Synthesis and Opioid Activity of 7-Oxygenated 2,3,4,4a,5,6,7,7a-Octahydro-1*H*-benzofuro[3,2-e]isoquinolin-9-ols

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 $3-(Cyclopropylmethyl)-9-hydroxy-7-oxo-2,3,4,4a\alpha,5,6,7,7a\alpha-octahydro-1H-benzofuro[3,2-e]iso$ quinoline (4b) containing the ACNO ring system of morphine and a 7-keto function on ring C has been synthesized and found to possess potent PQW ($ED_{50} = 0.15$ mg/kg sc) and anti-Straub tail (ED₅₀ = 0.02 mg/kg sc) activity. As compared to its 7-deoxy analog 1b, introduction of the 7-keto group did not significantly affect binding to any of the three opioid receptors $(\mu, \kappa,$ and δ), but caused a 34-fold reduction in σ -binding, suggesting reduced propensity to induce psychotomimetic effects. The C/D cis isomer of 4b (4c) was much less potent at the three opioid receptors, while displaying a slight increase in σ affinity. Both 7-hydroxy derivatives 4e and 4f were active in anti-Straub tail assay (ED₅₀ \leq 0.8 mg/kg sc), but only the α -isomer 4e demonstrated analysesic activity (PQW $ED_{50} = 0.37$ mg/kg sc) in the dose range tested. In guinea pig ileum preparations, **4e** was characterized as a selective full agonist at the κ opioid receptor (IC50 = 2.8 nM); while its β -isomer 4f was a partial agonist (78% at 1 μ M), with antagonist activity observed at both μ - and κ -opioid receptors.

Morphine is a potent analgesic alkaloid, its rigid structure consisting of five rings (ABCNO). Approaches based on simplification of the morphine skeleton for the discovery of novel analgesics have been adopted by generations of medicinal chemists.1 Like many substructural analogs of morphine, N-substituted $2,3,4,4a\alpha,5,6,7,7a\alpha$ -octahydro-1H-benzofuro[3,2-e]isoquinolin-9-ols (1), which contain the ACNO ring system of morphine, have been found to retain potent analgesic activity.² Among them, the N-cyclopropylmethyl analog (1b or J6549) is most interesting in that it possesses potent oral analgesic and narcotic-antagonism activity and is likely to have a low potential for addiction.^{2b} However, of concern is the fact that 1b showed significant binding to the σ receptor $(K_i, \sigma = 21 \text{ nM})^3$ which indicates potential psychotomimetic effects. In morphine, the related morphinan (2), and trans-4a-phenyldecahydroisoquinoline (3) series, C ring functionality such as 6-oxygenation, has been found to have beneficial effects on both analgesic potency and opioid receptor selectivity.4-6 As an effort to study the effect of placing an oxygen function in the corresponding position of the octahydrobenzofuroisoquinoline series represented by 1a and 1b, we report here the synthesis and opioid activity of a series of 7-oxygenated analogs of 1, namely 4a-g.

Chemistry

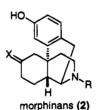
The first reported synthesis of 1a was achieved via the intramolecular Diels-Alder reaction of N-[2-(7methoxy-3-benzofuranyl)ethyl]-N-methyl-6α-pyronecarboxamide.² An alternative synthesis based on the

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Chart 1



1b: R = cyclopropylmethyl

1a: R = CH3

trans-4a-phenyldecahydroisoquinoilnes (3)

regioselective acylation of the anion derived from 4-(2,3dimethoxyphenyl)-1,2,3,6-tetrahydro-1-methylpyridine with y-butyrolactone was reported later. Neither of the above two methods can be easily adopted for the preparation of C-ring functionalized analogs of 1. Among the reported syntheses of the ACNO system of morphine with functionalized C-ring,8-10 the one reported by Weller et al. 10 was adopted for the preparation of our 7-keto analogs of 1 (Scheme 2). The key intermediate 4-[2-(phenylmethoxy)-3-methoxyphenyl]pyridine-3-carboxaldehyde (8) was synthesized by a new route (Scheme 1).11 The starting 7-methoxy[1]benzopyrano[3,4-c]pyridin-5(2H)-one (5)¹² was treated with sodium methoxide followed by benzyl bromide to give 4-arylnicotinate 6. Dibal reduction of 6 followed by oxidation of the resulting alcohol 7 with active manganese dioxide provided aldehyde 8, which was converted by literature procedures¹⁰ to give ester **9** in 58% overall yield from **5**. The conversion of 9 to the desired 7-keto analogs of 1, namely 4a-c, essentially followed the published se-

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Scheme 1a

^a Reagents, conditions, and yields: (a) NaOMe, $C_6H_5CH_2Br$, DMF; 92%; (b) DIBAL, toluene; 82%; (c) MnO₂, CH_2Cl_2 ; 92%; (d) monoethyl malonic acid ester, pyridine, piperidine(cat.), Δ ; 91%; (e) 5% Pd/C, EtOH, H_2 ; 92%.

Scheme 2a

^a Reagents, conditions, and yields: (a) BrCH₂COOEt, K₂CO₃, DMF; 93%; (b) NaOEt, EtOH; 90%; (c) NaH, 4-Å molecular sieve, THF, reflux; 72%; (d) PtO₂, EtOH, H₂; 53% (trans), 10% (cis); (e) 3 N HCl, reflux; 89%; (f) BBr₃-S(CH₃)₂, ClCH₂CH₂Cl, reflux.

quence for the N-methyl analog¹⁰ (Scheme 2). However, we found that the conversion of **11** to **12** was more efficiently carried out in ethanol. When dimethylformamide was used as the solvent, the reaction was very sluggish, probably due to the formation of a stable complex between the sodium enolate of **11** and its 3-methoxy group. In our hands, the catalytic hydrogenation of **13** over platinum afforded an **81%** yield of **14** and **15** in a ratio of **83:17**. The major trans isomer **14** was treated with HCl to give **4a**, which underwent

O-demethylation effected by BBr₃/S(CH₃)₂ complex to give **4b**. The minor isomer **15** underwent similar transformations to give **4c**, the cis isomer of **4b**. The N-methyl analog **4g** was prepared from 1-methyl-4-(2-hydroxy-3-methoxyphenyl)-3-(3-oxo-3-ethoxypropyl)-pyridinium iodide similarly as **4b** is prepared from **10**. Our assignment of the C/D ring junction stereochemistry (cis or trans) was supported by comparison of the ¹H-NMR data for **4a** and **16** with that reported for the N-methyl analogs of these two compounds. ^{9,10b} An

Figure 1. C/D cis ring junction in 15 as revealed by NOE experiment.

