, 50-60 °C; c = HCl.

only afforded mono(imino ether) 28 in 25% yield. The desired bis(imino ether) could not be obtained even when using higher reaction temperatures or longer reaction times. Similarly, attempts to obtain the piperazine from tert-butyldimethylsilyl ether 26 prepared in 63-64% yield from pure cis- or trans-24 was not successful.

Synthesis of benzyl-protected diol 27 had been previously reported by Russian investigators. ¹⁹ Our attempts to reproduce their procedure resulted in poor yields of intermediates. Alternatively, Fisher esterification of

⁽¹⁴⁾ Hino, T.; Sato, T. Tetrahedron Lett. 1971, 3127-3129.

⁽¹⁵⁾ Hino, T.; Sato, T. Chem. Pharm. Bull. 1974, 22, 2866-2874.

⁽¹⁶⁾ Schipper, E.; Day, A. R. J. Am. Chem. Soc. 1952, 74, 350-353.

⁽¹⁷⁾ Rao, K. V.; Ravindranath, B. J. Heterocycl. Chem. 1975, 12, 147-149.

⁽¹⁸⁾ Borch, R. F. Tetrahedron Lett. 1968, 61-65.

⁽¹⁹⁾ Kiryushkin, A. A.; Shchelokov, V. I.; Antonov, V. K.; Ovchinnikov, Yu A.; Shemyakin, M. M. Khim. Prir. Soedin. 1967, 3, 267-275; Chem. Abstr. 1968, 69, 77709V.

commercially available O-benzyl-L-serine followed by chromatography of the resultant methyl ester hydrochloride (Amberlite IRA-45/CH₃OH) and solvent removal yielded an oily eluate which dimerized, affording a cistrans mixture of 27 in 57% yield upon standing at room temperature for 7 days. Although these isomers could be separated by chromatography (silica gel/CHCl₃), for convenience, the mixture was employed in subsequent reactions. The dioxopiperazine geometry could not be assigned from their NMR spectra but could be confirmed following conversion to the corresponding piperazines 29. LiAlH₄ reduction of diastereomeric 27 afforded corresponding piperazines 29, which upon treatment with iodoacetamide/K₂CO₃ provided diastereomers 30 in 64% yield. Isomers were separated (silica gel/CHCl₃) and their structures confirmed by NMR analysis (see Experimental Section).

Attempted debenzylation of diamide 30 using catalytic hydrogenation (Pd/C), 20 transfer hydrogenation, 21,22 Na/liquid NH₃, 23 or Me₃SiI²⁴ were unsuccessful owing to competing N-dealkylation.

The second route provided intermediate piperazine-2,5-dicarboxylates 20-22 by reduction of the corresponding pyrazine precursors. Preparation of 20 had been previously reported, 25 but no evidence for the proposed geometry was presented. We prepared 20 from the corresponding pyrazine 31, which was obtained in 65-71% yield from SeO₂ oxidation of commercially available 2,5-dimethylpyrazine (Scheme II).²⁶ Catalytic hydrogenation of an alkaline solution of 31 using 10% Pd on carbon afforded an isomeric mixture of 20, which was separated by fractional crystallization under controlled pH. Gradual acidification of the concentrate initially crystallized trans-20 (pH ~ 5.5-6.5). Further reduction in pH crystallized cis-20. Geometric configurations were assigned by ¹H NMR analysis.

Piperazine tetraacids 14 were obtained by alkylation of the pure isomers of diacids 20 with use of bromoacetic acid and K₂CO₃ in water. Use of chloroacetamide under similar reaction conditions afforded the diamide diacid trans-15 from trans-20. cis-15 could not be obtained by this pro-Attempts to prepare the target bis(dioxopiperazines) 10 and 11 from the tetraacids 14 using $\mathrm{NH_{3}}^{27}$ urea,28 or formamide29 resulted in decomposition of 14.

Similarly, refluxing in Ac₂O,³⁰ heating neat, use of DCC, or heating in acidic solvents (CH₃CO₂H, H₂SO₄, or PPA) did not effect cyclization of trans-15 to 10. While Fisher esterification of trans-15 afforded tetraester trans-16, other methods of esterification³¹⁻³³ did not produce the desired amide ester trans-17 presumably owing to the insoluble nature of trans-15. Both cis and trans tetraesters 16 also were obtained in good yields (69-74%) from Fisher esterification of the corresponding tetraacids 14. Although reaction of tetraesters 16 with liquid NH3 in a sealed tube produced excellent yields of the corresponding tetraamides 18 and none of the bis(imides), reaction of either isomer of 16 with NH₃ in MeOH/NaOMe afforded small amounts (≤16%) of desired trans-10 in addition to the major product trans-18. Likewise, although heating 16 with formamide failed to produce the bis(dioxopiperazines) 10 or 11, reaction with formamide in the presence of NaH/ refluxing dimethoxyethane or dioxane afforded 10 as the exclusive product in about 25% yield. Heating either isomer of tetraamide 18 in PPA at 85 °C for 15-30 min afforded small amounts (10–15%) of 10; decomposition of the reactants was predominant.

In order to further improve the yield of 10 and also to obtain samples of the cis isomer 11, we investigated use of intermediates 21 and 22. Diacids 20 upon refluxing in methanolic HCl afforded the corresponding diesters 21. Attempts to prepare diester diamides 17 by alkylation of 21 with iodoacetamide was unsuccessful. Alternatively, reaction of 21 with liquid NH3 in a sealed tube afforded excellent yields of diamides 22. N-Alkylation of 22 using ethyl bromoacetate and K2CO3 in Me2SO afforded the diamide diesters 19, which upon refluxing in EtOH/NaOEt afforded the target bis(dioxopiperazines) 10 and 11 in 71-89% yield. Conversion to the corresponding bis-(morpholinomethyl) derivatives 12 and 13 took place in Me₂SO in the presence of morpholine and formaldehyde. Unlike morpholinomethyl analogues 8 and 9, these compounds were virtually insoluble in water.

NMR Spectral Analysis. Characterization of isomeric tricycles 10 and 11 was based in part upon analysis of the 500-MHz proton NMR spectrum of precursor piperazine-2,5-dicarboxylic acids cis- and trans-20. The piperazine ring proton resonance signals for trans-20 exhibited an AMX pattern. The axial HA resonance signal of the ring methylene group appeared as a deceptively simple triplet at δ 3.64. The geminal (14.1 Hz) and axial-equatorial (3.8 Hz) coupling constants were derived by analysis of the H_M equatorial doublet of doublets (δ 4.21) of the ring methylene group. The H_X methine doublet of doublets centered at δ 4.53 exhibited axial-equatorial and diaxial (12.4 Hz) couplings.

