Synthesis, NMDA Receptor Antagonist Activity, and Anticonvulsant Action of 1-Aminocyclobutanecarboxylic Acid Derivatives

Yehiel Gaoni,* Astrid G. Chapman,† Naila Parvez,† Peter C.-K. Pook,‡,§ David E. Jane,‡ and Jeffrey C. Watkins*,‡

Department of Organic Chemistry, Weizmann Institute of Science, Rehovot 76100, Israel, Institute of Psychiatry, De Crespigny Park, London, SE5 8AF U.K., and Department of Pharmacology, School of Medical Sciences, Bristol, BS8 1TD U.K.

Received July 27, 1994®

A range of cis- and trans-3-substituted 1-aminocyclobutane-1-carboxylic acids has been synthesized and evaluated for antagonism at excitatory amino acid receptor sites and for anticonvulsant activity. Potent and selective antagonist activity at N-methyl-D-aspartate (NMDA) receptor sites in neonatal rat motoneurones was shown by compounds in which the 3-substituent was, or contained, a 2'-carboxyethyl or 2'-phosphonoethyl moiety. Substances 4b, 24, 35, and 40 were more potent than the standard NMDA receptor antagonist, D-2-amino-5-phosphonopentanoate (D-AP5) as NMDA antagonists in this preparation, and about equipotent with $[3-(\pm)-2$ -carboxypiperazin-4-yl)-1-propyl]phosphonate (CPP). Anticonvulsant activity, as assessed following intracerebroventricular injection into audiogenic DBA/2 mice, generally paralleled NMDA receptor antagonist activity.

Introduction

Excitatory amino acid receptors represent the predominant excitatory synaptic transmitter receptors in the mammalian central nervous system. 1,2 There are two major families: metabotropic receptors, linked to second messenger systems through G proteins, and ionotropic receptors linked to ion-channels. It is likely that the transmitter activating these receptors is Lglutamate. In each family several major types have been recognized and, for the ionotropic excitatory amino acid receptors, these comprise the N-methyl-D-aspartate (NMDA), \alpha-amino-3-hydroxy-5-methylisoxazole-4-propionic acid (AMPA), and kainate receptors, named after their classical selective agonists. 1,3 Such receptors have been recongized as potential targets for therapeutic intervention in a number of neurological conditions.4-6 In particular, antagonists of the NMDA receptor have well-documented anticonvulsant, antispastic, and neuroprotective properties.7 The starting point for the present study was the finding that the glutamic acid analogue trans-1-aminocyclobutane-1,3-dicarboxylic acid^{8,9} (1a) is an extremely potent NMDA receptor agonist. Conventionally, NMDA receptor agonists can be converted to antagonists by increasing the length of the connecting chain between the α - and ω -acidic groups so that it becomes from one to three atoms longer than that in glutamic acid itself.^{3,10} Usually,^{3,10} but not always, 11,12 the α -amino carboxylic acid moiety in these compounds has the (R) configuration. The most effective ω-acidic group for NMDA receptor antagonist activity is the phosphonic group, and the prototypes for NMDA receptor antagonist activity are thus (R)-2amino-5-phosphonopentanoate (D-AP5; 5a) and (R)-2amino-7-phosphonoheptanoate (D-AP7; 5c).^{3,10} Surprisingly, however, neither trans- nor cis-1-amino-3-(phosphonomethyl)cyclobutane-1-carboxylic acid (3a. 4a) nor the propyl analogues (3c, 4c) showed potent

HOOC(CH₂)_n COOH

$$NH_2$$
 HOOC(CH₂)_n NH₂

1a: n = 0
1b: n = 1
1c: n = 2
1d: n = 3

(HO)₂OP(CH₂)_n COOH
NH₂ COOH
NH₂

3a: n = 1
3b: n = 2
3c: n = 3

(HO)₂OP-CH₂-(CH₂)_n COOH
(HO)₂OP-CH₂-(CH₂)_n COOH
NH₂

5a: n = 2
5b: n = 3
5c: n = 4

* Author to whom correspondence should be addressed.

Chemistry

The synthesis of the presently described cyclobutane amino acids is based on a general scheme for the preparation of substituted cyclobutanes from 1-(phenylsulfonyl)bicyclo[1.1.0]butanes. Up to four functional-

antagonist activity at NMDA receptors in previous studies.9 We nevertheless felt it worthwhile to explore the structure-activity relations of this series further, initially with the thought of retaining the two carboxyl groups of compound 1a (to link with agonist binding sites in the receptor) and introducing a further substituent in the 3-position to link with an additional antagonist binding site that has been postulated to exist in the receptor.^{3,13,14} We report here the synthesis of such a new series of potent NMDA antagonists, the crucial structural feature of which is not, however, the retention of the agonist 3-carboxy group (which can be omitted) but the introduction of a carboxyethyl or phosphonoethyl group, preferably in the cis configuration, and yielding analogues of 2-amino-6-phosphonohexanoic acid (AP6; 5b) rather than of AP5 (5a) or AP7 (5c) analogues.

[†] London.

[‡] Bristol.

[§] Present address, School of Health Sciences, Sunderland University, SR1 3SD U.K.

^{*} Abstract published in Advance ACS Abstracts, November 1, 1994.

Scheme 1^a

 $^{\alpha}$ (a) NaN3, DMF; (b) SOCl2; (c) piperidine, CH2Cl2; (d) H2, Pd/C; (e) PhCOCl, NEt3, CH2Cl2; (f) BuLi, CO2; (g) CH2N2; (h) 6% Na-Hg.

ized groups can be introduced into the original four-membered ring of the bicyclobutane by a sequence of reactions comprising (1) substitution of the bridghead proton, (2) addition of a nitrogen nucleophile across the central bond, (3) substitution α to the sulfone, mainly by a carboxyl, and (d) eventual substitution, after desulfonylation, α to the latter carboxyl. ^{15,16}

Scheme 1 outlines the synthesis of intermediates 18 and 19, used in the preparation of all cyclobutane amino acids described in this work. By carrying out the azidation of acid 6^{16} with sodium azide in DMF-tetramethylguanidine (TMG), product 7 was obtained regioselectively and stereoselectively in 86-92% yield. The stereoselectivity results, probably, from the kinetic protonation of an incipient α -sulfonyl anion by the TMG bound to the carboxyl group. Thus, while the azide ion adds from the endo side of 6, intramolecular protonation necessarily occurs from the exo side, leading to cis-7. Azidation of the piperidine amide of 6^{16} with tetramethylguanidinium azide in N-methyl-2-pyrrolidone produced both 7 and its geometrical trans isomer.

Adduct 7 was converted to the piperidine amide 9 via acid chloride 8, using standard procedures. Catalytic reduction of the azide group to the amine was more elaborate, since azide 9 produced a stable triazene intermediate which needed extra treatment for conversion to the amine. Following the hydrogenation by TLC, it is observed that while the starting azide disappears, two more polar spots appear, the less polar of which converts slowly to the more polar one. By stopping the hydrogenation after a few hours, solid, stable triazene 11 could be separated from amine 10 by chromatography and be fully characterized. Amine 10, free of 11,

was obtained in 93-96% yield after prolonged stirring under hydrogen over the palladium catalyst, followed by reflux under air atmosphere until total conversion of the triazene to the amine.

The benzoylated derivative 12, obtained from 6 in about 70% overall yield, is an important intermediate for the preparation of a large number of 3-substituted 1-aminocyclobutane-1-carboxylic acids via its dilithiated derivative. Carboxylation leads to a mixture of cis and trans acids 13. Separation of the isomers is not required before desulfonylation, since the geometry of the separated individual isomers is, anyway, not conserved in the desulfonylated product. Separations were, however, carried out in some cases for characterization purposes. Thus, after esterification of 13, isomers 14 and 15 were separated and individually characterized.

Key intermediates 18 and 19 were obtained by desulfonylation of total crude 13 followed by esterification and chromatographic separation, ester 19 being slightly more abundant. Alternatively, almost pure acids 16 and 17 could be obtained by fractional crystallization of the total acid product of desulfonylation and further purification by crystallization from ethanol. The total overall yield of 18 and 19 from 6 is 50-55%.

Configurational assignments of all compounds are based on correlations of certain intermediates with compounds the structure of which has been determined by X-ray crystallography¹⁸ and on spectral proton NMR considerations.¹⁶ In particular, the geometry of **18** and **19** was established by acid hydrolysis to the corresponding amino dicarboxylic acids. Thus, hydrolysis of **18** furnished the known atelia-herbert-smithii acid **2a**, namely, cis-1-aminocyclobutane-1,3-dicarboxylic acid, ^{18,20} while **19** furnished trans-1-aminocyclobutane-1,3-dicarboxylic acid **1a**, the structure of which has been established by X-ray crystallography. Amino acid **1a** was later found to be a potent and selective agonist of the NMDA subtype of glutamate receptors of the central nervous system. ^{8,9}

Intermediates 18 and 19, of well-established configuration, were used to prepare the NMDA antagonists which constitute the subject of this publication. Active products were obtained either by substitution α to the methyl carboxylate, using LDA and an electrophile, or by modifying the carboxylate group itself. The following schemes describe the specific transformations used in each case.

