



## SECOMULTIFLORANE-TYPE TRITERPENOID ACIDS FROM STEM BARK OF *SANDORICUM KOETJAPE*

SOLEH KOSELA, YOKI YULIZAR, CHAIRUL,\* MOTOO TORI† and YOSHINORI ASAKAWA†

Faculty of Science, University of Indonesia, Depok, Indonesia; \*LIPI, Bogor, Indonesia; †Faculty of Pharmaceutical Sciences, Tokushima Bunri University, Yamashiro cho, Tokushima 770, Japan

(Received 10 February 1994)

**Key Word Index**—*Sandoricum koetjape*; Meliaceae; stem bark; bryononic acid; secobryononic acid; secoisobryononic acid.

**Abstract**—Bryononic acid and two new ring-A secotriterpenoids were isolated from *Sandoricum koetjape* stem bark and their structures elucidated by NMR spectrometry.

### INTRODUCTION

The stem bark of *Sandoricum koetjape* was collected in Jakarta, Indonesia. Local people use this plant against colic and leucorrhoea [1]. In previous investigations bryononic acid, bryonolic acid, mesoinositol and dimethyl mucate were isolated from the fruit hulls [2], two triterpenoid acids, katiconic acid and indicic acid, were isolated from the heartwood [3], while limonoids were isolated from the seeds [4]. The stem bark has been investigated and koetjape acid, 3-oxo-olean-12-en-29-oic acid, katiconic acid, (–)-alloaromadendrene, (–)-caryophyllene oxide, and (+)-spathulenol [5] were isolated. Further fractionation of the petrol extract of stem bark of *S. koetjape* planted in Indonesia resulted in the isolation of two new secomultiflorane-type triterpenoids, named secobryononic acid (2) and secoisobryononic acid (3), along with bryononic acid (1), and we report here their structure elucidation.

### RESULTS AND DISCUSSION

Dried powdered stem bark of *S. koetjape* was extracted with petrol for a week. After removal of the solvent the crude extract was separated using column chromatography to yield 1 and a mixture of compounds 2 and 3 which were separated and purified further as the methyl esters (4 and 5) using HPLC.

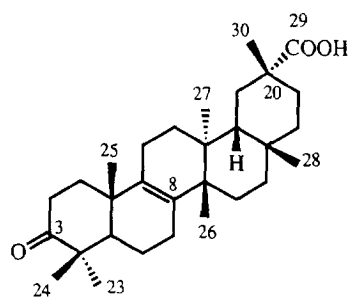
The molecular formula of 1 was determined as  $C_{30}H_{46}O_3$  from its HR-mass spectrum indicating that it was a triterpenoid. The presence of two carbonyl groups was inferred by a combination of the IR ( $\nu_{\max}$  1726 and 1676  $cm^{-1}$ ) and the  $^{13}C$ NMR ( $\delta$ 218.2 and 185.5) (Table 1) spectra. Seven methyl groups were indicated by both the  $^{13}C$ NMR [ $\delta$ 18.1, 19.4, 21.1, 21.6, 26.8, 31.2 and 32.7] and the  $^1H$ NMR [ $\delta$ 0.84 (3H), 0.96 (3H), 1.03 (6H), 1.06 (3H), 1.09 (3H) and 1.22 (3H)] spectra. The  $^1H$ NMR,

IR and mass spectra were identical with those of bryononic acid isolated from fruit hulls of the same plant [2]. Since we are interested in the conformation of this kind of triterpene, a crystal of 1 was analysed by X-ray spectrometry. The result is shown in Fig. 1. The A and E rings have chair conformations.

Compound 4 had the molecular formula  $C_{32}H_{50}O_4$  as determined by HR-mass spectrometry. The presence of two ester carbonyl groups was inferred from the  $^{13}C$ NMR ( $\delta$ 179.2 and 174.7) spectrum (Table 1). Six methyl and two methoxyl groups were shown by the  $^{13}C$ NMR ( $\delta$ 18.1, 21.6, 23.1, 23.3, 30.9, 31.3, 51.4 and 51.5) and the  $^1H$ NMR [ $\delta$ 0.79 (3H), 0.93 (3H), 0.96 (3H), 1.04 (3H), 1.18 (3H), 1.75 (3H), 3.62 (3H) and 3.63 (3H)] spectra. An exomethylene group ( $\delta$ 4.66 and 4.89) and a tetrasubstituted double bond were also present. Compound 4 was thus tetracyclic. The HMBC spectrum indicated the correlations of the six methyl groups as shown in Fig. 2. Thus compound 4 was revealed to be 3,4-secobryononic acid dimethyl ester. The stereochemistry is assumed to be the same as in 1.