For cis-20, which rapidly interconverts on the NMR time scale, an ABX pattern was observed for the piperazine proton resonance signals wherein calculated constants were $\delta_{\rm A}$ 3.93 and $\delta_{\rm B}$ 4.02 ($J_{\rm AX}$ = 4.4 Hz and $J_{\rm BX}$ = 7.1 Hz) with $\delta_{\rm X}$ (observed) 4.71 and $J_{\rm AB}$ (observed) = 14.5 Hz. Splitting patterns similar to those exhibited by cis- and trans-20 were identified, respectively, for cis and trans diesters 21 and diamides 22 and the diester diamide trans-19. The cis diester diamide 19 provided a first-order spectrum at 90 MHz which was in agreement with the proposed

⁽²⁰⁾ Heathcock, C. H.; Ratcliffe, R. J. Am. Chem. Soc. 1971, 93, 1746-1757.

Anantharamaiah, G. M.; Sivanandaiah, K. M. J. Chem. Soc., Perkin Trans. 1 **1977**, 490–491.

Felix, A. M.; Heimer, E. P.; Lambros, T. J.; Tzougraki, C.;

Meienhofer, J. J. Org. Chem. 1978, 43, 4194-4196. Reist, E. J.; Bartuska, V. J.; Goodman, L. J. Org. Chem. 1964,

Jung, M. E.; Lyster, M. A. J. Org. Chem. 1977, 42, 3761-3764.

⁽²⁵⁾ Felder, Von E.; Maffei, S.; Pietra, S.; Pitre, D. Helv. Chim. Acta 1960, 43, 888-896.

Schut, W. J.; Mager, H. I. X.; Berends, W. Recl. Trav. Chim. Pays-Bas 1961, 80, 391-398.

⁽²⁷⁾ Buckles, R. E.; Probst, W. J. J. Org. Chem. 1957, 22, 1728-1729.

Crockett, G. C.; Swanson, B. J.; Anderson, D. R.; Koch, T. H. Synth. Commun. 1981, 11, 447-454.

Creighton, A. M. Brit. Patent 1234935, 1971.

Henry, D. W. J. Heterocycl. Chem. 1966, 3, 503-511.

Arai, I.; Muramatsu, I. J. Org. Chem. 1983, 48, 121-123.

Mohacsi, E. Synth. Commun. 1982, 12, 453-456.

Rachele, J. R. J. Org. Chem. 1963, 28, 2898.

Table I. Lewis Lung Carcinoma (LL) Metastasis Study^a

					autopsy data ^b				
	survival data					av lung	av no. of metastasis		
compd	survival days	MSTc	% ILS ^d	N/T^e	av body wt, g	wt, mg	<2 mm	>2 mm	M/T^f
control	20, 23, 24, 25, 26, 28, 28, 28, 29, 32	27.0		0/10	16.3	741	0	49	10/10
10	18, 26, 28, 28, 31	28.0	4	0/5	16.9	853	0	48	4/4
11	17, 19, 28, 29, >40	28.0	4	1/5	19.3	640	0	22	2/3
12	17, 25, 26, 26, >40	26.0	<0	1/5	18.9	700	0	38	3/4
13	19, 32, >40, >40, >40	>40	>48	3/5	19.2	358	0	6	1/4

^a BDF₁ female (19-21 g) mice; implantation on day 0. Amputation on day 8. Post amputational schedule: 160 mg/kg from day 8, Q2Dx4 (ip). Compounds were administered in suspension (saline). b Autopsy data of dying mice and mice killed day 40. c Median survival time (days). d Increase in life span of 25% or greater indicates activity. Number of 40 days survivors/total mice. Number of mice with metastasis/total.

structure. Relative to tricyclic bis(dioxopiperazines), the spectrum of trans-10 exhibited a characteristic deceptively simple triplet for the H_A axial proton (δ 2.18) of the central ring methylene group not unlike that observed for the equivalent proton resonance signal in piperazine diacid precursor trans-20. The cis isomer 11 exhibited a spectrum (90 MHz) wherein the axial H_A and equatorial H_B resonance signals of the central ring methylene group appeared as broad multiplets between δ 2.7 and 2.9.

Biological Results and Discussion

The effects of compounds 10-13 in the postoperative LL carcinoma model is summarized in Table I. BDF₁ female mice weighing 19-21 g were used in the study. LL (10⁵ cells) was implanted im in the leg on day 0, and the tumor-bearing legs were amputated on day 8. Dying mice and the mice killed on day 40 following implantation were autopsied.

Among the four compounds evaluated (10–13), only the cis-bis(morpholinomethyl) analogue 13 markedly inhibited the metastasis. This isomer also provided for a significant increase in life span of the treated mice. This stereoselective antimetastatic effect is of particular interest in this series. Unlike the situation in the tetraazaperhydrophenanthrene series (6-9), wherein morpholinomethyl derivatives were considerably more water soluble than their parent imides, in the tetraazaanthracene series (10-13) all compounds were water insoluble. Since the parent bis-(dioxopiperazine) cis-11 did not exhibit any appreciable activity in this assay, it is unlikely that the activity of 13 is due to its metabolic conversion to 11. Rather, it seems likely that certain bis(morpholinomethyl) derivatives possess intrinsic antitumor activities. No obvious conclusions can be drawn to explain why cis ring junctures in both tricyclic systems provide the most effective morpholinomethyl derivatives in the LL model. Although morpholinomethyl derivatives are predictably unstable to hydrolysis, these data do not support the proposal that the activity of such derivatives is only attributable to the parent imide.34

Experimental Section

Melting points were determined in open capillaries with a Thomas-Hoover Uni-Melt apparatus and are uncorrected. Infrared spectra were recorded with a Beckman Model 4230 spectrophotometer. Nuclear magnetic resonance spectra were recorded with either a Bruker WP-80, HX-90E, 300-MHz or a Nicolet 500-MHz spectrophotometer. Me₄Si (CDCl₃, Me₂SO, pyridine-d₅) or TSP (D₂O) were used as internal standards unless otherwise specified. Chemical shifts are reported on the δ scale with peak multiplicities: c, complex; d, doublet; dd, doublet of doublets; m, multiplet; q, quartet; s, singlet; and t, triplet. Mass spectra were recorded with a Du Pont Model 21-491 mass spectrometer with a Model 21-094 data system. Elemental analyses were performed by Galbraith Laboratories, Inc., Knoxville, TN

cis-3,6-Bis[(acetyloxy)methyl]piperazine-2,5-dione (25) was prepared by using methodology similar to the methodology used for the prepartion of the (chloroacetyl)oxy analogue.¹⁷ A mixture of cis-24 (1.0 g, 5.75 mmol), AcCl (1.8 mL, 25.1 mmol), and glacial HOAc (2.5 mL) was stirred at room temperature for 5 h. An additional 2.5 mL of HOAc was added at the end of 1.5 h. The mixture was diluted with Me₂CO, filtered, and recrystallized from MeOH, yielding 1.15 g (77.7%) of white crystals: mp 227-228 °C; IR (KBr) 1755 (ester), 1685 (amide) cm⁻¹; NMR $(Me_2SO-d_6, 90 MHz) \delta 2.04 (s, 6 H, COCH_3), 4.21 (br s, 6 H,$ OCH₂CH), 8.42 (s, 2 H, CONH). Anal. (C₁₀H₁₄N₂O₆) C, H, N.

