ORIGINAL ARTICLES

College of Pharmacy, Chosun University, Seosuk-dong, Dong-gu, Gwangju, South Korea

Screening of new chemopreventive compounds from Digitalis purpurea

J. Y. LEE, E. WOO, K. W. KANG

Received April 27, 2005, accepted June 7, 2005

Keon Wook Kang, Ph.D.; College of Pharmacy, Chosun University, 375 Seosuk-dong, Dong-gu, Gwangju 501-759, South Korea kwkang@chosun.ac.kr

Pharmazie 61: 356-358 (2006)

Chemopreventive agents induce a battery of genes whose protein products can protect cells from chemical-induced carcinogenesis. In this study, we isolated four different glycosides (1 acteoside; 2 purpureaside A; 3 calceolarioside B; and 4, plantainoside D) from the leaves of *Digitalis purpurea* and studied their abilities to induce glutathione S-transferase (GST) and their protective efficiencies against aflatoxin B_1 -induced cytotoxicity in H4IIE cells. Of these four glycosides, acteoside significantly inhibited the cytotoxicity induced by aflatoxin B_1 (AFB₁) and also selectively increased GST α protein levels. Reporter gene analysis using an antioxidant response element (ARE) containing construct and subcellular fractionation assays, revealed that GST α induction by acteoside might be associated with Nrf2/ARE activation. The results suggest that acteoside possesses a potent hepatoprotective effect against AFB₁ and that it can be applied as a potential chemopreventive agent.

1. Introduction

Cancer chemoprevention is defined as the use of naturally occurring compounds to prevent, or reverse the process of carcinogenesis. Moreover, a large body of experimental data indicates the feasibility of this approach. A large number of phytochemicals can act as chemopreventive agents. These include diallyl sulfide from garlic and dithiolthiones from cruciferous plants, which protect tissues and prevent carcinogenesis (Wargovich et al. 1988;

356 Pharmazie **61** (2006) 4

ORIGINAL ARTICLES

Wilkinson et al. 1997). These sulfur-containing compounds have been extensively studied as potential chemopreventive agents (Wargovich et al. 1988; Kensler et al. 2004). The one known mode of chemoprevention by phytochemicals involves the induction of phase II detoxifying enzymes such as glutathione S-transferase (GST) (Kensler et al. 1986), and treatment with many chemopreventive agents increases hepatic GST activity by acting through the induction of the gene (Primiano et al. 1995; Guyonnet et al. 1999).

Digitalis purpurea is a perennial herb of the Scrophulariaceae family, and is commonly used to treat congestive heart failure (Willius and Keys 1941). Recently, it was also reported that Digitalis glycosides have potent anticancer effects (Lopez-Lazaro et al. 2003). As a part of our program to screen for potential cancer chemopreventive compounds from medicinal plants, we isolated four different glycosides (1 acteoside; 2 purpureaside A; 3 calceolarioside B; and 4, plantainoside D) from the leaves of Digitalis purpurea and studied their abilities to induce GST and their protective efficacies against aflatoxin B₁ (AFB₁)-induced cytotoxicity in a rat hepatocyte-derived cell line, H4IIE cells. Furthermore, we also monitored Nrf2/ARE activation in order to investigate the mechanistic basis underlying the induction of GST by the glycosides.

2. Investigations, results and discussion

We isolated four different glycosides from the leaves of *Digitalis purpurea*. First, the effect of each glycoside against AFB₁-induced cytotoxicity was examined in H4IIE cells. Pretreating the cells with acteoside (100 µM, 12 h) significantly inhibited the cell death induced by AFB₁, whereas the other glycosides did not protect cells from AFB₁ (Fig. 1). Earlier studies suggested that the AFB₁-induced formation of ROS or oxidant metabolites plays a critical role in its cytotoxic effect (Shen et al. 1995). This result suggests that the protective effect of acteoside could be due to a reduction in the formation of ROS. It has also been reported that acteoside protects CCl₄-induced hepatotoxicity possibly through the blocking of P450 or the scavenging of free radicals (Lee et al. 2004).