Scheme 3^a

^a Reagents, conditions, and yields: (a) L-Selectride, THF; 80%; (b) C_6H_5COOH , DEAD, $P(C_6H_5)_3$, THF; (c) NaSPr, DMF; 75%; (d) 1% NaOH, MeOH; 54% from 4d; (e) BBr₃-S(CH₃)₂, ClCH₂CH₂Cl, reflux; 83%.

added proof is provided by NOE experiment on compound 15, which showed strong dipolar coupling between the angular H-4a and H-12, thus establishing the spatial proximity or the cis relationship between H-4a and the phenyl ring in 15 (Figure 1). The 7-hydroxy analogs 4d-f were derived from 4a via stereospecific transformations as shown in Scheme 3. Thus, 4a was reduced with L-Selectride to give exclusively the α-hydroxy isomer 4d in 80% yield, which was O-demethylated to give 4e. Treatment of 4d with benzoic acid under Mitsunobu's condition¹³ resulted in inversion of the 7-hydroxyl function to provide the β -benzoate 17, which was hydrolysed and O-demethylated to give the β -hydroxy analog **4f**.

Pharmacology

Listed in Table 1 are the binding affinities (K_i, nM) of the title compounds $(4\mathbf{a}-\mathbf{g})$ for opioid receptors (μ, κ, κ) and δ) and the σ -receptor, and the in vivo data of these compounds for analgesic activity (PQW) and narcotic antagonism (AST). The data for compound 1b, morphine, and naloxone are included for comparison. As shown, the introduction of a 7-keto group (4b) did not significantly affect binding to any of the three opioid receptors but lowered the σ -binding by 34-fold over the corresponding C-ring unfunctionalized analog (1b). Compound 4b also retained good activity in PQW and anti-Straub tail assays, indicating its being a potent

 μ -antagonist and κ -agonist similar to **1b**. Compound **4c**, the cis isomer of 4b, is a much weaker ligand at the opioid receptors, while displaying a slight increase $(2\times)$ in σ -affinity. This is in agreement with the general observation that the degree of stereoselectivity at the σ -receptor is less than that at opioid receptors. ¹⁴ The significant anti-Straub tail activity demonstrated by 4c indicates its being a narcotic antagonist. Replacement of the N-cyclopropylmethyl group in **4b** with a methyl group (4g) resulted in much reduced binding at all four receptors assayed, with the largest drop observed at the κ -receptor. However, the analgesic activity was only decreased 3-fold with a concomitant 63-fold decrease in anti-Straub tail activity, indicating that the N-methyl group confers μ agonist activity instead of antagonist activity. Both 4e and 4f, the 7-hydroxy analogs derived from 4b, displayed good but reduced binding at the opioid receptors. However, some interesting stereoselectivity associated with their in vivo activity was observed. The α -hydroxy isomer **4e** was active in both PQW and anti-Straub tail assays, while its β -isomer 4f was only active in the anti-Straub tail assay and devoid of PQW activity in the dose range tested. In order to further delineate their pharmacological profile, compounds **4e** and **4f** were also evaluated at μ - and κ - opioid receptors in guinea pig ileum preparations. Compound 4e was found to produce a full agonist effect of inhibiting the electrically stimulated muscle contraction (IC₅₀ = 2.8 nM), while 4f behaved as a partial agonist with a maximal response of 78% at $1 \mu M$ (Table 2). The agonist activities of 4e and 4f in GPI preparations can be antagonized by naloxone, and the IC₅₀ values were shifted similarly as that of U-50488; the agonist effect of morphine was more sensitive to antagonism by naloxone (Table 3). The data indicates that the opioid agonist effects of 4e and 4f are mainly due to binding at the κ -receptor, in agreement with their in vivo PQW and AST activities. Opioid antagonist activity of the β -isomer 4f at μ - and κ -opioid receptors in guinea pig ileum was also examined. At 3 nM, 4f was found to be an effective antagonist at both μ - and κ -opioid receptors, with higher potency for the μ -receptor (Table 4). Methylation of the 3-hydroxyl group (4a and 4d) caused large reductions in opioid receptor affinities, with little effect on σ -binding. Compounds 4a and 4d also showed reduced potency in in vivo PQW and anti-Straub tail assays.

Conclusion

The opioid and sigma receptor binding affinities of compounds containing the ACNO partial structure of morphine as represented by 1 are sensitive to functionality in ring C. In particular, selective reduction in σ binding was realized by the introduction of a 7-keto group. Since the σ -receptor may mediate psychotomimetic effects, 15 compound 4b, being a weaker ligand at the σ -receptor while maintaining good analgesic and narcotic antagonism activity, is likely to be superior to 1b as a potential analgesic. Unlike binding to opioid receptors (μ , κ , and δ), binding affinity at the σ -receptor is not significantly affected by methylation of the phenolic hydroxyl group, modification of the 7-oxygen function, or stereochemistry at the C/D ring junction. As compared to the 7-keto compound 4b, the 7α -hydroxy derivative **4e** remains a μ -antagonist and κ -agonist, although with reduced potency; while its β -isomer **4f**

Table 1. Opioid Receptor Binding and in Vivo Analgesia and Narcotic Antagonism Activity of 7-Oxygenated 2,3,4,4a,5,6,7,7a-Octahydro-1*H*-benzofuro[3,2-e]isoquinolin-9-ols^a

				opioid receptor binding (K_i, nM)			σ -receptor	$ED_{50} \left(mg/kg \ sc \right)$	
no.	R_1	\mathbb{R}_2	X	μ	κ	δ	binding (K_i, nM)	PQW	AST
1b	Н	CH ₂ -c-C ₃ H ₅	H_2	0.57 $(0.4-0.7)^b$	1.2 (1-1.5)	5.3 (1.3-20)	21 (18-26)	0.025 (0.019-0.034)	0.018 (0.014-0.023)
4a	CH ₃	CH ₂ -c-C ₃ H ₅	0	76 (57–102)	1760 (406-26320)	1760 (169-13700)	440 (388-496)	0.93 $(0.38-2.3)$	0.34 $(0.17-0.68)$
4b	H	CH ₂ -c-C ₃ H ₅	0	0.25 (0.17-0.34)	1.73 (1.22-2.36)	2.15 $(1.04-3.97)$	710 (536-967)	0.15 $(0.068-0.34)$	0.022 (0.014-0.036)
4c		C/D cis isome	er of 4b	19 (17-22)	62 (51-77)	134 134 (77-280)	310 (113-613)	>5.4 (inactive)	1.0 (0.83-1.3)
4d	CH ₃	$\mathrm{CH}_2\text{-}\mathrm{c}\text{-}\mathrm{C}_3\mathrm{H}_5$	·····ОН & Н	29 (20-40)	140 (99–197)	150 (89-246)	410 (335-504)	3.0 (0.99-8.8)	1.4 (0.60-3.5)
4e	H	CH ₂ -c-C ₃ H ₅	·····ОН & Н	1.4 (0.8-2.1)	4.9 (2.7-8.0)	15 (12.1–17.8)	580 (236-1280)	0.37 $(0.21-0.64)$	0.55 $(0.23-1.3)$
4f	Н	CH ₂ -c-C ₃ H ₅	·····Н & — ОН	3.9 (2.3-5.9)	12 (11-13)	9.5 (7.4-11.9)	300 171-508)	>4.0 (inactive)	0.80 $(0.45-1.4)$
4g	Н	CH ₃	0	7.8 (4.5–13.4)	186 (103-345)	84 (12.8-159)	2090 (695-4945)	0.40 $(0.28-0.57)$	1.4 $(0.74-2.7)$
morphine				38 (25-61)	1870 (149-7080)	375 (28-3480)	>105	0.98 $(0.77-1.3)$,,
naloxone				1.1	12	19	>10 ⁵	>100 (inactive)	0.020 (0.015-0.027)

^a Data represents the mean of three experiments each performed in duplicate. ^b The 95% confidence limits are shown in parentheses.