trans-3,6-Bis[(acetyloxy)methyl]piperazine-2,5-dione (25) was prepared by using methodology similar to that employed for the preparation of the (chloroacetyl)oxy analogue.¹⁷ A mixture of trans-24 (1.0 g, 5.75 mmol), AcCl (1.8 mL, 25.1 mmol), HOAc (5 mL), and 10 drops of H₂SO₄ was stirred for 2 days at room temperature with occasional warming. The mixture was diluted with H₂O, filtered, and recrystallized from MeOH, yielding 0.53 g (35.8%) of white crystals: mp 237-238 °C; IR (KBr) 1740 (ester), 1680 (amide) cm⁻¹; NMR (Me₂SO- d_6 , 90 MHz) δ 2.02 (s, 6 H, $COCH_3$), 4.22 (s, br, 6 H, OCH_2CH), 8.37 (s, 2 H, CONH). Anal. $(C_{10}H_{14}N_2O_6)$ C, H, N.

cis-3,6-Bis[(acetyloxy)methyl]-5-ethoxy-3,6-dihydro-2-(1H)-pyrazinone (28). To a suspension of cis-25 (0.5 g, 1.94 mmol) in methylene chloride (10 mL) under nitrogen was added 4 mL of a 1 M solution of triethyloxonium tetrafluoroborate (Et₃O+BF₄-) in CH₂Cl₂ and the mixture stirred at room temperature. A second 4-mL portion of the Et₃O⁺BF₄⁻ solution was added after 6 h. The mixture was stirred at room temperature for 24 h and then carefully poured onto saturated aqueous NaHCO₃ solution (30 mL). The organic layer was separated, washed with H₂O, dried (Na₂SO₄), and concentrated under reduced pressure. The residue was recrystallized from petroleum ether, affording 0.13 g (25%) of a white solid: mp 118-120 °C; IR (KBr) 1740 (ester), 1700 (N=C), 1675 (amide) cm⁻¹; NMR $(CDCl_3, 90 \text{ MHz}) \delta 1.27 \text{ (t, 3 H, } OCH_2CH_3), 2.05 \text{ (s, 3 H, } COCH_3),$ 2.12 (s, 3 H, COCH₃), 4.03-4.50 (c, 8 H, OCH₂CH plus OCH₂CH₃), 6.60 (s, 1 H, CONH); MS (70 eV), m/e 286 (M⁺).

cis-3,6-Bis[[(tert-butyldimethylsilyl)oxy]methyl]piperazine-2,5-dione (26). A mixture of cis-24 (1.0 g, 5.75 mmol), t-BuMe₂SiCl (2.08 g, 13.8 mmol), and imidazole (1.96 g, 28.7 mmol) in DMF (15 mL) was stirred at room temperature for 10 h. The reaction mixture was diluted with H2O and the solid extracted with CHCl₃. The organic layer was washed with H₂O, dried (Na₂SO₄), and evaporated under reduced pressure. The solid obtained was recrystallized from EtOAc, yielding 1.47 g (63.5%) of white crystals: mp 188-190 °C; IR (KBr) 1680 (amide) cm⁻¹; NMR (CDCl₃, 90 MHz) δ 0.11 [s, 12 H, Si(CH₃)₂], 0.92 [s, 18 H, Si(t-Bu)], 3.7-4.17 (m, 6 H, OCH₂CH), 6.30 (s, br, 2 H, CONH). Anal. $(C_{18}H_{38}N_2O_4Si_2)$ C, H, N.

trans-3,6-Bis[[(tert-butyldimethylsilyl)oxy]methyl]piperazine-2,5-dione (26). A mixture of trans-24 (1.0 g, 5.75 mmol), t-BuMe₂SiCl (2.08 g, 13.8 mmol), and imidazole (1.96 g, 28.7 mmol) in DMF (5 mL was stirred at room temperature for 22 h. The reaction mixture was diluted with H₂O, and the solid was filtered and recrystallized from MeOH to yield 1.47 g (63.6%) of white crystals: mp 226-228 °C; IR (KBr) 1680 and 1690 (amide)

⁽³⁴⁾ Camerman, N.; Hempel, A.; Camerman, A. Science 1984, 225, 1165-1166

cm $^{-1}$; NMR (pyridine- d_5 , 90 MHz) δ 0.12 [s, 12 H, Si(CH₃)₂], 0.91 [s, 18 H, Si(*t*-Bu)], 4.1–4.47 (m, 6 H, OCH₂CH). Anal. (C₁₈-H₃₈N₂O₄Si₂) C, H, N.

cis- and trans-3,6-Bis[(phenylmethoxy)methyl]-2,5piperazinedione (27). To a suspension of O-benzyl-L-serine (12) g, 0.06 mol) in MeOH (100 mL) was passed HCl gas until the solution became clear. After refluxing for 6 h, the solvent was concentrated under reduced pressure. The residual O-benzyl-Lserine methyl ester hydrochloride was dissolved in MeOH (75 mL) and passed through a column (3 \times 46 cm) of weak base type ion exchange resin (Amberlite IRA-45, 16-50 mesh) which had been washed with 5% aqueous NaHCO3 solution (500 mL) followed by H₂O (600 mL) and MeOH (400 mL). The effluent and MeOH wash (500 mL) were concentrated to an oil, which was kept at room temperature for 7 days. The resulting semisolid was triturated with EtOAc and filtered to yield 6.17 g (56.7%) of cis- and trans-27. The isomers were separated on a silica gel column by elution with CHCl₃. The trans isomer eluted first and was recyrstallized from MeOH, affording white crystals: mp 205-207 °C; IR (KBr) 1675 (amide) cm⁻¹; NMR (CDCl₃, 300 MHz) δ 3.71 (dd, 2 H, OC H_2 CH, J_{AB} = 9.47 Hz, J_{AX} = 7.14 Hz), 3.80 (dd, 2 H, OCH_2CH , $J_{AB} = 9.47$ Hz, $J_{BX} = 3.34$ Hz), 4.18-4.22 (m, 2 H, OCH_2CH), 4.53 (d, 2 H, OCH_2Ph , $J_{AB} = 11.82$ Hz), 4.56 (d, 2 H, OCH_2Ph , $J_{AB} = 11.82$ Hz), 4.56 (d, 2 H, OCH_2Ph , $J_{AB} = 11.82$ Hz), 6.17 (s, 2 H, CONH), 7.27-7.38 (m, 10 H, Ph). Anal. (C₂₀H₂₂N₂O₄) C, H, N. Continued elution with CHCl₃ yielded the cis isomer as white crystals: mp (MeOH) 173-174 °C; IR (KBr) 1685 (amide) cm⁻¹; NMR (CDCl₃, 300 MHz) δ 3.67 (dd, 2 H, OCH2CH, $J_{\rm AM}$ = 9.24 Hz, $J_{\rm AX}$ = 8.28 Hz), 3.85 (dd, 2 H, OC H_2 CH, J_{AM} = 9.24 Hz, J_{MX} = 3.28 Hz), 4.18–4.22 (m, 2 H, OC H_2 CH), 4.44 (d, 2 H, OC H_2 Ph, J_{AB} = 11.69 Hz), 4.46 (d, $2 \text{ H, OCH}_2\text{Ph}, J_{AB} = 11.69 \text{ Hz}), 6.33 \text{ (s, } 2 \text{ H, CONH)}, 7.22-7.37$ (m, 10 H, Ph). Anal. $(C_{20}H_{22}N_2O_4)$ C, H, N.