Treatment of 18 or 19, or of a mixture of both with lithium diisopropylamide (LDA) followed by addition of methyl γ -bromocrotonate provided in a total 78% yield a mixture of adducts 21 and 22, produced probably via the conjugate addition intermediate product 20 (Scheme 2). No expected ethylenic product was detected. The relative geometry on the cyclopropane ring of 21 and 22 was unique, and most probably trans, because of an expected steric congestion in a cis isomer. A trans selectivity was observed also in sterically less demanding cases where a similar conjugate addition—cyclization sequence has been applied. 21,22 Acid hydrolysis of the separated product isomers provided amino acids 23 and 24 in about 70% yield.

The route followed from 18 to 2c and from 19 to 1c is shown in Scheme 3. It involves, in the *cis* series, the reduction of ester 18 with sodium borohydride to alcohol 25 followed by oxidation with activated DMSO to

Scheme 2a

^a (a) LDA; BrCH₂CH=CHCOOCH₃; (b) 6 M HCl.

Scheme 3^a

 $^{\alpha}$ (a) NaBH₄; (b) DMSO-TFAA; (c) Ph₃P=CHCOOCH₃; (d) H₂, cat.; (e) 6 M HCl.

aldehyde **26**, carboxyolefination to **27**, hydrogenation to **28**, and hydrolysis to **2c** (ca. 30% overall yield from **18**). In the *trans* series, amino acid **1c** was similarly obtained from **19**.

Intermediate **25** is also a direct precursor of the *atelia-herbert-smithii* acid *cis-*1-amino-3-(hydroxymethyl)cyclobutane-**1**-carboxylic acid, ²³ while **29** is a precursor of the corresponding, nonnatural *trans* isomer. ²⁴

Scheme 4 describes the preparation of amino acids 35, 38, 39, 40, and 41. The reaction of ester 18 with dimethyl (lithiomethyl)phosphonate produced ketone 33. Reduction of 33 with sodium borohydride in ethanol furnished racemic alcohol 34 which was hydrolized to the racemic amino acid 35. The overall yield from 18 was 50-53%.

Amino acid 38 was similarly obtained from 19. However, in this case, it was accompanied by the elimination product 39, which could be separated from 38 and fully characterized.

Hydrolysis of either **33** or **36** produced a similar mixture of amino acids **40** and **41**, in a ratio of ca. 3:2, respectively, and in a total 75-80% yield. Equilibration

Scheme 4^a

Scheme 5^a

^a (a) CH₃SO₂Cl, Et₃N; (b) BuLi; (c) H₂, cat.; (d) 6 M HCl.

of the isomers occured probably by enolization toward the ring. Amino acid 40 was obtained pure by crystallization of this mixture, or of chromatographically enriched mixtures, from water. Its geometry was established by catalytic hydrogenation in water and acetic acid to acid 35. Pure 41 was obtained in small amounts from the very last chromatography fractions.

Scheme 5 describes the preparation of 4b from 34 and of 3b from 37. In order to eliminate the hydroxyl group of 34 or 37 toward the phosphonate group and not toward the ring, it was first mesylated and then treated with butyllithium, producing 43 and 46, respectively. Hydrogenation and hydrolysis provided amino acids 4b and 3b in similar yields of ca. 50% from the corresponding alcohols.

Table 1. Antagonism of NMDA-Induced Depolarization and Anticonvulsant Activity of Cyclobutane Derivatives

		anticonvulsant action $(\mathrm{ED}_{50})^a$ (range^b)	
compd	\mathbf{EPMR}^{c}	μg	nmol
5a	1.0		
DL-CPP	0.20		
23	inact.		
	(1 mM)		
24	0.28	0.089 (0.059-0.134)	0.36(0.23-0.55)
2c	1.8	16.09 (14.39-18.01)	86.0 (77.0-96.3)
1c	7.9		
35	0.39	0.032 (0.024-0.048)	0.13(0.10-0.18)
38	1.0	0.124 (0.093-0.165)	0.52(0.39 - 0.69)
40	0.33	0.106 (0.071-0.157)	0.53 (0.035-0.78)
41	4.2	•	
4b	0.32	0.032 (0.024-0.048)	0.14(0.11 - 0.22)
3b	1.1	,	

^a Antagonist activity following intracerebroventricular administration against sound-induced clonic convulsions in DBA/2 mice; 7–11 mice per drug. ^b 95% confidence limits. ^c Equipotent molar ratio of cyclobutane derivatives relative to D-AP5 = 1.0 on neonatal rat motoneurones.

Pharmacology

Neonatal Rat Spinal Cord in Vitro. Previous studies have indicated that neither 3-(phosphonomethyl)- nor 3-(phosphonopropyl)-1-aminocyclobutane-1carboxylic acid had pronounced NMDA receptor antagonist activity.9 In the present study we confirmed this lack of potent activity for a mixture of cis and trans isomers of 3-(phosphonopropyl)-1-aminocyclobutane-1carboxylic acid (3c, 4c). However, marked antagonist activity was observed with a series of 1-aminocyclobutane-1-carboxylic acids bearing a carboxyethyl or phosphonoethyl substituent in the 3-position (Table 1). The most potent substances (24, 35, 40, and 4b) were approximately equipotent with (\pm) -CPP and about 3-4 times more potent than D-AP5 (5a). All of these compounds had the same configuration and were more active than their genometric isomers (23, 38, 41, and **3b**). The same isomeric preference (with the 1-amino and substituted 3-ethyl group cis to one another) was shown in another pair of moderately active isomers (1c, **2c**). Direct comparison of compounds with either carboxyethyl or phosphonoethyl substituents (1c, 3b; 2c, **4b**) indicated the latter compounds to be the more potent. However, among the most potent compounds was 24 which contained both a trans-3-carboxy and a cis-3-(2-carboxycyclopropyl) substituent relative to the 1-amino group. Interestingly, the corresponding cis-3carboxy-trans-3-(2-carboxycyclopropyl)-substituted compound 23 was completely inactive.

Anticonvulsant Activity. Six of the novel NMDA antagonists were tested for protection against audiogenic seizures in DBA/2 mice. Intracerebroventricular administration of these compounds led to dose-dependent suppression of sound-induced seizures. The ED₅₀ values for five of these substances (24, 35, 38, 40, 4b) all fell within 4-fold range of $0.032-0.124\,\mu\mathrm{g}$ ($0.13-0.52\,\mathrm{nmol}$); the other compound (2c) was 100-500-fold less potent (Table 1). The choice of pretreatment time (60 min) was based on preliminary results using compound 24, where 30 or 60 min pretreatment periods (icv) resulted in closely similar anticonvulsant ED₅₀ values.

Little or no acute overt behavioral effects of the drugs were observed at the pharmacologically effective doses used. Slight ataxia was observed at the highest doses used of **24** $(0.2 \mu g)$; the latter case was also associated

with a significant drug-induced decrease in rectal temperature. At doses 5–10-fold higher than those required for anticonvulsant protection (1 μ g of 24 and 50–100 μ g of 2c) the mice exhibited mild to moderate ataxia.

With one exception (compound **2c**), these studies on the suppression of audiogenic seizures in DBA mice showed a close parallelism between the NMDA receptor antagonist activity of the compounds and their anticonvulsant activity (Table 1). Although **2c** was the least active compound in both the *in vitro* and *in vivo* studies, the relative potency of this compound as an anticonvulsant was much lower than would have been predicted from its spinal cord NMDA receptor antagonist activity.

In these studies the compounds were injected intracerebroventricularly because of the small quantities of the substances available. It is not known how well the substances would pass the blood-brain barrier if injected systemically, but it would be expected that their ability to penetrate into the brain would resemble that of CGP 37849.²⁵ In this case, esterification of the α -carboxyl group may improve the ability of the compounds to pass the blood-brain barrier, though probably lowering its potency at the target sites, as shown in the case of CGP 37849.²⁵ Further studies are required to elucidate this question.