The HR-mass spectrum of 5 showed 498.3705 indicating the molecular formula  $C_{32}H_{50}O_4$ . The presence of two ester carbonyl groups was indicated by the  $^{13}C$ NMR ( $\delta$ 174.8 and 179.4) spectrum (Table 1).

Six methyl and two methoxyl groups were observed in the  $^{13}C$ NMR spectrum ( $\delta$ 15.9, 22.5, 23.9, 24.8, 31.2, 32.9, 51.6 and 51.7). The data were supported by the presence of six singlet methyl and two singlet methoxyl groups in the  $^1H$ NMR spectrum [ $\delta$ 0.82 (3H), 0.86 (3H), 0.97 (3H), 1.01 (3H), 1.19 (3H), 1.78 (3H), 3.64 (3H) and 3.68 (3H)]. The exomethylene group at C-23 had similar shifts in the  $^1H$  and  $^{13}C$ NMR spectra ( $\delta$ 4.78, 4.83 and 113.8) to those of 4. A trisubstituted double bond was present as indicated by the  $^1H$ NMR [ $\delta$ 5.44 (1H, t,  $J=2$  Hz)] and  $^{13}C$ NMR [ $\delta$ 117.2 (t) and 145.7 (s)] spectra. Thus 5 was also tetracyclic. From the HMBC spectrum, compound 5



Bryononic acid (1)

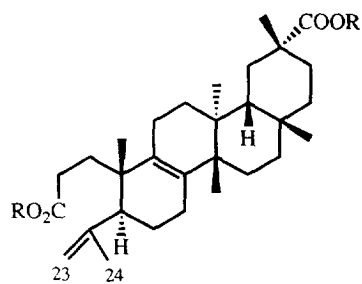
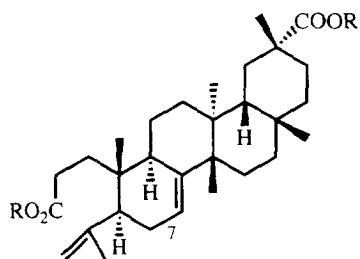
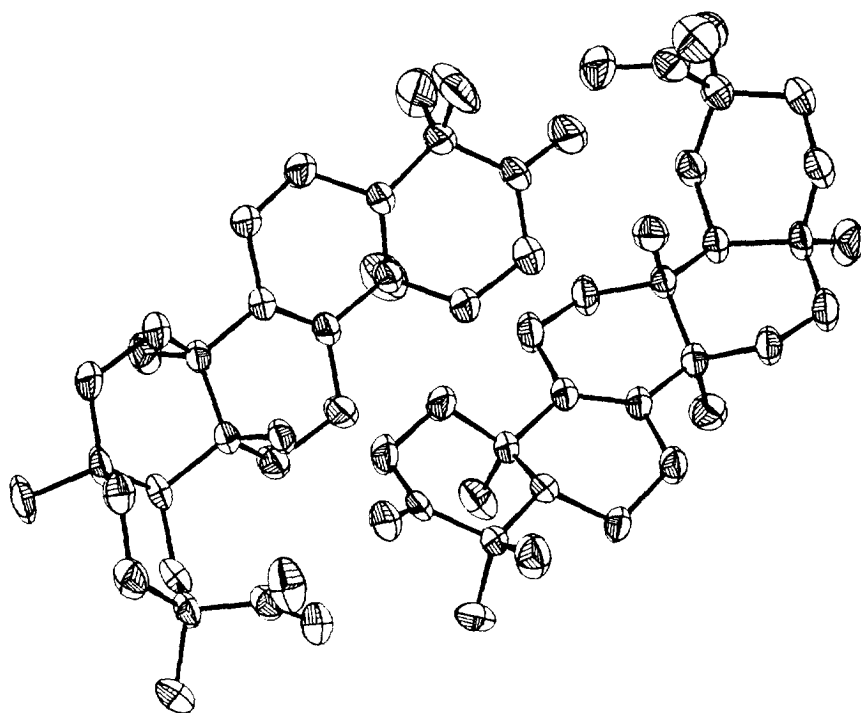
Secobryononic acid (2) R=H  
4 R=MeSecoisobryononic acid (3) R=H  
5 R=Me

Fig. 1. The ORTEP drawing for bryononic acid (1).

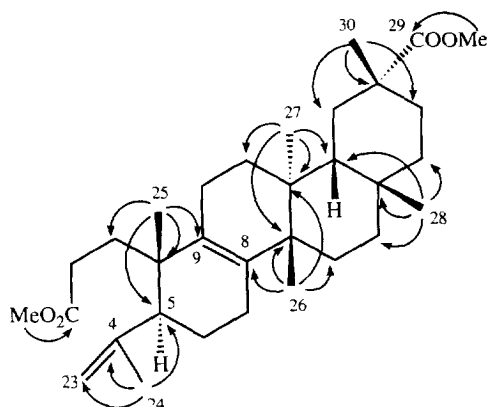


Fig. 2. The HMBC correlations for compound 4.

