

PHYTOTOXIC SESQUITERPENOIDS FROM *CANELLA WINTERANA*

BAI-PING YING,* GALEN PEISER, YI-YUAN JI, KRISTINA MATHIAS, DIANA TUTKO and YIH-SHEN HWANG

ISK Mountain View Research Center, Inc., 1195 W. Fremont Avenue, Sunnyvale, CA 94087, U.S.A.

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Key Word Index—*Canella winterana*; Canellaceae; drimane sesquiterpenoid; muzigadial; 3 α -acetoxypolygodial; 3 α -acetoxypolygodial 12-dimethyl acetal; 9-deoxymuzigadial 12 α -acetal; 9-deoxymuzigadial 12 β -acetal; 3 β -acetoxypolygodial 12 α -acetal; 3 β -acetoxypolygodial 12 β -acetal; 9-epideoxymuzigadial; 3 β -acetoxypolygonal; muzigaal; 3 β -acetoxypolygodial; 3 β -acetoxycinnamolide; isopolygodial; polygodial 12 α -acetal; drimenol; phytotoxicity.

Abstract—Nine new drimane-type sesquiterpenoids: 3 α -acetoxypolygodial, 3 α -acetoxypolygodial 12-dimethyl acetal, 9-deoxymuzigadial 12 α -acetal, 9-deoxymuzigadial 12 β -acetal, 3 β -acetoxypolygodial 12 α -acetal, 3 β -acetoxypolygodial 12 β -acetal, 9-epideoxymuzigadial, 3 β -acetoxypolygonal and muzigaal, were isolated from the leaves of *Canella winterana*, together with six previously known sesquiterpenoids: muzigadial, 3 β -acetoxypolygodial, 3 β -acetoxycinnamolide, isopolygodial, polygodial 12 α -acetal and drimenol. Unambiguous assignments of all ^1H and ^{13}C signals of 15 sesquiterpenoids were established by 2D NMR experiments.

INTRODUCTION

Canella winterana (L.) Gaertn. is a tree which grows in the Caribbean and subtropical area of Florida. Eight drimane sesquiterpenes, muzigadial (**1**) [1], warburganal, mukaadial, 9 α -hydroxycinnamolide [2], 3 β ,9 α -dihydroxycinnamolide [3], 9-deoxymuzigadial, 9-deoxyisomuzigadial and 3 β -acetoxypolygodial [4], have been found from the bark of this plant. 4,13 α -Epoxy muzigadial was isolated from its leaves [5]. Muzigadial and warburganal showed potent insect antifeedant [6], antifungal [7] and molluscicidal [8] activities.

In our search for bioactive compounds we found that the crude extract of the leaves of *C. winterana* exhibited phytotoxic activity. Bioassay-guided fractionation using the *Lemna minor* bioassay led to the isolation of nine new drimane sesquiterpenoids. In this paper, we report their isolation and structure determination.

RESULTS AND DISCUSSION

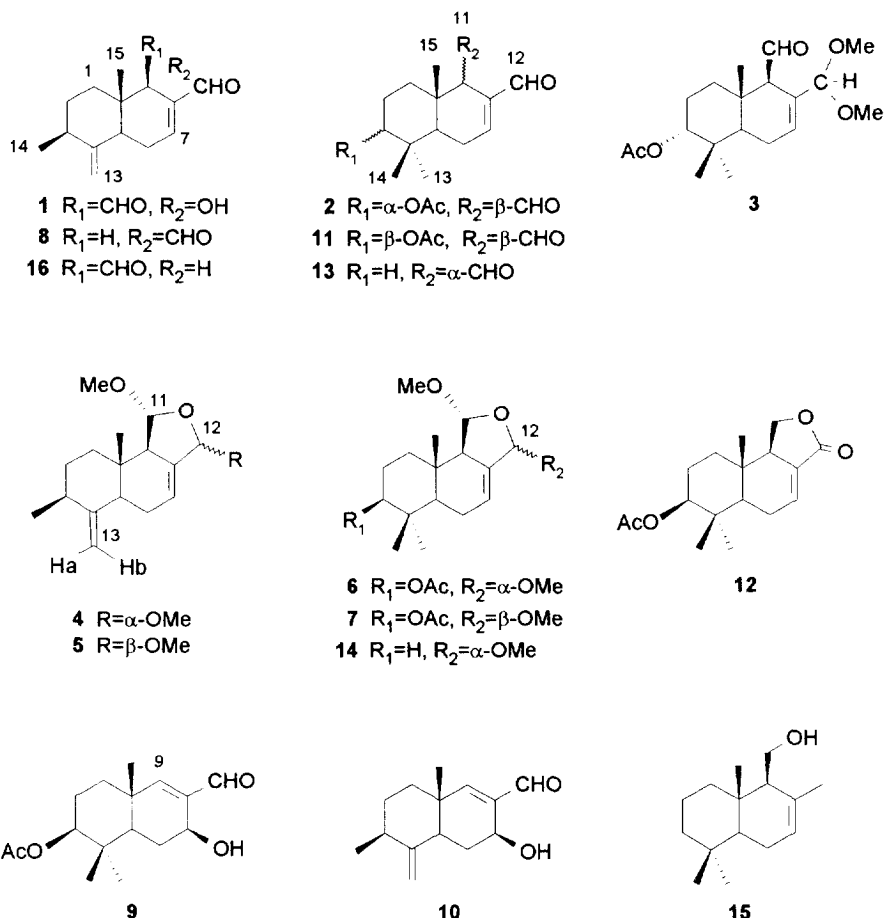
A combination of column chromatography on Sephadex LH-20, centrifugal partition chromatography (CPC) and recycling preparative HPLC of the pentane fraction of *C. winterana* resulted in the isolation of a major active compound, muzigadial (**1**) and nine new drimane sesquiterpenoids, 3 α -acetoxypolygodial (**2**), 3 α -acetoxypolygodial 12-dimethyl acetal (**3**), 9-deoxymuzig-

adial 12 α -acetal (**4**), 9-deoxymuzigadial 12 β -acetal (**5**), 3 β -acetoxypolygodial 12 α -acetal (**6**), 3 β -acetoxypolygodial 12 β -acetal (**7**), 9-epideoxymuzigadial (**8**), 3 β -acetoxypolygonal (**9**) and muzigaal (**10**), along with known 3 β -acetoxypolygodial (**11**) [4], 3 β -acetoxycinnamolide (**12**) [9], isopolygodial (**13**) [10–12], polygodial 12 α -acetal (**14**) [13] and drimenol (**15**) [11, 14, 15]. This is the first report of the isolation of **12–15** from this plant.

