



## FAGAROPSINE, A DEGRADED LIMONOID GLUCOSIDE FROM *FAGAROPSIS GLABRA*

JOËL BOUSTIE,\* MARIE-JOSÉ RESPAUD, CLAUDE MOULIS, CATHERINE LAVAUD,† JACQUELINE GLEYE and ISABELLE FOURASTÉ

Laboratoire de Pharmacognosie, Faculté de Pharmacie, Université de Toulouse III, 35 Chemin des Maréchaux-31062 Toulouse, Cedex, France; \*Laboratoire de Pharmacognosie et de Mycologie, Faculté de Pharmacie, Université de Rennes I, 2 Av. du Pr. Léon Bernard-35043 Rennes, Cedex, France; †Laboratoire de Pharmacognosie, Faculté de Pharmacie, Université de Reims, 51 Rue Cognacq-Jay-51096 Reims, Cedex, France

(Received in revised form 1 July 1994)

**Key Word Index**—*Fagaropsis glabra*; Rutaceae; limonoid; degraded limonoid glycoside; fagaropsine.

**Abstract**—Phytochemical studies of the alcoholic-soluble portion of *Fagaropsis glabra* have resulted in the isolation of fagaropsine, a degraded limonoid glycoside. Its absolute structure was elucidated as 1-*O*- $\beta$ -D-glucopyranosyl-4 $\alpha$ -(3'-furanyl)-7 $\beta$ -hydroxy-4 $\alpha$ ,8 $\alpha$ -dimethyl-4a,5,6,7,8-hexahydro-3-benzopyran-2-one on the basis of spectral data.

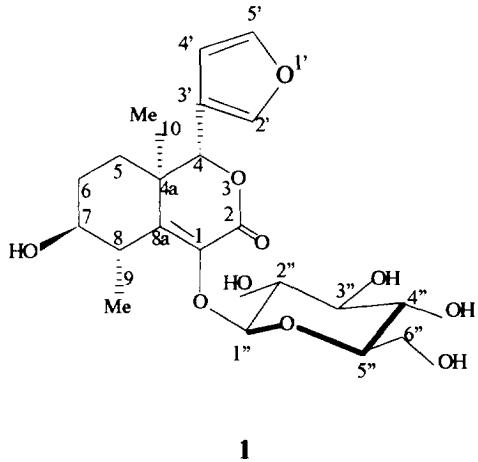
### INTRODUCTION

Previous investigations of *Fagaropsis* species related the presence in this genus of benzophenanthridine alkaloids [1, 2] and limonoids of the limonidic tetranortriterpenoid [1, 3] and degraded limonoid class [3–5]. The various biological activities of these limonoids are of agricultural and medicinal interest [6]. In the field of antifeedants research, structure–activity relationships [7] led chemists to focus on the C/D ring of these compounds [8, 9] which are naturally encountered as degraded limonoids. In addition to previous degraded limonoids found in *F. glabra* [3, 5], we report here the isolation and identification of the first glucosylated degraded limonoid glucoside (**1**) called fagaropsine, which differs in the D-ring glycosylation pattern from other reported  $\beta$ -D-limonoid glycosides [10].

### RESULTS AND DISCUSSION

As the electronic impact mass spectrum of **1** did not afford any positive result, the molecular weight was determined by FAB mass spectrometry. The positive ion FAB mass spectrum showed the pseudomolecular ion peak  $[M + H]^+$  at  $m/z$  441 corresponding to  $C_{21}H_{28}O_{10}$  and the base peak at  $m/z$  279 indicated the presence of a hexose  $[(M + H) - (180 + H_2O)]^+$ . The IR spectrum of **1** exhibited a carbonyl absorption at  $1712\text{ cm}^{-1}$  suggesting a pyrano-type aglycone with a conjugated  $\delta$ -lactonic ring.

These results were corroborated by an extensive analysis of the NMR data (Tables 1 and 2). The comparison with signals observed for degraded limonoids we previously isolated [3, 5], in addition to signals corresponding to a sugar moiety, indicated a glucosylated compound of the pyroangolenside type [8, 11]. The coupling con-



**1**

stant  $J = 6.5\text{ Hz}$  of the anomeric proton resonating at  $\delta 4.78$  and the  $^{13}\text{C}$  and  $^1\text{H}$  NMR signals were consistent with a  $\beta$ -D-glucose substitution. The molecular mass indicated a further oxygen substituent. Homodecoupling experiments and a 2D COSY-45 homonuclear spectrum clarified most of the ambiguous  $^1\text{H}$  coupling systems and confirmed the presence of a H-7 carbonylic signal located at  $\delta 3.86$  and partially overlapping with H-6A of the glucose. The broad singlet shape of H-7 suggested its equatorial position which was confirmed by ROEs observed between H-7, and the two H-6 and Me-8. In addition, the absence of ROEs between H-7 and the sugar protons suggested a C-1 *O*-glucosidic substitution. Conclusive evidence was given by the HMBC spectrum where a  $^3J_{\text{H}-\text{C}}$  cross-peak appeared between C-1 ( $\delta 139.3$ ) of the aglycone and H-1 ( $\delta 4.78$ ) of the sugar. Moreover, the