Glutathione (GSH), a nonprotein sulfhydryl intracellular molecule, plays a role as a protective substance and serves as an effective oxygen radical scavenger. GSH-conjugation

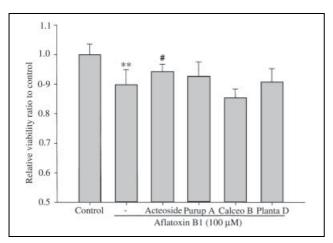


Fig. 1: Effects of the four glycosides on AFB₁-induced cytotoxicity. Cells were serum-starved and incubated in the presence or absence of each glycoside (100 $\mu M)$ for 12 h, and then treated with various concentrations of AFB₁ for 12 h. Cells viabilities were assessed by MTT assay. Data represent the means \pm S.D. of 8 different samples

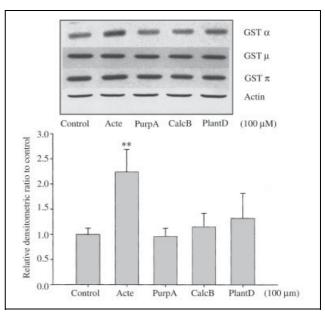


Fig. 2. Effects of the four glycosides isolated from the leaves of *Digitalis purpurea* on GST protein levels. GST subunit protein was monitored 18h after treating cells with each compound (100 μ M). Each lane was loaded with 10 μ g of cytosolic protein. Data represent the means \pm S.D. of 3 separate experiments (significant as compared to control, **p < 0.01; control level = 1)

with xenobiotics can be accelerated by GST, and increased GST activity can increase this conjugative reaction and thus lead to AFB_1 detoxification. Hence, the induction of GST is believed to be an important determinant of the characteristics of chemopreventive agents. In the present study, we determined the levels of GST subtypes in H4IIE cells treated with the four digitalis glycosides. Treatment of H4IIE cells with acteoside (100 μ M) for 18 h increased GST α levels, but the other glycosides caused no such increase (Fig. 2), which suggests that the selective induction of GST α by acteoside seems to be related with its cytoprotective effect against AFB₁. In addition, either GST μ or GST π was not induced by the glycosides (Fig. 2).

The role of antioxidant response elements (AREs) in the inducible expression of phase II enzymes (e.g. GSTa) by several antioxidants and chemopreventive agents has been extensively studied (Kang et al. 2001, 2003). Nrf2 is a key transcription factor that binds to ARE sequences, and which is implicated in the regulation of GST expression (Kang et al. 2001, 2002). To determine whether the induction of GST α by acteoside is mediated by the activation of Nrf2/ARE, a reporter gene assay was performed using H4IIE cells transfected with the mammalian expression vector pGL-797, which contained the luciferase structural gene and the ARE sequence of GSTA2 promoter (Kang et al. 2003). Exposure of cells transiently transfected with this plasmid, to acteoside resulted in a 6-fold increase in luciferase activity. However, luciferase activity was not noticeably induced by the other 3 glycosides (Fig. 3).

Next, we examined whether acteoside stimulates the translocation of Nrf2 to the nucleus. Subcellular fractionation and Western blot analyses confirmed that acteoside increased the level of Nrf2 in the nuclear fraction at $6-12\,h$ (Fig. 4). These data suggest that the induction of GST α by acteoside is associated with the activation of Nrf2-mediated ARE activation.

In summary, the present study demonstrates the protective effect of acteoside on AFB₁-induced cytotoxicity. The in-

Pharmazie **61** (2006) 4

ORIGINAL ARTICLES

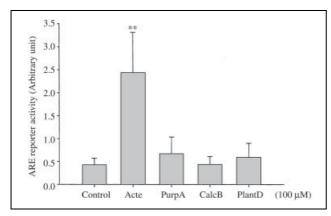


Fig. 3: Induction of luciferase activity by glycosides in H4IIE cells transiently transfected with the GSTA2 chimeric gene construct pGL-797 containing the ARE element. Dual luciferase reporter assays were performed on lysed H4IIE cells co-transfected with pGL-797 (firefly luciferase) and pRL-SV (Renilla luciferase) (in a ratio of 100:1) after exposure to each glycoside (100 μ M) for 18 h. Activation of the reporter gene was calculated as a relative change to Renilla luciferase activity. Data represent the means \pm S.D. of 4 separate experiments (significant as compared to control, $^{**}p < 0.01$; control level = 1)

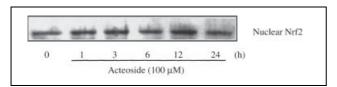


Fig. 4: Nuclear translocation of Nrf2 by acteoside. The subcellular localization of Nrf2 was immunochemically assessed in cells treated with acteoside (100 μM) for 1–24 h. All lanes contained 20 μg of nuclear extracts. Equal loading of protein in each sample was verified using Ponceau-S staining

duction of $GST\alpha$ by acteoside could contribute to its inhibitory effect on AFB_1 -induced hepatocyte damage, presumably by enhancing the GSH conjugation reaction or by enhancing ROS scavenging. Our results suggest that acteoside possesses hepatoprotective effect against AFB_1 , and raise the issue of its usefulness as a chemopreventive agent.