cis- and trans-2,5-Bis[(phenylmethoxy)methyl]piperazine (29). A mixture of cis- and trans-27 (0.5 g, 1.4 mmol) and LAH (0.21 g, 5.6 mmol) in dry THF (50 mL) was refluxed under N_2 for 6 h. The mixture was cooled to room temperature and the excess LAH carefully decomposed with $Na_2SO_4\cdot 10H_2O$. The mixture was filtered, the salts were extracted twice with hot THF, and the combined extracts were concentrated under reduced pressure, yielding a semisolid mixture of cis- and trans-29 (0.43 g), which was not further purified but used as such for the preparation of cis- and trans-30.

cis- and trans-2,5-Bis[(phenylmethoxy)methyl]-1,4piperazinediacetamide (30). A mixture of crude cis- and trans-29 (4.42 g, \sim 0.01 mol), K_2CO_3 (3.74 g, 0.027 mol), and iodoacetamide (5.52 g, 0.03 mol) in absolute EtOH (50 mL) was stirred at room temperature for 24 h. The mixture was diluted with H₂O (50 mL) and extracted three times with CHCl₃ (50 mL). The combined extracts were washed with H_2O , dried (MgSO₄), and evaporated under reduced pressure to afford 5.19 g of crude cis- and trans-30 which were purified by chromatography (silica gel/CHCl₃). The trans isomer eluted first (1.17 g, 19.63%): mp (MeOH) 200-201 °C; IR (KBr) 1640 (amide) cm-1; NMR (CDCl₃, 90 MHz) δ 2.41 [deceptively simple triplet (dd), 2 H, axial H of ring CH₂], 2.57-3.60 (c, 12 H, other ring H plus NCH₂CO and OCH₂), 4.44 (s, 4 H, OCH₂Ph), 5.43 (br s, 2 H, CONH₂), 7.08 (br s, 2 H, CONH₂), 7.31 (s, 10 H, Ph). Anal. $(C_{24}H_{32}N_4O_4)$ C, H, N. Continued elution afforded an intermediate fraction (0.91 g, 15.27%, cis-trans mixture) followed by pure cis isomer (1.73 g, 29.02%): mp (EtOAc) 160-162 °C; IR (KBr) 1685 (amide) cm⁻¹; NMR (CDCl₃, 90 MHz) δ 2.56–3.59 (c, 14 H, ring H plus NCH₂CO and OCH₂), 4.47 (s, 4 H, OCH₂Ph), 5.54 (br s, 2 H, CONH₂), 7.01 (br s, 2 H, CONH₂), 7.31 (s, 10 H, Ph). Anal. $(C_{24}H_{32}N_4O_4)$ C, H, N.

Pyrazine-2,5-dicarboxylic acid (31) was prepared according to the procedure of Schut et al.²⁶ A mixture of 2,5-dimethylpyrazine (25.0 g 0.23 mol), pyridine (500 mL), SeO₂ (125 g, 1.13 mol), and H₂O (50 mL) was refluxed for 12-13 h in a 1-L round-bottom flask equipped with a mechanical stirrer and a reflux condenser. The reaction mixture was cooled to room temperature and the precipitate A filtered and washed four times with hot pyridine-H₂O (10:1). The combined filtrate was evaporated to dryness under reduced pressure. The residue was dissolved in 2 N NH₄OH solution (50 mL) and concentrated under reduced pressure. This process was once repeated and the resulting brown colored residue was dissolved in 2 N NH₄OH so-

lution (180 mL) to which was added concentrated HCl solution (150 mL) to give a precipitate B. Precipitates A and B were combined and washed three times with 2 N HCl solution (70 mL) followed by ice-cold $\mathrm{H}_2\mathrm{O}$. The solid was placed in a 2-cm diameter column and centered so that the eluate would enter a second 7-cm-diameter column containing decolorising carbon (Norit A, 250 g, packed in H₂O). The top column was eluted with 2 N NH₄OH solution until such time that a neutralized sample of the eluent obtained from the charcoal column showed no strong color with FeSO₄. To 400-mL portions of eluent (generally totalling >3 L) was added 100 mL of concentrated HCl solution to yield a white precipitate of 31. The precipitate was filtered, washed with 2 N HCl solution followed by ice-cold H₂O, and dried at ca. 75 °C under reduced pressure to yield 27.65 g (71.1%) of a white solid: mp 271-272 °C dec (lit. 26 mp varied between 255 and 260 °C in evacuated capillary tubes); IR (KBr) 1725 (carboxylic acid) cm⁻¹; NMR (20% K_2CO_3/D_2O , 80 MHz) δ 9.11 (s, ring protons).

cis - and trans-Piperazine-2,5-dicarboxylic Acid (20). To a suspension of 31 (3.0 g, 0.02 mol) in H₂O (90 mL) was added KOH (3.0 g, 0.05 mol). The mixture was warmed and to the resulting solution was added 10% Pd-C (1.0 g). This mixture was hydrogenated (Parr shaker) under 40-42 psi at 50-60 °C for 12 h. The catalyst was filtered and the filtrate concentrated to about 30 mL under reduced pressure. Dropwise addition of concentrated HCl solution (pH 5.5-6.5) to the cooled concentrate (ice bath) afforded 1.19 g (38.5%) of white crystals: mp >280 $^{\circ}$ C (trans-20); IR (KBr) 1635 (carboxylic acid) cm $^{-1}$; NMR (15% CF_3CO_2H/D_2O , 500 MHz) δ 3.64 [deceptively simple triplet (dd), 2 H, axial H of ring CH₂], 4.21 (dd, 2 H, equatorial H of ring CH₂, $J_{\rm gem}=14.1~{\rm Hz}, J_{\rm ae}=3.8~{\rm Hz}), 4.53~({\rm dd}, 2~{\rm H}, {\rm ring~CH}, J_{\rm aa}=12.4~{\rm Hz}, J_{\rm ae}=3.8~{\rm Hz}).$ Anal. $({\rm C_6H_{10}N_2O_4})~{\rm C}, {\rm H}, {\rm N}.$ Continued addition of concentrated HCl solution (pH 5.0-5.5) afforded 1.09 g (35.3%) of solid cis-trans mixture. Further acidification (pH 4.0-5.0) yielded 0.77 g (24.9%) of white crystals (cis-20): mp > 280 °C; IR (KBr) 1652 (carboxylic acid) cm⁻¹; NMR (15% $\mathrm{CF_3CO_2H/D_2O},\,500$ MHz) δ 3.93 (dd, 2 H, H of ring CH₂, J_{AB} = 14.5 Hz, J_{AX} = 4.4 Hz), 4.02 (dd, 2 H, H of ring CH₂, J_{AB} = 14.5 Hz, $J_{BX} = 7.1$ Hz), 4.71 (q, 2 H, ring CH). Anal. (C₆H₁₀N₂O₄)