The ^1H and ^{13}C NMR (Table 1) spectra of **2** indicated the presence of two aldehyde groups (δ_{H} 9.43, 9.85, δ_{C} 192.5, 201.4), one methine group (δ_{H} 4.45, δ_{C} 79.8) bearing one acetoxy group (δ_{H} 2.06, δ_{C} 21.2, 170.6), three tertiary methyl groups (δ_{H} 0.92, 0.98, 0.98, δ_{C} 16.4, 21.6, 27.6), a trisubstituted conjugated olefinic bond (δ_{H} 7.10, δ_{C} 137.3, 152.5), three methylenes, one methine and two quaternary carbons. All these spectral features were very similar to those of **11**. The signals of C-1 and C-5 in **2** were significantly shifted upfield 3–5 ppm due to γ -interaction with the 3 α -acetoxy group, suggesting that **2** was 3 α -acetoxypolygodial. The complete assignment of ^1H and ^{13}C signals was achieved by ^1H – ^1H COSY, ^{13}C – ^1H COSY, long-range ^1H – ^1H COSY and long-range ^{13}C – ^1H COSY experiments.

There were 19 signals in the ^{13}C NMR spectrum (Table 1) of **3**: one saturated aldehyde group (δ_{C} 202.5), one acetoxy group (δ_{C} 21.2, 170.7), two methoxy groups (δ_{C} 53.5, 54.3), three tertiary methyl groups (δ_{C} 16.4, 21.5, 27.6), three methylenes, three methines, and two olefinic and three quaternary carbons. The chemical shifts of the AB ring carbons were similar to those of **2**, indicating that **3** was a derivative of **2**. The presence of two methoxy groups and an acetal carbon (δ_{H} 4.40, δ_{C} 105.5) suggested

*Author to whom correspondence should be addressed. Present address: Ricerca, Inc., 7528 Auburn Road, P.O. Box 1000, Painesville, OH 44077, U.S.A.



that **3** had an acetal structure at C-12 instead of an aldehyde as in **2**. Therefore, **3** was 3 α -acetoxypolygodial 12-dimethyl acetal. The assignment of all NMR signals was confirmed by ^{13}C - 1H COSY spectrum.

The 1H and ^{13}C NMR spectra (Table 1) of **4** and **5** were almost identical. There was one tertiary methyl, one secondary methyl and one exomethylene group in both **4** and **5**, indicating that they had similar structures to that of muzigadial. The lack of a carbon signal around δ_{77} and the presence of a tertiary carbon ($\delta_C 55.5$ and $\delta_H 2.64$ in **4**; $\delta_C 56.9$ and $\delta_H 2.40$ in **5**) indicated that there was no hydroxyl group at the 9 position. Both compounds had two methoxy groups ($\delta_C 54.3, 56.0, \delta_H 3.49$ and 3.42 in **4**; $\delta_C 55.5, 56.7, \delta_H 3.47$ and 3.52 in **5**), and two methine carbons bearing two oxygens [$\delta_C 107.2$ (C-11), 104.2 (C-12) and $\delta_H 5.13$ (H-12) in **4**; $\delta_C 105.7$ (C-11), 102.4 (C-12) and $\delta_H 5.44$ (H-12) in **5**] which were very similar to those of **14**. Therefore, **4** and **5** were acetals of 9-deoxymuzigadial. The assignment of all 1H and ^{13}C signals was supported by the ^{13}C - 1H COSY spectrum and confirmed by the long-range ^{13}C - 1H COSY spectrum. For example, in the long-range ^{13}H - 1H COSY spectrum of **4**, the cross-peaks between C-15 and H-9, C-9 and H-15, and C-9 and H-7 confirmed the structure around C-9.

The correlation between MeO-12 and H-12, C-12 and MeO-12, MeO-11 and H-11, and C-11 and MeO-11 confirmed the assignment of those signals. The stereochemistry of C-11 and C-12 was determined by an NOESY experiment. In the NOESY spectrum of **4**, the cross-peaks between H-11 and Me-15, and MeO-11 and H-5 indicated the α -configuration of the methoxy group at the 11-position. On the other hand, the correlation between MeO-12 and H-9, and MeO-12 and H-5 revealed the α -configuration of MeO-12. Thus, the structure of **4** was established to be 9-deoxymuzigadial 12 α -acetal. The assignment of H-13a and H-13b was decided based on the cross-peaks between Me-14 ($\delta 1.08$) and H-13a ($\delta 4.83$), and H-5 ($\delta 2.11$) and H-13b ($\delta 4.68$) in its NOESY spectrum.

In the NOESY spectrum of **5**, there were cross-peaks between MeO-11 and H-12, and MeO-12 and Me-15. The structure of **5** was determined as 9-deoxymuzigadial 12 β -acetal.

There were 19 signals in the ^{13}C NMR spectra (Table 1) of **6** and **7**, including one methine carbon bearing an acetoxy group, two methoxy groups, two acetal carbons and no aldehyde signal, indicating that **6** and **7** were acetals, similar to **4** and **5**. In addition, there

Table 1. ^{13}C NMR spectral data of **1–15** (75 MHz, CDCl_3)

was one double bond, three tertiary methyl groups, two other methine carbons and two quaternary carbons, suggesting that their carbon skeleton was the same as that of polygodial. The chemical shifts of C-1 and C-5 signals of **6** and **7** were very similar to those of **11** and **12**, but not to those of **2** and **3**, revealing that there was a β -acetoxy group in **6** and **7**. The β -configuration of the acetoxy group was supported by the coupling constants of H-3 (*dd*, 4.3, 11.0 Hz in **6**; *dd*, 5.5, 10.1 Hz in **7**) which indicated that H-3 had an axial configuration. The stereochemistry of C-11 and C-12 of **6** was established based on the similarity of the chemical shifts of C-11, MeO-11, H-11, C-12, MeO-12 and H-12 between **6** and **4** (δ_C 104.4, 56.0, δ_H 4.92, δ_C 106.8, 54.4 and δ_H 5.13 in **6**; δ_C 104.2, 56.0, δ_H 4.94, δ_C 107.2, 54.3 and δ_H 5.13 in **4**). Therefore **6** was determined to be β -acetoxy polygodial 12 α -acetal. Since a similar relationship could be observed between **7** and **5**, **7** was deduced to be 3 α -acetoxy polygodial 12 β -acetal.