Structure-Activity Discussion

The high activity of compound 24 (Table 1) as an NMDA receptor antagonist in neonatal rat motoneurones indicated the apparent success of our original strategy of retaining the trans-3-carboxyl group of the agonist trans-1-aminocyclobutane-1,3-dicarboxylic acid (1a). However, our subsequent exploration of related substances (Y. Gaoni, P. C.-K. Pook, and J. C. Watkins, unpublished observations) seemed to indicate that the crucial feature of this activity was not the presence of the agonist-essential trans-3-carboxyl moiety; for instance, the analogue of 2c containing an additional trans-3-carboxy group had the same activity as 2c. Indeed, the consistent structural feature of all the most active substances was the presence of a 2'-carboxyethyl or a 2'-phosphonoethyl group in the 3-position of the cyclobutane ring. The other structural features to emerge from the results reported here were (a) a phosphonoethyl group was more effective than a carboxyethyl group, (b) the cis orientation of the acidic 3-ethyl substituent was preferred to the trans orientation, and (c) a 1'-keto, 1-hydroxy or 1', 2'-methylene group were well tolerated in the 3-ethyl side chain.

The most unusual feature of these structure—activity relations is the finding that high activity was shown by "AP6-length" compounds. Allan and colleagues had previously reported only low activity of cis/trans mixtures of AP5- and AP7-length cyclobutane analogues, which, in other groups of compounds usually have higher activity than AP6-length compounds. We have confirmed this low activity of AP5-length and AP7-length cyclobutane analogues in our current study (unpublished observations of Y. Gaoni, P. C.-K. Pook, and J. C. Watkins). An explanation of these results were sought in computer-assisted molecular modeling studies.

Molecular Modeling Studies on AP5-AP7-Length Cyclobutane Analogues. Two template molecules

Figure 1. (-)-Gauche conformation of the CGS19755 template.

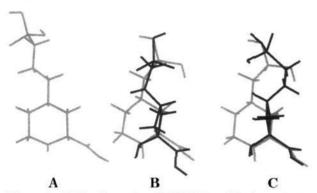


Figure 2. (A) Conformation of CGSAP7 used in the modeling study. (B) Superimposition of 4b and CGS19755. (C) Superimposition of 3b and CGS19755.

Table 2. Results of Superimposition of Cyclobutane Analogues on Both AP5- and AP7-Length Templates

	RMS deviation in aligned positions		
compd	CGS19755 template	CGSAP7 template	
4a	0.860	No Fit	
3a	0.515	0.490	
4b	0.155	0.131	
3b	0.147	0.182	
4c	0.403	0.359	
3c	No Fit	No Fit	

were selected to represent both AP5- and AP7-length analogues. In agreement with other molecular modeling studies of the NMDA receptor antagonist site, 11,26,27 the (-)-gauche conformation of CGS19755 (Figure 1) was used as the AP5-length template. The analogue of CGS19755 with the AP7-length (CGSAP7; Figure 2A)^{26,27} was chosen as the AP7-length template.

In this study all molecules were assembled using INSIGHT II (Version 2.2.0),33 and then low-energy conformations were obtained by performing energy minimization calculations using DISCOVER (steepest descents followed by the quasi-Newton-Raphson algorithm). The molecules were modelled in the un-ionized state. The N atom of the amino group, an oxygen atom of the carboxyl group, and an oxygen atom of the phosphonic acid group of the cyclobutane analogues (selected to represent relatively fixed sites of interaction with complementary sites in the receptor) were superimposed on the corresponding atoms in either the CGS19755 or the CGSAP7 template. The RMS deviation in aligned positions of these atoms was then recorded (see Table 2). The CGSAP7 template could be superimposed on to the CGS19755 template with a high degree of precision using the strategy outlined above (RMS deviation in aligned positions is 0.111).

The model suggests that both cis and trans forms of the AP6-length cyclobutane analogues (4b and 3b, respectively) might be expected to fit either AP5preferring or AP7-preferring subtypes of the NMDA receptor better than would the corresponding AP5 or AP7 cyclobutane analogues. In the case of an AP7preferring subtype, the model would also predict the higher activity of the cis relative to the trans form of the AP6 cyclobutane analogue, as observed (Table 1). In the case of an AP5-preferring receptor subtype, however, the trans form of the AP6-length analogue would have been predicted to be of similar potency to the cis form. If an AP5-preferring subtype of NMDA receptor is a major contributor to the excitatory amino acid receptor population in neonatal rat motoneurones, the observed lower activity of the trans cyclobutane AP6 analogue may possibly be explained by steric hindrance to accommodation of the ethyl chain in the trans form (Figure 2B.C). The current model differs from that of Ortwine and colleagues²⁷ in that, although a very similar conformation of the CGS19755 template was utilized, a more distal oxygen atom of the phosphonate group (relative to the carboxylate group) was required as the primary receptor interaction point in order to obtain good superimposition of the CGSAP7 template and the cis cyclobutane AP6 analogue (4b) on to the CGS19755 template. Limitations of this preliminary study may be more readily rationalized later when selective agonists and antagonists for specific subtypes of NMDA receptors become available. In this respect, it will be informative to test the compounds of the present investigation on various assemblies of cloned NMDA receptor subunits.²⁸

Experimental Section

Chemistry. General Procedures. Melting points were taken on a Fisher-Johns apparatus and were not corrected. Proton NMR spectra were measured with a Bruker AMX400 spectrometer in CDCl₃ at 400 MHz, unless otherwise indicated. Shifts are given in δ units downfield from internal TMS, and J values are given in hertz. For spectra taken in D₂O, shifts are given as above, based on locating the HDO signal at d 4.80 ppm. TLC was done on Merck Kieselgel precoated aluminum plates. Silica gel for column chromatograhy was Merck Kieselgel 60 (70–230 mesh). The solvent of crystallization is indicated in paretheses after the indicated melting points. Elemental analyses were performed at the Microanalytical Laboratory of the Hebrew University, Jerusalem. Analytical values were within $\pm 0.4\%$ of the calculated theoretical values.

cis-1-Azido-3-(phenylsulfonyl)cyclobutane-1-carboxylic Acid (7). The bicyclic acid 6 (2.38 g, 10 mmol) was warmed in DMF (30 mL) with TMG (2 mL) and NaN₃ (0.74 g, 15 mmol) at 90 °C during 6 h. The cooled solution was diluted with water, extracted twice with ether to remove nonacidic material, and then acidified with 3 M HCl, NaCl was added to saturation, and the mixture was extracted three times with ether. (Caution! Conduct these operations in a well ventilated hood-hydrazoic acid may be liberated). The ether solution was washed three times with water and dried over MgSO4. Evaporation of the solvent, trituration of the solid product with hexane, and filtration furnished from 2.4 to 2.6 g (86-92%) of pure 7. Azide 7: mp 102-103 °C (benzene); NMR (250 MHz) 2.76-2.94 (symmetrical m, 4 H), 3.86 (pent, 1 H), 7.6-7.9 (5 H, aromatic protons), 9.80 (br, acidic H). Anal. $(C_{11}H_{11}N_3O_4S)$ H, N; C: calcd, 46.98; found, 46.52.

cis-1-Azido-3-(phenylsulfonyl)cyclobutane-1-N,N-pentamethylenecarboxamide (9). Acid 7 was warmed with excess SOCl₂ (ca. 1 mL/mmol of 7) at 75 °C for 1 h. Excess SOCl₂ was then evaporated at reduced pressure, and the residue was treated twice by addition of hexane and reevaporation to dryness. The acid chloride 8 was dissolved in CH₂-Cl₂ (10 mL/mmol of 7) and cooled in ice. Piperidine (0.2 mL/mmol of 7; 2 equiv) was added to the solution, and the reaction was allowed to proceed for 1 h at room temperature. The reaction mixture was washed with dilute acid and water, dried, and evaporated to provide 9 in near quantitative yield. Azide

9: mp 94-95 °C (CH₂Cl₂-hexane); NMR (250 MHz) 1.60 (br, 6 H), 2.78-2.99 (symmetrical m, 4 H), 3.35 (br, 2 H, two of CH₂ NCH₂ protons), 3.48-3.61 (m, 3 H, two of CH₂ NCH₂ protons plus tertiary ring proton), 7.5-7.9 (5 H). Anal. $(C_{16}H_{20}N_4O_3S)$ C, H, N.