trans-2,5-Dicarboxy-1,4-piperazinediacetic Acid (14). A mixture of trans-20 (1.0 g, 5.7 mmol), bromoacetic acid (1.99 g, 14.3 mmol), and K_2CO_3 (5.13 g, 37.1 mmol) in H_2O (25 mL) was stirred at room temperature for 24 h. The reaction mixture was cooled (ice bath) and acidified (concentrated HCl solution) to pH 1-2, stored at 4 °C overnight, and filtered. The solid was washed with cold H_2O followed by Me_2CO and dried to afford 1.45 g (87.6%) of a white solid (trans-14): mp >280 °C (slow decomposition >230 °C); IR (KBr) 1655 (carboxylic acid) cm⁻¹; NMR (20% K_2CO_3/D_2O , 500 MHz) δ 2.26 [deceptively simple triplet (dd), 2 H, axial H of ring CH₂], 2.76 (d, 2 H, NCH₂CO, J_{AX} = 15.4 Hz), 3.04–3.09 (m, 4 H, other ring H), 3.31 (d, 2 H, NCH₂CO, J_{AX} = 15.4 Hz). Anal. ($C_{10}H_{14}N_2O_8$:4H₂O) C, H, N.

cis-2,5-Dicarboxy-1,4-piperazinediacetic acid (14) was prepared from cis-20 according to conditions identical with those described for the preparation of trans-14, affording 1.09 g (66.1%) of a white solid: mp 200–220 °C (slow decomposition); IR (KBr) 1740 and 1720 (carboxylic acid) cm⁻¹; NMR (20% K_2CO_3/D_2O , 500 MHz) δ 2.61 (dd, 2 H, equatorial H of ring CH₂, J = 11.3 and 3.5 Hz), 2.97 (d, 2 H, NCH₂CO, J = 15.7 Hz), 3.16–3.18 (m, 2 H, ring CH), 3.23–3.25 (m, 2 H, axial H of ring CH₂), 3.28 (d, 2 H, NCH₂CO, J = 15.8 Hz). This NMR spectrum changes with lower K_2CO_3 concentration. Anal. ($C_{10}H_{14}N_2O_8$) C, H, N.

trans-1,4-Bis(2-amino-2-oxoethyl)-2,5-piperazinedicarboxylic Acid (15). A mixture of trans-20 (1.0 g, 5.7 mmol), chloroacetamide (1.28 g, 13.7 mmol), and K_2CO_3 (3.48 g, 25.2 mmol) in H_2O (25 mL) was stirred at room temperature for 35 h. The reaction mixture was cooled (ice bath), acidified (concentrated HCl solution) to pH 1–2, stored at 4 °C overnight, and filtered. The solid was washed with cold H_2O followed by Me₂CO and dried to afford 1.08 g (65.8%) of a white solid: mp 247–248 °C dec; IR (KBr) 3440 and 3310 (NH), 1705 (carboxylic acid), 1650 (amide) cm⁻¹; NMR (20% K_2CO_3/D_2O , 80 MHz) δ 2.39 [deceptively simple triplet (dd), 2 H, axial H of ring CH₂], 2.86 (d, 2 H, NCH₂CO, J_{AB} = 16.2 Hz), 3.0–3.2 (m, 4 H, other ring H), 3.36 (d, 2 H, NCH₂CO, J_{AB} = 16.2 Hz). Anal. ($C_{10}H_{16}N_4O_6$) C, H N

Dimethyl trans-2,5-Bis(methoxycarbonyl)-1,4piperazinediacetate (16). HCl gas was bubbled (ca. 5 min) into a stirred suspension of trans-14 (4.0 g, 13.8 mmol) in MeOH (200 mL) and the mixture refluxed for 5 h. The solvent was removed under reduced pressure and the residual solid dissolved in 10% aqueous Na₂CO₂ solution (50 mL) and extracted with four 50-mL portions of CHCl₃. The combined extract was washed with H₂O and dried (MgSO₄) and the solvent removed under reduced pressure to yield 3.55 g (74.4%) of a white solid (trans-16): mp (MeOH) 152-153 °C; IR (KBr) 1740 (ester) cm⁻¹; NMR (CDCl₃, 300 MHz) δ 3.07 (dd, 2 H, equatorial H of ring CH₂, $J_{\rm gem}$ = 11.3 Hz, $J_{ae} = 4.5 Hz$), 3.44 (d, 2 H, NCH₂CO, $J_{AB} = 17.1 Hz$), 3.42–3.48 (m, 2 H, ring H), 3.60–3.62 (m, 2 H, ring H), 3.62 (d, 2 H, NCH₂CO, $J_{AB} = 17.1 \text{ Hz}$), 3.69 (s, 6 H, CO_2CH_3), 3.75 (s, 6 H, CO_2CH_3). Anal. $(C_{14}H_{22}N_2O_8)$ C, H, N.