The ^1H and ^{13}C NMR spectra (Table 1) of **8** were very similar to those of 9-deoxymuzigadial (**16**) [4]. The only differences were that the signals of C-1, C-5 and C-9 shifted upfield 2, 4 and 2.5 ppm, respectively. This indicated that the aldehyde group at C-9 in **8** had an α -configuration (pseudoaxial) and **8** was thus 9-epideoxymuzigadial. The assignment of all ^1H and ^{13}C NMR signals was confirmed by ^1H - ^1H COSY and ^{13}C - ^1H COSY experiments.

The ^{13}C NMR spectrum (Table 1) of **9** indicated that there was an acetoxy group (δ_C 21.3, 170.9) and 14 other signals, including one aldehyde (δ_C 196.1), three tertiary methyl, two olefinic, three methylene, three methine and two quaternary carbons. The ^1H - ^1H COSY, long-range ^1H - ^1H COSY and ^{13}C - ^1H COSY spectra clearly showed the relationship between all carbon and proton signals, indicating that **9** was a norsesquiterpenoid, either acetoxypolygonal or acetoxisopolygonal. The coupling constants (9.6 and 7.4 Hz) of the H-7 signal (δ_H 4.64, *dd*) clearly indicated that H-7 had an α -configuration; therefore, **9** was an acetoxisopolygonal. The similarity of chemical shifts of C-1 through C-5 between **9** and **7** indicated that the acetoxy group was located at the C-3 position. The β -configuration of the acetoxy group was evident from the *J* value (11.2 and 5.0 Hz). Thus, the structure of **9** was β -acetoxyisopolygonal.

The ^{13}C - ^1H COSY spectrum of **10** clearly revealed the relationship between its 14 carbon signals and proton signals: one aldehyde (δ_C 195.5, δ_H 9.49), one tertiary methyl group (δ_C 17.3, δ_H 0.82), one secondary methyl group (δ_C 18.1, δ_H 1.09), one exomethylene group (δ_C 104.5, δ_H 4.82 and 4.61, δ_C 152.4), one tri-substituted olefinic bond (δ_C 139.2 and 162.9, δ_H 6.71), three methylene groups (δ_C 29.1, δ_H 1.66 and 1.81; δ_C 31.8, δ_H 1.36 and 1.79; δ_C 37.4, δ_H 1.74 and 1.91), three methine groups (δ_C 38.2, δ_H 2.10; δ_C 43.1, δ_H 2.38; δ_C 61.6, δ_H 4.67) and one tertiary carbon (δ_C 39.2). The spectral data suggested that **10** was a derivative of muzigadial with one aldehyde group. The chemical shifts of the olefinic carbon and proton (δ_C 162.9, δ_H 6.71) indicated that this double bond was conjugated with the aldehyde group. Since the olefinic proton signal was a very sharp singlet, it had no adjacent proton. This

proton could only be located at the 9-position. In the ^1H - ^1H COSY spectrum there were cross-peaks among the signal at δ 2.38 and the signals at δ 1.74 and 1.91, which correlated with the signal at δ 4.67, suggesting that there

was a partial structure of $\begin{array}{c} | \\ -\text{CH}-\text{CH}_2-\text{CH}-\text{OH} \\ | \end{array}$. Therefore, the only position possible for the hydroxyl group was C-7. The broad doublet (*J* = 4.3 Hz) of the δ 4.67 signal indicated that the proton had a β -configuration. Thus, the structure of **10** was elucidated as 3,4,4a,5,6,7,8,8a-octahydro-3 α -hydroxy-6 β ,8a-dimethyl-5-methylene-2-naphthalenecarboxaldehyde. The trivial name muzigaal was given to this compound.

In addition to the nine new compounds (**2**-**10**), six known sesquiterpenoids (**1**, **11**-**15**) were identified by spectral analyses. The most abundant component from the leaves was **1** which had been previously isolated from the stem bark of this plant [1] and from the stem bark of the African plants *Warburgia stuhlmanni* and *W. ugandensis*. The ^1H and ^{13}C NMR spectral data of muzigadial in the literature were either incomplete or assigned incorrectly. By 2D NMR study, all ^1H and ^{13}C signals were assigned unambiguously.

Both ^1H and ^{13}C NMR spectral data of **11** exactly matched the literature data of β -acetoxy polygodial which was isolated from the stem bark of *C. winterana* [4].

Compound **12** showed the same ^1H and ^{13}C NMR spectral data (Table 1) as that of β -acetoxy cinnamolide, a component from the stem bark of *W. ugandensis* [9]. However, the assignment of the ^{13}C NMR signals needed to be corrected. On the ^{13}C - ^1H COSY spectrum, there were cross-peaks between the signals at δ_C 27.3 and δ_H 0.93, and at δ_C 13.5 and δ_H 0.84, indicating that the signal at δ_C 27.3 was C-13 and the signal at δ_C 13.5 was C-15. The signal at δ_C 24.6 correlated with the signals of δ_H 2.22 and 2.44, indicating that this signal should be assigned to C-6. Based on the correlation between the signals at δ_C 50.6 and δ_H 2.81, and δ_C 49.3 and δ_H 1.48, these two carbon signals could be assigned to C-9 and C-5, respectively. There has been no previous report of the isolation of β -acetoxy cinnamolide from *C. winterana*.

The identification of **13** was based on the comparison of its ^{13}C NMR spectral data with that of isopolygonal [12], which has been isolated from the seeds [10] and leaves [11] of *Polygonum hydropiper* and the leaves of *Pseudowinterana colorata* [12]. This is the first time that **13** was isolated from *C. winterana*.