cis-1-Amino-3-(phenylsulfonyl)cyclobutane-1-N,N-pentamethylenecarboxamide (10). Hydrogenation of 9 was carried out in ethyl acetate under atmospheric pressure with 10% Pd/C as the catalyst. The catalyst (10% by weight of 9) was first stirred in ethyl acetate under hydrogen for 0.25 h. A solution of 9 in the same solvent (10 mL/mmol of 9) was then added, and stirring under hydrogen was resumed and maintained for 20 h. If the reaction mixture still showed the triazene spot, it was refluxed until total conversion of 11 to 10. Filtration of the catalyst and evaporation of the solvent furnished solid 10 in 93-96% yield: mp 109-110 °C (CH₂- Cl_2 -hexane); NMR (250 MHz) 1.55 (br, 6 H), 1.89 (s, NH2), 2.44-2.52 and 3.03-3.11 (two m, 4 H), 3.44-3.53 (m, 5 H, CH_2 NCH₂ plus tertiary ring proton), 7.5-7.9 (5 H). Anal. $(C_{16}H_{22}N_2O_3S)\ C,\ H.$

cis-1-(Benzoylamino)-3-(phenylsulfonyl)cyclobutane-1-N,N-pentamethylenecarboxamide (12). Benzoylation of 10 was carried out in CH₂Cl₂ with Et₃N and PhCOCl (4 mL, 0.15 mL and 0.116 mL, respectively, per mmol, 322 mg, of 10). After addition of the benzoyl chloride at 0 °C, the reaction flask was warmed to room temperature for 1 h, during which time the product precipitated. Most of the solvent was then evaporated at reduced pressure, and the residue was taken in water, filtered, washed with water and with ethanol-water, and then air-dried. The highly insoluble product 12 thus obtained (80-86% yield relative to acid 6) is suitable for all further uses. An analytical sample was obtained by crystallization from ethanol: mp 277-278 °C; NMR 1.43 and 1.55 (two br s, 6H), 2.79 and 3.31 (two m, 4 H), 3.39 and 3.56 (two $br\,s,\,4\,H),\,3.73\,(pent,\,1\,H),\,7.38\,(s,\,NH),\,7.4-7.9\,(10\,H).\ \, Anal.$ $(C_{23}H_{26}N_2O_4S)$ C, H, N.

cis- and trans-3-(Benzoylamino)-3-(pentamethylenecarbamoyl)-1-(phenylsulfonyl)cyclobutane-1-carboxylic Acid (13) and cis- and trans-Methyl 3-(Benzoylamino)-3-(pentamethylenecarbamoyl)-1-(phenylsulfonyl)cyclobutane-1-carboxylate (14 and 15). A solutionsuspension of 12 (5 g, 11.7 mmol) in THF (140 mL) was stirred at -40 to -50 °C with 2.2 equiv of BuLi during 1 h. Pieces of solid CO2 were then added to the solution, and the reaction flask was allowed warm out of the cooling bath to room temperature. Water was then added, and most of the THF was evaporated under reduced pressure. The residue was dissolved in additional water (total volume ca. 100 mL) and extracted twice with CH2Cl2 for any nonacidic material, including unreacted 12 which may be recovered later. The water layer is then acidified and extracted several times with CH₂Cl₂ for the product. Crude 13, a mixture of cis and trans isomers, is usually obtained as a solid foam in 86-94% yield (4.8-5.2 g) and is ued as such for desulfonylation. For characterization purposes, a sample of crude 13 was esterified in THF with an ethereal solution of diazomethane and the esters were chromatographically separated (silica gel, 50 times the weight of product EtOAC-CH₂Cl₂-hexane, 1:1:1). The first eluted product was the cis-S,N compound 14: mp 256-257 °C dec (MeOH); NMR (80 MHz) 1.6 (br s, 6 H), 3.24 and 3.66 (ABq, 4 H), 3.55 (br s, 4), 3.66 (s, Me) 7.4-8.0 (10 H). Anal. $(C_{25}H_{28}N_2O_6S)$ C, H, N.

The second eluted product was the trans-S,N ester 15, mp 261-262 °C (dec.; MeOH); NMR (80 MHz) 1.53 (br s, 6 H), 3.09 and 3.72 (ABq, 4 H), 3.5 (br s, 4 H), 3.63 (s, Me), 7.4-8.0(10 H). Anal. $(C_{25}H_{28}N_2O_6S)$ C, H, N.

cis- and trans-1-(Benzoylamino)-1-(pentamethylenecarbamoyl)cyclobutane-3-carboxylic Acid (16 and 17) and cis- and trans-Methyl 1-(Benzoylamino)-1-(pentamethylenecarbamoyl)cyclobutane-3-carboxylate (18 and 19). Crude 13 (ca. 5 g), obtained by carboxylation of 5 g of 12, was stirred in a mixture of 30 mL of THF and 90 mL of MeOH with 12 g of 6% sodium amalgam during 3 h at room temperature. The reaction mixture was filtered on Celite, the solution was slightly acidified with 3 M HCl, and most of the solvent was evaporated at reduced pressure. Saturated aqueous NaCl (20 mL) was added to the residue, which was then acidified further with HCl and extracted three times with CH2-Cl₂ (total volume: ca. 75 mL). On washing the CH₂Cl₂ solution with water to remove excess HCl, the product begins to precipitate. After completion of the precipitation (ca. 1 h), the mixture is filtered to recover the solid product. This is washed on the filter with water and then air-dried to give 1.2-1.5 g of almost pure 17 (as indicated by esterification of a small sample). The filtrate is then further washed with water, dried, and evaporated to give a residue which can be induced to precipitate 0.3-0.4 g of almost pure 16, or else may be esterified and chromatographed for esters 18 and 19. Alternatively, the total acid product can be esterified with ethereal diazomethane in CH2Cl2-THF by stirring the reaction mixture until all the solid is dissolved, while ensuring the presence of excess diazomethane. The total yield in acids and esters is 74-79% relative to 12.

Pure acid 16 was obtained by crystallization of the above product from ethanol, mp 207-208 °C. Anal. $(C_{18}H_{22}N_2O_4)$

Pure acid 17 was similarly obtained by crystallization of the above described product from ethanol, mp 209-210 °C. Anal. $(C_{18}H_{22}N_2O_4)$ H; C: calcd. 62.05; found, 62.47.

For the separation of 18 and 19, the total product from carboxylation and desulfonylation of 6 g of 12 was esterified and chromatographed (200 g of silica gel; CH₂Cl₂-EtOAcether, 8:5:6). This provided 1.65 g of 18, 0.4 g of a mixture of 18 and 19, and 1.7 g of 19 (total: 3.75 g, 75% from 12).

Product 18: mp 185-186 °C (CH₂Cl₂-hexane); NMR 1.55 (br, 6 H), 2.49-2.56 and 3.22-3.30 (two unsymmetrical m, 4 H), 3.06 (m, 1 H), 3.50 (br, 4 H), 3.71 (s, Me), 7.08 (s, NH), 7.4-7.8 (5 H). Anal. ($C_{19}H_{24}N_2O_4$) C, H, N.

Product 19: mp 225-226 °C (CH₂Cl₂-hexane); NMR 1.57 (br, 6 H), 2.83-2.91 and 3.00-3.08 (two symmetrical m, 4 H), 3.71 (pent, 1 H), 3.53 (br s, 4 H), 3.69 (s, Me), 6.61 (s, NH), 7.4-7.8 (5 H). Anal. (C₁₉H₂₄N₂O₄) C, H, N.

cis- and trans-1-Aminocyclobutane-1,3-dicarboxylic **Acid (2a and 1a).** Product 18 (1.6 g, 4.65 mmol) was warmed in 6 M HCl (15 mL) at 130 °C for 24 h. Water (15 mL) was added to the cooled solution which was then extracted twice with ether to remove benzoic acid. The water solution was evaporated to dryness, and the residue was taken in water and reevaporated. The same operation was repeated twice more with water. The final hydrochloride residue was dissolved in ethanol (10 mL) and added with propylene oxide (2-3mL). Precipitation of 2a started after a while and was over after ca. 3 h. The crystalline solid was collected by filtration and recrystallized from water, yielding pure cis isomer 2a (0.7 g, 85%), identical with an authentic sample. 18 NMR (D₂O) 2.54-2.59 and 2.75-2.81 (symm. m, 4 H), 3.36 (pent, 1 H).

The trans isomer 1a was similarly obtained from 19, and in similar yield. It was identical in all respects with an authentic sample. 18 NMR (D₂O) 2.57-2.63 and 2.88-2.93 (m, 4 H), 3.41 (m, 1 H).