3. Experimental

H4IIE cells were obtained from the American Type Culture Collection (ATCC, Rockville, MD) and were maintained in Dulbecco's modified Eagle's medium containing 10% fetal calf serum, 50 units/ml penicillin, and 50 $\mu g/ml$ streptomycin at 37 °C in a humidified 5% CO $_2$ atmosphere. Cell viability was measured by MTT assay. Cytosolic and nuclear fraction isolation, immunoblot analysis, and reporter gene assays were performed as previously described (Kang et al. 2003). The paired Student's t-test was used to assess significant differences between the different treatment groups. The criterion for statistical significance was set at either p<0.05 or p<0.01.

References

Guyonnet D, Siess MH, Le Bon AM, Suschetet M (1999) Modulation of phase II enzymes by organosulfur compounds from allium vegetables in rat tissues. Toxicol Appl Pharmacol 154: 50–58.

Kang KW, Cho IJ, Lee CH, Kim SG (2003) Essential role of phosphatidylinositol 3-kinase-dependent CCAAT/enhancer binding protein beta activation in the induction of glutathione S-transferase by oltipraz. J Natl Cancer Inst 95: 53–66.

Kang KW, Cho MK, Lee CH, Kim SG (2001) Activation of phosphatidylinositol 3-kinase and Akt by tert-butylhydroquinone is responsible for antioxidant response element-mediated rGSTA2 induction in H4IIE cells. Mol Pharmacol 59:1147–1156.

Kang KW, Lee SJ, Park JW, Kim SG (2002) Phosphatidylinositol 3-kinase regulates nuclear translocation of NF-E2-related factor 2 through actin rearrangement in response to oxidative stress. Mol Pharmacol 62: 1001– 1010

Kensler TW, Egner PA, Davidson NE, Roebuck BD, Pikul A, Groopman JD (1986) Modulation of aflatoxin metabolism, aflatoxin-N7-guanine formation, and hepatic tumorigenesis in rats fed ethoxyquin: role of induction of glutathione S-transferases. Cancer Res 46: 3924–3931.

Kensler TW, Egner PA, Wang JB, Zhu YR, Zhang BC, Lu PX, Chen JG, Qian GS, Kuang SY, Jackson PE, Gange SJ, Jacobson LP, Munoz A, Groopman JD (2004) Chemoprevention of hepatocellular carcinoma in aflatoxin endemic areas. Gastroenterology 127: S310–S318.

Lee KJ, Woo ER, Choi CY, Shin DW, Lee DG, You HJ, Jeong HG (2004) Protective effect of acteoside on carbon tetrachloride-induced hepatotoxicity. Life Sci 74: 1051–1064.

Lopez-Lazaro M, Palma De La Pena N, Pastor N, Martin-Cordero C, Navarro E, Cortes F, Ayuso MJ, Toro MV (2003) Anti-tumour activity of *Digitalis purpurea* L. subsp. *heywoodii*. Planta Med 69: 701–704.
Primiano T, Egner PA, Sutter TR, Kelloff GJ, Roebuck BD, Kensler TW

Primiano T, Egner PA, Sutter TR, Kelloff GJ, Roebuck BD, Kensler TW (1995) Intermittent dosing with oltipraz: relationship between chemoprevention of aflatoxin-induced tumorigenesis and induction of glutathione S-transferases. Cancer Res 55: 4319–4324.

Shen HM, Ong CN, Shi CY (1995) Involvement of reactive oxygen species in aflatoxin B1-induced cell injury in cultured rat hepatocytes. Toxicology 99: 115–123.

Wargovich MJ, Woods C, Eng VW, Stephens LC, Gray K (1988) Chemoprevention of N-nitrosomethylbenzylamine-induced esophageal cancer in rats by the naturally occurring thioether, diallyl sulfide. Cancer Res 48: 6872–6875.

Wilkinson J, Clapper ML (1997) Detoxication enzymes and chemoprevention. Proc Soc Exp Biol Med 216: 192–200.

Willius FA, Keys TE (1941) Classics of Cardiology. New York, NY: Dover Publications Inc. 1: 231–252.

Pharmazie **61** (2006) 4