Compound **14** had characteristic ^1H and ^{13}C NMR signals of cyclic dimethyl acetal (δ_H 3.40 and 3.47, δ_C 54.3 and 56.0, and δ_H 4.91 and 5.12, δ_C 104.6 and 107.0) found from **4**-**7**. The presence of three tertiary methyl groups suggested that **14** was a derivative of polygodial. Comparison of the ^{13}C NMR spectral data of **14** with the published data of polygodial acetal (isolated from *P. hydropiper*) [13] revealed that **14** was an epimeric isomer of polygodial acetal. Fukuyama *et al.* [13] separated two isomers of polygodial acetals, but did not determine their stereochemistry. Recently, Tozyo *et al.* [14] prepared these two isomers, acetal-1 and acetal-2, from polygodial and determined their configurations. Therefore, the struc-

ture of **14** was elucidated as polygodial 12 α -acetal. The ^1H and ^{13}C signal assignments were established based on the ^1H - ^1H COSY and ^{13}C - ^1H COSY spectra. For example, the signal of H-12 (δ_{H} 5.12, s) correlated with the signal at δ_{H} 3.40, and the signal of H-11 (δ_{H} 4.91, d, $J = 3.3$ Hz) correlated with the signal of H-9 (δ_{H} 2.45). Therefore, the methoxy signal at δ_{H} 3.40 was assigned at the C-12 position. The cross-peaks between the signals at δ_{C} 104.6 and δ_{H} 5.12, δ_{C} 107.0 and δ_{H} 4.91, δ_{C} 56.0 and δ_{H} 3.47, and δ_{C} 54.3 and δ_{H} 3.40 confirmed their assignments. In the long range ^{13}C - ^1H COSY spectrum the signal at δ_{C} 33.3 correlated with the Me-15 signal (δ_{H} 0.77). Thus, this signal was assigned to C-10, and another quaternary carbon signal (δ_{C} 32.9) was assigned to C-4. All equivocal assignments on ^1H and ^{13}C NMR data were clarified.

In **15** there was one methyl group (δ_{C} 22.0 and δ_{H} 1.79) attached to an olefinic bond in addition to three tertiary methyl groups (δ_{C} 15.0 and δ_{H} 0.82, δ_{C} 22.1 and δ_{H} 0.89, and δ_{C} 33.4 and δ_{H} 0.87). Another characteristic of this compound was that there was a hydroxymethylene group (δ_{C} 61.0, and δ_{H} 3.76 and 3.86). Compound **15** was identified as drimenol which has been isolated from *Porella vernicosa* (a liverwort) [14], *Polygonum hydropiper* [11] and *Drimys winteri* [15].

There are several papers [16–19] that discuss the possible transformation of dialdehydes during extraction and separation processes. In our case, the dried plant material was extracted with methanol at room temperature and separated by two very mild methods (Sephadex LH-20 and centrifugal partition chromatography). Furthermore, the separation was monitored by LC-MS, ^{13}C and ^1H NMR, in addition to routine TLC. When the plant was extracted with methylene chloride, the ^{13}C and ^1H NMR spectra of the crude methylene chloride extract showed acetal signals clearly. The possibility of artifact formation cannot be completely excluded. At the very least the yield of each compound isolated may not be the same as the content of the original plant.

The phytotoxic activities of **1** through **14** in the *Lemna minor* bioassay are summarized in Table 2.

Table 2. Phytotoxic activities of **1**–**14**

Compound	IG ₅₀ (μM)
Muzigadial (1)	8–16
3 α -Acetoxypolygodial (2)	340
3 α -Acetoxypolygodial 12-dimethyl acetal (3)	> 150
9-Deoxymuzigadial 12 α -acetal (4)	18
9-Deoxymuzigadial 12 β -acetal (5)	8
3 β -Acetoxypolygodial 12 α -acetal (6)	60
3 β -Acetoxypolygodial 12 β -acetal (7)	30
9-Epideoxymuzigadial (8)	45–200
3 β -Acetoxypolygonal (9)	180–360
Muzigaal (10)	230–460
3 β -Acetoxypolygodial (11)	35–70
3 β -Acetoxycinnamolide (12)	> 700
Isopolygodial (13)	45
Polygodial 12 α -acetal (14)	18

EXPERIMENTAL

General. Mp: uncorr.; UV (MeOH); IR: film; NMR (300 MHz for ^1H and 75 MHz for ^{13}C) CDCl_3 , TMS as int. standard; CI-MS: CH_4 as reactant gas.

Plant material. The leaves of *C. winterana* were collected and identified in Puerto Rico by J. A. Cedeño and D. A. Kolterman. A voucher specimen was deposited in the Herbarium, University of Puerto Rico, Mayagüez.

Bioassay. The *Lemna minor* bioassay was conducted as described previously [20].

Extraction and isolation. The dried leaves (529 g) of *C. winterana* were extracted with MeOH at room temp. The MeOH extract was concd under red. pres. and partitioned with pentane and CH_2Cl_2 sequentially. The pentane phase was evapd to yield 50.26 g of oil which showed phytotoxic activity in the *Lemna minor* assay at 20 ppm. The pentane fr. was dissolved in CH_2Cl_2 and sepd on a Sephadex LH-20 column to provide 5 frs. The active fr. (no. 2) was further fractionated by centrifugal partition chromatography (Sanki NMF, hexane–EtOAc–MeOH– H_2O , 8:2:8:2, ascending mode) to give 11 frs. The final purification was carried out mainly by recycling prep. HPLC (GS-310P column/MeOH, ODS/75% MeOH, or Nucleosil/hexane–EtOAc) to obtain muzigadial (947 mg), 3 α -acetoxypolygodial (4 mg), 3 α -acetoxypolygodial 12-dimethyl acetal (26 mg), 9-deoxymuzigadial 12 α -acetal (744 mg), 9-deoxymuzigadial 12 β -acetal (101 mg), 3 β -acetoxypolygodial 12 α -acetal (24 mg), 3 β -acetoxypolygodial 12 β -acetal (26 mg), 9-epideoxymuzigadial (96 mg), 3 β -acetoxypolygonal (11 mg), muzigaal (33 mg), 3 β -acetoxypolygodial (27 mg), 3 β -acetoxycinnamolide (14 mg), isopolygodial (39 mg), polygodial 12 α -acetal (263 mg) and drimenol (13 mg).