Methyl r-3-(Benzoylamino)-t-1-[trans-2-(methoxycarbonyl)cyclopropyl]-3-(pentamethylenecarbamoyl)cyclobutane-1-carboxylate (21) and Methyl r-3-(Benzoylamino)-c-1-[trans-2-(methoxycarbonyl)cyclopropyl]-3-(pentamethylenecarbamoyl)cyclobutane-1-carboxylate (22). To a solution of a mixture of 18 and 19 (1.71 g, 4.97 mmol) in THF (80 mL) was added at -78 °C a solution of LDA, prepared at 0 °C from BuLi (8.4 mL of 1.47 M solution, 12.5 mmol; 2.5 equiv) and disopropylamine (2 mL, 13 mmol) in THF (15 mL). After stirring at -78 °C for 0.5 h, excess methyl γ -bromocrotonate (1.0 mL, ca. 9.0 mmol) was added, and stirring was continued in an ice bath for 1 h. Saturated aqueous ammonium chloride solution (5 mL) was added, and most of the THF was evaporated under reduced pressure. Water was added to the residue, slightly acidified, and extracted with EtOAc. After washing and drying, the solution was added with ethereal diazomethane to account for any hydrolized acidic product. Evaporation of the solvent and chromatography (200 g of silica gel; CH₂Cl₂-ether-EtOAc, 4:2: 1) separated 21 (1.14 g) from 22 (0.43 g), with intermediate mixed fractions (0.15 g; total: 1.72 g, 78%).

Compound **21**: mp 212–213 °C (EtOH); NMR 1.01–1.06, 1.18–1.23, 1.63–1.68, and 1.79–1.84 (four m, 4 H, cyclopropane protons), 1.50–1.56 (br, 6 H), 2.58–2.80 (m, 4 H, cyclobutane protons), 3.39 and 3.52 (br, 4 H), 3.68 and 3.73 (two s, two Me), 7.17 (s, NH), 7.4–7.8 (5 H). Anal. ($C_{24}H_{30}N_2O_6$) C, H.

Compound **22**: mp 230–231 °C (EtOH); NMR 1.07–1.12, 1.20–1.25, 1.76–1.80, and 1.85–1.90 (four m, 4 H, cyclopropane protons), 1.50 and 1.59 (br, 6 H), 2.56–2.60 and 3.25–3.30 (two m, 4 H, cyclobutane protons), 3.47 (br s, 4 H), 3.63 and 3.70 (two s, two Me), 6.80 (s, NH), 7.4–8.0 (5 H). Anal. ($C_{24}H_{30}N_{2}O_{6}$) C, H.

r-1-Amino-c-3-(trans-2-carboxycyclopropyl)cyclobutane-1,3-dicarboxylic Acid (24). Product 22 (86 mg) was treated with 6 M HCl (10 mL) as described above for the conversion of 18 to 2a to yield acid 24 (32 mg, 68%): mp darkening from above 200 °C, no melting (H₂O); NMR (D₂O) 1.09-1.14, 1.21-1.26, 1.66-1.70, and 1.81-1.86 (four m, 4 H, cyclopropane protons), 2.30-2.38 and 2.90-3.00 (dd and symm. m, respectively; two AB systems of the cyclobutane 4 H, the lower field protons showing long range coupling with two of the cyclopropane protons). Anal. ($C_{10}N_{13}NO_6$ - H_2O) C, H, N.

r-1-Amino-*t*-3-(*trans*-2-carboxycyclopropyl)cyclobutane-1,3-dicarboxylic Acid (23) was similarly prepared, in a similar yield, from 21: mp decomposition at ca. 295 °C (H_2O); NMR (D_2O) 1.17–1.22, 1.25–1.30, 1.72–1.76, and 1.94–2.00 (four m, 4 H, cyclopropane protons), 2.5–2.7 (unsymm. m, 4 H, cyclobutane protons). Anal. ($C_{10}H_{13}NO_6$) C, H.

cis- and trans-1-(Benzoylamino)-3-(hydroxymethyl)cyclobutane-1-N,N-pentamethylenecarboxamide (25 and 29). To a solution of ester 18 (3.44 g, 10 mmol) in dioxanewater (100 mL each) were added 3.8 g (100 mmol) of NaBH₄, and the mixture was stirred for 20 h at room temperature. To the ice-cooled flask was then added carefully 3 M HCl until the solution was slightly acidic. The volume of the solution was reduced by about two-thirds at reduced pressure, whereby the product started to precipitate or was induced to precipitate. It was collected by filtration, washed with water, and air-dried to provide 2.28 g of crude product. Further concentration of the filtrate provided a further crop of solid which was similarly treated. The total solid (2.75 g) was purified by chromatography (60 g of silica gel; CH₂Cl₂-10% MeOH) to furnish 2.2 g of pure cis-25 (70% yield): mp 228-229 °C (EtOAc-MeOH); NMR 1.49 and 1.57 (br, 4 and 2 H), 2.44 (m, 3 H), 2.64 (br, OH), 3.00 (m, 2 H), 3.50 (br, 4 H), 3.68 (br s, 2 H), 7.22 (s, NH), 7.4-7.8 (5 H). Anal. (C₁₈H₂₄N₂O₃) C, H.

Alcohol **29** was similarly prepared from ester **19**, in a similar yield: mp 212–213 °C (EtOAc–MeOH); NMR 1.46 and 1.54 (br, 4 and 2 H), 2.5–2.7 (m, 5 H), 2.75 (OH), 3.47 (br s, 4 H), 3.56 (d, J=5.7, CH_2OH), 7.4-7.8 (NH and aromatic protons, 6 H). Anal. ($C_{18}H_{24}N_2O_3$) C, H.

cis- and trans-1-(Benzoylamino)-3-formylcyclobutane-1-N,N-pentamethylenecarboxamide (26 and 30). To a solution of 0.25 mL of DMSO in CH₂Cl₂ (1.5 mL) cooled to -75 °C was added dropwise via syringe a solution of 0.2 mL of trifluoroacetic anhydride (TFAA) in 1 mL of CH2Cl2. Stirring was continued for 10 min, and then a solution of 25 (200 mg) in CH₂Cl₂ (ca. 20 mL) was added to the reaction flask (the highly insoluble 25 was dissolved with warming in 40 mL of CH₂Cl₂, and the solution was then concentrated to ca. 20 mL). Stirring was continued for 3 h at that temperature and then for 0.5 h out of the cold bath. Triethylamine (0.5 mL) was then added, and stirring was continued for 5 min. Workup involved washing with water, with dilute acid, and with a saturated sodium chloride solution. The water layer was saturated with NaCl and extracted three times with CH₂Cl₂. The combined organic extracts were dried on MgSO₄, filtered, and evaporated to furnish 181 mg of crude aldehyde. Chromatography (20 g of silica gel; CH₂Cl₂-5% MeOH) provided pure 26 (125 mg, 63%): mp 176-177 °C (EtOAc); NMR 1.51, 1.59, and 1.68 (three br s, 6 H), 2.67 (m, 2), 3.15 (m, 3), 3.51 (br s, 4), 6.70 (s, NH), 7.4-7.8 (5 H), 9.85 (s, 1). Anal. $(C_{18}H_{22}N_2O_3)$ C, H.

Aldehyde 30 was similarly prepared from 29, in a similar yield: mp 195-196 °C (EtOAc); NMR 1.60 (br, 6 H), 2.85 and

3.13 (d AB q, $J_{A,B}$ = 13.4, $J_{A,X}$ = 7.3, $J_{B,X}$ = 9.6, 4 H), 3.36 (m, 1 H), 3.52 (br s, 4 H), 6.53 (br s, NH), 7.4–7.8 (5 H). Anal. (C₁₈H₂₂N₂O₃) C, H.

cis- and trans-1-(Benzoylamino)-3-[2-(methoxycarbonyl)ethenyl]cyclobutane-1-N,N-pentamethylenecarboxamide (27 and 31). Aldehyde 26 (110 mg) was warmed in benzene (20 mL) with methyl (triphenylphosphoranylidene)-acetate (160 mg) at 80 °C for 20 h. The solvent was evaporated to dryness, and the residue was triturated twice with ether which dissolved the phosphorus-containing compounds and left behind crude solid 27. Recrystallization from EtOAc-hexane provided pure 27 (100 mg, 77%): mp 218-219 °C; NMR 1.52 and 1.59 (br, 2 and 4 H) 2.49 and 3.17 (two m, 4 H), 2.99 (m, 1 H), 3.53 (br, 4 H), 3.72 (s, Me), 5.80 (d, J=15.6, 1 H) and 7.07 (dd, J=15.6 and 7.2, 1 h), 6.37 (br s, NH), 7.4-7.8 (5 H). Anal. (C₂₁H₂₆N₂O₄) C, H.