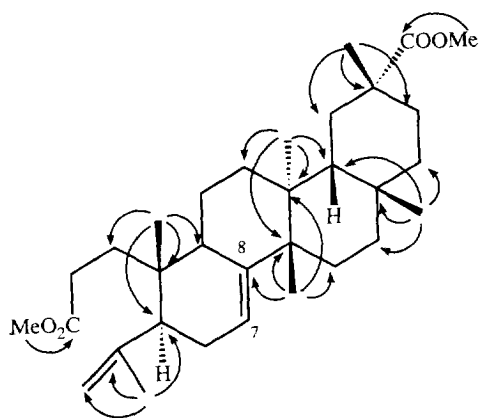


Fig. 3. The HMBC correlations for compound 5.

Table 1.  $^{13}\text{C}$  NMR (100 MHz) data for compounds 1, 4 and 5 ( $\text{CDCl}_3$ )

C	1	4	5
1	34.2 <sup>a</sup>	31.6	31.7
2	35.4 <sup>a</sup>	29.7	28.1
3	218.2	174.7	174.8
4	47.1	147.4	147.5
5	51.1	46.4	49.0
6	20.5 <sup>b</sup>	25.1 <sup>a</sup>	30.3 <sup>a</sup>
7	27.7 <sup>b</sup>	26.5 <sup>a</sup>	117.2
8	134.9	138.9	145.7
9	132.7	129.5	40.3
10	30.8	41.2	37.1
11	20.6 <sup>b</sup>	20.9	17.5 <sup>a</sup>
12	29.9	30.3	35.7
13	42.2	37.1 <sup>b</sup>	36.0
14	37.0	47.8 <sup>b</sup>	42.5
15	25.2	24.8	29.2
16	34.4 <sup>a</sup>	34.4	32.9
17	37.4	30.9	31.2
18	44.4	44.6	47.3
19	30.4	30.0 <sup>c</sup>	30.7
20	40.4	40.4	40.4
21	29.5	30.7 <sup>c</sup>	29.6
22	36.8 <sup>a</sup>	36.9	36.9
23	21.1	113.8	113.8
24	26.8	23.1	22.5
25	19.4	23.3	15.9
26	21.6	21.6	23.9
27	18.1	18.1	24.8
28	31.2	31.3	31.4
29	185.5	179.2	179.4
30	32.7	32.7	33.2
OMe		51.4	51.4
OMe		51.5	51.4

<sup>a-c</sup>Assignments may be changed within each vertical column.

was revealed to be 3,4-secooleana-4(23),7-diene-3,29-dioic acid dimethyl ester as shown in Fig. 3.

#### EXPERIMENTAL

Mps: uncorr. Spectral data: NMR, 100 MHz for  $^{13}\text{C}$  and 400 MHz for  $^1\text{H}$ ; TMS,  $\text{CDCl}_3$ .

**Plant material.** *Sandoricum koetjape* was collected in Jakarta, Indonesia in 1993. The voucher specimen is deposited at Bogor Herbarium and identified by M.S. Nani.

**Extraction and isolation.** The dried powdered stem bark of *S. koetjape* (1.05 kg) was extracted with 5 l petrol at room temp. After removal of the solvent under red. pres. a residue (36.4 g) was obtained. The residue (18.1 g) was subjected to CC on silica gel (160 g) with petrol and petrol-EtOAc as eluents. Frs 6–9 were combined, kept overnight and yielded 1 (1.5 g) as crystals. Frs 15–17 were a mixture of 2 and 3 (240 mg). The mixture was methylated with  $\text{CH}_2\text{N}_2$  and sepd using HPLC to give the dimethyl esters 4 (16.5 mg) and 5 (9.2 mg).

**Bryononic acid (1).** 250–253°, prism; IR  $\nu_{\text{max}}$   $\text{cm}^{-1}$ : 1726, 1676, 1458, 1382;  $^1\text{H}$  NMR:  $\delta$  0.84 (3H, s), 0.96 (3H, s), 1.03 (6H, s), 1.06 (3H, s), 1.09 (3H, s) and 1.22 (3H, s);  $^{13}\text{C}$  NMR

(Table 1); EI-MS  $m/z$  (rel. int.): 454  $[\text{M}]^+$  (51), 439 (29), 410 (13), 395 (10), 271 (7), and Int. 257 (76), 245 (73), 235 (100), 189 (25), 133 (15), 123 (7), 121 (19), 109 (16), 95 (26), 81 (11); HR-MS  $m/z$ : 454.3458  $[\text{M}]^+$  (calcd. for  $\text{C}_{30}\text{H}_{46}\text{O}_3$ , 454.3435).