Dimethyl cis-2,5-Bis(methoxycarbonyl)-1,4-piperazinediacetate (16). HCl gas was bubbled through a stirred suspension of cis-14 (0.5 g, 1.72 mmol) in MeOH (50 mL) until all solids dissolved. The solution was refluxed for 6 h, the solvent removed under reduced pressure, and the residual solid dissolved in 10%aqueous Na₂CO₃ solution (25 mL). The aqueous solution was extracted with four 25-mL portions of CHCl3 and the combined extract was washed with H₂O, dried (MgSO₄), and concentrated under reduced pressure to afford 0.41 g (69.4%) of cis-16 as a viscous oil: IR (neat) 1745 (ester) cm⁻¹; NMR (CDCl₃, 300 MHz) δ 3.01 (dd, 2 H, equatorial H of ring CH₂, $J_{\rm AM}$ = 11.1 Hz, $J_{\rm MX}$ = 3.6 Hz), 3.30 (dd, 2 H, axial H of ring CH₂, J_{AM} = 11.1 Hz, J_{AX} = 6.6 Hz), 3.53 (s, 4 H, NCH₂CO), 3.67–3.70 (m, 2 H, other ring H), 3.70 (s, 6 H, CO_2CH_3), 3.72 (s, 6 H, CO_2CH_3). Anal. (C_{14} - $H_{22}N_2O_8\cdot 0.8H_2O)$ C, H, N.

trans-1,4-Bis(2-amino-2-oxoethyl)-2,5-piperazinedicarboxamide (18). NH₃ was condensed (5–10 mL) into a pressure bottle containing trans-16 (0.2 g, 0.58 mmol) in MeOH (3 mL) held at -78 °C (Me₂CO, dry ice). The bottle was stoppered and let stand at room temperature for 2 days. NH3 was released slowly after cooling the bottle (ice bath). The solid was filtered, washed with MeOH, and dried to afford 0.14 g (85.4%) of a white solid (trans-18): mp >260 °C (discoloration >230 °C); IR (KBr) 3420 and 3300 (NH), 1660 and 1630 (amide) $cm^{-1};\ NMR\ (15\%$ CF_3CO_2H/D_2O , 80 MHz) δ 3.1-3.4 (m, 2 H), 3.5-4.0 (m, 6 H), 4.1-4.3 (m, 2 H). Anal. ($C_{10}H_{18}N_6O_4$) C, H, N.

cis-1,4-Bis (2-amino-2-oxoethyl)-2,5-piperazinedicarboxamide (18). NH₃ was passed (5-10 mL) into a pressure bottle containing cis-16 (0.5 g, 1.44 mmol) in MeOH (2 mL) held at -78°C (Me₂CO, dry ice) and the reaction mixture worked up as in the preparation of trans-18, affording 0.38 g (92.7%) of a white solid (cis-18): mp 233-235 °C dec; IR (KBr) 3420 and 3300 (NH), 1660 (amide) cm⁻¹; NMR (15% CF₃CO₂H/D₂O, 80 MHz) δ 3.35-3.90 (c, 4 H, ring CH₂), 3.75 (s, 4 H, NCH₂CO), 4.09-4.26 (m, 2 H, ring CH). Anal. (C₁₀H₁₈N₆O₄) C, H, N.

Dimethyl trans-Piperazine-2,5-dicarboxylate (21). A suspension of trans-20 (1.0 g, 5.7 mmol) in saturated HCl-MeOH (100 mL) was refluxed with stirring for 24 h. The solvent was removed under reduced pressure and the residual solid dissolved in 20% Na₂CO₃ solution (35 mL). The solution was extracted with seven 30-mL portions of CHCl₃, and the combined extracts were washed with cold H₂O, dried (MgSO₄), and evaporated under reduced pressure to yield 0.55 g (47.7%) of a white solid. A small portion was recrystallized from Me₂CO-hexane, affording colorless needles: mp 116-118 °C; IR (KBr) 3280 and 3200 (NH), 1730 (ester) cm⁻¹; NMR (CDCl₃, 90 MHz) δ 2.06 (s, 2 H, NH), 2.80 (dd, 2 H, axial H of ring CH₂, $J_{\text{gem}} = 12.5$ Hz, $J_{\text{aa}} = 10.0$ Hz), 3.15-3.54 (m, 4 H, other ring H), 3.74 (s, 6 H, CO₂CH₃). Anal. (C₈H₁₄N₂O₄) C, H, N.

Acidification (concentrated HCl solution) of the cooled (ice bath) aqueous mother liquor afforded 0.29 g of crystalline trans-20, which was recycled.

Dimethyl cis-Piperazine-2,5-dicarboxylate (21). HCl gas was bubbled into a stirred suspension of cis-20 (1.0 g, 5.7 mmol) in MeOH (50 mL) until all solids dissolved. The solution was refluxed with stirring for $12\ h$ during which time it became turbid. The mixtures was stirred at room temperature overnight and the solids filtered. The solids were neutralized (20% Na₂CO₃ solution) and extracted (CHCl₃) to afford 56 mg (4.87%) of a white solid (trans-21) identical in all respects with the material prepared from trans-20.

The filtrate was evaporated under reduced pressure. The residual solid was dissolved in 20% Na₂CO₃ solution (35 mL) and extracted with six 30-mL portions of CHCl₃. The combined extracts were washed with cold H₂O, dried (MgSO₄), and evaporated under reduced pressure to yield a colorless oil, which crystallized in vacuo over several hours, affording 0.79 g (69.1%) of cis-21: mp 65-67 °C; IR (KBr) 3360 and 3340 (NH), 1740 and 1720 (ester) cm⁻¹; NMR (CDCl₃, 90 MHz) δ 2.16 (s, 2 H, NH), 2.97-3.38 (m, 4 H, ring CH₂), 3.51 (q, 2 H, ring CH), 3.74 (s, 6 H, CO_2CH_3). Anal. $(C_8H_{14}N_2O_4)$ C, H, N.

trans-Piperazine-2,5-dicarboxamide (22). NH₃ was passed (5-10 mL) into a pressure bottle containing trans-21 (0.82 g, 4.07 mmol) in MeOH (6 mL) held at -78 °C (Me₂CO, dry ice). The bottle was stoppered and let stand at room temperature for 2 days. NH₃ was released slowly after cooling (ice bath). The solid was filtered, washed with MeOH, and dried to afford 0.67 g (95.4%) of a white solid: mp >260 °C (slow discoloration >230 °C); IR (KBr) 3290 and 3170 (NH), 1655 (amide) cm⁻¹; NMR (15% CF_3CO_2H/D_2O , 80 MHz) δ 3.60 (dd, 2 H, axial H of ring CH_2 , $J_{\text{gem}} = 13.9 \text{ Hz}, J_{\text{aa}} = 12.5 \text{ Hz}), 4.15 \text{ (dd, 2 H, equatorial H of ring CH}_2, J_{\text{gem}} = 13.9 \text{ Hz}, J_{\text{ae}} = 3.7 \text{ Hz}), 4.53 \text{ (dd, 2 H, ring CH, } J_{\text{aa}} = 12.5 \text{ Hz}, J_{\text{ae}} = 3.7 \text{ Hz}).$ Anal. $(C_6H_{12}N_4O_2)$ C, H, N.

cis-Piperazine-2,5-dicarboxamide (22). NH₃ was passed (5-10 mL) into a pressure bottle containing cis-21 (0.75 g, 3.70 mmol) in MeOH (5 mL) held at -78 °C (Me₂CO, dry ice) and the reaction mixture worked up as in the preparation of trans-22, affording 0.52 g (81.5%) of a white solid: mp 201-203 °C dec. The filtrate was evaporated under reduced pressure and the residual solid washed with MeOH and dried to yield an additional 44 mg of white solid, providing a combined yield of 88.4%; IR (KBr) 3450 and 3300 (NH), 1695 and 1665 (amide) cm⁻¹; NMR $(D_2O, 90 \text{ MHz}) \delta 2.68-3.08 \text{ (m, 4 H, ring CH}_2), 3.29 \text{ (q, 2 H, ring CH}_2)$ CH), with 4.61 (s, HOD). Anal. $(C_6H_{12}N_4O_2)$ C, H, N.