Ester 31 was similarly prepared from 30 in a similar yield: mp 232–233 °C (EtOAc); NMR 1.53 and 1.61 (br, 4 and 2 H), 2.70 and 2.83 (two symm. m, 4 H), 3.31 (sextet, 1 H), 3.52 (br s, 4 H), 3.73 (s, Me), 5.79 (d, J=15.6,1 H) and 7.00 (dd, J=15.6 and 7.2, 1 H), 7.4–7.8 (5 H). Anal. ($C_{21}H_{26}N_2O_4$) C, H.

cis- and trans-1-(Benzoylamino)-3-[2-(methoxycarbonyl)ethyl]cyclobutane-1-N,N-pentamethylenecarboxamide (28 and 32). Compound 27 (90 mg) in EtOH (20 mL) was introduced into prereduced PtO₂ catalyst (20 mg in 5 mL EtOH), and the mixture was stirred under hydrogen for 2 h. Filtration of the catalyst and evaporation of the solvent furnished solid 28 in quantitative yield (91 mg): mp 169-170 °C (EtOAc-hexane); NMR 1.51 and 1.58 (br, 4 and 2 H), 1.84 and 2.26 (q and t, 4 H, side-chain methylenes), 2.14 and 3.06 (two m, 3 and 2 H), 3.51 (br, 4 H), 3.66 (s, Me), 6.35 (s, NH), 7.4-7.8 (5 H). Anal. (C₂₁H₂₈N₂O₄) C, H.

Hydrogenation of 31 similarly furnished 32: mp 197–198 °C ($\rm CH_2Cl_2$ -hexane); NMR 1.51 and 1.60 (br, 4 and 2 H), 1.76 and 2.25 (q and t of side-chain methylene protons), 2.45 and 2.60 (two m, 3 and 2 ring protons), 3.51 (br, 4 H), 3.65 (s, Me), 6.54 (s, NH), 7.4–7.8 (5H). Anal. ($\rm C_{21}H_{28}N_2O_4$) C, H.

cis-1-Amino-3-(2-carboxyethyl)cyclobutane-1-carboxylic Acid (2c). Hydrolysis of 28 (75 mg) was carried out as described above for 18 providing crystalline 2c (32 mg, 85%), pure by TLC (cellulose plates; $CH_3CN-H_2O-AcOH-pyridine$, 90:20:5:1.5) and by NMR: mp ca. 260 °C dec; NMR (D₂O) 1.76 and 2.30 (q and t, 4 H, side chain methylenes), 1.97–2.03 and 2.59–2.65 (two symmetrical m, 4 H), 2.47 (sept, 1 H). Anal. (2C₈H₁₃NO₄·H₂O) C, H, N.

trans-1-Amino-3-(2-carboxyethyl)cyclobutane-1-carboxylic Acid (1c) was similarly prepared by hydrolysis of 32: mp darkening from about 240 °C, no melting up to 300 °C; NMR 1.54 and 1.76 (two m, 4 H, side-chain methylenes), 2.43 (d, J=8.2, 4 H), 2.64 (pent, 1 H). Anal. (2C₈H₁₃NO₄·H₂O) C, H, N.

cis- and trans-1-(Benzoylamino)-3-[2-(dimethoxyphosphoryl)acetyl]cyclobutane-1-N,N-pentamethylenecarboxamide (33 and 36). A solution of dimethyl (lithiomethyl)phosphonate was prepared at -78 °C from dimethyl methylphosphonate (3.5 mL, 31 mmol) in THF (160 mL) by stirring for 1 h with BuLi (12.4 mL of a 2.5 M solution in hexane, 31 mmol). Solid ester 18 (3.2 g, 9.4 mmol) was introduced all at once into the reaction flask, and stirring was continued for 40 min at -78 °C and for 1 h in an ice bath. Addition of water, evaporation of the THF, acidification of the water layer, and extraction with CH2Cl2 provided a crude product that was purified by chromatography (65 g of silica gel; $CH_2Cl_2-5\%$ MeOH) to furnish pure cis-33 (3.76 g, 92%): mp 173-174 °C (CH₂Cl₂-hexane); NMR 1.4-1.6 (br, 6 H), 2.63 and 3.22 (two m, 4 H), 3.13 (d, J = 22.6, POC H_2 CO), 3.30 (m, 1 H), 3.4-3.6 (br, 4 H), 3.73 and 3.76 (two s, two Me), 7.37 (s, NH), 7.4-7.8 (5 H). Anal. (C₂₁H₂₉N₂PO₆) C, H.

Ketone **36** was similarly prepared from **19** in a similar yield: mp 196-197 °C (CHCl₃-hexane); NMR 1.52 and 1.59 (br, 4 and 2 H), 2.77 and 3.05 (two m, 4 H), 3.09 (d, J=22.5, 2 H), 3.49 (br, 4 H), 3.55 (pent, 1 H), 3.74 and 3.77 (two s, two Me), 7.08 (s, NH), 7.4-4.8 (5 H). Anal. ($C_{21}H_{29}N_2PO_6$) C, H.

cis- and trans-1-(Benzoylamino)-3-[2-(dimethoxyphosphoryl)-1-hydroxyethyl]cyclobutane-1-N,N-pentamethylenecarboxamide (34 and 37). Ketone 33 (1090 mg, 2.5

mmol) was dissolved with warming in ethanol (20 mL). The solution was cooled back to room temperature, and solid NaBH₄ (70 mg, 1.84 mmol) was added. After 0.25 h at room temperature, AcOH was added dropwise until gas evolution stopped. The solvent was evaporated to dryness under reduced pressure, and the residue was taken in CH₂Cl₂, filtered on Celite, and chromatographed (30 g of silica gel; CH₂Cl₂-7% MeOH) to provide pure cis-34 (1020 mg, 93%): mp 169-170 °C (EtOAc); NMR 1.50 and 1.56 (br, 4 and 2 H), 1.74-1.86 (m, 2 H, POCH₂), 2.25-2.42 and 2.99-3.09 (two m, 3 and 2 H), 3.50 (br. 4 H), 3.74 and 3.77 (two d, J = 5, two Me), 4.09(m, 1 H CHOH), 4.20 (s, OH), 7.32 (s, NH), 7.4-7.8 (5 H). Anal. $(C_{21}H_{31}N_2PO_6)$ C, H.

Alcohol 37 was similarly prepared from 36 in a similar yield: mp 191-192 °C (EtOAc-MeOH); NMR 1.49 and 1.57 (br, 4 and 2 H), 1.75-1.95 (m, 2 H, POCH₂), 2.45-2.79 (m, 5 H), 3.48 (br, 4 H), 3.72 and 3.75 (two d, J = 7.7, two Me), 3.93(m, 1 H, CHOH), 7.4-7.8 (5 H). Anal. (C₂₁H₃₁N₂PO₆) C, H.

cis-1-Amino-3-(2-phosphono-1-hydroxyethyl)cyclobutane-1-carboxylic Acid (35). Hydrolysis of 34 (160 mg; 6 M HCl, 130 °C, 24 h) and workup with propylene oxide gave after filtration an amorphous, sticky solid. After being induced to crystallize in some water, ethanol was added to complete precipitation. Recrystallization from water-ethanol provided pure 35 (66 mg, 61%): mp 236-237 °C (decomposition); NMR (D_2O) 1.69–1.85 (m, 2 H), 2.20–2.34 and 2.64–2.72 (two m, 2 and 3 H), 3.95 (br, 1 H, CHOH). Anal. (C₇H₁₄NPO₆) C, H, N.

trans-1-Amino-3-(2-phosphono-1-hydroxyethyl)cvclobutane-1-carboxylic Acid (38) and 1-Amino-3-(2-phosphonoethylidene)cyclobutane-1-carboxylic Acid (39). Similar hydrolysis of 37 provided 38, accompanied by 39. Partial separation of 38 from 39 was achieved by crystallization from water, whereby 39 precipitated free of 38. Acid 38 was further purified by chromatography, as described above for 35, and by crystallization from water-ethanol: mp, decomposition above 200 °C; NMR (D₂O) 1.68-1.86 (m, 2 H), 2.35–2.4 and 2.6–2.8 (two m, 2 and 3 H), 4.03 (br, 1 H, CHOH). Anal. ($C_7H_{14}NPO_6$) C, H, N.

Acid 39 was obtained pure by a second crystallization from water: mp, rapid decomposition above 250 °C; NMR (D₂O) 1.95-2.06 and 2.17-2.29 (two symm. m, 2 H, PCH2), 2.33, 2.61, 2.71, and 3.01 (four m, 4 H), 5.05 (m, 1H, CH₂CH=C). Anal. (C₇H₁₂NPO₅) C, H, N.

cis- and trans-1-Amino-3-(2-phosphonoacetyl)cyclobutane-1-carboxylic Acid (40 and 41). Hydrolysis of either 33 or 36 (6 M HCl, 130 °C, 24 h; work up with propylene oxide) gave a similar mixture of 40 and 41 in a ratio of ca. 3:2, respectively, and in a total 75-80% yield. By crystallization of the mixture from water, a varying amount pure isomer 39 could be obtained, as evidenced by 400 MHz ¹H NMR (TLC did not distinguish between the isomers). Further separation of the isomers was achieved by chromatography of the mixture on Dowex 1 ion-exchange resin (acetate form; elution with 1 M AcOH) and fractional crystallization from water of the solid obtained from the first half of product-containing fractions. This provided again isomer 40 free of 41. Mixtures rich in 41, which are obtained from the second half of productcontaining fractions, can be re-equilibrated to the initial 3:2 composition of the mixture by warming in 3 M HCl at 130 $^{\circ}\mathrm{C}$ for 20 h. Pure isomer 41 could be obtained from the very last chromatography fractions by crystallization from water.