Table 1.  $^1\text{H}$  NMR (300 MHz) data of fagaropsine in  $\text{CD}_3\text{OD}$ 

H	$\delta$ J (Hz)
<b>Aglycone</b>	
4	5.26 s
5 <sub>ax</sub>	1.85 <i>td</i> (12.5, 6.0)
5 <sub>eq</sub>	1.13*
6 <sub>ax</sub>	1.95 <i>tt</i> (12.5, 2.5)
6 <sub>eq</sub>	1.68 <i>br dd</i> (12.5, 6.0)
7	3.84 <i>br s</i> *
8	3.53 <i>qt</i> (7.6, 1.5)
Me-4a	1.16 s
Me-8	1.20 <i>d</i> (7.6)
2'	7.59 <i>dd</i> (1.7, 0.8)
4'	6.50 <i>dd</i> (2.0, 0.8)
5'	7.53 <i>dd</i> (2.0, 1.7)
$\beta$ -D-glucose	
1''	4.78 <i>d</i> (6.5)
2''	3.41*
3''	3.39*
4''	3.38*
5''	3.34*
6''A	3.84* <i>dd</i> (12.0, 2.1)
6''B	3.68 <i>dd</i> (12.0, 5.0)

\*Overlapped signals.

Table 2.  $^{13}\text{C}$  NMR (75 MHz) data of fagaropsine in  $\text{CD}_3\text{OD}$ 

C	$\delta$	Gated Dec. $J_{\text{C}-\text{H}}$ (Hz)
<b>Aglycone</b>		
1	139.3 s	
2	165.2 s	
4	82.6 <i>d</i>	(149.5)
4a	39.6 s	
5	28.9 <i>t</i>	(129.5)
6	23.5 <i>t</i>	(130.0)
7	71.2 <i>d</i>	(145.3)
8	37.0 <i>d</i>	(133.3)
8a	155.0 s	
9	19.7 <i>q</i>	(129.5)
10	19.5 <i>q</i>	(129.5)
2'	142.9 <i>dd</i>	(199.0, 10.5)
3'	121.2 <i>br s</i>	
4'	111.2 <i>dd</i>	(174.0, 14.0)
5'	144.5 <i>ddd</i>	(204.2, 10.0, 9.0)
$\beta$ -D-glucose		
1''	105.2 <i>d</i>	(134.4)
2''	75.4 <i>d</i>	(145.7)
3''	77.8 <i>d</i>	(141.9)
4''	71.1 <i>d</i>	(145.3)
5''	78.5 <i>d</i>	(141.3)
6''	62.5 <i>t</i>	(142.2)

assignment of the  $^1\text{H}$  and  $^{13}\text{C}$  NMR resonances of **1** was supported by the HMBC spectrum and the sequence of the sugar protons which appeared as unresolved signals at  $\delta$  3.30–3.45 in the 1D  $^1\text{H}$  NMR, can be determined as H-2'', 3'', 4'', 5'' (Table 1).

The high field position of the angular methyl group signal in the  $^1\text{H}$  NMR spectrum indicated its position *cis* to the furan ring ( $\simeq +0.4$  ppm in *trans*-isomers) [8] and supported the normal limonoid stereochemistry. The comparison with NMR spectra of previously identified degraded limonoids [3–5, 11] and particularly dictamdiol, an aglycone isomer [3, 12], suggested that ring C had a chair conformation. A W-like coupling (1.5 Hz) was present between H-6<sub>eq</sub> ( $\delta$  1.68) and H-8 ( $\delta$  3.53) which inferred the axial ( $\alpha$ )-position of the 8-methyl group. Thus, the structure of fagaropsine (**1**) was deduced to be 1-*O*- $\beta$ -D-glucopyranosyl-(4*R*,4*aR*,7*S*,8*S*)-4-(3'-furanyl)-7-hydroxy-4*a*,8-dimethyl-4,4*a*,5,6,7,8-hexahydro-2*H*-3-benzopyran-2-one as the nomenclatural system adopted refers to its limonoid structure.

This compound is the first natural degraded limonoid isolated in a glucosylated form. It suggests the presence in this species of another biosynthetic route in limonoid metabolic glycosylation in contrast to the numerous 17-*O*- $\beta$ -glucopyranosyl limonoids found in *Citrus* [10, 13] or *Tetradium* [14] species.