Table 2. Opioid Agonist Activity of 4e and 4f in GPI Preparation

compd	IC ₅₀ (nM)	% max response
4e	2.8 ± 0.7 (4)	_
4f	_	$78 \pm 3 (3)$
morphine	$79 \pm 6 (4)$	_
U-50488	2.2 ± 0.2 (4)	

^a Partial agonist potency (\pm SE) at 1 μ M.

Table 3. Effect of Naloxone on the Opioid Agonist Activity of **4e** and **4f** in GPI Preparation^a

compd	IC ₅₀ ratio ^b	$K_{\rm e}({ m nM})^c$
4e	8.1 ± 1.4 (3)	14.1
4f	$8.5 (2)^d$	13.3
U-50488	$9.1 \pm 1.3(3)$	12.3
morphine	$38 \pm 2.5 (4)$	2.7

 $[^]a$ Concentration of naloxone, 100 nM. b IC₅₀ of the agonist in the presence of antagonist divided by the control IC₅₀ in the same preparation. c K_e = [antagonist]/(IC₅₀ ratio - 1). d This ratio is an estimate because the maximum agonist response of **4f** was 78%.

Table 4. Evaluation of Opioid Antagonist Activity of 4f in GPI Preparation^a

compd	IC ₅₀ ratio ^b	$K_{\rm e} ({ m nM})^{\rm c}$
μ , morphine κ , U-50488	6.3 ± 0.7 (4) 2.3 ± 0.7 (3)	0.57 2.3

^a Concentration of 4f, 3 nM. ^b See Table 3.

was found to be only a partial agonist at the κ -receptor and an effective antagonist at both μ - and κ -receptors.

Experimental Section

Synthesis. Melting points were taken in a capillary tube by using a Yamato MP-21 melting point apparatus and are uncorrected. IR spectra were determined with a Perkin-Elmer 1760-X FT-IR spectrometer. NMR spectra were recorded on a Bruker AM-300 spectrometer; chemical shifts were recorded in parts per million downfield from Me $_4$ Si. Mass spectra were recorded on a JEOL JMS-D300 mass spectrometer; HRMS was obtained with a JEOL JMS-HX110 spectrometer. Elemental analyses were performed with a Perkin-Elmer 240C instrument. TLC was performed on Merck (Art. 5717) silica gel plates and visualized with UV light (254 nm) or upon heating after treatment with 2% phosphomolybdic acid in ethanol. Flash column chromatography was performed with Merck 40–63- μ m silica gel. Reagent grade THF was distilled from sodium benzophenone prior to use. Other anhydrous solvents were distilled from CaH $_2$ and stored over 4-Å molecular sieves until use.

3-(Methoxycarbonyl)-4-[2-(phenylmethoxy)-3-methoxyphenyl]pyridine (6). Lactone 5 (14.0 g, 61.6 mmol) was dissolved in DMF (180 mL) and cooled to -65 °C (CO₂/CHCl₃). Sodium methoxide (4.66 g, 86.3 mmol) in DMF (40 mL) was added dropwise, the mixture was stirred for 1 h, and benzyl bromide (10.2 mL, 85.9 mmol) in DMF (30 mL) was added over 30 min. The mixture was warmed to room temperature over 3 h and stirred for 2 h. The solvent was removed by vaccum distillation. To the residue were added saturated NH₄Cl(aq) (100 mL) and water (400 mL), and the resulting mixture was extracted with CH_2Cl_2 (250 mL \times 3). The combined organic layer was dried (MgSO₄) and evaporated. The residue was chromatographed (silica gel; ether:n-hexane = 1:1) to afford 6as a white solid (19.8 g, 92%): mp 76–76.5 °C (EtOAc/n-hexane); R_f 0.53 (ether); ¹H NMR (300 MHz, CDCl₃) δ 3.66 (s, 3H), 3.90 (s, 3H), 4.73 (s, 2H), 6.75-7.19 (m, 9H), 8.56 (d, J = 1)5.1 Hz, 1H), 8.99 (s, 1H); 13 C NMR (75 MHz, CDCl₃) δ 51.0, 55.2, 73.9, 112.7, 120.5, 123.4, 124.8, 126.6, 126.9, 127.2, 127.4, 132.9, 136.2, 143.9, 145.9, 149.4, 150.9, 152.0, 165.6; IR (neat) 2955, 1730, 1582, 1265 cm⁻¹; MS (EI, 70 eV) m/e calcd for $C_{21}H_{19}NO_4^+$ 349.1314, found 349.1304; 349 (M⁺), 91 (base). Anal. $(C_{21}H_{19}NO_4)$ C, H, N.

[4-[2-(Phenylmethoxy)-3-methoxyphenyl]pyridin-3-yl]-methanol (7). To a solution of ester 6 (14.0 g, 40.1 mmol) in dry toluene (300 mL) at -78 °C was added DIBALH (100 mL, 1.0 M in toluene) over 30 min. The mixture was stirred and

warmed to 0 °C during 3 h. HOAc(aq) (500 mL, 1 M) was added to the reaction mixture, and the toluene layer was separated. The aqueous layer was basified to a pH of 10 with 20% NaOH(aq) and then extracted with CH₂Cl₂/2-propanol (6/ 1, 200 mL \times 3). The organic layers were combined, washed with saturated NaHCO₃(aq), dried (MgSO₄), and evaporated. The residue was chromatographed (silica gel; ether) to afford 7 as a white solid (10.54 g, 82%): mp 94-94.5 °C (from EtOAc/ n-hexane); R_f 0.23 (ether); ¹H NMR (300 MHz, CDCl₃) δ 2.85 (s, 1H, OH), 3.92 (s, 3H), 4.37 (s, 2H), 4.72 (s, 2H), 6.71-7.21 (m, 9H), 8.42 (d, $J=5.0~{\rm Hz},~1{\rm H}),~8.67$ (s, 1H); $^{13}{\rm C}$ NMR (20 MHz, CDCl₃) δ 55.7, 60.5, 75.0, 112.8, 121.7, 124.3, 124.4, 127.8, 128.0, 128.2, 132.8, 134.9, 136.2, 144.2, 145.2, 147.6, 149.5, 152.7; IR (neat) 3238, 1467, 1218 cm⁻¹; MS (EI, 70 eV) m/e calcd for $C_{20}H_{19}NO_3^+$ 321.1365, found 321.1367; 321 (M⁺), 304, 212, 91 (base). Anal. (C₂₀H₁₉NO₃) C, H, N.