Acid 40: mp 230-231 °C (decomposition; H₂O); NMR (D₂O), measured ca. 1 h after dissolution) 2.60-2.66 and 2.81-2.87 (two symm. m, 4 H), 3.05 and 3.07 (two d of unequal intensity, $J_{\rm H.P}=21.5$, unequal distribution of the partly deuteriumexchanged P-CH2-CO protons; the two protons are totally exchanged with deuterium after 24 h), 3.76 (pent, 1 H). Anal. $(C_7H_{12}NPO_6)$ C, H, N.

Acid 41: mp, decomposition above 175 °C (H₂O); NMR (D₂O) 2.60-2.66 asnd 3.00-3.06 (two m, 4 H; two d due to PCH₂-CO, and partly superimposed on the latter m, disappear slowly by exchange with deuterium), 3.80 (m, 1 H). Anal. (C7H12-NPO₆) C, H, N.

The ratio of the two isomers in their mixtures can be determined by the integration ratio of the 2.81-2.87 multiplet of 40 to the 2.60-2.66 multiplet common to 40 and 41.

Acid 40 (180 mg) was hydrogenated in water (30 mL) and acetic acid (10 mL) over prereduced PtO2 hydrogenation catalyst (33 mg). The mixture was stirred under hydrogen for 20 h and then filtered and evaporated to provide acid 35 (175 mg), pure by NMR.

cis- and trans-1-(Benzovlamino)-3-[2-(dimethoxyphosphoryl)-1-[(methylsulfonyl)oxy]ethyl]cyclobutane-1-N,Npentamethylenecarboxamide (42 and 45). Alcohol 34 (400 mg) was mesylated in CH₂Cl₂ by treatment with CH₃SO₂Cl (0.33 mL) in the presence of Et₃N (0.6 mL) for 20 h. The crude product was chromatographed (20 g of silica gel; CH₂Cl₂-5% MeOH) to yield pure 42 (343 mg, 70%): mp 197-198 °C (CH₂-Cl₂-MeOH); NMR 1.98-2.05 and 2.15-2.22 (two ddd, 2 H, ABq of POCH₂, further split by CHOMs and by P), 2.50-2.56 and 3.01-3.06 (two m, 4 H), 2.91 (m, 1 H), 3.13 (s, SO₂CH₃), 3.74 and 3.77 (two d, 2 Me), 5.05 (m, 1 H, CH OMs), 7.36 (s, NH), 7.4-7.9 (5 H). Anal. (C₂₂H₃₃N₂PO₈S) C, H.

Mesylate 45 was similarly prepared from alcohol 37: mp 160-161 °C (CH₂Cl₂-hexane); NMR 1.5 and 1.7 (br, 4 and 2 H), 2.1-2.3 (m, 2 H, PC H_2), 2.56-2.69, 2.76-2.85, and 3.00-3.06 (three m, 2, 2, and 1 H), 3.15 (s, OSO₂CH₃), 3.50 (br, 4 H), 3.74 and 3.77 (two d, two Me), 4.93 (m, 1 H, CHOMs), 6.85 (br, NH), 7.4-7.9 (5 H). Anal. (C₂₂H₃₃N₂PO₈S) C, H.

cis-1-(Benzoylamino)-3-[2-(dimethoxyphosphoryl)ethenyl]cyclobutane-1-N,N-pentamethylenecarboxamide (43) and cis-1-(Benzoylamino)-3-[2-(dimethoxyphosphoryl)ethyl]cyclobutane-1-N,N-pentamethylenecarboxamide (44). Mesylate 42 (320 mg) was treated in THF (20 mL), at -78 °C, with BuLi (2.2 equiv) for 3 h. After quenching with aqueous NH₄Cl and extractive workup with EtOAc, the crude product was chromatographed (15 g of silica gel; CH₂Cl₂-5% MeOH) to yield 43 (230 mg, 93%), which was used in the following hydrogenation step. A sample was recrystallized from ethyl acetate to give pure 43: mp 183-184 °C; NMR 2.42-2.48 and 3.06-3.11 (two m, 4 H), 3.32 (m, 1 H), 3.66 and 3.69 (two Me), 5.62 and 6.76 (t and m, 2 H; olefinic protons), $7.4 - 7.9 \; (5 \; H), \; 8.37 \; (N \textit{H}). \; \; \; Anal. \; \; (C_{21} H_{29} N_2 PO_5) \; C, \; H.$

Product 43 (200 mg) was hydrogenated in ethanol (10 mL) over prereduced PtO₂ (40 mg) for 3 h. Filtration of the catalyst and evaporation of the solvent furnished oily 44 (160 mg) which showed no unsaturation (NMR) and which was used in the following hydrolysis step (see below).

trans-1-(Benzoylamino)-3-[2-(dimethoxyphosphoryl)ethenyl]cyclobutane-1-N,N-pentamethylenecarboxamide (46) and trans-1-(benzoylamino)-3-[2-(dimethoxyphosphoryl)ethyl]cyclobutane-1-N,N-pentamethylenecarboxamide (47) were similarly prepared from 45, in similar yields. Phosphonate 46: mp 165-166 °C; NMR 1.5 and 1.6 (br, 6 H), 2.71-2.74 and 2.79-2.82 (two symm. m, 4 H), 3.32 (m, 1 H), 3.52 (br s, 4 H), 3.70 and 3.73 (two s, two Me), 5.61 and 6.85 (dd and ddd, $J_{AB} = 17.1$, $^2J_{PH} = 20.2$, 2 H), 6.66 (s, NH), 7.4-7.8 (5 H). Anal. (C₂₁H₂₉N₂PO₅) C, H.

Hydrogenation of 46, as described for 43, furnished again an oily product which was used as such for hydrolysis to 3b.

cis-1-Amino-3-(2-phosphonoethyl)cyclobutane-1-carboxylic Acid (4b). Hydrolysis of 44 (140 mg; 6 M HCl, 130° C, 24 h) and workup with propylene oxide gave a sticky solid which was dissolved in water and purified by chromatography (20 cm column of Dowex 1, acetate form; elution with 1 M AcOH) to provide 4b (70 mg, 75%): mp decomposition from ca. 250 °C (H₂O); NMR (D₂O) 1.49-1.58 and 1.66-1.74 (two m, 4 H, side chain protons), 2.04-2.10 and 2.69-2.74 (two m, 4 H), 2.55 (pent, 1 H). Anal. (C₇H₁₄NPO₅) C, H, N.

trans-1-Amino-3-(2-phosphonoethyl)cyclobutane-1-carboxylic acid (3b) was similarly obtained from 47: mp, no melting below 300 °C, decomposition (H₂O); NMR (D₂O) 1.46-1.55 and 1.69-1.77 (two m, 4 H, side chain protons), 2.39 (d, J = 8.5, 4 H), 2.61 (m, 1 H). Anal. (2C₇H₁₄NPO₅·H₂O) C, H,

References

(1) Monaghan, D. T.; Bridges, R. J.; Cotman, C. W. The excitatory amino acid receptors: their classes, pharmacology and distinct properties in the function of the central nervous system. Annu. Rev. Pharmacol. Toxicol. 1989, 29, 365-402.