**Secobryononic acid methyl ester (4)** (dimethyl 3,4-secomultiflora-4(23),8-diene-3,29-dioate).  $[\alpha]_D^{22} + 18.2^\circ$ , ( $\text{CHCl}_3$ ;  $c$  1.65); IR  $\nu_{\text{max}}$   $\text{cm}^{-1}$ : 1730, 890;  $^1\text{H}$  NMR:  $\delta$  0.79 (3H, s, H-27), 0.93 (3H, s, H-25), 0.96 (3H, s, H-26), 1.04 (3H, s, H-28), 1.18 (3H, s, H-30), 1.75 (3H, s, H-24), 3.62 (3H, s, OMe-29), 3.63 (3H, s, OMe-3), 4.66 (1H, s, H-23), 4.89 (1H, s, H-23);  $^{13}\text{C}$  NMR (Table 1); EI-MS  $m/z$  (rel. int.): 498  $[\text{M}]^+$  (90), 483 (10), 411 (100), 275 (20), 249 (50), 235 (15), 189 (45), 161 (40), 147 (20); HR-MS  $m/z$ : 498.3690  $[\text{M}]^+$  (calcd. for  $\text{C}_{32}\text{H}_{50}\text{O}_4$ , 498.3698).

**Secoisobryononic acid methyl ester (5)** (dimethyl 3,4-secomultiflora-4(23),7-diene-3,29-dioate).  $[\alpha]_D^{22} - 23.2^\circ$ , ( $\text{CHCl}_3$ ;  $c$  0.92); IR  $\nu_{\text{max}}$   $\text{cm}^{-1}$ : 1730, 890;  $^1\text{H}$  NMR:  $\delta$  0.82 (3H, s, H-25), 0.86 (3H, s, H-27), 0.97 (3H, s, H-26), 1.01 (3H, s, H-28), 1.19 (3H, s, H-30), 1.78 (3H, s, H-24), 3.64 (3H, s, OMe-29), 3.68 (3H, s, OMe-3), 4.78 (1H, s, H-23),

4.83 (1H, s, H-23), 5.44 (1H, t,  $J = 2$  Hz, H-7);  $^{13}\text{C}$  NMR (Table 1); EI-MS  $m/z$  (rel. int.): 498  $[\text{M}]^+$  (100), 483 (13), 411 (72), 275 (18), 249 (43), 235 (10), 189 (48), 161 (27), 147 (20); HR-MS  $m/z$ : 498.3705  $[\text{M}]^+$  (calcd. for  $\text{C}_{32}\text{H}_{50}\text{O}_4$ , 498.3696).

*Single-crystal X-ray diffraction analysis of bryononic acid (1).* The crystals were prepared from  $\text{CHCl}_3$ –MeOH. The crystal data for **1** were as follows: triclinic; space group P1 with  $a = 6.678$  (1),  $b = 13.756$  (3),  $c = 14.625$  (2) Å,  $\alpha = 100.48$  (2),  $\beta = 98.72$  (2),  $\gamma = 92.42$  (2)°,  $V = 102.4$  (4) Å<sup>3</sup>, and  $Z = 2$ . The empirical formula was  $\text{C}_{30}\text{H}_{46}\text{O}_3$ ,  $M_r = 454$ , and calculated density was  $1.16 \text{ g cm}^{-3}$ . 3D X-ray data were collected using a graphite-monochromated  $\text{CuK}_\alpha$  radiation ( $\lambda = 1.54178$ ) on a Mac Science MXC18 automatic four-circle diffractometer. Of 4267 total unique reflections, 4225 were used. The structure was solved by a direct method (Shelxs). The crystal has two molecules in a unit cell. The final  $R$  factor was  $R = 0.050$ ,  $R_w = 0.067$ .

Atomic coordinates, bond lengths and angles, and thermal parameters have been deposited at the Cambridge Crystallographic Data Centre.

#### REFERENCES

1. Naito, Y. (1986) *Medicinal Herb Index in Indonesia*, p. 222. P. T. Eisei.
2. Sim, K. Y. and Lee, H. T. (1972) *Phytochemistry* **11**, 3341.
3. King, F. E. and Morgan, J. W. W. (1960) *J. Chem. Soc.* 4738.
4. Powell, R. G., Mikolajczak, K. L., Zilkowski, B. W., Mantus, E. K., Cherry, D. and Clardy, J. (1991) *J. Nat. Prod.* **54**, 241.
5. Kaneda, N., Pezzuto, J. M., Kinghorn, A. D., Farnsworth, N. R., Santisuk, T., Tuchinda, P., Udchachon, J. and Reutrakul, V. (1992) *J. Nat. Prod.* **55**, 654.