4-[2-(Phenylmethoxy)-3-methoxyphenyl]pyridine-3carboxaldehyde (8). To a solution of alcohol 7 (11.9 g, 37.0 mmol) in CH₂Cl₂ (150 mL) was added MnO₂ (20.9 g, 240 mmol). The mixture was brought to reflux for 14 h and then cooled to room temperature. The suspension was filtered, and the filtrate was evaporated. The residue was chromatographed (silica gel, ether/n-hexane = 3:1) to afford 8 as pale yellow crystals (10.9 g, 92%): mp 95.5-96 °C; R_f 0.53 (ether); ¹H NMR (300 MHz, CDCl₃) δ 3.95 (s, 3H), 4.79 (s, 2H), 6.75 $7.19 \text{ (m, 9H)}, 8.61 \text{ (d, } J = 5.0 \text{ Hz, 1H)}, 9.00 \text{ (s, 1H)}, 9.70 \text{ (s, } 3.00 \text{ (s,$ 1H); ¹³C NMR (20 MHz, CDCl₃) δ 56.1, 75.0, 113.9, 122.4, 125.5, 128.1, 128.2, 128.5, 128.8, 130.2, 136.2, 144.5, 148.5, 148.7, 152.9, 153.1, 191.1; IR (neat) 1698, 1589, 1476 cm⁻¹ MS (EI, 70 eV) m/e calcd for $C_{20}H_{17}NO_3^+$ 319.1209, found 319.1202; 319 (M+), 290, 91 (base). Anal. (C₂₀H₁₇NO₃) C, H,

1-(Cyclopropylmethyl)-4-(2-hydroxy-3-methoxyphenyl)-3-(3-oxo-3-ethoxypropyl)pyridinium Bromide (10). Cyclopropylmethyl bromide (4.42 mL, 45.6 mmol) was added to a solution of 9 (6.06 g, 20.1 mmol) in DMF (15 mL), and the resulting mixture was stirred for 9 h at 78 °C. DMF and excess cyclopropylmethyl bromide were removed by Kugelrohr distillation (1 mmHg, 60 °C). The crude product was purified by recrystallization from butanone to afford 10 (6.75 g. 77%) as a white solid: mp 117-120 °C; ¹H NMR (300 MHz, CDCl₃) $\delta 0.66-0.72$ (m, 4H), 1.07 (t, J = 7.0 Hz, 3H), 1.42 (m, 1H), 2.50 (t, J = 7.2 Hz, 2H), 3.02 (t, J = 7.2 Hz, 2H), 3.85 (s, 3H),3.93 (q, J = 7.1 Hz, 2H), 4.61 (d, J = 7.5 Hz, 2H), 6.62-6.65(m, 1H), 6.85-6.92 (m, 2H), 7.73 (d, J = 6.1 Hz, 1H), 9.04-9.12 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 4.5, 11.9, 13.8, 25.8, 32.8, 56.0, 60.7, 65.2, 112.5, 120.3, 120.6, 121.6, 129.5, 140.8, 141.3, 142.7, 143.7, 147.5, 156.0, 172.1; IR (KBr) 3327, 2842, 1729, 1636 cm $^{-1}$. Anal. (C₂₁H₂₆NO₄Br H₂O) C, H, N.

1-(Cyclopropylmethyl)-4-[2-(2-ethoxy-2-oxoethoxy)-3methoxyphenyl]-3-(3-oxo-3-ethoxypropyl)pyridinium Bromide (11). A mixture of 10 (5.41 g, 12.4 mmol), ethyl bromoacetate (2.48 g, 14.8 mmol), and finely powdered K₂CO₃ (2.57 g, 18.6 mmol) in DMF (15 mL) was stirred for 1 h at 25 °C. The solvent was removed by kugelrohr distillation. The residue was treated with CH2Cl2 and filtered, and the filtrate was evaporated to give 11 as an amber-colored oil (6.03 g, 93%): ¹H NMR (300 MHz, CDCl₃) δ 0.72-0.81 (m, 4H), 1.11-1.27 (m, 6H), 1.52 (m, 1H), 2.63 (m, 2H), 3.05 (t, J = 7.1 Hz,2H), 3.86 (s, 3H), 3.96-4.11 (m, 4H), 4.51-4.54 (m, 2H), 4.87 (d, J = 7.6 Hz, 2H), 6.73-6.76 (m, 1H), 7.02-7.18 (m, 2H), 7.81 (d, J = 6.2 Hz, 1H), 9.27 (s, 1H), 9.46 (d, J = 6.5 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 4.8, 12.3, 14.1, 25.9, 32.9, 55.9, 60.7, 60.9, 64.9, 69.0, 114.4, 120.9, 124.8, 129.0, 129.5, 141.0, 141.6, 143.3, 143.7, 151.6, 155.9, 168.6, 172.0; IR (neat) 2981, 2938, 1756, 1733, 1640 cm⁻¹.

Spiro[2-(ethoxycarbonyl)-7-methoxybenzofuran-3(2H), 4'(1'H)-1'-(cyclopropylmethyl)-3'-(3-oxo-3-ethoxypropyl)pyridine] (12). To a solution of 11 (2.76 g, 5.28 mmol) in anhydrous EtOH (20 mL) was added 20% NaOEt/ EtOH (6.6 mL, 17.0 mmol). The mixture was stirred at 19 °C for 30 min and quenched with saturated NH₄Cl(aq). After evaporation of EtOH, H2O (80 mL) was added, and the mixture was extracted with ether (100 mL \times 3). The combined extracts were washed with brine, dried (MgSO₄), and evaporated to give **12** as a yellow solid (2.1 g, 90%): R_f 0.67 (ether); ¹H NMR (300

MHz, CDCl₃) δ 0.12-0.17 (m, 2H), 0.46-0.52 (m, 2H), 0.81-0.93 (m, 1H), 1.11-1.25 (m, 6H), 1.86-2.28 (m, 4H), 2.95 (d, J = 6.5 Hz, 2H, 3.83 (s, 3H), 3.94 - 4.36 (m, 5H), 4.98 (s, 1H),5.90-5.96 (m, 2H), 6.69-6.92 (m, 3H); 13 C NMR (75 MHz, CDCl₃) δ 3.0, 3.1, 3.2, 10.9, 14.0, 14.1, 14.2, 24.8, 25.5, 33.7, 34.2, 55.3, 55.5, 55.9, 57.8, 58.0, 59.9, 60.1, 60.6, 60.8, 91.1, 95.5, 97.5, 99.3, 107.9, 111.6, 117.8, 122.0, 127.3, 129.2, 130.7, 134.8, 144.1, 145.8, 169.1, 172.9; IR (neat) 2981, 2937, 1752, 1734, 1681, 1611 cm⁻¹; MS (EI, 70 eV) m/e calcd for $C_{25}H_{31}$ - NO_6^+ 441.2152, found 441.2156; 441 (M⁺), 368 (base), 340, 55.