## EXPERIMENTAL

**General.** Mps: uncorr. Analyt. TLC was on silica gel GF<sub>254</sub> and furanyl compounds were visualized with Ehrlich reagent spray followed by immersing in HCl vapour.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were obtained in

$\text{CD}_3\text{OD}$  at 300.13 and 75.45 MHz, respectively, using TMS as int. standard. FAB-MS were recorded in glycerol and glycerol + KI matrix.

**Plant material.** *Fagaropsis glabra* Capuron trunk bark was collected in the Sambava country (NE of Malagasy Republic) and authenticated at source by the ORSTOM centre of Tananarive where a voucher specimen is deposited with the reference number : 59.

**Extraction and isolation.** Dried powdered trunk bark (900 g) of *F. glabra* was defatted with petrol and then successively extracted with  $\text{CH}_2\text{Cl}_2$  (12 l) and EtOH 90° (15 l). The ethanolic extract was chromatographed over Amberlite XAD-4. The extract eluted with  $\text{MeOH}-\text{H}_2\text{O}$  (3 : 1) contained compounds reacting with Ehrlich reagent and was flash-chromatographed over silica gel. Crude fagaropsine was eluted with  $\text{EtOAc}-\text{MeOH}$  (9 : 1) and purified over a Bond Elut C<sub>18</sub> eluted with  $\text{MeOH}-\text{H}_2\text{O}$  (3 : 2) to afford 12 mg of **1**.

**Fagaropsine 1.** Mp 165–170°; UV  $\lambda_{\text{max}}^{\text{MeOH}}$  nm (log  $\epsilon$ ): 204 (3.8), 238sh; FT-IR  $\nu_{\text{max}}^{\text{KBr}}$  cm<sup>-1</sup>: 3430 (hydroxy), 1712, 1650 (conjugated  $\delta$ -lactone), 1504, 1070, 1026, 928, 876, 764, 728 (3-substituted furan); FAB-MS  $m/z$  (rel. int.): 441 ([M + H]<sup>+</sup>;  $\text{C}_{21}\text{H}_{28}\text{O}_{10}$ ) (100), 279 [(M + H) – (glucose +  $\text{H}_2\text{O}$ )<sup>+</sup>] (90);  $^1\text{H}$  and  $^{13}\text{C}$  NMR: Tables 1 and 2.

**Acknowledgements**—We sincerely thank Dr P. Uriac (Rennes) for his structural advice. We also thank Dr Montserrat B. (CNRS Toulouse) for recording the mass spectra.

## REFERENCES

1. Waterman, P. G. and Khalid, S. A. (1981) *Biochem. Syst. Ecol.* **9**, 45.
2. Blaise, A. J., Marion, C. and Winternitz, F. (1986) *J. Nat. Prod.* **49**, 724.
3. Boustie, J., Gleye, J., Blaise, A. and Fourasté, I. (1992) *Planta Med.* **58**, 228.
4. Blaise, A. J. and Winternitz, F. (1985) *Phytochemistry* **24**, 2379.
5. Boustie, J., Moulis, C., Gleye, J., Fourasté, I., Servin, P. and Bon, M. (1990) *Phytochemistry* **29**, 1699.
6. Champagne, D. E., Koul, O., Isman, M. B., Scudder, G. G. E. and Towers, G. H. N. (1992) *Phytochemistry* **32**, 377.
7. Bentley, M. D., Rajab, M. S., Mendel, M. J. and Alford, A. R. (1990) *J. Agric. Food Chem.* **38**, 1400.
8. Mateos, A. F. and De La Fuente Blanco, J. A. (1991) *J. Org. Chem.* **56**, 7084.
9. Grappin, M. R., Vanucci, C. and Lhommet, G. (1993) *J. Chem. Soc. Perkin Trans. 1*, 995.
10. Hasegawa, S., Fong, C. H., Herman, Z. and Miyake, M. (1992) *ACS Symp. Ser.* **490**, 87.
11. Cassady, J. M. and Liu, C. S. (1972) *J. Chem. Soc. Chem. Commun.* 86.
12. Hu, C., Han, J., Zhao, J., Song, G., Li, Y. and Yin, D. (1989) *Zhiwu Xuebao* **31**, 453.
13. Hasegawa, S., Bennett, R. D., Herman, Z., Fong, C. H. and Ou, P. (1989) *Phytochemistry* **28**, 1717.
14. Ozaki, Y., Miyake, M., Maeda, H., Ifuku, Y., Bennett, R. D. and Hasegawa, S. (1991) *Phytochemistry* **30**, 2365.