Diethyl trans-2,5-Dicarbamoyl-1,4-piperazinediacetate (19). To a suspension of trans-22 (0.2 g, 1.16 mmol) and anhydrous K₂CO₃ (0.32 g, 2.32 mmol) in 2 mL of Me₂SO was added dropwise ethyl bromoacetate (0.28 mL, 2.55 mmol) and the mixture stirred at room temperature for ca. 24 h. The reaction mixture was diluted with cold H₂O. The white solids were filtered and washed several times with cold H2O and once with Me2CO and dried, affording 0.336 g (84.2%) of trans-19: mp >260 °C (slow discoloration >200 °C); IR (KBr) 3340 and 3200 (NH), 1730 (ester), 1665 (amide) cm⁻¹; NMR (15% CF₃CO₂H/D₂O, 500 MHz) δ 1.19 (t, 6 H, CO₂CH₂CH₃), 3.38 [deceptively simple triplet (dd), 2 H, axial H of ring CH₂], 3.67 (q, 4 H, CO₂CH₂CH₃), 3.82-3.86 (m, 4 H, equatorial H of ring CH₂ plus NCH₂CO), 3.97 (d, 2 H, NCH_2CO , $J_{AB} = 17.5 Hz$), 4.28 (dd, 2 H, ring CH, $J_{aa} = 10.6 Hz$, $J_{ae} = 2.7 \text{ Hz}$). Anal. $(C_{14}H_{24}N_4O_6) C$, H, N.

Diethyl cis-2,5-Dicarbamoyl-1,4-piperazinediacetate (19). To a suspension of cis-22 (0.5 g, 2.91 mmol) and anhydrous K₂CO₃ (0.8 g, 5.81 mmol) in Me₂SO (5 mL) was added ethyl bromoacetate (0.71 mL, 6.4 mmol) dropwise. The mixture was stirred at room temperature for ca. 24 h and filtered. The filtrate was diluted with EtOAc, filtered, and concentrated at 50-60 °C under reduced pressure. The resulting oily residue, which contained traces of inorganic material, was crystallized from MeOH-EtOAc-hexane following filtration of the initially precipitated inorganic substances to afford 0.40 g (40%) of transparent crystals: mp 130-132 °C; IR (KBr) 3410 and 3180 (NH), 1750 and 1730 (ester), 1690 and 1665 (amide) cm⁻¹; NMR (D_2O , 90 MHz) δ 1.11 (t, 6 H, $CO_2CH_2CH_3$), 2.92 (d, 4 H, ring CH_2 , J = 4.8 Hz), 3.29 (t, 2 H, ring CH, J = 4.8 Hz), 3.34 (s, 4 H, NCH₂CO), 4.06 (q, 4 H, $CO_2CH_2CH_3$), with 4.61 (s, HOD). Anal. ($C_{14}H_{24}N_4O_6$) C, H, N.

trans-Tetrahydrodipyrazino[1,2-a:1',2'-d]pyrazine-1,3,7,9(2H,4H,8H,10H)-tetrone (10). Sodium metal (31 mg, 1.35 mmol) was added to 5 mL of absolute EtOH and the mixture stirred under N₂. To the solution was added diamide diester trans-19 (0.2 g, 0.58 mmol). After the solution was refluxed under N_2 for 5 h, the solvent was removed under reduced pressure and the residual solid dissolved in cold $\rm H_2O$ (5 mL) and acidified to pH 6-7 (concentrated HCl solution). The crystallized solid was stored at 4 °C overnight, filtered, washed with cold H₂O followed by Me₂CO, and dried to afford 0.104 g (71.23%) of a white solid: mp >280 °C (slow distortion >270 °C); IR (KBr) 3200 and 3100 (NH), 1730 and 1700 (imide) cm⁻¹; NMR (Me₂SO- d_6 , 90 MHz) δ 2.18 [deceptively simple triplet (dd), 2 H, axial H of central ring

CH₂], 2.97–3.75 (m, 8 H, other ring H), 11.22 (s, br, 2 H, imide); MS (70 eV), m/e 252 (M⁺). Anal. (C₁₀H₁₂N₄O₄) C, H, N.

cis-Tetrahydrodipyrazino[1,2-a:1,2'-d]pyrazine-1,3,7,9-(2H,4H,8H,10H)-tetrone (11). Na metal (46 mg, 2 mmol) was added to 7 mL of absolute EtOH and the mixture stirred under N_2 . Diamide diester cis-19 (344 mg, 1 mmol) was added to the resulting solution and the mixture refluxed under N_2 for 6 h. The solvent was evaporated under reduced pressure, and the residual solid dissolved in cold H_2O (5 mL) and acidified (concentrated HCl solution) to pH 5 (approximate). The crystallized solid was stored at 4 °C overnight, filtered, washed with cold H_2O followed by M_2CO , and dried to afford 226 mg (89.68%) of a white solid: mp >260 °C (slow decomposition >200 °C); IR (KBr) 3250 and 3105 (NH), 1730 and 1690 (imide) cm⁻¹; NMR (Me₂SO-d₆, 90 MHz) δ 2.7-2.9 (m, br, 4 H, central ring CH₂), 3.3-3.5 (m, 6 H, other ring H), 3.55 (s, 2 H, imide); MS (70 eV), m/e 252 (M⁺). Anal. ($C_{10}H_{12}N_4O_4$) C, H, N.

trans-Tetrahydro-2,8-bis (4-morpholinylmethyl) dipyrazino[1,2-a:1',2'-d]pyrazine-1,3,7,9(2H,4H,8H,10H)-tetrone (12). To a suspension of trans-10 (315 mg, 1.25 mmol) in Me₂SO (5 mL) was added morpholine (0.38 mL, 4.37 mmol) and HCHO (0.37 mL of a 37% solution, 5.0 mmol). The mixture was stirred at 55-65 °C for 5 h and then at room temperature overnight. Me₂SO was removed by distillation under reduced pressure and the residual solid triturated with EtOH, filtered, washed (EtOH), and dried to afford 488 mg (86.83') of a white solid which underwent slow decomposition above 225 °C: IR (KBr) 1735 and 1685 (imide) cm⁻¹; NMR (CDCl₃, 90 MHz) δ 2.35 [deceptively simple triplet (dd), 2 H, axial H of central ring CH₂], 2.54-2.65 (m, 8 H, NCH₂ of morpholine), 3.05-3.91 (c, 16 H, other ring H), 4.78 (s, 4 H, NCH₂N). Anal. (C₂₀H₃₀O₆N₆) C, H, N.