Ethyl 3-(Cyclopropylmethyl)-7-hydroxy-9-methoxy-5,7a-dihydro-3H-benzofuro[3,2-e]isoquinoline-6-carboxylate (13). To a mixture of NaH (0.816 g, 34 mmol) and 4-Å molecular sieve (3.64 g) in THF (50 mL) was added a solution of 12 (3.33 g, 7.54 mmol) in THF (60 mL). The resulting mixture was brought to reflux for 2.5 h and then poured into saturated NH₄Cl(aq) (200 mL). The mixture was evaporated and redissolved in a mixture of water (50 mL) and ether (200 mL). The ether layer was separated, and the aqueous layer was extracted further with ether (200 mL \times 2). The combined organic layers were washed with brine, dried (MgSO₄), and evaporated to give 14 as a yellow crystalline solid (2.13 g, 72%): $R_f 0.48$ (ether:hexane, 1:1); ¹H NMR (300 MHz, $CDCl_3$) δ 0.15-0.20 (m, 2H), 0.51-0.57 (m, 2H), 0.95 (m, 1H), 1.24 (t, J = 7.1 Hz, 3H), 2.73 (s, 2H), 2.88-3.01 (m, 2H), 3.83 $(\mathbf{s}, 3\mathbf{H}), 4.15 \ (\mathbf{q}, J = 7.1 \ \mathbf{Hz}, 2\mathbf{H}), 4.42 \ (\mathbf{d}, J = 7.7 \ \mathbf{Hz}, 1\mathbf{H}), 4.76$ (s, 1H), 5.85-5.91 (m, 1H), 6.11 (s, 1H), 6.68-6.88 (m, 3H), 11.86 (s, 1H); 13 C NMR (75 MHz, CDCl₃) δ 3.1, 3.2, 10.9, 14.1, 26.2, 50.0, 55.9, 57.8, 60.7, 88.9, 101.7, 102.4, 103.2, 111.6, 117.1, 122.0, 126.9, 127.6, 136.3, 144.6, 144.9, 165.0, 171.4; MS (EI, 70 eV) m/e calcd for $C_{23}H_{25}NO_5^+$ 395.1733, found 395.1736; 395 (M⁺), 267, 212, 55 (base).

Ethyl 3-(Cyclopropylmethyl)-7-hydroxy-9-methoxy- $2,3,4,4a\alpha,5,7a\alpha$ -hexahydro-1H-benzofuro[3,2-e]isoquinoline-6-carboxylate (14) and Ethyl 3-(Cyclopropylmethyl)-7-hydroxy-9-methoxy-2,3,4,4a α ,5,7a β -hexahydro-1Hbenzofuro[3,2-e]isoquinoline-6-carboxylate (15). A mixture of 13 (520 mg, 1.31 mmol) and PtO₂ (100 mg) in EtOH (20 mL) was shaken in a Parr hydrogenator under 45 psi of H₂ for 11 h. The catalyst was removed via filtration through Celite, and the filtrate was evaporated. The residue was chromatographed (silica gel; MeOH:CH2Cl2, 1:30) to afford 350 mg (67%) of C/D trans isomer 14 and 72 mg (14%) of C/D cis isomer 15. 14: pale-yellow crystals; mp 119-120 °C; R_f 0.47 (MeOH:CH₂Cl₂, 8:92); ¹H NMR (300 MHz, CDCl₃) δ 0.17-0.3 (m, 2H), 0.54-0.56 (m, 2H), 0.93 (m, 1H), 1.22 (t, J = 7.2 Hz,3H), 1.88-2.01 (m, 3H), 2.18-2.29 (m, 2H), 2.38-2.46 (m, 3H), 2.62 (d, J = 11.6 Hz, 1H), 2.95 (d, J = 12 Hz, 1H), 3.08-3.13(m, 1H), 3.82 (s, 3H), 4.06-4.18 (m, 2H), 4.78 (s, 1H), 6.75-6.81 (m, 2H), 7.03-7.05 (m, 1H), 11.90 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 3.99, 4.04, 8.1, 14.1, 24.5, 37.1, 37.4, 48.3, 54.5, 56.0, 60.8, 63.6, 87.5, 101.4, 112.3, 118.7, 120.7, 131.2, 145.8, 148.1, 165.7, 171.6; IR (neat) 3331, 2937, 1661, 1651, 1622, 1616 cm^{-1} ; MS (EI, 70 eV) m/e calcd for $C_{23}H_{29}NO_5^+$ 399.2046, found 399.2043; 399 (M⁺, base), 55. **15**: oil; R_f 0.59 (MeOH: CH_2Cl_2 , 8:92); ¹H NMR (300 MHz, CDCl₃) δ 0.06-0.11 (m, 2H), 0.47-0.53 (m, 2H), 0.84 (m, 1H), 1.18-1.27 (m, 3H), 1.74- $1.87 \text{ (m, 2H)}, 1.96-2.33 \text{ (m, 6H)}, 2.52 \text{ (m, 1H)}, 2.94 \text{ (dd, } J = 1.87 \text{ (m, 2H)}, 1.96-2.33 \text{ (m, 6H)}, 2.52 \text{ (m, 1H)}, 2.94 \text{ (dd, } J = 1.87 \text{ (m, 2H)}, 1.96-2.33 \text{ (m, 6H)}, 2.52 \text{ (m, 1H)}, 2.94 \text{ (dd, } J = 1.87 \text{ (m, 2H)}, 2.94 \text{ (dd, } J = 1.87 \text{ (dd, 3H)}, 2.94 \text{ (dd, 3H$ 11.7, 3.9 Hz, 1H), 3.06 (d, J = 12.0 Hz, 1H), 3.81 (s, 3H), 4.08-4.18 (m, 2H), 4.96 (s, 1H), 6.69-6.85 (m, 3H), 11.88 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 3.8, 4.0, 8.3, 14.0, 24.3, 35.6, 36.7, 48.2, 50.4, 55.7, 55.9, 60.8, 63.9, 82.1, 98.9, 112.0, 114.5, 121.8, 134.2, 145.2, 146.7, 163.8, 171.9; MS (EI, 70 eV) m/e calcd for $C_{23}H_{29}NO_5^+$ 399.2046, found 399.2052; 399 (M⁺), 353, 55 (base).