Muzigadial (1). Needles (from hexane– CH_2Cl_2), mp 128–129°. UV λ_{max} nm: 225 (log ϵ 3.73). IR ν_{max} cm^{-1} : 3460, 2958, 2914, 2872, 1716, 1674, 1657, 1408, 1339, 1207, 1137, 1022, 992, 900, 844, 810, 791. ^1H NMR: δ 0.88 (3H, s, Me-15), 1.07 (1H, m, H-2), 1.08 (3H, s, Me-14), 1.10 (1H, m, H-1) 1.68 (1H, m, H-1), 2.00 (2H, m, H-2 and H-3), 2.51 (2H, m, CH_2 -6), 2.62 (1H, m, H-5), 4.77 (1H, s, H-13b), 4.94 (1H, s, H-13a), 7.25 (1H, br t, $J = 3.3$ Hz, H-7), 4.45 (1H, m, H-3), 9.45 (1H, s, H-12), 9.66 (1H, s, H-11). EI-MS m/z (rel. int.): 249 [$\text{M} + 1$]⁺ (1), 219 (100), 187 (12), 177 (16), 159 (17), 135 (42), 107 (36), 107 (36), 91 (38); positive CI-MS m/z (rel. int.): 248 [M]⁺ (28), 220 (20), 219 [$\text{M} - \text{CHO}$] (100).

3 α -Acetoxypolygodial (2). Oil. UV λ_{max} nm: 220. IR ν_{max} cm^{-1} : 2996, 2912, 1725, 1437, 1407, 1312, 1250, 955, 701, 671. ^1H NMR: δ 0.92 (3H, s, Me-13), 0.98 (6H, s, Me-14 and 15), 1.62 (1H, m, H-5), 1.7 (2H, m, CH_2 -2), 1.8 (2H, m, CH_2 -1), 2.06 (3H, s, MeCO), 2.31 (1H, dd, $J = 11.4$, 20.4 Hz, H-6 β), 2.60 (1H, ddd, $J = 4.8$, 5.1, 20.4 Hz, H-6 α), 3.32 (1H, br s, H-9), 4.45 (1H, m, H-3), 7.10 (1H, dd, $J = 2.8$, 4.8 Hz, H-7), 9.43 (1H, s, H-12), 9.85 (1H, d, $J = 2.5$ Hz). EI-MS m/z (rel. int.): 264 [$\text{M} - 28$]⁺ (5), 218 (14), 204 [$\text{M} - 28 - 60$]⁺ (100), 189 [$\text{M} - 28 - 60 - 15$]⁺ (87), 174 (32), 133 (36), 122 (72), 107 (83), 69 (66), 55 (71). Negative CI-MS m/z (rel. int.): 308 [$\text{M} + \text{CH}_4$][−] (80), 306 (82), 294 (97), 278 (100), 218 (97), 166 (86), 112 (46), 98 (58).

3 α -Acetoxypolygodial 12-dimethyl acetal (3). Oil. IR ν_{\max} cm^{-1} : 2967, 2878, 1728, 1688, 1459, 1442, 1396, 1375, 1247, 1032, 983, 736. ^1H NMR: δ 0.91 (3H, s, Me-13), 0.95 (3H, s, Me-15), 0.97 (3H, s, Me-14), 1.6 (2H, m, CH_2 -1), 1.7 (3H, m, H-5 and CH_2 -2), 2.1 (1H, m, H-6), 2.05 (3H, s, MeCO), 2.35 (1H, ddd, $J = 4.7, 5.1, 18.5$ Hz, H-6), 2.76 (1H, br d, $J = 4.1$ Hz, H-9), 3.24 (3H, s, MeO-11), 3.29 (3H, s, MeO-12), 4.40 (1H, s, H-12), 4.47 (1H, br t, $J = 7.7$ Hz, H-3), 6.12 (1H, dd, $J = 2.6, 4.4$ Hz, H-7), 9.65 (1H, d, $J = 4.2$ Hz, H-11). EI-MS m/z (rel. int.): 309 [$\text{M} - \text{CHO}$] $^+$ (20), 277 [$\text{M} - 60 - 1$] $^+$ (32), 218 (45), 204 (76), 189 (64), 173 (47), 122 (79), 107 (88), 69 (93), 57 (100). Negative CI-MS m/z (rel. int.): 278 [$\text{M} - 60$] $^-$ (100).

9-Deoxymuzigadial 12 α -acetal (4). Mp 69–71 $^\circ$. IR ν_{\max} cm^{-1} : 2958, 2855, 2820, 1718, 1674, 1640, 1456, 1384, 1326, 1253, 1191, 1084, 980, 949, 833. ^1H NMR: δ 0.62 (3H, s, Me-15), 1.08 (3H, d, $J = 6.5$ Hz, Me-14), 1.23 (1H, ddd, $J = 3, 12, 25$ Hz, H-2), 1.53 (1H, ddd, $J = 3, 13, 13$ Hz, H-1), 1.65 (1H, m, H-2), 1.74 (1H, ddd, $J = 3, 3, 13$ Hz, H-1), 2.02 (1H, m, H-3), 2.11 (3H, m, H-5 and CH_2 -6), 2.64 (1H, br t, $J = 3$ Hz, H-9), 3.42 (3H, s, MeO-12), 3.49 (3H, s, MeO-11), 4.68 (1H, br s, H-13b), 4.83 (1H, br s, H-13a), 4.94 (1H, d, $J = 4.7$ Hz, H-11), 5.13 (1H, s, H-12), 5.93 (1H, br t, $J = 2$ Hz, H-7). EI-MS m/z (rel. int.): 263 [$\text{M} - 15$] $^+$ (4), 247 [$\text{M} - \text{OMe}$] $^+$ (26), 218 (100), 203 (49), 189 (64), 173 (48), 171 (47), 131 (51), 109 (77), 91 (87), 65 (76). Positive CI-MS m/z (rel. int.): 279 [$\text{M} + 1$] $^+$ (38), 263 [$\text{M} - 15$] $^+$ (60), 261 (64), 247 [$\text{M} - \text{OMe}$] $^+$ (100).