cis-Tetrahydro-2,8-bis(4-morpholinylmethyl)dipyrazino[1,2-a:1',2'-d]pyrazine-1,3,7,9(2H,4H,8H,10H)-tetrone (13). To a solution of cis-11 (100 mg, 0.39 mmol) in Me₂SO

(2 mL) was added morpholine (0.12 mL, 1.39 mmol) and HCHO (0.12 mL of a 37% solution, 1.59 mmol). The solution was stirred at 55–65 °C for 5 h and at room temperature overnight. Me₂SO was removed by distillation under reduced pressure and the residual oil crystallized from Et₂O–Me₂CO, affording 153 mg (85.95%) of a white solid: mp 179–181 °C dec; IR (KBr) 1735 and 1680 (imide) cm⁻¹; NMR (CDCl₃, 90 MHz) δ 2.50–2.62 (m, 8 H, NCH₂ of morpholine), 2.83–3.89 (c, 18 H, ring H), 4.79 (s, 4 H, NCH₂N). Anal. (C₂₀H₃₀O₆N₆) C, H, N.

Acknowledgment. We gratefully acknowledge partial support of this work through U.S. Public Health Service Research Grant CA25445 from the National Cancer Institute. FT NMR 500-MHz spectra were obtained at The Ohio State University Chemical Instrumentation Center with use of a NT-500 spectrometer funded in part by NIH Grant No. 1-S10-RR01458-01A1. Spectra were produced by Dr. C. E. Cottrell.

Registry No. 10, 92927-70-3; 11, 92927-69-0; 12, 96705-80-5; 13, 96705-81-6; cis-14, 96705-82-7; trans-14, 96705-83-8; trans-15, 96705-84-9; cis-16, 96705-85-0; trans-16, 96705-86-1; cis-18, 96705-87-2; trans-18, 96705-88-3; cis-19, 96705-89-4; trans-19, 96705-90-7; cis-20, 96705-91-8; trans-20, 96705-92-9; cis-21, 96705-93-0; trans-21, 96728-88-0; cis-22, 96705-94-1; trans-22, 96705-95-2; cis-24, 15996-17-5; trans-24, 15996-16-4; cis-25, 96789-11-6; trans-25, 96789-12-7; cis-26, 96705-96-3; trans-26, 96705-98-5; cis-29, 96705-99-6; trans-27, 96843-75-3; trans-27, 96843-76-4; cis-28, 96705-98-5; cis-29, 96705-99-6; trans-29, 96706-00-2; cis-30, 96706-01-3; trans-30, 96706-02-4; 31, 122-05-4; 31-2NH₃, 96728-89-1; K_2 CO₃, 79-07-2; HCHO, 50-00-0; O-benzyl-L-serine, 4726-96-9; O-benzyl-L-serine methyl ester hydrochloride, 19525-87-2; iodo-acetamide, 144-48-9; 2,5-dimethylpyrazine, 123-32-0; bromoacetic acid, 79-08-3; ethyl bromoacetate, 105-36-2; morpholine, 110-91-8.

Antiparasitic Agents. 6.1 Synthesis and Anthelmintic Activities of Novel Isothiocyanatophenyl-1,2,4-oxadiazoles

R. D. Haugwitz,*^{†,§} A. J. Martinez,[†] J. Venslavsky,[†] R. G. Angel,[†] B. V. Maurer,[†] G. A. Jacobs,[†] V. L. Narayanan,[†] L. R. Cruthers,[‡] and J. Szanto[‡]

The Squibb Institute for Medical Research, Princeton, New Jersey 08540, and Squibb Agricultural Research Center, Three Bridges, New Jersey 08867. Received September 5, 1984

The syntheses and anthelmintic activities of 31 3- and 5-(isothiocyanatophenyl)-1,2,4-oxadiazoles are reported. In the primary anthelmintic screen, 3-(4-isothiocyanatophenyl)-1,2,4-oxadiazole (39) showed 100% nematocidal activity and 3-(2-furanyl)-5-(4-isothiocyanatophenyl)-1,2,4-oxadiazole (63), 3-(2-furanyl)-5-(2-chloro-4-isothiocyanatophenyl)-1,2,4-oxadiazole (64), and 3-(2-furanyl)-5-(4-chloro-3-isothiocyanatophenyl)-1,2,4-oxadiazole (66) showed 100% taeniacidal activity when administered orally to mice. The two most active members of this series, 39 and 63, were active against the gastrointestinal nematodes of sheep at 100 mg/kg. In addition, 39 was also found to be active against hookworms in dogs at a single, oral dose of 200 mg/kg.

During the early stages of our anthelmintic development program, 1,2,4-oxadiazole Ia emerged as a potential lead.² This finding was shortly followed by the rediscovery of the antiparasitic oxadiazoles Ib³ and Ic⁴ (Figure 1). Several other 1,2,4-oxadiazoles had been reported to possess antiparasitic activity.⁵⁻⁸ We decided to focus our investigation on isothiocyanatophenyl-substituted 1,2,4-oxadiazoles. Our earlier work on heterocyclic isothiocyanates had yielded compounds II and III equivalent to thiabendazole in anthelmintic activity^{1,9} (Figure 2). While the main thrust of this study was directed toward the evaluation of 3- and 5-(isothiocyanatophenyl)-1,2,4-oxadiazoles¹⁰ (Table

III), numerous nitro intermediates¹¹ and various 3- and 5-substituted 1,2,4-oxadiazoles were also screened for an-

- For paper 5 in this series, see: Haugwitz, R. D.; Angel, R. G.; Jacobs, G. A.; Maurer, B. V.; Narayanan, V. L.; Cruthers, L. R.; Szanto, J. J. Med. Chem. 1982, 25, 969.
- (2) U.S. Patent 3575997.
- (3) U.S. Patents 3 279 988 and 3 356 684.
- (4) Ger. Offen, 2050346.
- (5) Breuer, H. J. Med. Chem. 1969, 12, 708.
- (6) British Patent 1 228 142.
- (7) Ger. Offen. 2310287 and 2312500.
- (8) U.S. Patents 2312411 and 3897447.
- (9) Haugwitz, R. D.; Maurer, B. V.; Jacobs, G. A.; Narayanan, V. L.; Cruthers, L. R.; Szanto, J. J. Med. Chem. 1979, 22, 1113.
- (10) U.S. Patents 3853893, 3910940, 3910940, 3929814, and 4022901.
- (11) Jefford, C. W.; Cadby, P. A.; Smith, L. C.; Pipe, D. F. Pharmazie 1982, 37, 395.

[†]Squibb Institute for Medical Research.

[‡]Squibb Agricultural Research Center.

[§] Present address: National Cancer Institute, Bethesda, Maryland 20205.