3-(Cyclopropylmethyl)-9-methoxy-7-oxo- $2,3,4,4a\alpha,5,6,7,7a\alpha$ -octahydro-1H-benzofuro[3,2-e]isoquinoline (4a). A solution of 14 (526 mg, 1.32 mmol) in CH₃CN (1.5 mL) and 3 N HCl(aq) (30 mL) was refluxed for 7 h. When cooled, the reaction mixture was basified with 20% NaOH-(aq) and extracted with CH_2Cl_2 (50 mL \times 3). The combined extracts were washed with brine, dried (MgSO₄), and evaporated to give 4a (384 mg, 89%) as an amber oil. An analytical sample was obtained via chromatography (silica gel, MeOH: CH_2Cl_2 , 7:93): mp 176.5-179 °C (HCl salt); R_f 0.37 (MeOH: CH_2Cl_2 , 10:90); ¹H NMR (300 MHz, $CDCl_3$) δ 0.11-0.16 (m, 2H), 0.51-0.57 (m, 2H), 0.92 (m, 1H), 1.55-1.61 (m, 1H), 1.73-

- 3-(Cyclopropylmethyl)-9-methoxy-7-oxo-2,3,4,4ac,5,6,7,7a β -octahydro-1H-benzofuro[3,2-e]isoquinoline (16). 15 was subjected to the reaction condition described above to give the C/D cis ketone 16: R_f 0.56 (10:90 MeOH:CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃) δ 0.06-0.11 (m, 2H), 0.46-0.52 (m, 2H), 0.85 (m, 1H), 1.77-1.92 (m, 4H), 2.13-2.46 (m, 7H), 2.97 (m, 2H), 3.84 (s, 3H), 4.58 (s, 1H), 6.73 (m, 2H), 6.85 (dd, J=8.4, 7.2 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 3.8, 4.0, 8.2, 24.5, 34.5, 36.5, 36.8, 50.6, 52.5, 55.6, 56.0, 63.8, 88.8, 112.3, 114.6, 122.4, 134.5, 144.8, 147.2, 207.2; MS (EI, 70 eV) m/e calcd for $C_{20}H_{25}NO_3^+$ 327.1835, found 327.1835; 327 (M⁺), 55 (base).
- 3-(Cyclopropylmethyl)-9-hydroxy-7-oxo-2,3,4,4aa,5,6,7,7aa-octahydro-1H-benzofuro[3,2-e]isoquinoline (4b). A mixture of 4a (10 mg, 0.031 mmol), 1-propanethiol (23 mg, 0.31 mmol), and NaH (6 mg, 0.25 mmol) in DMF (4 mL) was heated at 110 °C for 2 h. NH₄Cl (50 mg) was added to the mixture, and DMF was removed by Kugelrohr distillation. The residue was chromatographed (silica gel; MeOH: CH₂Cl₂, 15:85) to give **4b** as an oil (5 mg, 52%): ¹H NMR (300 MHz, $CDCl_3$) δ 0.15 (m, 2H), 0.52-0.58 (m, 2H), 0.94 (m, 1H), 1.23-1.28 (m, 1H), 1.74-7.83 (m, 1H), 1.87 (dt, J = 12.9, 2.5Hz, 1H), 2.07 (td, J = 12.8, .3.9 Hz, 1H), 2.32-2.54 (m, 6H), 2.67 (t, J = 11.7 Hz, 1H), 2.98 (d, J = 12.3 Hz, 1H), 3.12 (dd, J = 12.3 Hz, 1H), 3.J = 11.4, 3.8 Hz, 1H), 4.41 (s, 1H), 6.72-6.81 (m, 2H), 6.92 (dd, J = 6.9, 1.6 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 4.0, 4.1, 8.1, 25.7, 38.2, 39.9, 40.1, 48.2, 52.7, 54.5, 63.5, 91.8, 116.7, 118.0, 121.8, 130.2, 141.9, 147.6, 208.5; MS (EI, 70 eV) m/e calcd for $C_{19}H_{23}NO_3^+$ 313.1678, found 313.1670; 313 (M⁺, base), 272.

3-Methyl-9-hydroxy-7-oxo-2,3,4,4aα,5,6,7,7aα-octahydro-1*H*-benzofuro[3,2-e]isoquinoline (4g). similar to the preparation of 4b from 10, 1-methyl-4-(2-hydroxy-3-methoxyphenyl)-3-(3-oxo-3-ethoxypropyl)pyridinium iodide was converted to 4g (overall yield 15%): 1 H NMR (200 MHz, CDCl₃) δ 1.49–2.99 (m, 11H), 2.43 (s, 3H), 4.41 (s, 1H), 6.78–6.83 (m, 2H), 7.05 (dd, J = 6.9, 1.6 Hz, 1H); IR (neat) 1720 cm $^{-1}$; MS (EI, 70 eV) m/e 273 (M $^{+}$, base).

- 3-(Cyclopropylmethyl)-9-hydroxy-7-oxo-2,3,4,4aa,5,6,7,7a β -octahydro-1H-benzofuro[3,2-e]isoquinoline (4c). 16 was demethylated as described above to give 4c as an oil (41%): ^{1}H NMR (200 MHz, CDCl₃) δ 0.03-0.27 (m, 2H), 0.40-0.66 (m, 2H), 0.78-1.03 (m, 1H), 1.65-2.70 (m, 11H), 2.86-3.22 (m, 2H), 4.64 (s, 1H), 6.60-6.89 (m, 2H); MS (EI, 70 eV) m/e calcd for $C_{19}H_{23}NO_{3}^{+}$ 313.1678, found 313.1687; 313 (M^{+} , base), 272.
- 3-(Cyclopropylmethyl)-7a-hydroxy-9-methoxy- $2,3,4,4a\beta,5,6,7,7a\beta$ -octahydro-1H-benzofuro[3,2-e]isoquinoline (4d). To a solution of ketone 4a (655 mg, 2.00 mmol) in dry THF (50 mL) at -78 °C was added dropwise L-Selectride (5 mL, 1.0 M in THF). The mixture was stirred for 1.5 h, treated with water (1.5 mL), and let warm to room temperature. After evaporation of THF, the residue was treated with 1 N NaOH (50 mL), and the resulting mixture was extracted with 2-propanol/CH₂Cl₂, 1:8 (50 mL \times 3). The combined extracts were washed with brine, dried (MgSO₄), and evaporated. The residue was chromatographed (silica gel; MeOH: CH₂Cl₂, 15:85) to afford **4d** as a white solid (528 mg, 80%): mp 171–173 °C (HCl salt); R_f 0.39 (15:85 MeOH:CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃) δ 0.07-0.12 (m, 2H), 0.46-0.52 (m, 2H), 0.88 (m, 1H), 1.05 (m, 1H), 1.19–1.27 (m, 1H), 1.50–1.64 (m, 2H), 1.84-1.96 (m, 2H), 2.13 (m, 1H), 2.36-2.58 (m, 4H), 2.83-2.93 (m, 2H), 3.18 (s, 1H), 3.80 (s, 3H), 3.91-3.98 (m, 1H), 4.53 (d, J = 3.8 Hz, 1H), 6.69–6.75 (m, 2H), 7.02 (dd, J = 13.1, 7.0 Hz, 1H); 13 C NMR (75 MHz, CDCl₃) δ 3.85, 3.88, 8.0, 22.2, 23.2, 34.7, 39.6, 48.2, 53.5, 54.7, 55.6, 63.4, 67.0, 90.1, 111.4, 119.0, 120.0, 132.0, 144.1, 148.9; IR (KBr) 3367, 2943,