9-Deoxymuzigadial 12 β -acetal (5). Oil. ^1H NMR: δ 0.67 (3H, s, Me-15), 1.08 (3H, d, $J = 6.5$ Hz, Me-14), 1.23 (1H, ddd, $J = 3.3, 12.7, 25.6$ Hz, H-2), 1.49 (1H, ddd, $J = 3, 3, 13.3$ Hz, H-1), 1.66 (1H, ddd, $J = 3.8, 6.9, 12.7$ Hz, H-2), 1.82 (1H, ddd, $J = 3.6, 13.3, 13.3$ Hz, H-1), 2.02 (1H, m, H-3), 2.07 (1H, m, H-5), 2.11 (2H, dd, $J = 11.4, 20.4$ Hz, H-6), 2.40 (1H, dd, $J = 3.3, 6.7$ Hz, H-9), 3.47 (3H, s, MeO-11), 3.52 (3H, s, MeO-12), 4.68 (1H, br s, H-13b), 4.83 (1H, br s, H-13a), 4.92 (1H, d, $J = 6.7$ Hz, H-11), 5.44 (1H, br s, H-12), 5.84 (1H, br d, $J = 3$ Hz, H-7).

3 β -Acetoxypolygodial 12 α -acetal (6). Oil. IR ν_{\max} cm^{-1} : 2920, 1732, 1465, 1444, 1389, 1368, 1246, 1191, 1091, 1023, 964, 923, 826. ^1H NMR: δ 0.79 (3H, s, Me-15), 0.88 (3H, s, Me-13), 0.96 (3H, s, Me-14), 1.41 (1H, dd, $J = 5.5, 11.3$ Hz, H-5), 1.44 (1H, m, H-1), 1.67 (2H, m, CH_2 -2), 2.1 (1H, m, H-6), 2.06 (3H, s, MeCO), 2.20 (1H, m, H-6), 2.44 (1H, br t, $J = 3.5$ Hz, H-9), 3.41 (3H, s, MeO-12), 3.46 (3H, s, MeO-11), 4.53 (1H, dd, $J = 4.2, 10.9$ Hz, H-3), 4.92 (1H, d, $J = 3.9$ Hz, H-11), 5.13 (1H, s, H-12), 5.81 (1H, br d, $J = 3.0$ Hz, H-7). EI-MS m/z (rel. int.): 337 [$\text{M} - 1$] $^+$ (2), 307 [$\text{M} - \text{OMe}$] $^+$ (52), 278 [$\text{M} - 60$] $^+$ (56), 218 [$\text{M} - 60 - 60$] $^+$ (99), 203 (100), 187 (10), 175 (10), 173 (16), 171 (27). Positive CI-MS m/z (rel. int.): 337 [$\text{M} - 1$] $^+$ (1.5), 307 [$\text{M} - \text{OMe}$] $^+$ (52), 278 [$\text{M} - 60$] $^+$ (57), 218 [$\text{M} - 60 - 60$] $^+$ (100), 203 (99), 171 (26), 105 (22), 91 (28).

3 β -Acetoxypolygodial 12 β -acetal (7). Mp. 110–113 $^\circ$. IR ν_{\max} cm^{-1} : 2930, 1732, 1444, 1395, 1374, 1246, 1193, 1102, 1022, 986, 931. ^1H NMR: δ 0.87 (3H, s, Me-15), 0.89 (3H, s, Me-13), 0.96 (3H, s, Me-14), 1.33 (1H, dd, $J = 5.3, 11.4$ Hz, H-5), 1.45 (1H, m, H-1), 1.6 ~ 1.85 (5H, m), 2.06 (3H, s, MeCO), 2.21 (1H, m, H-9), 3.46 (3H, s, MeO-11), 3.48 (3H,

s, MeO-12), 4.53 (1H, dd, $J = 5.5, 10.1$ Hz, H-3), 4.92 (1H, d, $J = 5.9$ Hz, H-11), 5.39 (1H, s, H-12), 5.80 (1H, br s, H-7).

9-Epideoxymuzigadial (8). Oil. ^1H NMR: δ 0.77 (3H, s, Me-15), 1.08 (3H, d, $J = 6.5$ Hz, Me-14), 1.29 (1H, ddd, $J = 4, 12, 24$ Hz, H-2 α), 1.61 (1H, m, H-1 α), 1.70 (1H, m, H-2 β), 1.82 (1H, m, H-1 β), 1.95 (1H, m, H-3), 2.38 (1H, br s, H-5), 2.48 (2H, m, CH_2 -6), 3.45 (1H, br d, $J = 2.7$ Hz, H-9), 4.73 (1H, s, H-13b), 4.89 (1H, s, H-13a), 7.14 (1H, dd, $J = 2.7, 4.2$ Hz, H-7), 9.44 (1H, s, H-12), 9.91 (1H, d, $J = 2.7$ Hz, H-11); EI-MS m/z (rel. int.): 232 [M] $^+$ (100), 217 (39), 214 (90), 203 (59), 199 (57), 189 (48), 173 (70), 159 (58); negative CI-MS m/z (rel. int.): 249 [$\text{M} + \text{CH}_4 + 1$] $^-$ (34), 235 (100), 219 (79).

3 β -Acetoxypolygonal (9). Oil. UV λ_{\max} nm: 226 (log ϵ 4.05). IR ν_{\max} cm^{-1} : 3484, 2968, 2872, 1732, 1676, 1458, 1374, 1247, 1029, 736. ^1H NMR: δ 0.93 (3H, s, Me-15), 0.94 (3H, s, Me-13), 1.12 (1H, m, H-5), 1.18 (3H, s, Me-14), 1.47 (1H, m, H-1 α), 1.54 (1H, m, H-6), 1.70 (1H, m, H-1 β), 1.81 (2H, m, CH_2 -2), 2.07 (3H, s, MeCO), 2.18 (1H, dd, $J = 7.0, 12.8$ Hz, H-6), 3.65 (1H, br s, OH), 4.50 (1H, dd, $J = 5.0, 11.3$ Hz, H-3), 4.64 (1H, dd, $J = 7.4, 9.6$ Hz, H-7), 6.46 (1H, s, H-9), 9.40 (1H, s, H-12).