- (2) Lodge, D., Collingridge, G. L., Eds. The Pharmacology of Excitatory Amino Acids. A Special Report. Trends Pharmacol.
- Watkins, J. C. Evans, R. H. Excitatory amino acid transmitters. Annu. Rev. Pharmacol. Toxicol. 1981, 21, 165-204
- (4) Herrling, P. L. Clinical implications of NMDA receptors. In The NMDA Receptor, 2nd ed.; Watkins, J. C., Collingridge, G. L., Eds.; Oxford University Press: Oxford, 1994, in press.
- (5) Meldrum, B. A.; Chapman, A. G. Competitive NMDA antagonists as drugs. In The NMDA Receptor, 2nd ed.; Watkins, J. C. Collingridge, G. L., Eds.; Oxford University Press: Oxford, 1994,
- (6) Iversen, L. L.; Kemp, J. A. Non-competitive NMDA antagonists as drugs. In *The NMDA Receptor*, 2nd ed.; Watkins, J. C., Collingridge, G. L., Eds.; Oxford University Press: Oxford, 1994,
- Watkins, J. C., Collingridge, G. L., Eds. The NMDA Receptor,
- 2nd ed.; Oxford University Press: Oxford, 1994, in press. Lanthorn, T. H.; Hood, W. F.; Watson, G. B.; Compton, R. P.; Rader, R. K.; Gaoni, Y.; Monahan, B. cis-2,4-Methanoglutamate is a potent and selective N—methyl-D-aspartate receptor agonist. Eur. J. Pharmacol. 1990, 182, 397–404.
- (9) Allan, R. D.; Hanrahan, J. R.; Hambley, T. W.; Johnston, G. A. R.; Mewett, K. N.; Mitrovic, A. D. Synthesis and activity of a potent N-methyl-D-aspartic acid agonist, trans-1-aminocyclobutane-1,3-dicarboxylic acid, and related phosphonic and carboxylic
- acids. J. Med. Chem. 1990, 33, 2905-2915. (10) Jane, D. E.; Olverman, H. J.; Watkins, J. C. Agonists and competitive antagonists: structure-activity and molecular modelling studies. In The NMDA Receptor, 2nd ed.; Watkins, J. C. Collingridge, G. L., Eds.; Oxford University Press: Oxford, 1994, in press.
- (11) Ornstein, P. L.; Klimkowski, V. J. Competitive NMDA receptor
- (11) Ornstein, P. L.; Klimkowski, V. J. Competitive NMDA receptor antagonists. In Excitatory amino acid receptors: design of agonists and antagonists; Krogsgaard-Larsen, P., Hansen, J. J., Eds.; Ellis Horwood Ltd.: Chichester, UK, 1992; pp 183-201.
 (12) Müller, W.; Lowe, D. A.; Nejt, H.; Urwyler, S.; Herrling, P. L.; Blaser, D.; Seebach, D. Synthesis and N-methyl-D-aspartate (NMDA) antagonist properties of the enantiomers of α-amino-5-(phosphonomethyl)[1,1'-biphenyl]-3-propanoic acid. Use of a new chiral glycine derivative. Helv. Chim. Acta 1992, 75, 855-864.
 (13) Olverman, H. J.; Jones, A. W.; Mewett, K. N.; Watkins, J. C. Structure/activity relations of NMDA receptor ligands as studied by their inhibition of 3H-D-AP5 hinding in rat brain membranes.
- by their inhibition of ³H-D-AP5 binding in rat brain membranes.
- Neuroscience 1988, 26, 17-31.

 (14) Honoré, T.; Drejer, J.; Nielsen, M.; Watkins, J. C.; Olverman, H. J. Molecular target size of NMDA-antagonist binding sites. Eur. J. Pharmacol. 1987, 136, 137-138.
- (15) Gaoni, Y.; Tomažič, A. Bridgehead reactivity, nucleophilic and radical additions, and lithium aluminum hydride reduction of 1-(arylsulfonyl)bicyclobutanes: general access to substituted, functionalized cyclobutanes. Synthesis of (±)-citrilol acetate, (±)-junionone, and the tricyclo[3.3.0.0^{1.4}]octane and tricyclo- $[4.3.0.0^{1.7}]$ nonane ring systems. J. Org. Chem. 1985, 50, 2948-
- (16) Gaoni, Y. New bridgehead-substituted 1-(arylsulfonyl)bicyclo-[1.1.0] butanes and some novel addition reactions of the bicyclic system. Tetrahedron 1989, 45, 2819–2840. (17) Gaoni, Y.; Tomažič, A.; Potgieter, E. Stereochemistry of addition
- of organocopper reagents and of the hydride ion to 1-(arylsulfonyl)bicyclo[1.1.0]butanes. J. Org. Chem. 1985, 50, 2943-2947.
- Gaoni, Y. Regiospecific additions of hydrazoic acid and benzylamine to 1-(arylsulfonyl)bicyclo[1.1.0]butanes. Application to the synthesis of cis and trans 2,4-methanoglutamic acids. Tetrahedron Lett. 1988, 29, 1591-1594.

- (19) Gaoni, Y. J. Org. Chem., in press.
- (20) Bell, E. A.; Qureshi, M. Y.; Pryce, R. J.; Janzen, D. H.; Lemke, P.; Clardy, J. 2,4-Methanoproline (2-carboxy-2,4-methanopyrrolidine) and 2,4-methanoglutamic acid (1-amino-1,3-dicarboxycyclobutane) in seeds of ateleia herbert smithii Pittier (leguminosae). J. Am. Chem. Soc. 1980, 102, 1409-1412.
- (21) Yamaguchi, M.; Torisu, K.; Minami, T. threo-Selective Michael Addition of N,N-dibenzylglycinate and alaninate enolates to α,β unsaturated esters. A concise and stereoselective synthesis of (±)-CCG-II. Chem. Lett. 1990, 377-380.
- (22) Zindel, J.; de Meijere, A. A short and efficient diastereoselective synthesis of 2'-substituted 2-cyclopropylglycines. Synthesis 1994, 190-194.
- (23) Austin, G. N.; Baird, P. D.; Chow, H. F.; Fellows, L. E.; Fleet, G. W. J.; Nash, R. J.; Peach, J. M.; Pryce, R. J.; Stirton, C. H. Isolation from atelia herbert smithii Pittier (sophoreae, leguminosae) and X-ray structure of cis-1-amino-(hydroxymethyl)cyclobutane-1-carboxylic acid, an achiral non-protein amino acid. Tetrahedron 1987, 43, 1857-1861.
- (24) Fleet, G. W. J.; Seijas, J. A.; Vazquez Tato, M. P. Synthesis of cis- and trans-1-amino-3-(hydroxymethyl)cyclobutane-1-carboxylic acids. Tetrahedron 1988, 44, 2077-2080.
- (25) Fagg, G. E.; Olpé, H. R.; Pozza, M. F.; Baud, J.; Steinmann, M.; Schmutz, M.; Portet, C.; Baumann, P.; Thedinga, K.; Bittiger, H.; Allgeier, H.; Heckendorn, R.; Angst, C.; Brundish, D.; Dingwall, J. G. CGP 37849 and CGP 39551: novel and competitive N-methyl-D-aspartate receptor antagonists with oral activ-
- ity. Br. J. Pharmacol. 1990, 99, 791-797.
 (26) Hutchison, A. J.; Williams, M.; Angst, C.; de Jesus, R.; Blanchard, L.; Jackson, R. H.; Wilusz, E. J.; Murphy, D. E.; Bernard, P. S.; Schneider, J.; Campbell, T.; Guida, W.; Sills, M. A. 4-(Phosphonoalkyl)- and 4-(phosphonoalkenyl)-2-piperidinecarboxylic acids: synthesis, activity at N-methyl-D-aspartic acid receptors, and anticonvulsant activity. J. Med. Chem. 1989, 32, 2171-8.
- (27) Ortwine, D. F.; Malone, T. C.; Bigge, C. F.; Drummond, J. T.; Humblet, C.; Johnson, G.; Pinter, G. W. Generation of N-methyl-D-aspartate agonist and competitive antagonist pharmacophore models. Design and synthesis of phosphonoalkyl-substituted tetrahydroisoquinolines as novel antagonists. J. Med. Chem. 1**992**, *35*, 1345-1370.
- (28) McBain, C. J.; Mayer, M. L. NMDA receptor structure and function. Physiol. Rev. In press.
- Evans, R. H.; Francis, A. A.; Jones, A. W.; Smith, D. A. S.; Watkins, J. C. The effects of a series of ω -phosphonic α -carboxylic amino acids on electrically evoked and amino acid induced responses in isolated spinal cord preparations. Br. J. Pharmacol. **1982**, 75, 65-75.
- (30) Jones, A. W.; Smith, D. A. S.; Watkins, J. C. Structure-activity relations of dipeptide antagonists of excitatory amino acids. Neuroscience 1984, 13, 573-581.
- (31) Patel, S.; Chapman, A. G.; Graham, J. L.; Meldrum, B. S.; Frey, P. Anticonvulsant activity of the NMDA antagonists, D(-)4-(3phosphonopropyl)piperazine-2-carboxylic acid (D-CPP) and D(-(D-4-(3-phosphonoprop-2-enyl)piperazine-2-carboxylic acid (D-CPP-ene) in a rodent and primate model of reflex epilepsy. Epilepsy Res. 1990, 7, 3-10.
- (32) Litchfield, J. T.; Wilcoxon, F. A simplified method of evaluating dose-effect experiments. J. Pharmacol. Exp. Ther. 1949, 96, 99-
- (33) INSIGHT II Version 2.2.0 can be obtained from Biosym Technologies, 9685 Scranton Road, San Diego, CA 92121-2777.