1618, 1585, 1489, 1461 cm $^{-1}$; MS (EI, 70 eV) m/e calcd for $\rm C_{20}H_{27}NO_3^+$ 329.1991, found 329.1987; 329 (M $^+$), 55 (base). Anal. ($\rm C_{20}H_{27}NO_3^+HCl^1/_2H_2O)$ C, H, N.

- 3-(Cyclopropylmethyl)-7a,9-dihydroxy- $2,3,4,4a\beta,5,6,7,7a\beta$ -octahydro-1H-benzofuro[3,2-e]isoquinoline (4e). A mixture of 4d (280 mg, 0.85 mmol), 1-propanthiol (0.6 mL, 6.6 mmol), and NaH (100 mg, 4.17 mmol) in DMF (7 mL) was heated at 110 °C for 3.5 h. DMF was removed by Kugelrohr distillation, and the residue was dissolved in CH₂Cl₂ (15 mL). The soluton was extracted with 1 N NaOH (20 mL \times 2). The aqueous extracts were acidified to pH = 8 with 6 N HCl and extracted with 2-propanol/CHCl₃ (1:4). The combined organic extracts were washed with brine, dried (MgSO₄), evaporated, and recrystallized (2-propanol/ ethyl acetate) to give 4e as a white solid (202 mg, 75%): mp 223-225 °C (HCl salt, dec); R_f 0.31 (20:80 MeOH:CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃) δ 0.04-0.09 (m, 2H), 0.43-0.49 (m, 2H), 0.80-1.01 (m, 2H), 1.14-1.22 (m, 1H), 1.45-1.54 (m, 2H), 1.84 (m, 2H), 2.04 (m, 1H), 2.34-2.41 (m, 3H), 2.50-2.59 (m, 1H), 2.77-2.89 (m, 2H), 3.90 (m, 3H), 4.42 (d, J = 3.8 Hz, 1H), 6.53-6.58 (m, 1H), 6.65 (d, J = 7.7 Hz, 1H), 6.81 (d, J = 7.6Hz, 1H); 13 C NMR (75 MHz, CDCl₃) δ 3.8, 7.4, 22.1, 22.5, 34.1, 38.8, 48.2, 54.5, 63.2, 66.6, 89.5, 116.0, 118.0, 120.2, 131.3, $140.6, 148.3; IR (KBr) 3521, 3440, 3377, 2953, 2744, 1481 cm^{-1}$ MS (EI, 70 eV) m/e calcd for $C_{19}H_{25}NO_3^+$ 315.1835, found 315.1838; 315 (M⁺, base), 274, 55. Anal. (C₁₉H₂₅NO₃HClH₂O) C, H, N.
- 7β -(Benzoyloxy)-3-(cyclopropylmethyl)-9-methoxy- $2,3,4,4a\beta,5,6,7,7a\beta$ -octahydro-1H-benzofuro[3,2-e]isoquinoline (17). To a stirred solution of alcohol 4d (250 mg, 0.759 mmol), triphenylphosphine, (398 mg, 1.518 mmol), and benzoic acid (185 mg, 1.518 mmol) in dry THF (25 mL) was added dropwise diethyl azodicarboxylate (0.24 mL, 1.518 mmol) in THF (1 mL) at room temperature. After 1 h, the solvent was evaporated and the residue was chromatographed (silica gel; MeOH:CH₂Cl₂, 8:92) to afford 17 as an oil: R_f 0.57 (MeOH: CH₂Cl₂, 10:90); ¹H NMR (300 MHz, CDCl₃) δ 0.10-0.15 (m, $2H),\,0.50-0.54\,(m,\,2H),\,0.91\,(m,\,1H),\,1.41\,(m,\,1H),\,1.52-1.62$ $(m,2H),1.84-1.89\,(m,2H),2.00-2.04\,(m,1H),2.17-2.41\,(m,1H),2.18-2.41\,(m,1H),2.21\,(m,1H),2.21\,(m,1H),2.21\,(m,1H),2.21\,(m,1H),2.21\,(m,1H),2.21\,(m,1H),2.21\,(m,1H),2.21\,(m,1H),2.2$ 4H), 2.64 (t, J = 11.8 Hz, 1H), 2.86 (m, 1H), 2.98-3.03 (m, 1H), 3.80 (s, 3H), 4.45 (d, J = 6.2 Hz, 1H), 5.01-5.08 (m, 1H), 6.76-6.82 (m, 2H), 7.05-7.11 (m, 1H), 7.38-7.43 (m, 2H), 7.50-7.55 (m, 1H), 8.03-8.06 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 3.9, 4.0, 8.4, 23.5, 27.6, 39.1, 39.7, 48.3, 49.3, 54.9, 56.2, 63.7, 74.7, 91.2, 112.6, 119.2, 120.4, 128.2, 129.7, 130.3, 132.9, 133.1, 146.0, 148.0, 165.8; MS (EI, 70 eV) m/e 433 (M⁺), 392, 312, 105, 55 (base).
- 3-(Cyclopropylmethyl)-7β-hydroxy-9-methoxy- $2,3,4,4a\beta,5,6,7,7a\beta$ -octahydro-1H-benzofuro[3,2-e]isoquinoline (18). A solution of benzoate 17 and NaOH (120 mg, 3 mmol) in MeOH (25 mL) was stirred for 3 h at room temperature. The solvent was evaporated, and to the residue was added 0.5 N aqueous Na₂CO₃ (15 mL). The resulting mixture was extracted with 2-propanol/CHCl₃, 1:4 (25 mL × 2). The combined extracts were dried (MgSO₄), evaporated, and chromatographed (silica gel; MeOH:CH2Cl2, 20:80) to give **18** as a white solid (135 mg, 54% from **4d**): mp 215-216.5 °C (HCl salt); R_f 0.38 (15:85 MeOH:CH₂Cl₂); ¹H NMR (300 MHz, $CDCl_3$) $\delta 0.09-0.14$ (m, 2H), 0.49-0.55 (m, 2H), 0.91 (m, 1H), 1.22-1.52 (m, 3H), 1.71-1.89 (m, 3H), 2.11 (m, 1H), 2.25-2.42 (m, 3H), 2.64 (t, J = 11.8 Hz, 1H), 2.85-2.89 (m, 1H),2.97-3.02 (m, 1H), 3.51-3.58 (m, 1H), 3.82 (s, 3H), 4.11 (d, J) = 7.1 Hz, 1H), 6.75-6.78 (m, 2H), 6.97-7.03 (m, 1H); $^{13}\text{C NMR}$ $(75 \text{ MHz}, \text{CDCl}_3) \delta 3.9, 4.0, 8.1, 24.5, 30.8, 38.4, 40.5, 48.1,$ $49.3,\ 54.7,\ 55.8,\ \underline{63}.4,\ 71.8,\ 95.5,\ 111.6,\ 119.0,\ 120.3,\ 133.9,$ $146.0,\,147.7;\,IR\,(KBr)\,3514,\,3392,\,2938,\,1489,\,1450,\,1273\;cm^{-1}$ MS (EI, 70 eV) m/e calcd for $C_{20}H_{27}NO_3^+$ 329.1991, found 329.1974; 329 (M⁺, base), 288, 55. Anal. $(C_{20}H_{27}NO_{3}-$ HCl·0.5H₂O) C, H, N.
- 3-(Cyclopropylmethyl)-7 β , 9-dihydroxy-2,3,4,4a β ,5,6,7,7a β -octahydro-1H-benzofuro[3,2-e]isoquinoline (4f). To a solution of BBr₃-(CH₃)₂S (1.08 mmol) in ClCH₂CH₂Cl (25 mL) was added alcohol 18 (79 mg, 0.24 mmol) in ClCH₂CH₂Cl (5 mL). The resulting mixture was stirred for 2 h at reflux and then cooled to room temperature. The cooled mixture was treated with water (5 mL), basified to pH = 10