Muzigaal (10). Mp 100–101 $^\circ$. UV λ_{\max} nm: 225 (log ϵ 4.15). IR ν_{\max} cm^{-1} : 3528, 3460, 3385, 2957, 2917, 1667, 1636, 1450, 1405, 1385, 1369, 1242, 1224, 1181, 1128, 1050, 1023, 999, 960, 908, 882, 723, 634. ^1H NMR: δ 0.82 (3H, s, Me-15), 1.09 (3H, d, $J = 6.4$ Hz, Me-14), 1.36 (1H, ddd, $J = 8.4, 12.6, 22.3$ Hz, H-1), 1.66 (1H, m, H-2), 1.74 (1H, m, H-6), 1.79 (1H, m, H-1), 1.81 (1H, m, H-2), 1.91 (1H, br d, $J = 13.8$ Hz, H-6), 2.10 (1H, dq, $J = 5.8, 6.4$ Hz, H-3), 2.38 (1H, br d, $J = 12.8$ Hz, H-5), 4.61 (1H, s, H-13b), 4.67 (1H, br d, $J = 4.3$ Hz, H-7), 4.82 (1H, s, H-13a), 6.71 (1H, s, H-9), 9.49 (1H, s, H-12).

3 β -Acetoxypolygodial (11). Oil. IR ν_{\max} cm^{-1} : 3434, 2967, 1732, 1682, 1644, 1372, 1247, 1033, 736. ^1H NMR: δ 0.93 (3H, s, Me-13), 0.97 (3H, s, Me-15), 1.01 (3H, s, Me-14), 1.36 (1H, dd, $J = 5.1, 11.4$ Hz, H-5), 1.64 (1H, m, H-1), 1.70 (2H, m, CH_2 -2), 1.90 (1H, m, H-1), 2.50 (2H, m, CH_2 -6), 2.82 (1H, br s, H-9), 4.57 (1H, dd, $J = 4.3, 10.8$ Hz, H-3), 7.14 (1H, ddd, $J = 2.2, 4.2, 6.1$ Hz, H-7), 9.48 (1H, s, H-12), 9.54 (1H, d, $J = 4.2$ Hz, H-11). EI-MS m/z (rel. int.): 264 [$\text{M} - 28$] $^+$ (6), 204 [$\text{M} - 28 - 60$] $^+$ (100), 189 [$\text{M} - 28 - 60 - 15$] $^+$ (92), 133 (34), 122 (54), 107 (64), 69 (77), 57 (95). Negative CI-MS m/z (rel. int.): 308 [$\text{M} + \text{CH}_4$] $^-$ (100), 306 (90), 291 [$\text{M} - 1$] $^-$ (76), 278 (68), 251 (82), 235 (65).

3 β -Acetoxycinnamolide (12). Oil. IR ν_{\max} cm^{-1} : 2970, 2955, 1754, 1728, 1682, 1482, 1461, 1444, 1422, 1396, 1376, 1244, 1201, 1140, 1095, 1041, 1040, 1010, 967, 742. ^1H NMR: δ 0.84 (3H, s, Me-15), 0.93 (3H, s, Me-13), 1.00 (3H, s, Me-14), 1.42 (1H, m, H-1), 1.48 (1H, dd, $J = 5.5, 11.4$ Hz, H-5), 1.64 (1H, m, H-1), 1.73 (2H, m, CH_2 -2), 2.08 (3H, s, MeCO), 2.22 (1H, ddd, $J = 4.7, 11.4, 16$ Hz, H-6), 2.44 (1H, ddd, $J = 5.5, 9, 16$ Hz, H-6), 4.05 (1H, t, $J = 9$ Hz, H-11), 4.40 (1H, t, $J = 9$ Hz, H-11), 4.55 (1H, dd, $J = 5, 11$ Hz, H-3), 6.89 (1H, dd, $J = 3.3, 6.8$ Hz, H-7). EI-MS m/z (rel. int.): 293 [$\text{M} + 1$] $^+$ (1), 232 [$\text{M} - 60$] $^+$ (5), 122 (100), 107 (44), 96 (17), 57 (95); negative CI-MS m/z (rel.

int.): 291 $[M - 1]^-$ (100), 251 (38), 249 (27), 235 (42), 233 (27), 67 (65). Positive CI-MS m/z (rel. int.): 293 $[M + 1]^+$ (95), 249 (3), 233 (14), 122 (100), 107 (19).

Isopolygodial (**13**). Oil. IR $\nu_{\max} \text{ cm}^{-1}$: 2928, 1722, 1682, 1657, 1461, 1418, 1391, 1368, 1211, 1034, 970, 953, 735. $^1\text{H NMR}$: δ 0.91 (3H, s, Me-13), 0.94 (3H, s, Me-15), 0.97 (3H, s, Me-14), 1.16 (1H, *ddd*, $J = 3.8, 13.4, 13.4 \text{ Hz}$, H-3 α), 1.47 (1H, *m*, H-3 β), 1.5 (2H, *m*, H-1 and H-2), 1.6 (1H, *m*, H-5), 1.64 (1H, *m*, H-2 β), 1.79 (1H, *br d*, $J = 11 \text{ Hz}$, H-1 β), 2.22 (1H, *dd*, $J = 11.6, 20.5 \text{ Hz}$, H-6 β), 2.57 (1H, *ddd*, $J = 4.7, 5.1, 20.5 \text{ Hz}$, H-6 α), 3.26 (1H, *br s*, H-9), 7.11 (1H, *dd*, $J = 2.5, 4.7 \text{ Hz}$, H-7), 9.41 (1H, s, H-12), 9.86 (1H, *br s*, H-11). EI-MS m/z (rel. int.): 206 $[M - 28]^+$ (84), 191 (31), 121 (69), 109 (100), 91 (52). Negative CI-MS m/z (rel. int.): 251 $[M + \text{CH}_4 + 1]^-$ (16), 237 (100), 221 (77), 128 (53); positive CI-MS m/z (rel. int.): 235 $[M + 1]^+$ (16), 221 (74), 205 $[M - \text{CHO}]^+$ (100), 139 (43), 109 (70).