(2) (a) Ciganek, E. 2,3,4,4a,5,6,7,7a-Octahydro-1H-benzofuro-[3,2-e]-isoquinoline: A New Morphine Fragment. J. Am. Chem. Soc. 1981, 102, 6261-6262. (b) Ciganek, E. Octahydro-1H-benzo-[4,5]furo[3,2-e]-isoquinoline Analgesic and Narcotic Antagonistic

Compounds. U.S. Patent 4 243 668, 1981; Chem. Abstr. 1981, 95, 809241.

(3) Tam, S. W. Unpublished results.

with aqueous Na₂CO₃, and extracted with 2-propanol/CHCl₃, 1:4 (25 mL \times 2). The combined extracts were washed with brine, dried (MgSO₄), and evaporated. The residue was chromatographed (silica gel; MeOH:CH₂Cl₂, 1:4) to give 4f as a white solid (62 mg, 83%): mp 265 °C (dec, HCl salt); R_f 0.30 (MeOH:CH₂Cl₂, 20:80); ¹H NMR (300 MHz, CDCl₃) δ 0.10 (m, $2H),\,0.50\,(m,\,2H),\,0.87\,(m,\,1H),\,1.14-1.45\,(m,\,3H),\,1.66-1.85$ (m, 3H), 2.01 (m, 1H), 2.28-2.43 (m, 3H), 2.64 (t, J = 12.0 Hz,1H), 2.86 (d, J = 12.2 Hz, 1H), 2.97 (dd, J = 11.5, 3.6 Hz, 1H),3.45 (m, 1H), 4.02 (d, J = 7.3 Hz, 1H), 6.60 - 6.70 (m, 2H), 6.81 $(d, J = 6.8 \text{ Hz}, 1\text{H}); ^{13}\text{C NMR} (75 \text{ MHz}, \text{CDCl}_3) \delta 3.9, 4.0, 7.5,$ 24.6, 31.3, 37.5, 40.2, 48.1, 50.0, 54.3, 63.2, 71.5, 94.9, 116.3, 117.6, 120.6, 133.5, 143.0, 146.6; IR (KBr) 3415, 3198, 3145, 2935, 1055 cm⁻¹; MS (EI, 70 eV) m/e calcd for $C_{19}H_{25}NO_3^+$ 315.1835, found 315.1825; 315 (M+).

Opioid Receptor Binding Assay. Brain membranes were prepared from male Hartley guinea pigs, 250-300 g,16 and binding was performed with 100 mM NaCl by the method of Tam.¹⁷ The following labeled ligands were used: 0.5 nM [3H]naloxone (μ -binding);1 nM (-)-[3H]ethylketocyclazocine with 500 nM 2-D-Ala-5-D-Leu-enkephalin (DADLE) and 20 nM sufentanil (κ -binding); 1 nm [3H]DADLE with 4 nM sufentanil (δ -binding) and 1 nM (+)-[³H]SKF 10,047 (σ binding). IC₅₀s were calculated from log-logit plots. Apparent K_i 's were calculated from the equation $K_i = IC_{50}/[1 + (L/K_d)]$

Mouse Antiphenylquinone Writhing (PQW) Test. The methods of Siegmund et al.18 and Blumberg et al.19 were used after modification. Fasted male CFI mice, 18-23 g, are injected with coded doses of test compound and then challenged with 1.25 mg/kg ip phenylquinone 5 min prior to the designated test time. Analgesia is indicated by a complete blockade of the writhing response during a 10-min observation period starting at the designated test time.

Anti-Straub tail (AST) Test. The test was modified from the methods of Shemano and Wendel²⁰ and Blumberg and Dayton.²¹ Fasted male CFI mice, 18-23 g, are injected with coded doses of test compound and then challenged with 0.08 mg/kg ip etonitazene 5 min prior to the designated test time. Narcotic antagonism is indicated by a complete blockade of the narcotic Straub tail response during a 5-min observation period starting at the designated test time.

Stimulated Guinea Pig Ileum Bioassay. Male Hartley guinea pigs weighing between 250 and 400 g were used. The longitudinal muscle strips were prepared by the method of Rang.²² Tissues were mounted in 10-mL organ baths between vertically-spaced platinum electrodes and bathed with 37 °C oxygenated Krebs solution. The composition of the Krebs solution was (mM): NaCl 117, KCl 5.9, CaCl₂ 2.4, MgCl₂·6H₂O 1.2, D-glucose 11.8, Tris base 20. Preparations were stimulated electrically (0.1 Hz, 0.8 ms duration, supramaximal voltage at a resting tension of 1 g. Contractions were recorded isometrically on a polygraph. The preparation was allowed to equilibrate with continuous stimulation for 90 min. Cumulative dose-response curves for the test compounds were measured. After complete washout and reequilibration, tissue preparations were incubated with an antagonist for 30 min, and then cumulative dose-response curves for the agonists were measured again.

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