Polygodial 12 α -acetal (**14**). Oil. IR $\nu_{\max} \text{ cm}^{-1}$: 2940, 2923, 2846 1460, 1443, 1390, 1365, 1326, 1278, 1257, 1205, 1192, 1113, 1094, 1077, 1017, 963, 923, 824. $^1\text{H NMR}$: δ 0.77 (3H, s, Me-15), 0.87 (3H, s, Me-14), 0.91 (3H, s, Me-13), 1.22 (1H, *dddd*, $J = 3.5, 3.5, 10, 13 \text{ Hz}$, H-1), 1.32 (1H, *dd*, $J = 5.5, 11.5 \text{ Hz}$, H-5), 1.40 (2H, *m*, CH₂-3), 1.5 (2H, *m*, CH₂-2), 1.66 (1H, *m*, H-1), 1.93 (1H, *dddd*, $J = 3.3, 3.5, 11.5, 18.4 \text{ Hz}$, H-6 β), 2.16 (1H, *ddd*, $J = 3.5, 8.4, 18.4 \text{ Hz}$, H-6 α), 2.45 (1H, *br t*, $J = 3.2 \text{ Hz}$, H-9), 3.40 (3H, s, MeO-12), 3.47 (3H, s, MeO-11), 4.91 (1H, *d*, $J = 3.9 \text{ Hz}$, H-11), 5.12 (1H, s, H-12), 5.80 (1H, *br d*, $J = 3.0 \text{ Hz}$, H-7). EI-MS m/z (rel. int.): 279 $[M - 1]^+$ (0.4), 249 $[M - \text{OMe}]^+$ (17), 220 $[M - 60]^+$ (75), 205 $[M - 60 - 15]^+$ (100), 173 (11), 135 (55), 111 (53), 91 (24). Positive CI-MS m/z (rel. int.): 279 $[M - 1]^+$ (1), 249 $[M - \text{OMe}]^+$ (30), 220 $[M - 60]^+$ (100), 205 $[M - 60 - 15]^+$ (89), 135 (42), 111 (38), 91 (29).

Drimenol (**15**). Oil. IR $\nu_{\max} \text{ cm}^{-1}$: 3405, 2922, 2848, 1443, 1383, 1250, 1213, 1175, 1155, 1032, 784, 742, 641. $^1\text{H NMR}$: δ 0.82 (3H, s, Me-15), 0.87 (3H, s, Me-13), 0.89 (3H, s, Me-14), 1.07 (1H, *m*, H-1), 1.20 (2H, *m*, H-3 and H-5), 1.41 (1H, *m*, H-3), 1.48 (2H, *m*, CH₂-2), 1.79 (3H, s, Me-12), 1.86 (1H, *m*, H-9), 1.92 (1H, *m*, H-6), 1.95 (1H, *m*, H-1), 1.97 (1H, *m*, H-6), 3.76 (1H, *m*, H-11), 3.86 (1H, *m*, H-11), 5.55 (1H, *br t*, $J = 2.1 \text{ Hz}$, H-7). EI-MS m/z (rel. int.): 222 $[M]^+$ (17), 191 $[M - \text{CH}_2\text{OH}]^+$ (10), 124 $[M - 60]^+$ (65), 109 (100).

REFERENCES

1. El-Ferally, F. S. (1978) *J. Chem. Soc., Chem. Commun.* 75.
2. Kioy, D., Gray, A. I. and Waterman, P. G. (1989) *J. Nat. Prod.* **52**, 174.
3. Kioy, D., Gray, A. I. and Waterman, P. G. (1990) *J. Nat. Prod.* **53**, 1372.
4. Al-Said, M. S., El-Khawaja, S. M., El-Ferally, F. S. and Hufford, C. D. (1990) *Phytochemistry* **29**, 975.
5. Al-Said, M. S., Khalifa, S. I. and El-Ferally, F. S. (1989) *Phytochemistry* **28**, 297.
6. Kubo, I., Miura, I., Pettei, M. J., Lee, Y.-W., Pilkievicz, F. and Nakanishi, K. (1977) *Tetrahedron Letters* 4553.
7. Taniguchi, M., Adachi, T., Oi, S., Kimura, A., Katsumura, S., Isoe, S. and Kubo, I. (1984) *Agric. Biol. Chem.* **48**, 73.
8. Nakanishi, K. and Kubo, I. (1977) *Isr. J. Chem.* **16**, 28.
9. Kioy, D., Gray, A. I. and Waterman, P. G. (1990) *Phytochemistry* **29**, 3535.
10. Asakawa, Y. and Takemoto, T. (1979) *Experientia* **35**, 1420.
11. Fukuyama, Y., Sato, T., Asakawa, Y. and Takemoto, T. (1982) *Phytochemistry* **21**, 2895.
12. McCallion, R. F., Cole, A. L. J., Walker, J. R. L., Blunt, J. W. and Munro, M. H. G. (1982) *Planta Med.* **44**, 134.
13. Fukuyama, Y., Sato, T., Miura, I. and Asakawa, Y. (1985) *Phytochemistry* **24**, 1521.
14. Tozyo, T., Yasuda, F., Nakai, H. and Tada, H. (1992) *J. Chem. Soc., Perkin Trans I.* 1859.
15. Asakawa, Y., Toyota, M. and Takemoto, T. (1978) *Phytochemistry* **17**, 457.
16. Sierra, J. R., Lopez, J. T. and Cortes, M. J. (1986) *Phytochemistry* **25**, 253.
17. Okuda, K. and Scheuer, P. J. (1983) *J. Org. Chem.* **48**, 1866.
18. Butter, M. S. and Capon, R. J. (1993) *Aust. J. Chem.* **46**, 1255.
19. Cimino, G., De Rosa, S., De Stefano, S., Morrone, R. and Sodano, G. (1985) *Tetrahedron* **41**, 1093.
20. Einhellig, A. F., Leathers, R. G. and Hobbs, L. L. (1985) *J. Chem. Ecol.* **2**, 65.